CANOPS-GRB v1.0: a new Earth system model for simulating the evolution of ocean-atmosphere chemistry over geologic timescales

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Abstract

A new Earth system model of intermediate complexity — CANOPS-GRB v1.0— is presented for use in quantitatively assessing the dynamics and stability of atmospheric and oceanic chemistry on Earth and Earth-like planets over geologic timescales. The new release is designed to represent the coupled major element cycles of C, N, P, O, and S, as well as the global redox budget (GRB) in Earth's exogenic (ocean-atmosphere-crust) system, using a process-based approach. This framework provides a mechanistic model of the evolution of atmospheric and oceanic O_2 levels on geologic timescales and enables comparison with a wide variety of geological records to further constrain the processes driving Earth's oxygenation. A complete detailed description of the resulting Earth system model and its new features are provided. The performance of CANOPS-GRB is then evaluated by comparing a steady-state simulation under present-day conditions with a comprehensive set of oceanic data and existing global estimates of bio-element cycling. The dynamic response of the model is also examined by varying phosphorus availability in the exogenic system. CANOPS-GRB reliably simulates the short- and long-term evolution of the coupled C-N-P-O_2-S biogeochemical cycles and is generally applicable across most period of Earth's history given suitable modifications to boundary conditions and forcing regime. The simple and adaptable design of the model also makes it useful to interrogate a wide range of problems related to Earth's oxygenation history and Earth-like exoplanets more broadly. The model source code is available on GitHub, and represents a unique community tool for investigating the dynamics and stability of atmospheric and oceanic chemistry on long timescales.

Short summary:

A new biogeochemical model (CANOPS-GRB v1.0) for assessing the redox stability and dynamics of the ocean-atmosphere system on geologic timescales has been developed. In this paper, we present a full description of the model and its performance. CANOPS-GRB is a useful tool for understanding the factors regulating atmospheric O_2 level and has the potential to greatly refine our current understanding of Earth's oxygenation history.

1 Introduction

A quarter century has passed since the first discovery of exoplanets (Mayor and Queloz, 1995). In the next quarter century, a full-scale search for signs of life — biosignatures — on Earth-like exoplanets is one of the primary objectives of the next generation of exoplanetary observational surveys (National Academies of Sciences and Medicine, 2019; The LUVOIR Team, 2019). The definition of biosignatures includes a variety of signatures that require biological activity for their origin (Des Marais et al., 2002; Lovelock, 1965; Sagan et al., 1993; Schwieterman et al., 2018; National Academies of Sciences and Medicine, 2019), but atmospheric composition has received the most interdisciplinary attention since the dawn of the search for life beyond our own planet (Hitchcock and Lovelock, 1967; Lovelock, 1965, 1972, 1975; Sagan et al., 1993) because of its potential for remote detectability. Indeed, it is likely that deciphering of exoplanetary atmospheric composition based on spectroscopic information will, at least for the foreseeable future, be our only promising means for life detection beyond our solar system. However, the detection of atmospheric composition cannot immediately answer the question of the presence or absence of a surface biosphere because significant gaps remain in our understanding of the relationships between atmospheric composition and biological activity occurring at the surface on life-bearing exoplanets. Many of these gaps arise from a lack of robust theoretical and quantitative frameworks for the emergence and maintenance of remotely detectable atmospheric biosignatures in the context of planetary biogeochemistry.

It is also important to emphasize that the abundance of atmospheric biosignature gases of living planets will evolve via an intimate interaction between life and global biogeochemical cycles of bio-essential elements across a range of timescales. Indeed, the abundances of biosignature gases such as molecular oxygen (O_2)

and methane (CH_4) in Earth's atmosphere have evolved dramatically through coevolutionary interaction with Earth's biosphere for nearly 4 billion years — through remarkable fluctuations in atmospheric chemistry and climate (Catling and Kasting, 2017; Lyons et al., 2014; Catling and Zahnle, 2020). To the extent that the coupled evolution of life and the atmosphere is a universal property of life-bearing planets that maintain robust atmospheric biosignatures, the construction of a biogeochemical framework for diagnosing atmospheric biosignatures should be a subject of urgent interdisciplinary interest.

Establishing a mechanistic understanding of our own planet's evolutionary history is also an important milestone for the construction of a search strategy for life beyond our solar system, as it provides the first step towards understanding how remotely detectable biosignatures emerge and are maintained on a planetary scale. While numerous atmospheric biosignature gases have been proposed, the most promising candidates have been 'redox-based' species, such as O_2, ozone (O_3), and CH_4 (Meadows, 2017; Meadows et al., 2018; Reinhard et al., 2017a; Krissansen-Totton et al., 2018). In particular, O_2 is of great interest to astrobiologists because of its crucial role in metabolism on Earth. Thus, a considerable effort has been devoted over recent decades toward quantitatively and mechanistically understanding Earth's oxygenation history. In particular, a recent surge in the generation of empirical records for Earth's redox evolution has yielded substantial progress in our 'broad stroke' understanding of Earth's oxygenation history and has shaped our view of biological evolution (Kump, 2008; Lyons et al., 2014). One of the intriguing insights obtained from the accumulated geochemical records is that atmospheric O_2 levels might have evolved more dynamically than previously thought — our current paradigm of Earth's oxygenation history suggests that atmospheric O_2 levels may have risen and then plummeted during the early Proterozoic, then remained low (probably <10% of the present atmospheric level; PAL) for much of the ~1 billion years leading up to the catastrophic climate system perturbations and the initial diversification of complex life during the late Proterozoic.

The possibility of low but 'post-biotic' atmospheric O_2 levels during the mid-Proterozoic has important ramifications not only for our basic theoretical understanding of long-term O_2 cycle stability on a planet with biological O_2 production, but also for biosignature detectability (Reinhard et al., 2017a). However, our quantitative and mechanistic understanding of the Earth's O_2 cycle in deep time is still rudimentary at

present. For example, one possible explanation for low atmospheric O_2 levels during the mid-Proterozoic is simply a less active or smaller biosphere (Crockford et al., 2018; Derry, 2015; Laakso and Schrag, 2014; Ozaki et al., 2019a). However, mechanisms for regulating biotic O_2 generation rates and stabilizing atmospheric O_2 levels at low levels on billion-year timescales remain obscure. As a result, the level of atmospheric O_2 and its stability during the early-mid Proterozoic are the subject of vigorous debate (Bellefroid et al., 2018; Canfield et al., 2018; Cole et al., 2016; Planavsky et al., 2018; Planavsky et al., 2016; Tang et al., 2016; Zhang et al., 2016). Perhaps even more importantly, a relatively rudimentary quantitative framework for probing the dynamics and stability of the oxygen cycle leads to the imprecision of geochemical reconstructions of ocean-atmosphere O_2 levels.

Planetary atmospheric O_2 levels are governed by a kinetic balance between sources and sinks. Feedback arises because the response of source/sink fluxes to changes in atmospheric O_2 levels is intimately interrelated to each other. Since the biogeochemical cycles of C, N, P, and S exert fundamental control on the redox budget through non-linear interactions and feedback mechanisms, a mechanistic understanding of these biogeochemical cycles is critical for understanding Earth's O_2 cycle. However, the wide range of timescales that characterize C, N, P, O_2 and S cycling through the reservoirs of the Earth system makes it difficult to fully resolve the mechanisms governing the dynamics and stability of atmospheric O_2 levels from geologic records. From this vantage, developing new quantitative tools that can explore biogeochemical cycles under conditions very different from those of the present Earth is an important pursuit.

This study is motivated by the conviction that an ensemble of 'open' Earth system modeling frameworks with explicit and flexible representation of the coupled C-N-P-O_2-S biogeochemical cycles will ultimately be required to fully understand the dynamics and stability of Earth's O_2 cycle and its controlling factors. In particular, a coherent mechanistic framework for understanding the global redox (O_2) budget (GRB) is critical for filling remaining gaps in our understanding of Earth's oxygenation history and the cause-and-effect relationships with an evolving biosphere. Here, we develop a new Earth system model, named CANOPS-GRB, which implements the coupled biogeochemical cycles of C-N-P-O_2-S within the Earth's surface system (ocean-atmosphere-crust). The core of this model is an ocean biogeochemical model,

112 CANOPS (Ozaki et al., 2011; Ozaki and Tajika, 2013; Ozaki et al., 2019a). This model has been used to
113 examine conditions for the development of widespread oceanic anoxia/euxinia during the Phanerozoic
114 (Ozaki et al., 2011; Ozaki and Tajika, 2013; Kashiyama et al., 2011) and to quantitatively constrain
115 biogeochemical cycles during the Precambrian (Cole et al., 2022; Ozaki et al., 2019a; Ozaki et al., 2019b;
116 Reinhard et al., 2017b). In this study, we extend this model to simulate the biogeochemical dynamics of the
117 coupled ocean-atmosphere-crust system. The model design (such as the complexity of the processes and
118 spatial-temporal resolution of the model) is constrained by the requirement of simulation length (>100
119 million years) and actual model run-time. A lack of understanding of biogeochemistry in deep-time and
120 availability and quality of geologic records also limit the model structure. With this in mind, we aim for a
121 comprehensive, simple, yet realistic representation of biogeochemical processes in the Earth system,
122 yielding a unique tool for investigating coupled biogeochemical cycles within the Earth system over a wide
123 range of time scales. We have placed particular emphasis on the development of a global redox budget in the
124 ocean-atmosphere-crust system given its importance in the secular evolution of atmospheric O_2 levels.
125 CANOPS-GRB is an initial step towards developing the first large-scale biogeochemistry evolution model
126 suited for the wide range of redox conditions, including explicit consideration of the coupled C-N-P-O_2-S
127 cycles and the major biogenic gases in planetary atmospheres (O_2 and CH_4).

2 Model description

Here we present a full description of a new version of the Earth system model CANOPS — CANOPS-GRB v1.0 — which is designed to facilitate simulation for a wide range of biogeochemical conditions so as to permit quantitative examination of evolving ocean-atmosphere chemistry throughout Earth's history. Below we first describe the concept of model design (Sect. 2.1). Next, we describe the overall structure of the model and the basic design of global biogeochemical cycles (Sect. 2.2 and 2.3). That is followed by a detailed description of each sub-model.

2.1 CANOPS-GRB in the hierarchy of biogeochemical models

A full understanding of Earth's evolving O_2 cycle requires a quantitative framework that includes mechanistic links between biological metabolism, ocean-atmosphere chemistry, and geologic processes. Such a framework must also represent the feedbacks between ocean-atmosphere redox state and biogeochemical cycles of redox-dependent bio-essential elements. Over recent decades, considerable progress has been made in quantifying the feedbacks between atmospheric O_2 levels and the coupled C-N-P-O_2-S biogeochemical cycles over geological timescales (Berner, 2004a; Lasaga and Ohmoto, 2002; Betts and Holland, 1991; Holland, 1978; Bolton et al., 2006; Slomp and Van Cappellen, 2007; Van Cappellen and Ingall, 1994; Colman et al., 2000; Belcher and Mcelwain, 2008). Refinements to our understanding of mechanisms regulating Earth's surface redox state have been implemented in low-resolution box models where the ocean-atmosphere system is expressed by a few boxes (Bergman et al., 2004; Laakso and Schrag, 2014; Lenton and Watson, 2000b, a; Van Cappellen and Ingall, 1996; Handoh and Lenton, 2003; Petsch and Berner, 1998; Claire et al., 2006; Goldblatt et al., 2006; Alcott et al., 2019). These models offer insights into basic system behavior and can illuminate the fundamental mechanisms that exert the most leverage on biogeochemical cycles because of their simplicity, transparency and low computational demands. However, these model architectures also have important quantitative limitations. For example, with low spatial resolution the modeler needs to assume reasonable (but *a priori*) relationships relating to internal biogeochemical cycles in the system. For instance, because of a lack of high vertical resolution, oceanic box models (Knox and Mcelroy, 1984; Sarmiento and Toggweiler, 1984; Siegenthaler and Wenk, 1984) usually overestimate the sensitivity of atmospheric CO_2 levels to biological activity at high-latitude surface ocean

155 relative to projections by general circulation models (Archer et al., 2000). Oceanic biogeochemical cycles
156 and chemical distributions are also characterized by strong vertical and horizontal heterogeneities, which
157 have the potential to affect the strength of feedback processes (Ozaki et al., 2011). In other words, the low-
158 resolution box modeling approach might overlook the strength and response of the internal feedback loops.
159 Thus, the development of an ocean model with high resolution of ocean interior and reliable representation
160 of water circulation is preferred to investigate the mechanisms controlling atmospheric O_2 levels under
161 conditions very different from those of the modern Earth.

162

163 In the last decade, comprehensive Earth system models of intermediate complexity (EMICs) have also been
164 developed and extended to include ocean sediments and global C cycling (Ridgwell and Hargreaves, 2007;
165 Lord et al., 2016). Such models can be integrated over tens of thousands of years, allowing experimentation
166 with hypothetical dynamics of global biogeochemical cycles in the geological past (Reinhard et al., 2020;
167 Olson et al., 2016). However, a key weakness of existing EMICs is the need to parameterize (or ignore)
168 boundary (input/output) fluxes — either due to the computational expense of explicitly specifying boundary
169 conditions or due to poorly constrained parameterizations. For example, the oceanic P cycle is usually
170 treated as a closed system, limiting the model's applicability to timescales less than the oceanic P residence
171 time (~15–20 kyr). Further, boundary conditions such as continental configuration and oceanic bathymetry
172 are variable or poorly constrained in deep time and the use of highly complex models is difficult to justify
173 the computational cost. Finally, exploration of hypotheses concerning the biogeochemical dynamics in deep
174 time often require large model ensembles across broad parameter space given the scope of uncertainty. This
175 makes the computational cost of EMICs intractable at present for many key questions.

176

177 The CANOPS-GRB model is designed to capture the major components of Earth system biogeochemistry
178 on timescales longer than ~10^3 years, but is simple enough to allow for runs on the order of 10^9 model years.
179 The model structure is also designed so that the model captures the essential biogeochemical processes
180 regulating the global O_2 budget, while keeping the calculation cost as moderate as possible. For example, the
181 simple relationships of biogeochemical transport processes at the interface of the Earth system (hydrogen
182 escape to space, early diagenesis in marine sediments, and weathering) are employed based on the

183 systematic application of 1-D models in previous studies (Bolton et al., 2006; Daines et al., 2017;
184 Middelburg et al., 1997; Claire et al., 2006; Wallmann, 2003), providing a powerful, computationally
185 efficient means for exploring the Earth system under a wide range of conditions. The resultant CANOPS-
186 GRB model can be run on a standard personal computer on a single CPU with an efficiency of
187 approximately 6 million model years per CPU hour. In other words, model runs in excess of 10^9 model years
188 are tractable with modest wall times (approximately 7 days). The model is thus not as efficient as simple box
189 models, but is highly efficient relative to EMICs, making sensitivity experiments and exploration of larger
190 parameter space over a billion years feasible, particularly with implementation on a high-performance
191 computing cluster (see (Cole et al., 2022)). CANOPS-GRB thus occupies a unique position within the
192 hierarchy of global biogeochemical cycle models, rendering it a useful tool for the development of more
193 comprehensive, low- to intermediate-complexity models of Earth system on very long timescales.

194 **2.2 Overall model structure**

195 The overall structure of the model is shown in Fig. 1. The model consists of ocean, atmosphere and
196 sedimentary reservoirs. The core of the model is an ocean model, comprising a high-resolution 1-D
197 intermediate-complexity box model of the global ocean (Sect. 2.4). The ocean model is coupled to a
198 parameterized marine sediment module (Sect. 2.4.4) and a one box model of the atmosphere (Sect. 2.6). The
199 atmospheric model includes O_2 and CH_4 as chemical components, and abundances of these molecules are
200 calculated based on the mass balance between sources and sinks (e.g., biogenic fluxes of O_2 and CH_4 from
201 the ecosystems and photochemical reactions). The net air-sea gas exchange of chemical species (O_2, H_2S,
202 NH_3 and CH_4) is quantified according to the stagnant film model (Liss and Slater, 1974; Kharecha et al.,
203 2005) (Sect. 2.4.5). The ocean and atmosphere models are embedded in a 'rock cycle' model that simulates
204 the evolution of sedimentary reservoir sizes on geologic timescales (Sect. 2.5). Three sedimentary reservoirs
205 (organic carbon, *ORG*; pyrite sulfur, *PYR*; and gypsum sulfur, *GYP*) are considered in the CANOPS-GRB
206 model. These reservoirs interact with the ocean-atmosphere system through weathering, outgassing, and
207 burial.

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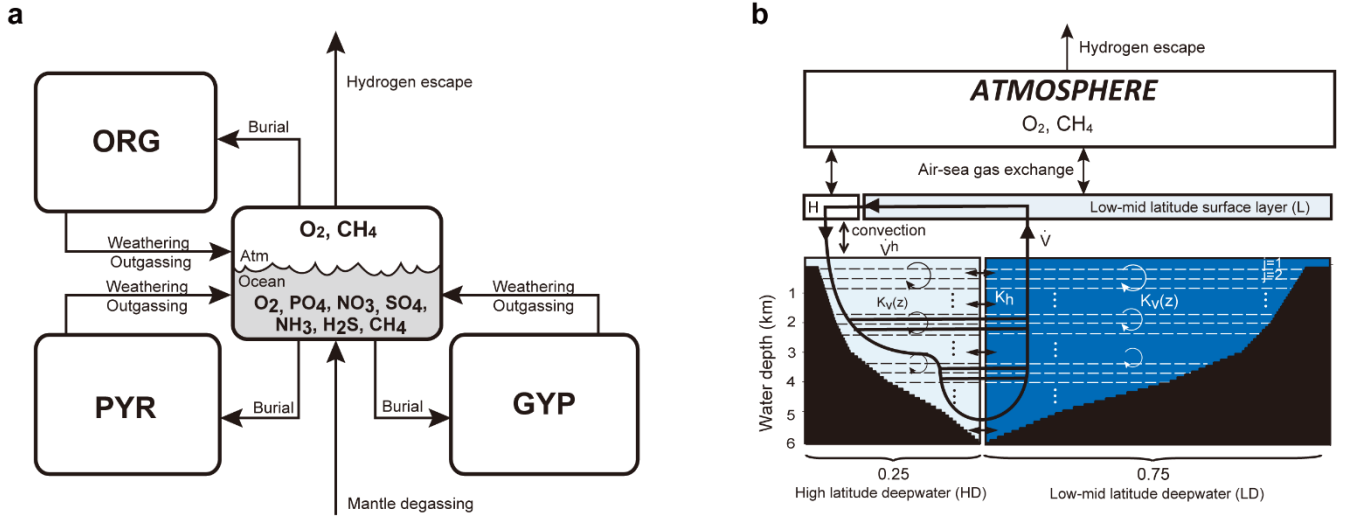
Figure 1. CANOPS-GRB model configuration. (**a**) The schematic of material cycles in the surface (ocean-atmosphere-crust) system. Three sedimentary reservoirs, organic carbon (*ORG*), pyrite sulfur (*PYR*), and gypsum sulfur (*GYP*), are considered. Sedimentary reservoirs interact with the ocean-atmosphere system via weathering, volcanic degassing, and burial. No interaction with the mantle is included, except for the input of reduced gases from the mantle. Total mass of sulfur is conserved in the surface system. (**b**) Schematic of ocean and atmosphere modules. "L" and "H" denote the low-mid latitude mixed surface layer and high-latitude surface layer, respectively. An ocean area of 10% is assumed for H. River flux for each region is proportional to the areal fraction. Ocean interior is divided into two sectors, high-latitude deep water (HD) and low-mid latitude deep water (LD), which are vertically resolved. The area of HD is 25% of the whole ocean. The deep overturning circulation, $\dot{V}$, equals the poleward flow in the model surface layer (from L to H). $K_v^l(z)$ and $K_v^h(z)$ are the vertical eddy diffusion coefficients in the LD and HD regions, respectively. K_{hor} and $\dot{V}_h$ are the horizontal diffusion coefficient and polar convection, respectively. The black hatch represents the seafloor topography assumed. The parameters regarding geometry and water transport are tabulated in Table 3.

225 The ocean model is a vertically resolved transport-reaction model of the global ocean, which was originally

226 developed by Ozaki et al. (2011) and Ozaki and Tajika (2013). The model consists of 122 boxes across two

227 regions — a low-mid latitude region and a high-latitude region (Fig. 1b). The ocean model describes water

228 transport processes as exchange fluxes between boxes and via eddy diffusion terms. More specifically,

229 ocean circulation is modelled as an advection-diffusion model of the global ocean — a general and robust

230 scheme that is capable of producing well-resolved modern profiles of circulation tracers using realistic

231 parameter values (the physical set up of the model can be found in Sect. 2.4.1 and 2.4.2). The

232 biogeochemical sub-model provides a mechanistic description of the marine biogeochemical cycles of C, P,
233 N, O_2, and S (Sect. 2.4.3). This includes explicit representation of a variety of biogeochemical processes
234 such as biological productivity in the sunlit surface oceans, a series of respiration pathways and secondary
235 redox reactions under oxic and anoxic conditions (Sect. 2.4.3), and deposition, decomposition, and burial of
236 biogenic materials in marine sediments (Sect. 2.4.4), allowing a mechanistically based examination of
237 biogeochemical processes. The suite of metabolic reactions included in the model is listed in Table 1. Ocean
238 biogeochemical tracers considered in the CANOPS-GRB model are phosphate (PO_4^{3-}), nitrate (NO_3^-), total
239 ammonia (ΣNH_3), dissolved oxygen (O_2), sulfate (SO_4^{2-}), total sulfide (ΣH_2S), and methane (CH_4). Note that
240 biogeochemical cycling of trace metals (e.g., Fe and Mn) is not included in the current version of the model.
241 All H_2S and NH_3 degassing from the ocean to the atmosphere is assumed to be completely oxidized by O_2 to
242 SO_4^{2-} and NO_3^- and returns to the ocean surface. These simplifications limit application of the model to very
243 poorly oxygenated Earth system states ($pO_2 < 10^{-5}$ PAL). Ocean model performance was tested for the
244 modern-day ocean field observational data (Sect. 3). Simulation results were also compared to previously
245 published integrated global flux estimates.

246

247 The CANOPS model has been extended and altered a number of times since first publication. The
248 description of biogeochemical cycles in the original version of CANOPS (Ozaki and Tajika, 2013; Ozaki et
249 al., 2011) does not include the S and CH_4 cycles because of their aims to investigate the conditions for the
250 development of oceanic anoxia/euxinia on timescales less than a million years during the Phanerozoic. More
251 recently, Ozaki et al. (2019a) implemented an open system modeling approach for the global S and CH_4
252 cycles, enabling quantitative analysis of global redox budget for given atmospheric O_2 levels and crustal
253 reservoir sizes. In this version of CANOPS atmospheric O_2 levels and sedimentary reservoirs are treated as
254 boundary conditions because imposing them simplifies the model and significantly reduces computing time.
255 However, this approach does not allow exploration of the dynamic behavior of atmospheric O_2 in response
256 to other boundary conditions. In the newest version presented here, significant improvements in the
257 representation of global biogeochemistry were achieved by (1) an explicit calculation of atmospheric O_2
258 levels based on atmospheric mass balance (Sect. 2.6), (2) expansion of the model framework to include
259 secular evolution of sedimentary reservoirs (Sect. 2.5.5), and (3) simplification of the global redox budget

260 between the surface (ocean-atmosphere-crust) system and the mantle (Sect. 2.3.5). These improvements are
261 in line with the requirement of an 'open' Earth system model, which is necessary for a systematic,
262 quantitative understanding of Earth's oxygenation history.

263

264 **Table 1.** Biogeochemical reactions considered in the CANOPS-GRB model.

Process	Stoichiometry[*]	Reaction #
Ammonia assimilation	$\alpha CO_2 + \beta NH_4^+ + H_3PO_4 + \alpha H_2O \rightarrow OM + \alpha O_2$	R1
Nitrate assimilation	$\alpha CO_2 + \beta NO_3^- + H_3PO_4 + (\alpha + \beta)H_2O + 2\beta H^+ \rightarrow OM + (\alpha + 2\beta)O_2$	R2
Nitrogen fixation	$N_2 + 5H_2O \rightarrow 2NH_4^+ + 2OH^- + \frac{3}{2}O_2$	R3
Aerobic respiration	$OM + \alpha O_2 \rightarrow \alpha CO_2 + \beta NH_4^+ + H_3PO_4 + \alpha H_2O$	R4
Denitrification	$OM + \frac{4}{5}\alpha NO_3^- + \frac{4}{5}\alpha H^+ \rightarrow \alpha CO_2 + \beta NH_4^+ + H_3PO_4 + \frac{7}{5}\alpha H_2O + \frac{2}{5}\alpha N_2$	R5
Sulfate reduction	$OM + \frac{1}{2}\alpha SO_4^{2-} + \alpha H^+ \rightarrow \alpha CO_2 + \beta NH_4^+ + H_3PO_4 + \alpha H_2O + \frac{1}{2}\alpha H_2S$	R6
Methanogenesis	$OM \rightarrow \frac{1}{2}\alpha CO_2 + \frac{1}{2}\alpha CH_4 + \beta NH_4^+ + H_3PO_4$	R7
Nitrification	$NH_4^+ + 2O_2 \rightarrow NO_3^- + H_2O + 2H^+$	R8
Aerobic H$_2$S oxidation[†]	$\Sigma H_2S + 2O_2 \rightarrow SO_4^{2-} + 2H^+$	R9
Aerobic CH$_4$ oxidation	$CH_4 + 2O_2 \rightarrow CO_2 + 2H_2O$	R10
Anaerobic CH$_4$ oxidation	$CH_4 + SO_4^{2-} \rightarrow HS^- + HCO_3^- + H_2O$	R11
Photooxidation of CH$_4$	$CH_4 + 2O_2 \rightarrow CO_2 + 2H_2O$	R12
Hydrogen escape to space	$CH_4 + O_2 \rightarrow CO_2 + 4H \uparrow$	R13

265 [*]OM denotes organic matter, $(CH_2O)\alpha(NH_4^+)\beta H_3PO_4$
266 [†]$\Sigma H_2S = H_2S + HS^-$

2.3 Global biogeochemical cycles

268 We construct a comprehensive biogeochemical model in order to investigate the interaction between
269 dynamic behaviors of Earth's oxygenation history and its biogeochemical processes, as well as redox
270 structure of the ocean. Here we provide the basic implementation of global biogeochemical cycles of C, P, N,
271 and S, with particular emphasis on processes of mass exchange between reservoirs that play a critical role in

272 global redox budget (Fig. 2). Our central aim here is to describe the overall design of the biogeochemical

273 cycles. The details of each sub-model are provided in the following sections.

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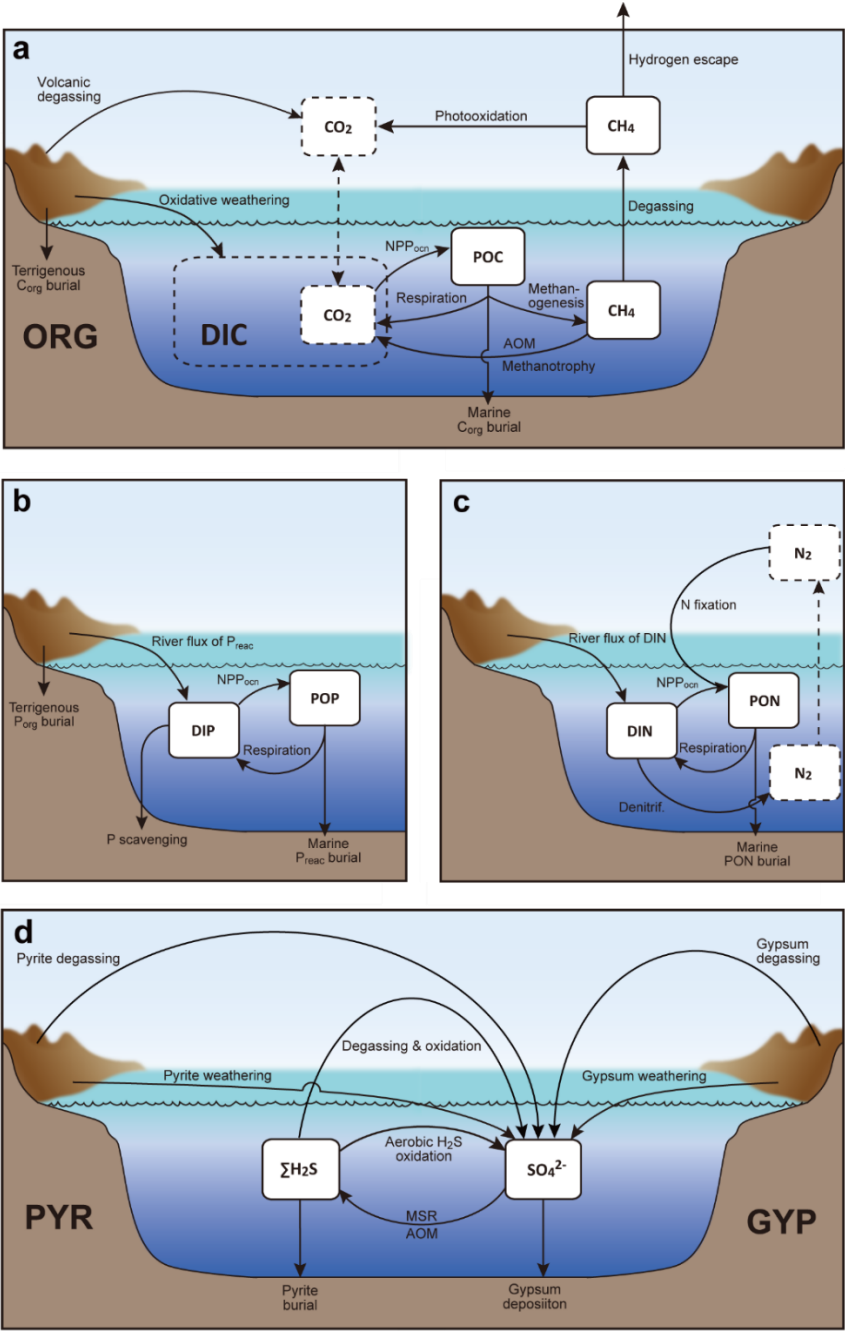
Figure 2. Schematics of global biogeochemical cycling in CANOPS-GRB. (**a**) Global C cycle. The primary source of C for the ocean-atmosphere system is volcanic degassing and oxidative weathering of sedimentary organic carbon, whereas primary sink is burial of marine and terrigenous organic matter into sediments. Inorganic carbon reservoirs (depicted as dashed boxes) and DOC are not considered. NPP_{ocn} = marine net primary production. DIC = dissolved inorganic carbon. POC = particulate organic carbon. MSR = microbial sulfate reduction. AOM = anaerobic oxidation of methane. CANOPS-GRB includes CH_4 generation via methanogenesis and its oxidation reactions via methanotrophy and AOM in the ocean interior, as well as CH_4 degassing flux to the atmosphere and its photooxidation. The rates of CH_4 photooxidation and hydrogen escape to space are calculated based on parameterizations proposed by previous studies (Goldblatt et al., 2006; Claire et al., 2006). Note that CH_4 flux from land biosphere is not shown here. (**b**) Global P cycle schematic. Weathering of reactive P (P_{reac}) is the ultimate source, whereas burial in sediments is the primary sink. A part of the weathered P is buried as terrigenous organic P, and the remaining is delivered to the ocean. The redox-dependent P burial in marine sediments is modelled by considering three phases (organic P, Fe-bound P, and authigenic P). DIP = dissolved inorganic phosphorus. POP = particulate organic phosphorus. The hypothetical P scavenging via Fe-species in anoxic-ferruginous waters is depicted, but it is not modelled in our standard model configuration. (**c**) Global N cycle schematic. The abundance of inorganic nitrogen species (ammonium and nitrate), which are lumped into DIN (dissolved inorganic nitrogen), is affected by denitrification and nitrification. The primary source is nitrogen fixation and riverine flux, whereas primary sink is denitrification and burial in marine sediments. PON = particulate organic nitrogen. The nitrogen weathering/riverine flux is assumed to be equal to the burial flux so that there is no mass imbalance in global N budget. Aeolian delivery of N from continent to the ocean is not included. (**d**) Global S cycle schematic. The reservoir sizes of sedimentary sulfur (pyrite sulfur, PYR, and gypsum sulfur, GYP) and two sulfur species (SO_4^{2-} and ΣH_2S) in the ocean are controlled by volcanic outgassing, weathering, burial, MSR, AOM, and sulfide oxidation reactions. Weathering and volcanic inputs are the primary source of S to the ocean, and burial of pyrite and gypsum in marine sediments is the primary sink. It is assumed that hydrogen sulfide escaping from the ocean to the atmosphere is completely oxidized and returns to the ocean as sulfate. The organic sulfur cycle is ignored in this study.

2.3.1 Carbon cycle

The CANOPS-GRB model includes particulate organic carbon (POC), atmospheric CH_4, dissolved CH_4 in the ocean, and sedimentary organic carbon (ORG) as carbon reservoirs (Fig. 2a). The primary sources of carbon for the ocean-atmosphere system are volcanic degassing and oxidative weathering of sedimentary organic carbon, while the primary sink is burial of marine and terrigenous organic matter in sediments. Atmospheric CO_2, dissolved inorganic carbon (DIC), and dissolved organic carbon (DOC) are not explicitly modelled in the current version of the model, and the full coupling of the inorganic carbon cycle within CANOPS-GRB is left as an important topic for future work. Neglecting the inorganic carbon cycle means that there are no climatic feedbacks in the system, and because of this simplification, the CANOPS-GRB

311 model cannot be applied to problems such as those in which the Earth's climate and redox states of the

312 ocean-atmosphere system are closely related each other or to validate model predictions based on geologic

313 records (such as $\delta^{13}C$), but this allows us to avoid introducing the additional complexities and uncertainties

314 in the model.

Organic carbon cycle

316 The biogeochemical model is driven by the cycling of the primary nutrient phosphorus, which is assumed to

317 be the ultimate limiting factor for biological productivity (see Sect. 2.4.3). Previous versions of CANOPS do

318 not take into account the impact of the activity of terrestrial ecosystem on the global O_2 budget. In the

319 CANOPS-GRB model, we improve on this by evaluating the activity levels of terrestrial and marine

320 ecosystems separately: The global net primary production (NPP), J_{NPP} (in terms of organic C), is given as a

321 sum of the oceanic (J_{NPP}^{ocn}) and terrestrial (J_{NPP}^{lnd}) NPP:

$$J_{NPP} = J_{NPP}^{ocn} + J_{NPP}^{lnd} \quad . \tag{1}$$

322 Biological production in the ocean surface layer depends on P availability while nutrient assimilation

323 efficiency is assumed to be lower in the high latitude region (Sect. 2.4.3). Terrestrial NPP is affected by the

324 atmospheric O_2 level (Sect. 2.5.1). In this study, the flux (in terms of moles per year) is expressed with a

325 capital J, whereas the flux density (in terms of moles per square meter per year) is expressed with a

326 lowercase j.

327

328 In our standard model configuration, oceanic primary production follows canonical Redfield stoichiometry

329 (C:N:P = 106:16:1) (Redfield et al., 1963). Flexible C:N:P stoichiometry of particulate organic matter

330 (POM) can be explored by changing a user-flag. Nutrients (P and N) are removed from seawater in the

331 photic zone via biological uptake, and exported as POM to deeper aphotic layers. The exported POM sinks

332 through the water column with a speed of v_{POM} (the reference value is 100 m d^{-1}). As it settles through the

333 water column, POM is subject to decomposition via a series of respiration pathways dependent on the redox

334 state of proximal seawater (Sect. 2.4.3). This gives rise to the release of dissolved constituent species back

335 into seawater. Within each layer a fraction of POM is also intercepted by a sediment layer at the bottom of

336 each water depth. Fractional coverage of every ocean layer by seafloor is calculated based on the prescribed

337 bathymetry (Sect. 2.4.1). Settling POM reaching the seafloor undergoes diagenetic alteration (releasing

338 additional dissolved species into seawater) and/or permanent burial. The ocean model has 2×60 sediment

339 segments (HD and LD have 60 layers, respectively), and for each segment the rates of organic matter

340 decomposition and burial are calculated by semi-empirical relationships extracted from ocean sediment data

341 and 1-D modelling of early diagenesis (Sect. 2.4.4). Specifically, the organic C (C_{org}) burial at each water

342 depth is calculated based on the burial efficiency (BE_{org}), which is defined as the fraction of POC buried in

343 sediments relative to that deposited on the seafloor at each water depth and is also a function of

344 sedimentation rate and bottom water O_2 levels. Organic matter not buried is subject to decomposition.

345

346 The key biogeochemical fluxes of our reference state (mimicking the present condition) are summarized in

347 Table 2. The reference value for burial rate of terrigenous C_{org} is set at 3 Tmol C yr^{-1}, assuming that burial of

348 terrigenous organic matter accounts for ~20% of the total burial. Combined with the burial rate of marine

349 C_{org} in our standard run, the total burial rate is 14.3 Tmol C yr^{-1}, representing the dominant O_2 source flux to

350 the modern ocean-atmosphere system. At steady-state, this is balanced by oxidative weathering and volcanic

351 outgassing of sedimentary C_{org}: The reference value of oxidative weathering of organic matter is determined

352 as 13.0 Tmol C yr^{-1} based on the global O_2 budget (Sect. 2.3.5). Previous versions of CANOPS (Ozaki et al.,

353 2019a) treat sedimentary reservoirs as a boundary condition. This model limitation is removed in the

354 CANOPS-GRB model — the reservoir size of sedimentary C_{org} (*ORG*) freely evolves based on the mass

355 balance through burial, weathering, and volcanic outgassing (Sect. 2.5.5). We adopted an oft-quoted value of

356 1250 Emol (E = 10^{18}) for our reference value of the *ORG*, based on literature survey (Berner, 1989; Garrels

357 and Lerman, 1981).

358

Table 2. Key biogeochemical fluxes obtained from the reference run. * denotes the reference value. Tmol =
 10^{12} mol.

Fluxes	Label	Value	Comments
Carbon cycle (Tmol C yr^{-1})			
Marine NPP	$J_{NPP}^{ocn,*}$	3794	Simulated (Eq. 24)
Terrestrial NPP	$J_{NPP}^{lnd,*}$	5000	Prescribed (Prentice et al., 2001)
Marine organic C burial	$J_{org}^{b,ocn,*}$	11.28	Simulated (Eq. 40)
Terrestrial organic C burial	$J_{org}^{b,lnd,*}$	3.0	Prescribed (This study)
Oxidative organic C weathering	$J_{org}^{w,*}$	13.03	Tuned (Eq. 15)
Organic C degassing	$J_{org}^{m,*}$	1.25	Prescribed (Bergman et al., 2004)
Phosphorus cycle (Tmol P yr^{-1})			
Reactive P weathering	$J_{P}^{w,*}$	0.158	Eqs. (4) and (84)
Terrestrial organic P burial	$J_{P}^{b,lnd,*}$	0.003	Eq. (85)
Riverine reactive P flux	$J_{P}^{r,*}$	0.155	Tuned (This study)
Marine organic P burial	$J_{Porg}^{b,*}$	0.0438	Simulated (Eq. 51)
Fe-bound P burial	$J_{PFe}^{b,*}$	0.0323	Simulated (Eq. 53)
Ca-bound P burial	$J_{PCa}^{b,*}$	0.0788	Simulated (Eq. 54)
Nitrogen cycle (Tg N yr^{-1})			
Nitrogen fixation	J_{Nfix}^{*}	180.5	Simulated
Denitrification in the water column	$J_{deni}^{wc,*}$	102.5	Simulated
Benthic denitrification	$J_{deni}^{sed,*}$	62.4	Simulated (Eq. 55)
Marine organic N burial	$J_{Norg}^{b,*}$	15.8	Simulated (Eq. 56)
Organic N weathering	$J_{Norg}^{w,*}$	15.8	$= J_{Norg}^{b,*}$
Sulfur cycle (Tmol S yr^{-1})			
Pyrite weathering	$J_{pyr}^{w,*}$	1.0	Prescribed (This study)
Gypsum weathering	$J_{gyp}^{w,*}$	1.6	Prescribed (This study)
Pyrite degassing	$J_{pyr}^{m,*}$	0.3	Prescribed (This study)
Gypsum degassing	$J_{gyp}^{m,*}$	0.5	Prescribed (This study)
Pyrite burial	$J_{pyr}^{b,*}$	1.3	Simulated
Gypsum burial	$J_{gyp}^{b,*}$	2.1	Prescribed (This study)

Methane cycle

The ocean model includes biogenic CH_4 generation via methanogenesis and its oxidation reactions via methanotrophy and anaerobic oxidation of methane (AOM) in the ocean interior (R10 and R11 in Table 1), as well as a CH_4 degassing flux to the atmosphere. The land model also calculates the biogenic CH_4 flux from the terrestrial ecosystem to the atmosphere using a transfer function (Sect. 2.5.2). The abundance of CH_4 in the atmosphere is explicitly modelled as a balance of its source (degassing from marine and terrestrial ecosystems) and sink (photooxidation and hydrogen escape), where CH_4 sink fluxes are calculated according to parameterized O_2 dependent functions proposed by previous studies. More specifically, the oxidation rate of CH_4 in the upper atmosphere is calculated based on the empirical parameterization obtained from a 1-D photochemistry model (Claire et al., 2006). The rate of hydrogen escape to space is evaluated with the assumption that it is diffusion limited and that CH_4 is a major H-containing chemical compound carrying hydrogen to the upper atmosphere (Goldblatt et al., 2006). No continental abiotic or thermogenic CH_4 fluxes are taken into account, because previous estimates of the modern fluxes are negligible relative to the biogenic flux, although we realize that it could have played a role in the global redox budget (<0.3 Tmol yr^{-1}; (Fiebig et al., 2009)). We also note that the current version of the model does not include the possibility of aerobic CH_4 production in the sea (Karl et al., 2008). Our reference run calculates atmospheric CH_4 to be 0.16 ppmv (Sect. 3.3), slightly lower than that of the preindustrial level of 0.7 ppmv (Raynaud et al., 1993; Etheridge et al., 1998), but we consider this to be within reasonable error given unknowns in the CH_4 cycle.

2.3.2 Phosphorus cycle

Phosphorus is an essential element for all life on Earth and it is regarded as the 'ultimate' bio-limiting nutrient for primary productivity on geologic time scales (Tyrrell, 1999). Thus, the P cycle plays a prominent role in regulating global O_2 levels. In the CANOPS-GRB model, we model the reactive (i.e., bioavailable) P (P_{reac}) cycling in the system and ignore non-bioavailable P. Specifically, dissolved inorganic P (DIP) and particulate organic P (POP) are explicitly modelled (Fig. 2b), whereas dissolved organic P (DOP) is ignored.

On geologic timescales, the primary source of P to the ocean-atmosphere system is continental weathering: a phosphorus is released through the dissolution of apatite which exists as a trace mineral in silicate and carbonate rocks (~0.1wt%; (Föllmi, 1996)). The total P_{reac} flux via weathering, J_P^w, is given as follows:

$$J_P^w = f_P f_R J_P^{w,*} ,$$

$$(2)$$

where * denotes the reference value, and f_P and f_R are parameters that control the availability of P in the system. Specifically, f_R is a global erosion factor representing the impact of tectonic activity on total terrestrial weathering rate, and f_P represents the availability of P_{reac}, which is used in a sensitivity experiment to assess the response of atmospheric O_2 levels to changing P_{reac} availability (Sect. 4.1). A fraction of the weathering flux J_P^w is removed via burial on land, while the remainder is transported to the ocean (Sect. 2.5.2):

$$J_P^{b,lnd} = k_{11} V J_P^w ,$$

$$(3)$$

$$J_P^r = (1 - k_{11} V) J_P^w$$

$$(4)$$

where $J_P^{b,lnd}$ and J_P^r denote the burial rate of terrigenous organic P and riverine P_{reac} flux to the ocean, respectively, k_{11} is a reference value for the fraction of the total P flux removed by the terrestrial biosphere, and V denotes the vegetation mass normalized to the modern value. These treatments are based on the Earth system box model COPSE (Bergman et al., 2004; Lenton et al., 2016; Lenton et al., 2018; Lenton and Watson, 2000b) which has been extensively tested and validated against geologic records during the Phanerozoic. In the CANOPS-GRB model, J_P^r is tuned so that modelled oceanic P inventory of the reference state is consistent with modern observations of the global ocean (Sect. 3.2.4). Our resulting tuned value is 0.155 Tmol P yr^{-1} falling in the mid-range of published estimates of 0.11–0.33 Tmol P yr^{-1}, although previous estimates of the riverine P_{reac} flux show large uncertainty (Sect. 3.2.4).

Note that our representation of P weathering ignores the effect of climate (Eq. 2). In the current version of the model the rate of P weathering is treated as one of the model forcings. Although ignoring the climate feedback on P mobility makes interpretation of the model results more straightforward, the incorporation of a climate-sensitive crustal P cycle is an important avenue for future work.

413 Since atmospheric P inputs are equivalent to less than 10% of the continental P supply to the modern oceans

414 and much of this flux is not bioavailable (Graham and Duce, 1979), we neglect the aeolian flux in this study.

415 Therefore, riverine input is the primary source of P_{reac} to the ocean. We highlight that open-system

416 modelling is crucial for realistic simulations of ocean biogeochemistry on timescales longer than the

417 residence time of P in the ocean (15–20 kyr for the modern ocean) (Hotinski et al., 2000), and in this

418 framework the riverine input of P_{reac} must be balanced over the long-term by loss to sediments via burial.

419 The change in total marine P_{reac} inventory, M_P, is given as follows:

$$\frac{dM_P}{dt} = J_P^r - J_P^{b,ocn} \ , \tag{5}$$

420 where $J_P^{b,ocn}$ denotes the total burial flux of P_{reac} in the marine system which is the sum of the burial fluxes

421 of three reactive phases, i.e. organic P (P_{org}), Fe-bound P (P_{-Fe}), and Ca-bound P (P_{-Ca}) (Sect. 2.4.4):

$$J_P^{b,ocn} = J_{Porg}^b + J_{P-Fe}^b + J_{P-Ca}^b \ . \tag{6}$$

422 O_2-dependent P burial is taken into account using empirical relationships from previous studies (Slomp and

423 Van Cappellen, 2007; Van Cappellen and Ingall, 1996, 1994). The burial of P_{org} at each water depth is a

424 function of burial efficiency, which is controlled by the burial efficiency of organic matter, C/P

425 stoichiometry of POM, sedimentation rate and bottom water $[O_2]$. We note that the strength of anoxia-

426 induced P recycling in marine sediments is very poorly constrained, especially in the Precambrian oceans

427 (Reinhard et al., 2017b). Recent studies also suggest that the P retention potential in marine sediments could

428 be affected not only by bottom water O_2 levels but by redox states (sulfidic vs. ferruginous) and the Ca^{2+}

429 concentration of bottom waters, as well as various environmental conditions such as temperature, and pH

430 (Zhao et al., 2020; Algeo and Ingall, 2007; Papadomanolaki et al. 2022). These are fruitful topics for future

431 research.

432

433 We do not explicitly account for P removal via hydrothermal processes, because it is estimated that this

434 contribution is secondary in the modern marine P cycle (0.014–0.036 Tmol P yr^{-1}; (Wheat et al., 1996;

435 Wheat et al., 2003)). We note, however, that the hydrothermal contribution to the total P budget in the

436 geologic past remains poorly constrained. We also note that in anoxic, ferruginous oceans, P scavenging by

437 Fe-minerals could also play an important role in controlling P availability and the overall budget (Reinhard

438 et al., 2017b; Derry, 2015; Laakso and Schrag, 2014). Modern observations (Dellwig et al., 2010;
439 Turnewitsch and Pohl, 2010; Shaffer, 1986) and modeling efforts (Yakushev et al., 2007) of the redoxcline
440 in the Baltic Sea and the Black Sea suggest an intimate relationship between Mn, Fe, and P cycling.
441 Trapping efficiencies of DIP by settling authigenic Fe and Mn-rich particles were found to be as high as
442 0.63 (the trapping efficiency is defined as the downward flux of P in Mn-, and Fe-oxides divided by the
443 upward flux of DIP) (Turnewitsch and Pohl, 2010). Although coupled Mn-Fe-P dynamics might have been a
444 key aspect of the biogeochemical dynamics in the Precambrian oceans, we exclude this process in our
445 standard model due to poor constraints and provide a clear and simplified picture of basic model behavior.
446 The key features between the P availability and atmospheric O_2 levels are explored by changing f_P in this
447 study (Sect. 4).

2.3.3 Nitrogen cycle

449 In the CANOPS-GRB model, two dissolved inorganic nitrogen (DIN) species (total ammonium ΣNH_4^+ and
450 nitrate NO_3^-) and particulate organic nitrogen (PON) are explicitly calculated (Fig. 2c). Atmospheric
451 nitrogen gas is assumed to never limit biospheric carbon fixation, and is not explicitly calculated. Dissolved
452 organic N (DON) and terrestrial N cycling (e.g., N fixation by terrestrial ecosystems and riverine-
453 terrigenous organic N transfer) are ignored.

454

455 In the surface ocean N assimilation via nitrate and ammonium depends on the availability of these
456 compounds. If the N required for sustaining a given level of biological productivity is not available, the
457 additional N required is assumed to be provided by atmospheric N_2 via nitrogen fixers. The ocean model
458 explicitly calculates denitrification and nitrification reactions in the water column and marine sediments (R5
459 and R8 in Table 1). The benthic denitrification rate is estimated using a semi-empirical parameterized
460 function obtained from a 1-D early diagenetic model (see Sect. 2.4.4), while nitrification is modelled as a
461 single step reaction (R8). N_2O and its related reactions, such as anammox, are not currently included.

462

463 The oceanic N cycle is open to external inputs of nitrogen. While the ultimate source of N to the ocean-
464 atmosphere system is weathering of organic N, nitrogen fixation represents the major input flux to the ocean

465   with the capacity to compensate for N loss due to denitrification. The time evolution of DIN inventory, M_N,

466   in the ocean can be written as follows:

$$\frac{dM_N}{dt} = \left(J_{Nfix} - J_{deni}^{wc} - J_{deni}^{sed} \right) + \left(J_{Norg}^{w} - J_{Norg}^{b} \right),$$

(7)

467   where J_{Nfix} denotes the N fixation rate, and J_{deni}^{wc} and J_{deni}^{sed} are denitrification rates in the water column

468   and sediments, respectively. The first set of terms on the right-hand side represent the internal N cycle in the

469   ocean-atmosphere system, while the second set of terms represent the long-term N budget which interacts

470   with sedimentary reservoir. Ultimately, loss of fixed N from the ocean-atmosphere system only occurs via

471   burial of organic N (N_{org}) in sediments, J_{Norg}^{b}. This loss is compensated for by continental weathering, J_{Norg}^{w},

472   which is assumed to be equal to the burial rate of N_{org} so that the N cycle has no impact on the global redox

473   budget. In the current version of the model, we ignore aeolian flux and all riverine N fluxes other than

474   weathering since these are minor relative to N fixation (Wang et al., 2019). As a result, modelled N fixation

475   required for oceanic N balance can be regarded as an upper estimate.

2.3.4 Sulfur cycle

477   The original CANOPS ocean model (Ozaki and Tajika, 2013; Ozaki et al., 2011) treated two sulfur species,

478   SO_4^{2-} and ΣH_2S, in a closed system: Neither inputs to the ocean from rivers, hydrothermal vents, and

479   submarine volcanoes, nor outputs due to evaporite formation and sedimentary pyrite burial were simulated.

480   This simplification can be justified when the timescale of interest is less than the residence time of the S

481   cycle (~10–20 Myr). The recently-revised CANOPS model (Ozaki et al., 2019a) extends the framework by

482   incorporating the S budget in the ocean. In their model framework, the sedimentary S reservoirs are treated

483   as boundary conditions: The size of sedimentary gypsum and pyrite reservoirs are prescribed and no explicit

484   calculations of mass balance are performed. In CANOPS-GRB, we removed this model limitation and the

485   sedimentary reservoirs are explicitly evaluated based on mass balance which is controlled by burial,

486   outgassing and weathering (see Sect. 2.4). Specifically, seawater SO_4^{2}, ΣH_2S, and sedimentary sulfur

487   reservoirs of pyrite sulfur (*PYR*) and gypsum sulfur (*GYP*) are explicitly evaluated in the current version of

488   the model. No atmospheric sulfur species are calculated—all H_2S degassing from the ocean to the

atmosphere is assumed to be oxidized to sulfate and return to the ocean. The organic sulfur cycle is not considered in this study.

Sulfur enters the ocean mainly from river runoff, J_S^r, with minor contributions from volcanic outgassing of sedimentary pyrite, J_{pyr}^m and gypsum, J_{gyp}^m. The reference value for the riverine flux is set at 2.6 Tmol S yr^{-1}, consistent with the published estimate of 2.6±0.6 Tmol S yr^{-1} (Raiswell and Canfield, 2012). The riverine flux is written as the sum of gypsum weathering and oxidative weathering of pyrite: $J_S^r = J_{gyp}^w + J_{pyr}^w$. Sulfur weathering fluxes are also assumed to be proportional to the sedimentary reservoir size. Estimates of modern volcanic input fall within the range of 0.3–3 Tmol S yr^{-1} (Catling and Kasting, 2017; Kagoshima et al., 2015; Raiswell and Canfield, 2012; Walker and Brimblecombe, 1985). We adopted a value of 0.8 Tmol S yr^{-1} for this flux (Kagoshima et al., 2015). Our total input of 3.4 Tmol S yr^{-1} is also within the range of the previous estimate of 3.3±0.7 Tmol S yr^{-1} (Raiswell and Canfield, 2012). Sulfur is removed from the ocean either via pyrite burial, J_{pyr}^b, or gypsum deposition, J_{gyp}^b (Fig. 2d). The time evolution of the inventory of total S in the ocean can thus be written, as follows:

$$\frac{d\left(M_{SO_4} + M_{H_2S}\right)}{dt} = \left(J_S^r + J_{pyr}^m + J_{gyp}^m\right) - \left(J_{pyr}^b + J_{gyp}^b\right), \tag{8}$$

where M_{SO4} and M_{H2S} denote the inventory of sulfate and hydrogen sulfide in the ocean, respectively. Two sulfur species (SO_4^{2-} and ΣH_2S) are transformed via microbial sulfate reduction (MSR) (R6), AOM (R11), and aerobic sulfide oxidation reactions (R9). The above equation thus can be divided into following equations:

$$\frac{dM_{SO_4}}{dt} = J_S^r + J_{pyr}^m + J_{gyp}^m + J_{H_2S}^{ox} - J_{MSR\&AOM} - J_{gyp}^b, \tag{9}$$

$$\frac{dM_{H_2S}}{dt} = -J_{H_2S}^{ox} + J_{MSR\&AOM} - J_{pyr}^b, \tag{10}$$

where J_{H2S}^{ox} denotes the oxidation of hydrogen sulfide and $J_{MSR\&AOM}$ is sulfate reduction via MSR and AOM. Pyrite burial is represented as the sum of pyrite precipitation in the water column and sediments: $J_{pyr}^b = J_{pyr}^{b,wc} + J_{pyr}^{b,sed}$, where the pyrite burial rate in marine sediments is assumed to be proportional to the rate of benthic sulfide production. The proportional coefficient, pyrite burial efficiency (e_{pyr}), is one of the

511 tunable constants of the model: For normal (oxic) marine sediments e_{pyr} is tuned such that the seawater SO_4^{2-}

512 concentration for our reference run is consistent with modern observations (Sect. 2.4.3). Pyrite precipitation

513 in the water column is assumed to be proportional to the concentration of $\Sigma H_2 S$.

514

515 Although the present-day marine S budget is likely out of balance because of a lack of major gypsum

516 formation, the S cycle can be considered to operate at steady state on timescales longer than the residence

517 time of sulfur in the ocean. According to S isotope mass balance calculations, $\sim$10–45% of the removal flux

518 is accounted for by pyrite burial, and the remainder is removed via formation of gypsum/anhydrite in the

519 near-modern oceans (Tostevin et al., 2014). Although gypsum deposition would have been strongly

520 influenced by tectonic activity (Halevy et al., 2012), we assume that the rate of gypsum deposition on

521 geologic time scales is proportional to the ion product of Ca^{2+} and SO_4^{2-} (Berner, 2004b) in the low- to mid-

522 latitude surface layer (L), and is defined as follows:

$$J_{gyp}^{b} = \left(\frac{[Ca^{2+}]_l [SO_4^{2-}]_l}{[Ca^{2+}]^* [SO_4^{2-}]^*} \right) J_{gyp}^{b,*} = f_{Ca} \left(\frac{[SO_4^{2-}]_l}{[SO_4^{2-}]^*} \right) J_{gyp}^{b,*} \tag{11}$$

523 where l denotes the low- to mid-latitude surface layer and f_{Ca} is a parameter that represents the seawater Ca^{2+}

524 concentration normalized by the present value ($f_{Ca} = 1$ for the reference run). The reference value of gypsum

525 burial $J_{gyp}^{b,*}$ is determined by assuming that gypsum deposition accounts for $\sim$60% of the total S removal

526 from the near-modern ocean.

2.3.5 Global redox budget

528 In the previous version of the CANOPS model (Ozaki et al., 2019a), the atmospheric O_2 level was

529 prescribed as a boundary condition, rather than modeled in order to limit computational demands. In this

530 study, we remove this model limitation by introducing an explicit mass balance calculation of atmospheric

531 O_2 (Sect. 2.6.3). This improvement allows us to explore the dynamic response of O_2 levels in the ocean-

532 atmosphere system (Sect. 4).
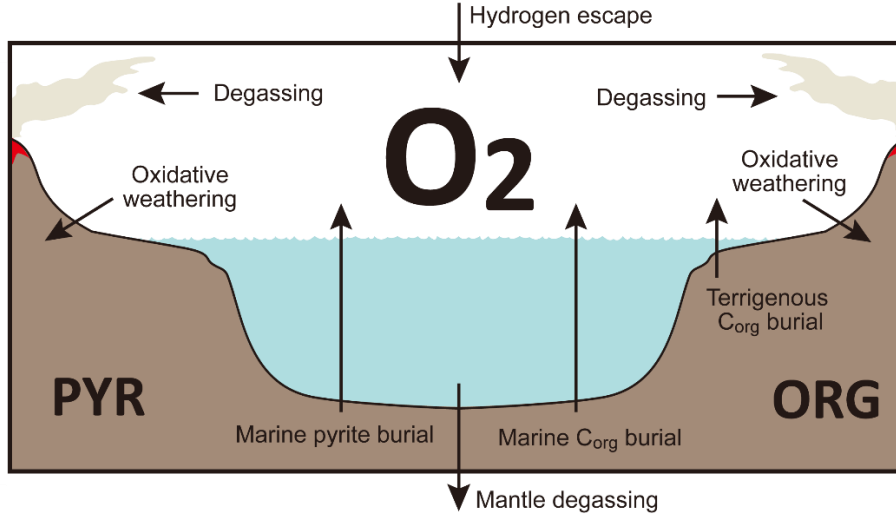
Figure 3. Schematics of global redox (O₂) budget. Arrows represent the O₂ flux. The primary source is burial of organic carbon and pyrite sulfur in sediments and hydrogen escape to space. The primary sink is volcanic outgassing and weathering of crustal organic matter and pyrite. PYR = sedimentary reservoir of pyrite sulfur. ORG = sedimentary reservoir of organic carbon. CANOPS-GRB tracks the global redox (O₂) budget for each simulation.

CANOPS-GRB v1.0 is designed to be a part of a comprehensive global redox budget (GRB) framework (Fig. 3) (Catling and Kasting, 2017; Ozaki and Reinhard, 2021). Here GRB is defined for the combined ocean-atmosphere system. In this study we track GRB in terms of O₂ equivalents. The ultimate source of O₂ is the activity of oxygenic photosynthesis (and subsequent burial of reduced species, such as organic matter and pyrite sulfur, in sediments), whereas the primary sink of O₂ is the oxidative weathering of organic carbon and pyrite which are assumed to be O₂-dependent (Sect. 2.5.3). On timescales longer than the residence time of O₂ in the ocean-atmosphere system, O₂ source fluxes should be balanced by sink fluxes. Specifically, the O₂ budget in the coupled ocean-atmosphere system can be expressed as follows:

$$\mathrm{GRB} = \left(J_{\mathrm{org}}^{\mathrm{b,ocn}} + J_{\mathrm{org}}^{\mathrm{b,lnd}} - J_{\mathrm{org}}^{\mathrm{w}} - J_{\mathrm{org}}^{\mathrm{m}} \right) + 2\left(J_{\mathrm{pyr}}^{\mathrm{b}} - J_{\mathrm{pyr}}^{\mathrm{w}} - J_{\mathrm{pyr}}^{\mathrm{m}} \right) + \left(J_{\mathrm{Hesc}} - J_{\mathrm{man}} \right), \tag{12}$$

where the first and second set of terms on the right-hand side represent the redox balance via organic carbon and pyrite sulfur subcycles, respectively. J_{Hesc} in the third term denotes hydrogen escape to space, representing the irreversible oxidation of the system. For well-oxygenated atmospheres this process plays a minor role in the redox budget, but for less oxygenated atmospheres with high levels of CH₄ this flux could lead to redox imbalance. In this study we include the input of reducing power (e.g., H₂ and CO) from the

553 Earth's interior to the surface, J_{man}, which is assumed to be equal to the value of J_{Hesc} ($J_{man} = J_{Hesc}$) to avoid

554 redox imbalance in the exogenic system. In reality, mantle degassing and the rate of hydrogen escape are not

555 necessarily equal, resulting in redox imbalance that may exert a fundamental control on atmospheric redox

556 chemistry on geologic timescales (Hayes and Waldbauer, 2006; Ozaki and Reinhard, 2021; Canfield, 2004;

557 Eguchi et al., 2020), however to maintain simplicity we have left this as a topic for future work. As a result,

558 the terms on the right-hand side must be balanced at steady state. Our model can meet this criterion. Note

559 that the effects of the Fe cycle on the O_2 budget (e.g., the oxidative weathering of Fe(II)-bearing minerals;

560 (Ozaki et al., 2019a)) are not included in the core version of the CANOPS-GRB v1.0 code and in the

561 analyses presented here for the sake of simplicity.

562

563 The CANOPS-GRB model also tracks the O_2 budgets for the atmosphere (*ARB*) and ocean (*ORB*)

564 independently:

$$ARB = \Phi_{ex}^{air\text{-}sea} + \left(J_{org}^{b,lnd} - J_{org}^{w} - J_{org}^{m} \right) - 2\left(J_{pyr}^{w} + J_{pyr}^{m} \right) + \left(J_{Hesc} - J_{man} \right), \tag{13}$$

$$ORB = -\Phi_{ex}^{air\text{-}sea} + J_{org}^{b,ocn} + 2J_{pyr}^{b}, \tag{14}$$

565 where $\Phi_{ex}^{air\text{-}sea}$ represents the net exchange of oxidizing power between the ocean and atmosphere via gas

566 exchange (O_2 with minor contributions of NH_3, H_2S and CH_4). These separate redox budgets are also

567 tracked in order to validate global budget calculations.

568

569 For our reference condition, we obtain the reference value for the oxidative weathering rate of C_{org} ($J_{org}^{w,*}$)

570 using the redox budget via C_{org} subcycle:

$$J_{org}^{w,*} = J_{org}^{b,ocn,*} + J_{org}^{b,lnd,*} - J_{org}^{m,*}. \tag{15}$$

571 Given flux values based on the calculated ($J_{org}^{b,ocn,*} = 11.28$ Tmol C yr^{-1}) and prescribed ($J_{org}^{b,lnd,*} = 3$ Tmol

572 C yr^{-1}, $J_{org}^{m,*} = 1.25$ Tmol C yr^{-1}) values on the right-hand side, $J_{org}^{w,*}$ is estimated as 13.03 Tmol C yr^{-1}

573 (Table 2).

2.4 Ocean model

Here we undertake a thorough review, reconsideration and revision (where warranted), of all aspects of the ocean model, including bringing together developments of the model following the original papers describing the CANOPS ocean model framework (Ozaki and Tajika, 2013; Ozaki et al., 2011).

The ocean model includes exchange of chemical species with external systems via several processes such as air-sea exchange, riverine input, and sediment burial. The biogeochemical model also includes a series of biogeochemical processes, such as the ocean biological pump and redox reactions under oxic-anoxic-sulfidic conditions. Our ocean model is convenient for investigating Earth system changes on timescales of hundreds of years or longer and it can be relatively easily integrated, rendering the model unique in terms of biogeochemical cycle models. CANOPS is also well suited for sensitivity studies and can be used to obtain useful information upstream of more complex models.

Development of the ocean model included two initial goals: First, to adopt a general and robust ocean circulation scheme capable of producing well-resolved modern distributions of circulation tracers using realistic ventilation rates with a limited number of free parameters. The model outputs for circulation tracers are validated by comparison with modern observations (see Sect. 3). This confirms that our ocean circulation scheme is adequate for representing the global patterns of water mass transport. A second key goal was to couple the circulation model with an ocean biogeochemical model, and to evaluate performance by comparison with modern ocean biogeochemical data (see Sect. 3.2). Examination of the distributions and globally-integrated fluxes of C, N, P, S, and O_2 for the modern ocean reveals that the ocean model can capture the fundamentals of marine biogeochemical cycling.

2.4.1 Structure

CANOPS ocean model is a 1-D (vertically resolved) intermediate complexity box model of ocean biogeochemistry (see Fig. 1b for the schematic structure) originally developed by (Ozaki and Tajika, 2013; Ozaki et al., 2011). Our model structure is an improved version of the HILDA model (Joos et al., 1991; Shaffer and Sarmiento, 1995). Unlike simple one-dimensional global ocean models (e.g, (Southam et al.,

601 1982)), the HILDA-type model includes explicit high-latitude dynamics whereby the high-latitude surface
602 layer exchanges properties with the deep ocean. This treatment is crucial for simulating preformed
603 properties and observed chemical distributions, especially for phosphate and dissolved O_2 in a self-
604 consistent manner. Unlike simple box-type global ocean models, the model has high vertical resolution. This
605 is needed for representing proper biogeochemical processes which show strong depth dependency.
606 Furthermore, HILDA type models (Arndt et al., 2011; Shaffer et al., 2008), unlike multi-box-type global
607 ocean models (Hotinski et al., 2000), use a small number of free parameters to represent ocean physics and
608 biology. The simple and adaptable structure of the model should make it applicable to a wide range of
609 paleoceanographic problems. The ocean model couples a diffusion-advection model of the global ocean
610 surface and interior with a biogeochemical model (Sect. 2.4.3) and a parameterized sediment model (Sect.
611 2.4.4).

612

613 The ocean surface consists of a mixed layer at low-mid latitude (L) and high-latitude (H). Below the surface
614 layers, we adopt the present-day averaged seafloor topography of (Millero, 2006), with the hypsometric
615 profile shown in Fig. 4a. Below the surface water layers, the ocean interior comprises two regions: the high-
616 mid latitude region (HD) and low-mid latitude region (LD). Each region is subdivided vertically, with high
617 resolution ($\Delta z = 100$ m). Each of the 60 ocean layers in each latitude region (120 total) is assigned ocean
618 sediment properties. The cross-sectional area, volume, and sediment surface area of each box is calculated
619 from the benthic hypsometry. Inclusion of the bathymetry allows evaluation of the flux of biogenic materials
620 which settle on, and are buried in, seafloor sediments at each water depth (Sect. 2.3.3 and 2.3.4).

621

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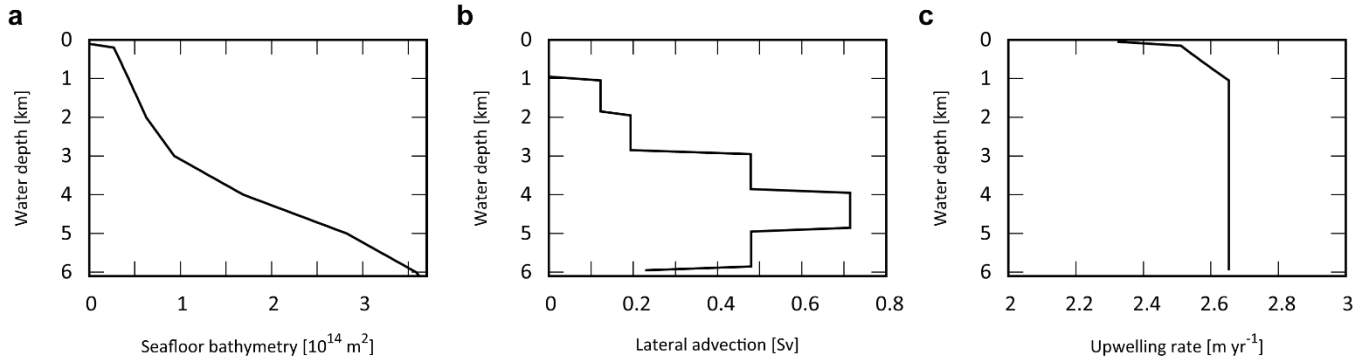


Figure 4. Ocean bathymetry and water transport. (**a**) Seafloor topography (cumulative seafloor area) (Millero, 2006) adopted in the CANOS-GRB model. (**b**) Lateral water advection from HD to LD section assumed in the standard run (in Sv). Total advection rate $\dot{V}$ was set at 20 Sv. (**c**) Upwelling rate in the LD region (in m yr^{-1}) of the standard run.

2.4.2 Transport

The ocean circulation model represents a general and robust scheme that is capable of producing well-resolved modern profiles of circulation tracers using realistic parameter values, and the coupled biogeochemical model (Sect. 2.4.3) and the parameterized sediment model (Sect. 2.4.4).

The time-space evolution of model variables in the ocean is described by a system of horizontally integrated vertical diffusion equations for non-conservative substances. The tracer conservation equation establishes the relationship between change of tracer concentration at a given grid point and the processes that can change that concentration. These processes include water transport by advection and mixing as well as sources and sinks due to biological and chemical transformations. The temporal and spatial evolution of the concentration of a dissolved component in the aphotic zone is described by a horizontally integrated vertical diffusion equation, which relates the rate of change of tracer concentration at a given point to the processes that act to change the tracer concentration:

$$\frac{\partial [X]}{\partial t} = \left.\frac{\partial [X]}{\partial t}\right|_{trans} + \Theta_{bio} + \Theta_{react} , \tag{16}$$

where [X] represents horizontally integrated physical variables (such as potential temperature, salinity or
^{14}C) or concentration of a chemical component, t denotes time, and Θ_{bio} and Θ_{react} represents internal sources
and sinks associated with the biological pump and chemical reactions, respectively. An external source/sink
term Θ_{ex}, which represents riverine input and/or air-sea gas exchange, is added to the surface layers. The
first term on the right-hand side of equation (16) represents the physical transport:

$$\left.\frac{\partial[X]}{\partial t}\right|_{trans} = -A^{l,h}(z)w^{l,h}(z)\frac{\partial[X]}{\partial z} + \frac{\partial}{\partial z}\left(A^{l,h}(z)K_v^{l,h}(z)\frac{\partial[X]}{\partial z}\right) + K_{hor}\frac{\partial^2[X]}{\partial y^2} \ . \tag{17}$$

The terms on the right-hand side express (from left to right) the advection, vertical diffusion, and horizontal
diffusion. Here, l and h indicate the LD and HD, respectively. The factors $K_v^{l,h}(z)$, K_{hor}, $A^{l,h}(z)$, and $w^{l,h}(z)$
denotes the vertical and horizontal diffusion coefficients, the areal fraction of the water layer at water depth
z to the sea surface area, and upwelling (for LD) or downwelling (for HD) velocity, respectively.

In the CANOPS ocean model, ocean circulation and mixing are characterized by five physical parameters:
(1) water transport via thermohaline circulation, $\dot{V}$, associated with high latitude sinking and low-mid
latitude upwelling; (2) constant horizontal diffusion between the aphotic zones, K_{hor}; (3) strong, depth-
dependent vertical diffusion between the aphotic zones in the high latitude region, $K_v^h(z)$; (4) high latitude
convection, $\dot{V}_h$; and (5) depth-dependent vertical diffusion in the low-mid latitude region, $K_v^l(z)$. These
parameters are tuned to give tracer distributions consistent with present-day observations. Thermohaline
circulation and high-latitude convection are considered to be general physical modes on any rotating planet,
and our simplified water transport scheme allows us to represent them with a limit number of free
parameters. However, we emphasize that the water transport scheme explored here is designed to represent
the modern ocean circulation on Earth. As a result, some of these parameterizations may need to be
modified when applying to ancient oceans or oceans on exoplanets. Nevertheless, given our simple design,
our water transport scheme is relatively flexible to modify the water circulations that are markedly different
from the modern ocean.

Advection

Advective water transport in the ocean model represents the major features of modern meridional overturning circulation. The rate of production of ventilated ocean waters ranges from 14 to 27 Sv (1 Sv = 10^6 m^3 s^{-1}) in the North Atlantic and from 18 to 30 Sv in the Southern Ocean (e.g., (Doney et al., 2004; Lumpkin and Speer, 2007)). The formation of deepwater effectively supplies "fresh" ventilated water to the abyss. We choose $\dot{V}$ = 20 Sv as a reference value, giving a mean overturning time of about 2,140 yr, consistent with the ventilation time estimated from observations (Broecker and Peng, 1982).

The downwelling of the surface waters at H forms HD that flows into the intermediate to deep oceanic layers of LD, which, in turn, upwells over L (Fig. 1b). In many one-dimensional ocean models, downwelling water enters the ocean interior via the deepest model layer (e.g., (Southam et al., 1982; Shaffer and Sarmiento, 1995; Volk and Hoffert, 1985)). In the real ocean, downwelling waters are transported along isopycnal layers below approximately 1,000 m (e.g., (Doney et al., 2004; Lumpkin and Speer, 2007; Shaffer and Sarmiento, 1995; Volk and Hoffert, 1985)). Hence, we assume that high-latitude deep water flows into each ocean layer below 1,100 m. While there is some uncertainty in the pattern of lateral advection, the flow is determined in our model assuming a constant upwelling rate below a depth of 1,100 m in the LD region. The upwelling/downwelling rate $w^{l,h}(z)$ is then determined by the seafloor topography and the deep water lateral inflow, assuming continuity. Figure 4b shows the lateral advection of deep waters with a reference circulation rate $\dot{V}$ of 20 Sv. This assumption provides a plausible upwelling rate, which is consistent with the oft-quoted value of 2–3 m yr^{-1} (Broecker and Peng, 1982) (Fig. 4c).

Vertical mixing

Ocean circulation is dominated by turbulent processes driven by wind and tidal mixing. These processes occur as eddies which occur at a wide range of spatial scales, from centimeters to whole ocean basins. In numerical models of ocean circulation, turbulent mixing in the ocean interior is commonly represented as a diffusion process, characterized by an eddy diffusion coefficient. The vertical eddy diffusion coefficient $K_v(z)$ is typically on the order of 10^{-5} to 10^{-4} m^2 s^{-1} and it is common to assume a depth-dependence which

690   smoothly increases from the thermocline ($\sim 10^{-5}$ m^2 s^{-1}) to the abyss ($\sim 10^{-4}$ m^2 s^{-1}) using an inverse or

691   hyperbolic tangent function (e.g., (Shaffer et al., 2008; Yakushev et al., 2007)). To account for thermocline

692   ventilation, we assumed a relatively high vertical diffusion coefficient in mid-water depth ($K_l = 6.3 \times 10^{-5}$ m^2

693   s^{-1} for water depth 500–1500 m). We also adopted a higher value for the vertical diffusion coefficient ($K_u =$

694   1.6×10^{-4} m^2 s^{-1}) in the upper most 500 m of the ocean in order to represent the highly convective Ekman

695   layer in the upper part of the ocean.

$$K_v^1(z) = \begin{cases} K_u & (z \geq \text{-500 m}) \\[2ex] K_l & (\text{-500} \geq z \geq \text{-1500 m}) \\[2ex] \kappa^s + \dfrac{\kappa^d - \kappa^s}{2}\left(1 + \tanh\left(\dfrac{z - z^1}{z^1}\right)\right) & (\text{otherwise}) \end{cases} \tag{18}$$

696   where κ^s and κ^d are vertical mixing coefficients, and z^1 is the transition length scale (Romaniello and Derry,

697   2010). In the high latitude region where no permanent thermocline exists, more rapid communication with

698   deepwaters can occur. Previous studies have pointed out that the vertical diffusivities at high latitude can be

699   very high (up to $O(10^{-2}$ m^2 s^{-1})) (e.g., (Sloyan, 2005)). To account for this we include high-latitude

700   convection between H and YD ($\dot{V}_h = 57.4$ Sv) and higher vertical diffusion ($K_v^h(z) = 2 \times K_v^1(z)$).

701   **Horizontal diffusion**

702   The horizontal diffusivity is included according to Romaniello and Derry (2010). On basin scales, the

703   horizontal (isopycnal) eddy diffusivity is 10^7–10^8 times larger than the vertical (diapycnal) eddy diffusivity

704   due to anisotropy of the density field. For a spatial scale of 1,000 km, horizontal eddy diffusion is estimated

705   to be $O(10^3$ m^2 s^{-1}) (e.g., (Ledwell et al., 1998)). We adopt this value. As Romaniello and Derry (2010) did,

706   we assume horizontal mixing follows the pathways of advective fluxes between laterally adjacent regions.

707   The reciprocal exchange fluxes may be written as

$$J_{hor}^{ex} = K_{hor} A_\perp \frac{\partial [X]}{\partial y} = \frac{K_{hor} A_\perp}{L} \Delta [X], \tag{19}$$

708 where $J_{\text{hor}}^{\text{ex}}$ denotes the exchange fluxes between the layers (in mol yr^{-1}), $A_\perp$ represents the cross-sectional

709 area separating two adjacent reservoirs, L is a characteristic spatial distance separating the reservoirs, $\Delta[X]$ is

710 the difference in concentration between two reservoirs (Romaniello and Derry, 2010). By assuming that L is

711 of the same order as the length of the interface separating the two regions, we can approximate

712 $A_\perp \approx \Delta z \times O(L)$, where Δz is the thickness of the interface separating the two regions. Then we obtain

$$J_{\text{hor}}^{\text{ex}} = K_{\text{hor}} \Delta z \Delta[X]. \tag{20}$$

713 Therefore, when we discretize the ocean interior at 100 m spacing approximately 0.1 Sv of reciprocal

714 mixing occurs between adjacent layers.

715 **Ocean circulation tracers**

716 We use potential temperature θ, salinity S, and radioactive carbon ^{14}C, as physical tracers. Distributions of

717 these tracers are determined by the transport mechanisms described above. In this study, we adopt the values

718 at the surface layers (L and H) as upper boundary conditions: $\theta^l = 15°C$, $\theta^h = 0°C$, $S^l = 35$ psu, $S^h = 34$ psu,

719 $\Delta^{14}C^l = -40‰$, and $\Delta^{14}C^h = -100‰$. The radioactive decay rate for ^{14}C is 1.21×10^{-4} yr^{-1}. Although ^{14}C can be

720 incorporated in the biogenic materials and transported into deep water, we ignore this biological effect for

721 simplicity. The associated error is ~10% of the profiles produced by circulation and radioactive decay

722 (Shaffer and Sarmiento, 1995). The parameter values used in the ocean circulation model are listed in Table

723 3.

724

725

Table 3. Physical set-up of the ocean circulation model.

Parameters	Label	Value	Unit	Ref.
Ocean surface area	A	3.62×10^{14}	m^2	(Ozaki and Tajika, 2013)
Surface area of high-latitude layer (H)	A^h	0.362×10^{14}	m^2	(Ozaki and Tajika, 2013)
Depth of mixed layer	h_m	100	m	(Ozaki and Tajika, 2013)
Grid spacing	Δz	100	m	(Ozaki and Tajika, 2013)
Water depth of ocean bottom	z_b	6,100	m	(Ozaki and Tajika, 2013)
Ocean overturning rate	$\dot{V}$	20	Sv	(Ozaki and Tajika, 2013)
Vertical mixing coefficient ($z < 500$ m)	K_u	5,000	$m^2\ yr^{-1}$	(Ozaki and Tajika, 2013)
Vertical mixing coefficient (500 m $< z <$ 1,500 m)	K_l	2,500	$m^2\ yr^{-1}$	(Ozaki and Tajika, 2013)
Mixing coefficient	κ_s	473	$m^2\ yr^{-1}$	(Romaniello and Derry, 2010)
Mixing coefficient	κ_d	3,154	$m^2\ yr^{-1}$	(Romaniello and Derry, 2010)
Transition depth for vertical mixing coefficient	z_l	1,000	m	(Romaniello and Derry, 2010)
High-latitude convection rate	$\dot{V}_h$	57.4	Sv	(Ozaki and Tajika, 2013)
Horizontal diffusion coefficient	K_{hor}	1,000	$m^2\ s^{-1}$	(Romaniello and Derry, 2010)

727

2.4.3 Ocean biogeochemical framework

The ocean circulation model is coupled to a biogeochemical model, which includes an explicit representation of a variety of biogeochemical processes in the ocean. The parameters used in the oceanic biogeochemical model are listed in Table 4.

732 **Table 4.** Parameter values used in the oceanic biogeochemistry module of CANOPS-GRB.

Parameter	Label	Value	Unit	Ref.
Efficiency factor for phosphate uptake at L	ε^l	1.0	–	(Ozaki and Tajika, 2013)
Efficiency factor for phosphate uptake at H	ε^h	0.15	–	(Ozaki and Tajika, 2013)
Phosphate half saturation constant	K_P	1×10^{-6}	mM	(Ozaki and Tajika, 2013)
Export ratio	f_{exp}	0.2	–	(Ozaki et al., 2019a)
Redfield C/P ratio	α^*	106	mol mol^{-1}	(Redfield et al., 1963)
Redfield N/P ratio	β^*	16	mol mol^{-1}	(Redfield et al., 1963)
POM sinking velocity	v_{POM}	100	m d^{-1}	(Ozaki et al., 2011)
Mass fraction of G_1	m_1	0.72	–	(Ozaki and Tajika, 2013)
Mass fraction of G_2	m_2	0.25	–	(Ozaki and Tajika, 2013)
Mass fraction of G_3	m_3	0.03	–	(Ozaki and Tajika, 2013)
Decomposition rate of G_1	k_1	0.6	d^{-1}	(Ozaki et al., 2011)
Decomposition rate of G_2	k_2	0.1	d^{-1}	(Ozaki et al., 2011)
Decomposition rate of G_3	k_3	0.0	d^{-1}	(Ozaki et al., 2011)
Half saturation constant for aerobic respiration	K_{O2}	8×10^{-3}	mM	(Boudreau, 1996)
Half saturation constant for denitrification	K_{NO3}	3×10^{-2}	mM	(Boudreau, 1996)
Half saturation constant for MSR	K_{MSR}	0.2	mM	This study
Half saturation constant for AOM	K_{AOM}	0.093	mM	(Beal et al., 2011)
Aerobic oxidation rate of ammonium	k_{R8}	1.825×10^{4}	mM^{-1} yr^{-1}	(Oguz et al., 2001)
Aerobic oxidation rate of sulfide	k_{R9}	3.65×10^{3}	mM^{-1} yr^{-1}	(Oguz et al., 2001)
Aerobic methane oxidation rate	k_{R10}	1×10^{7}	mM^{-1} yr^{-1}	(Van Cappellen and Wang, 1996)
Anaerobic methane oxidation rate	k_{R11}	3×10^{-4}	yr^{-1}	(Ozaki et al., 2019a)
Pyrite formation rate in the water column	k_{pyr}^{wc}	0.01	yr^{-1}	This study
Reference value of seawater sulfate concentration	$[SO_4^{2-}]^*$	28.9	mM	(Ozaki et al., 2019a)

733

Biological production

The overall biogeochemical cycling scheme is based on the cycling of primary nutrient (phosphate; PO_4^{3-}), which limits biological productivity — export production is related to the availability of P within the euphotic zone (Maier-Reimer, 1993; Yamanaka and Tajika, 1996; Shaffer et al., 2008):

$$j_{exp}^{l,h} = \alpha^{l,h} h_m \varepsilon^{l,h} [PO_4^{3-}]^{l,h} \frac{[PO_4^{3-}]^{l,h}}{[PO_4^{3-}]^{l,h} + K_P} , \tag{21}$$

where j_{exp} represents new/export production of POC (in unit of mol C m^{-2} yr^{-1}), α denotes C:P stoichiometry of POM, h_m is the mixed layer depth, ε denotes the assimilation efficiency factor for P uptake, and K_P denotes the half-saturation constant. The value of ε for the low-mid latitude region is assumed to be 1. In contrast, we assume a lower efficiency for high latitude region because biological production tends to be limited by environmental factors other than phosphate availability (e.g., amount of solar radiation, mixed layer thickness, sea-ice formation, and iron availability). This is used as one of the fitting parameters in the model. Downwelling waters contain a certain level of nutrients (i.e., preformed nutrients).

In our standard run, the stoichiometry of organic matter is parameterized using the canonical Redfield ratio (C:N:P = 106:16:1) (Redfield et al., 1963). However, we note that flexible C:N:P stoichiometry has been the subject of recent discussion. In the modern oceans, C:N:P ratios of exported POM vary across latitude, reflecting ecosystem structure (Galbraith and Martiny, 2015). Local observations (and laboratory experiments) suggest that the C:N:P ratio of cyanobacteria is a function of seawater PO_4^{3-} concentration (Larsson et al., 2001). The evolutionary perspective has also been discussed (Quigg et al., 2003; Sharoni and Halevy, 2022). In the previous version of the CANOPS model, the C-N-P stoichiometry of primary producers responds dynamically to P availability in the surface layer (Reinhard et al., 2017b):

$$\alpha = \alpha^* + \frac{\alpha_{max} - \alpha^*}{2} \left(1 + \tanh \left(\frac{\gamma_{P0} - [PO_4^{3-}]}{\gamma_{P1}} \right) \right), \tag{22}$$

$$\beta = \beta^* + \frac{\beta_{max} - \beta^*}{2} \left(1 + \tanh \left(\frac{\gamma_{P0} - [PO_4^{3-}]}{\gamma_{P1}} \right) \right), \tag{23}$$

754 where α and β represent the C/P ratio and N/P ratio of POM, * denotes the canonical Redfield ratios, max

755 denotes the maximum value ($\alpha_{max} = 400$ and $\beta_{max} = 60$), and γ_{P0} and γ_{P1} are tunable constants ($\gamma_{P0} = 0.1$ μM

756 and $\gamma_{P1} = 0.03$ μM) (Kuznetsov et al., 2008). In the CANOPS-GRB model, this dynamic response of POM

757 stoichiometry can be explored by changing the user-flag from the standard static response. In this study, we

758 do not explore the impacts of flexible POM stoichiometry on global biogeochemistry (i.e., $\alpha_{max} = \alpha^*$ and

759 $\beta_{max} = \beta^*$).

760

761 Biological production in the surface mixed layer increases the concentration of dissolved O_2 and reduces the

762 concentrations of DIP and DIN according to the stoichiometric ratio (R1 and R2; Table 1). DIN

763 consumption is partitioned between nitrate and ammonium, assuming that ammonium is preferentially

764 assimilated. CANOPS-GRB evaluates the availability of fixed N in the surface ocean, and any N deficiency

765 required for a given level of productivity is assumed to be compensated for on geologic time scales by N

766 fixers. In other words, it is assumed that biological N fixation keeps pace with P availability, so that P (not

767 N) ultimately determines oceanic biological productivity.

768

769 To date, models of varying orders of complexity have been developed to simulate oceanic primary

770 production and nutrient cycling in the euphotic layer, from a single nutrient and single phytoplankton

771 component system to the inclusion of multiple nutrients and trophic levels in the marine ecosystem, usually

772 coupled to physical models (e.g., (Yakushev et al., 2007; Oguz et al., 2000)). To avoid this level of

773 complexity, we introduce a parameter, f_{exp}, called export ratio (Sarmiento and Gruber, 2006), which relates

774 the flux densities of export production and NPP, as follows:

$$j_{NPP}^{ocn} = \frac{j_{exp}}{f_{exp}}, \tag{24}$$

775 where j_{NPP}^{ocn} denotes the NPP in terms of mol C m^{-2} yr^{-1}. In the modern ocean globally averaged value of f_{exp}

776 is estimated at 0.2 (Laws et al., 2000), and we assumed this value in this study. The rate of recycling of

777 organic matter in the photic zone is thus given by

$$j_{\text{recy}} = j_{\text{NPP}}^{\text{ocn}} - j_{\text{exp}} = \frac{1 - f_{\text{exp}}}{f_{\text{exp}}} j_{\text{exp}}. \qquad (25)$$

778 The respiration pathway of j_{recy} depends on the availability of terminal electron acceptors (O_2, NO_3^- and

779 SO_4^{2-}). Following exhaustion of these species as terminal electron acceptors, organic matter remineralization

780 occurs by methanogenesis (R7). See below for the treatment of organic matter remineralization in the water

781 column.

Biological pump

783 Most POM exported to the deep sea is remineralized in the water column before reaching the seafloor (e.g.,

784 (Broecker and Peng, 1982)). Nutrients returning to seawater at intermediate depths may rapidly return to the

785 surface ocean and support productivity. The remaining fraction of POM that reaches the sediment ultimately

786 exerts an important control on oceanic inventories of nutrients and O_2. An adequate representation of the

787 strength of biological pump is therefore critical to any descriptions of global biogeochemical cycles.

788

789 The governing equation of the concentration of biogenic particles G is

$$\frac{\partial G}{\partial t} + v_{\text{POM}} \frac{\partial G}{\partial z} = -rG \qquad (26)$$

790 where r is a decomposition rate and v_{POM} is the settling velocity of POM in the water column. We assume a

791 settling velocity of 100 m d^{-1} for our reference value (e.g., (Suess, 1980)), although a very wide range of

792 values and depth-dependency have been reported (e.g., (Berelson, 2001a)). Therefore, the settling velocity is

793 fast enough to neglect advective and diffusive transport of biogenic particles. Note that the settling velocity

794 would affect the intensity of biological pump and chemical distribution in the ocean interior. Considering the

795 ballast hypothesis in the modern ocean (Armstrong et al., 2001; Francois et al., 2002; Ittekkot, 1993; Klaas

796 and Archer, 2002), the settling velocity of POM in the geological past would very likely have been different

797 from the modern ocean. As (Kashiyama et al., 2011) pointed out, there would be a critical aspect among

798 sinking rate of POM, intensity of biological pump and chemical distribution in the ocean. The quantitative

799 and comprehensive evaluation of their effect is an important issue for the future work (Fakhraee et al., 2020).

800

801 In order to solve equation (26) explicitly, a relatively small time step (~1 day) would be required. However,
802 because the sinking velocity and remineralization of biogenic material are fast processes, we assume that the
803 POM export and remineralization occurs in the same time step (ignoring the term $\partial G/\partial t$). Then the
804 concentration of biogenic particles can be solved as follows:

$$G(z + \Delta z) = G(z)\exp\left(-\frac{r\Delta z}{v_{POM}}\right). \tag{27}$$

805 where Δz is a spatial resolution of the model.

Organic matter decomposition

807 As POM settles through the water column, it is nearly entirely decomposed back to dissolved tracers.
808 Therefore, decomposition of POM is a key process for modelling biogeochemistry in the ocean. To avoid
809 the complex treatment of this process (such as repackaging and aggregation/dispersal of particles), various
810 empirical schemes for POM sinking flux have been proposed, such as exponential (Volk and Hoffert, 1985)
811 or power law (Martin et al., 1987) functions (Fig. 5). However, the estimation of Volk and Hoffert generally
812 tends to overestimate in the upper water column (<1.5 km) and underestimate at depth. It is important to
813 note that data series of sediment trap measurements were obtained from a limited geographic and depth
814 range. Berelson (2001b) and Lutz et al. (2002) conducted further estimates of the sediment flux and found
815 regional variability in the sinking flux. Broadly, these data indicate that commonly applied flux relationships
816 generally tend to overestimate flux to depth.

817

818

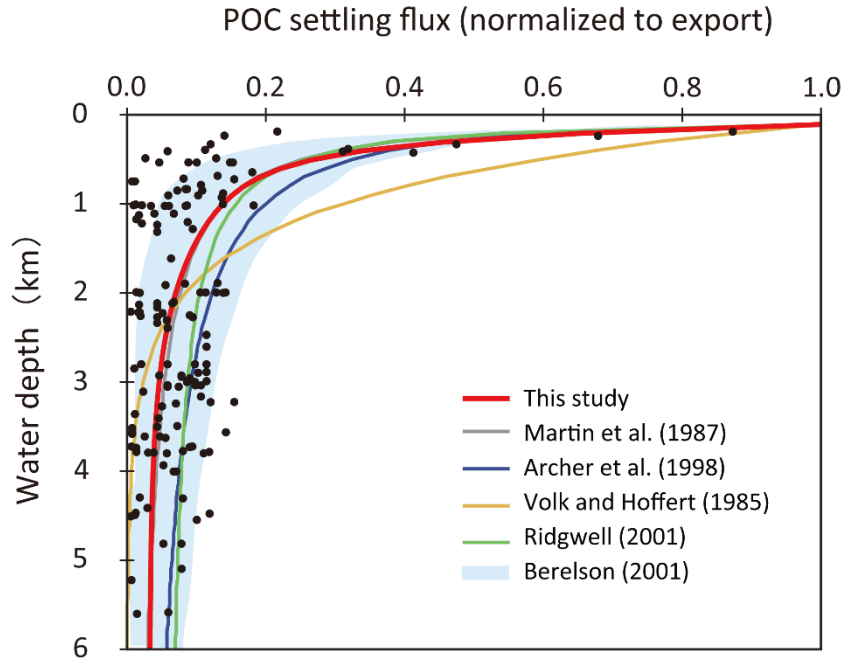



Figure 5. Empirical relationships between POC settling flux normalized to export production (Lutz et al., 2002) and water depth (Archer et al., 1998; Berelson, 2001b; Martin et al., 1987; Volk and Hoffert, 1985). The profile of the CANOPS-GRB model is depicted as a red line. The black dots represent observational data (Honjo and Manganini, 1993; Lutz et al., 2002; Tsunogai and Noriki, 1991; Honjo, 1980) (and references therein).

824

The microbial degradation of different groups of organic matter with different labilities differs over timescales ranging from hours to millions of years. In order to represent the decrease in POM lability with time and water depth, we adopt the so-called multi-G model (Westrich and Berner, 1984) that describes the detailed kinetics of organic matter decomposition (Ozaki and Tajika, 2013; Ozaki et al., 2011). In the CANOPS model, POM is described using two degradable fractions (G_1 and G_2) and one inert (G_3) fraction using different rate constants k_i (i=1, 2, 3) for each component. Rate constants are tuned on the basis of consistency with the typical profile of the POM sinking flux estimated from sediment trap studies (Fig. 5). In this study, constant stoichiometries between C, N, and P during the remineralization of POM are assumed throughout the water column, taking values equal to those characterizing mean export production.

834

835   The electron acceptor used in the respiration reaction changes from dissolved O_2 to other oxidants (e.g.,

836   NO_3^- and SO_4^{2-}) as O_2 becomes depleted. The respiration pathway is controlled by the free energy change

837   per mole of organic carbon oxidized. The organic matter decomposition is performed by the oxidant which

838   yields the greatest free energy change per mole of organic carbon oxidized. When the oxidant is depleted,

839   further decomposition will proceed utilizing the next most efficient (i.e., the most energy producing) oxidant

840   until either all oxidants are consumed or oxidizable organic matter is depleted (e.g., (Froelich et al., 1979;

841   Berner, 1989)). In oxic waters, organic matter is remineralized by an aerobic oxidation process (R4). As

842   dissolved O_2 is depleted, NO_3^- and/or SO_4^{2-} will be used (R5 and R6). Denitrification is carried out by

843   heterotrophic bacteria under low concentrations of dissolved O_2, if there is sufficient nitrate. For anoxic,

844   sulfate-lean oceans, the methanogenic degradation of organic matter will occur (R7). In the CANOPS-GRB

845   model, we parameterized the dependence of decomposition of POM with a Michaelis-Menten type

846   relationship with respect to the terminal electron acceptors:

$$R_4 = \frac{[O_2]}{K_{O_2} + [O_2]}\left(\sum k_i G_i\right) \tag{28}$$

$$R_5 = \frac{K'_{O_2}}{K'_{O_2} + [O_2]}\frac{[NO_3^-]}{K_{NO_3} + [NO_3^-]}\left(\sum k_i G_i\right) \tag{29}$$

$$R_6 = \frac{K'_{O_2}}{K'_{O_2} + [O_2]}\frac{K'_{NO_3}}{K'_{NO_3} + [NO_3^-]}\frac{[SO_4^{2-}]}{K_{MSR} + [SO_4^{2-}]}\left(\sum k_i G_i\right) \tag{30}$$

$$R_7 = \frac{K'_{O_2}}{K'_{O_2} + [O_2]}\frac{K'_{NO_3}}{K'_{NO_3} + [NO_3^-]}\frac{K'_{MSR}}{K'_{MSR} + [SO_4^{2-}]}\left(\sum k_i G_i\right)$$
$$= \left(1 - R_{O_2} - R_{NO_3} - R_{SO_4}\right)\left(\sum k_i G_i\right) \tag{31}$$

847   where K_{O2}, K_{NO3}, and K_{MSR} are Monod constants, and K'_{O2}, K'_{NO3}, K'_{MSR} are inhibition constants. The

848   Monod-type expressions are widely used in mathematical models of POM decomposition processes (e.g.,

849   (Boudreau, 1996)). The oxidants for organic matter decomposition change with the availability of each

850   oxidant, which vary with time and water depth. The parameter values are based on previous studies on early

851   diagenetic processes in marine sediments (Boudreau, 1996; Van Cappellen and Wang, 1996). SO_4^{2-} has been

852   one of the major components of the Phanerozoic oceans and has been an important oxidizing agent in

853 anaerobic systems. In the original CANOPS model (Ozaki and Tajika, 2013; Ozaki et al., 2011), it was

854 assumed that the saturation constant K_{MSR} is zero, meaning that the SO_4^{2-} is never a limiting factor. In

855 contrast, during the Precambrian, seawater SO_4^{2-} could have been extremely low (Lyons and Gill, 2010).

856 The half saturation constant for MSR (K_{MSR}) determines the degree to which MSR contributes to the total

857 respiration rates. However, estimates for K_{MSR} in natural environments and pure cultures vary over several

858 orders of magnitude (~0.002–3 mM) (Tarpgaard et al., 2011; Pallud and Van Cappellen, 2006). We assume

859 a reference value of 0.2 mM for this study.

860

861 Finally, temperature may also have played an important role in organic matter decomposition rates. The

862 dependence of ammonification on temperature is sometimes described by an exponential function or Q_{10}

863 function (e.g., (Yakushev et al., 2007)). While we recognize that the temperature dependency of organic

864 matter decomposition might have played an important role in oceanic biogeochemical cycles in the

865 geological past (Crichton et al., 2021), these dynamics are not included in CANOPS-GRB v1.0.

866 **Secondary redox reactions**

867 Total ammonia (ΣNH_3), total sulfide (ΣH_2S), and methane (CH_4), produced during organic matter

868 degradation, are subject to oxidation to NO_3^-, SO_4^{2-}, and CO_2 via a set of secondary redox reactions (Table

869 1). Rate constants for these reactions are taken from the literature. The ocean model includes nitrification

870 (R8), total sulfide oxidation by O_2 (R9), aerobic oxidation of CH_4 by O_2 (R10), and AOM by SO_4^{2-} (R11).

871 Nitrification, the oxidation of ammonium to nitrate, occurs in several stages and is accomplished mainly by

872 chemolithotrophic bacteria (Sarmiento and Gruber, 2006). In this study, we treat all nitrification reactions as

873 a combined reaction (R8). The rate of this process is assumed to depend on the concentration of both oxygen

874 and ammonia as follows:

$$R_8 = k_{R8}[NH_4^+][O_2]. \qquad (32)$$

875

876 The oxidation of sulfide formed in anoxic waters by MSR can also be written as a series of reactions (e.g.,

877 (Yakushev and Neretin, 1997)), but we treat it as an overall reaction (R9). The rate of this secondary redox

878 reaction is also formulated using a bimolecular rate law:

$$R_9 = k_{R9}[\Sigma H_2S][O_2].\tag{33}$$

879 The rate constant for this process has been shown to vary significantly as a function of several redox-
880 sensitive trace metals which act as catalysts (Millero, 1991). Here we assume k_{R9} = 3650 mM^{-1} yr^{-1} based on
881 the observations of the chemocline of the Black Sea (Oguz et al., 2001).

882

883 In the original CANOPS model (Ozaki et al., 2019a; Ozaki and Tajika, 2013), syngenetic pyrite formation in
884 the water column was not considered. In a more recent revision of the model, this process was added (Cole
885 et al., 2022) and parameterized such that iron sulfide formation is assumed to be proportional to the
886 hydrogen sulfide concentration:

$$R_{pyr}^{wc} = k_{pyr}^{wc}[\Sigma H_2S],\tag{34}$$

887 where k_{pyr}^{wc} is a model constant (its reference value is set at 0.01 yr^{-1}). This constant is a function of the
888 ferrous iron concentration in seawater, but it is the subject of large uncertainty. The total flux (in mol S yr^{-1})
889 can be obtained by integrating the precipitation flux density over the whole ocean:

$$J_{pyr}^{wc} = \int R_{pyr}^{wc} \frac{dV}{dz} dz.\tag{35}$$

890

891 The aerobic oxidation of CH$_4$ is formulated using a bimolecular rate law:

$$R_{10} = k_{R10}[CH_4][O_2].\tag{36}$$

892 The rate of AOM is formulated using a Monod-type law (Beal et al., 2011):

$$R_{11} = k_{R11}[CH_4]\frac{[SO_4^{2-}]}{K_{AOM} + [SO_4^{2-}]}.\tag{37}$$

893 Rate constants for above reactions are taken from the literature (Table 4). Secondary redox reactions were
894 calculated implicitly with an operator splitting scheme (Steefel and Macquarrie, 1996) so as to maintain
895 numerical stability.

 2.4.4 Sediment-water exchange

 The burial of biogenic material in marine sediments plays a critical role in global biogeochemical cycles,

 especially with respect to the marine budgets of nutrients, carbon, and sulfur. This is intimately linked to

 atmospheric O_2 levels on geologic timescales. Specifically, the burial rate of C_{org} in marine sediments exerts

 a primary control on the evolution of atmospheric O_2 levels throughout Earth's history. Given the

 complexity of biogeochemical processes within sediments and our limited knowledge on many of the early

 diagenetic processes, we adopt some semi-empirical relationships extracted from ocean sediment data. This

 approach, rather than explicit modelling, is also required to reduce the computational cost of the simulation

 on timescales >100 Myr. The related parameter values are listed in Table 5.

 Table 5. Parameters used in the sediment-water interface module of CANOPS-GRB.

Parameters	Label	Value	Unit	Ref.
Scale constant for benthic aerobic respiration	k	0.02	yr^{-1}	(Ozaki et al., 2019a)
Burial efficiency of C_{org} at zero sediment accumulation rate	be_1	5.0	%	(Dale et al., 2012)
Burial efficiency of C_{org} at infinite sediment accumulation rate	be_2	75	%	(Dale et al., 2012)
Centre of the regression for burial efficiency of organic C	a	0.01	$g\ cm^{-2}\ yr^{-1}$	(Dale et al., 2012)
Pyrite burial efficiency in oxic sediments	e_{pyr}^{oxic}	0.117	–	This study
Pyrite burial efficiency in anoxic sediments	e_{pyr}^{anox}	1	–	(Ozaki et al., 2019a)
O_2 threshold value for P burial efficiency	$oxic$	0.25	mM	(Ozaki et al., 2011)
C_{org}/N_{org} ratio of buried sediments	$(C_{org}/N_{org})_b$	10	$mol\ mol^{-1}$	(Ozaki and Tajika, 2013)

 POM deposition

 The fraction of settling POM that reaches the sediment surface, J_{org}^{dep} (in mol C yr^{-1}) is a function of both

 the settling flux density, j_{org}^{dep} (in mol C $m^{-2}\ yr^{-1}$), and topography (Fig. 4a):

$$J_{org}^{dep} = \int_{z_1}^{z_2} j_{org}^{dep}(z)\frac{dA}{dz}dz, \tag{38}$$

 where the settling flux density can be written as follows:

$$j_{org}^{dep} = v_{POM}G. \tag{39}$$

 where G is the concentration of POM, and v_{POM} denotes the sinking velocity.

Carbon cycling

Interactions between the ocean and underlying sediments play an important role in influencing whole-ocean chemical and nutrient inventories on geologic time scales. POM deposited to the seafloor is subject to decomposition during diagenetic processes associated with burial in marine sediments. Only a small fraction of organic matter will ultimately be buried and removed from the surface environment. However, understanding what factors control the preservation of organic matter in marine sediments has been a controversial topic, and we still lack a robust understanding of this process. With this issue in mind, we adopt an empirical approach obtained using the observational data from previous studies.

The burial flux density of C_{org} at each water depth, $j_{org}^{b,ocn}$ (in terms of mol C m^{-2} yr^{-1}), is calculated based on burial efficiency, BE_{org}:

$$j_{org}^{b,ocn} = BE_{org}\, j_{org}^{dep}. \tag{40}$$

Burial efficiency is defined as the fraction of organic matter buried in sediments relative to the total depositional flux. Burial efficiency is described by simplified parametric laws based on empirical relationships from modern day observations. Previous studies demonstrate strong dependency of this term on total sedimentation rate, SR (e.g., (Henrichs and Reeburgh, 1987)). Figure 6 demonstrates the relationship between BE_{org} and SR compiled from literature surveys. The sedimentation rate in the modern ocean varies over about five orders of magnitude, with a primary dependence on material supplied from the continents. There is a strong relationship, especially for SR less than 0.01 cm yr^{-1}. In contrast to the strong SR dependence under oxic conditions, anoxic settings show a much weaker dependence of BE_{org} on SR (Betts and Holland, 1991; Henrichs and Reeburgh, 1987) (Fig. 6). In this study, the following relationship proposed by (Henrichs and Reeburgh, 1987) is adopted for sediments underlying well-oxygenated bottom water (O$_2$ concentration of bottom water, $[O_2]_{bw} > 200$ μM):

$$BE_{org} = \frac{SR^{0.4}}{2.1}. \tag{41}$$

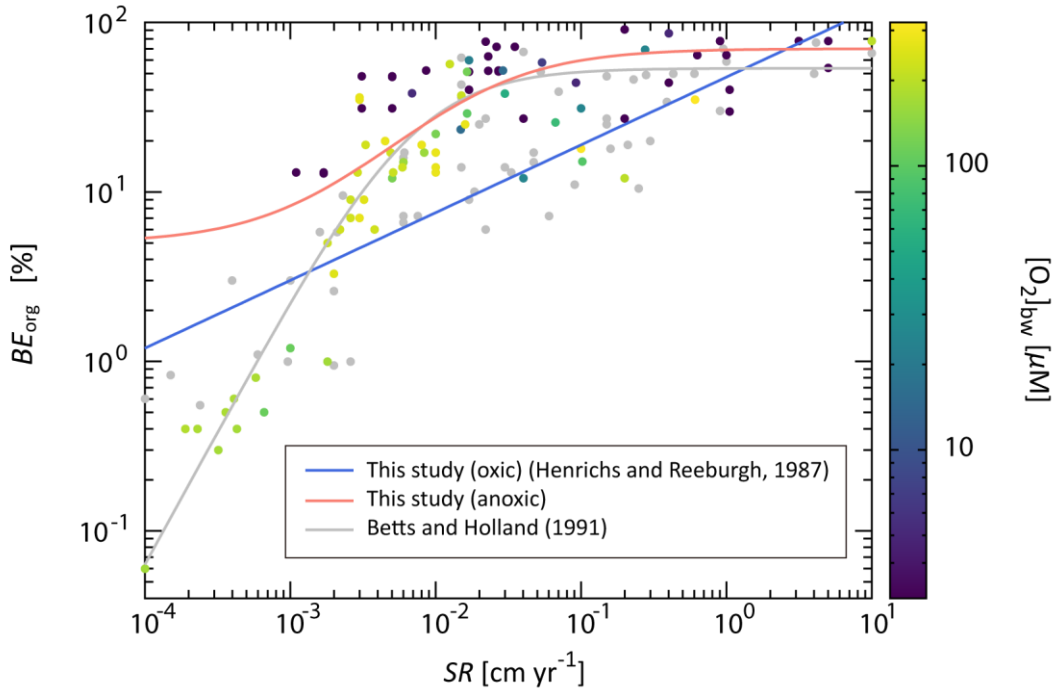
Given that BE_{org} depends on the $[O_2]_{bw}$ (Lasaga and Ohmoto, 2002; Katsev and Crowe, 2015), we adopt the following formulation for sediments underlying less-oxygenated bottom waters ($[O_2]_{bw} < 30$ μM) (Dale et al., 2012):

$$BE_{org} = b_2 + \frac{b_1 - b_2}{1 + SR/a},$$
(42)

937 where $a = 0.019$, $b_1 = 0.05$, and $b_2 = 0.7$, respectively. For intermediate $[O_2]_{bw}$ levels, BE_{org} is evaluated as a
938 function of $[O_2]_{bw}$ with a log-linear interpolation method. Note that the original CANOPS model (Ozaki and
939 Tajika, 2013; Ozaki et al., 2011) adopted Eq. (41) without considering the O_2 dependency, whereas more
940 recent versions employ Eq. (42) for both oxic and anoxic sediments with different values of a, b_1 and b_2. In
941 CANOPS-GRB v1.0, we adopted both equations, because of the sake of more accurate reproduction of C_{org}
942 burial distribution in the modern ocean (Sect. 3.2.2).

943

944

945

946 **Figure 6.** Burial efficiency of organic carbon (BE_{org}) as a function of sedimentation rate (SR). The dots
947 denote the observational data compiled from literature survey (Betts and Holland, 1991; Canfield, 1993;
948 Henrichs and Reeburgh, 1987; Tromp et al., 1995; Hartnett et al., 1998). The color represents the O_2
949 concentration of bottom water, $[O_2]_{bw}$, with grey dots for the unknown $[O_2]_{bw}$ value. Blue and red lines are
950 the relationship for well-oxygenated ($[O_2]_{bw} > 200\ \mu M$) (Henrichs and Reeburgh, 1987) and anoxic ($[O_2]_{bw}$
951 $< 30\ \mu M$) marine sediments adopted in the CANOPS-GRB model, whereas grey solid line is a previously
952 proposed empirical relationship proposed by (Betts and Holland, 1991).

953

Sedimentation rate depends strongly on water depth and distance from shore (Hedges et al., 1999), and we apply the relationship between water depth, z, and the reference value of SR shown by (Tromp et al., 1995) (Fig. 7).

$$z = 2700 \times erfc\left(2.1 + \log SR^*\right).$$
(43)

Using these formulas with seafloor topography (Fig. 4a) and j_{org}^{dep} (Eq. 39), we can calculate $j_{org}^{b,ocn}$ for each ocean depth. In the CANOPS-GRB model, we also introduce an erosion factor, f_R, representing the global weathering/sedimentation rate (Sect. 2.4.3). Given the intimate coupling between global erosion rate and mass transfer from continents to the ocean, SR scales with the erosion factor ($f_R = 1$ for our reference run):

$$SR(z) = f_R \, SR(z)^*.$$
(44)

961

962

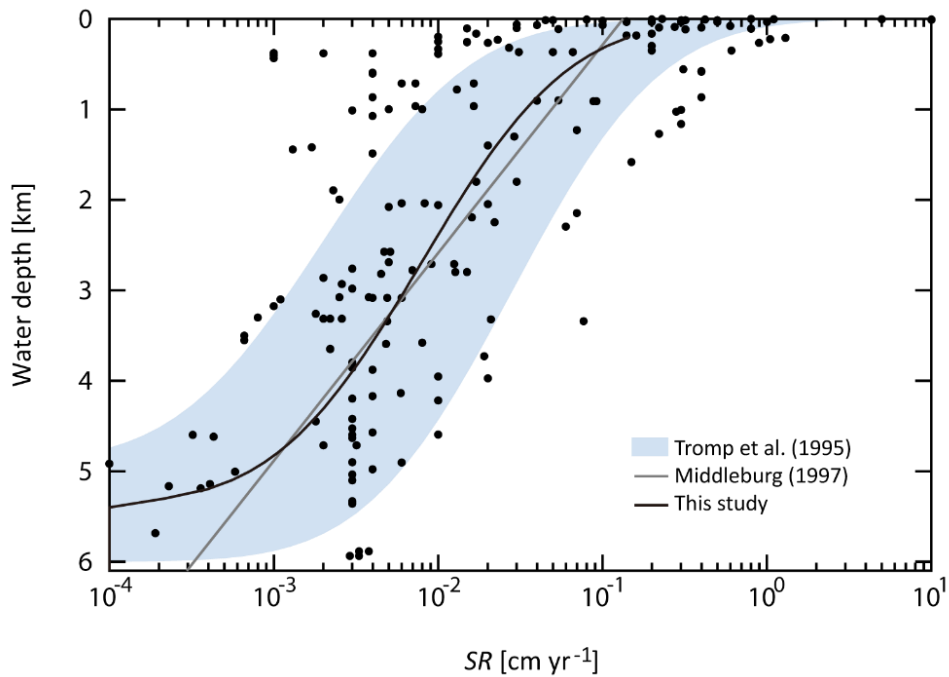



Figure 7. Sedimentation rate as a function of water depth. Data (black dots) were compiled from literature survey (Colman et al., 2000; Baturin, 2007; Betts and Holland, 1991; Tromp et al., 1995; Cha et al., 2005; Reimers et al., 1992). Black line represents the relationship assumed in the CANOPS-GRB model. Previously estimated empirical relationships (Middelburg et al., 1997; Tromp et al., 1995) are also shown.

967

Organic matter that is not buried is subject to decomposition. The benthic decomposition rate at each water depth is given as follows:

$$j_{recy}^{sed} = j_{org}^{dep} - j_{org}^{b,ocn} = \left(1 - BE_{org}\right) j_{org}^{dep}.$$ (45)

The respiration pathway used in the benthic decomposition is evaluated based on semi-empirical relationships obtained by 1-D early diagenesis models (see below). The fraction of aerobic degradation in total sedimentary respiration, f_{aero}, is calculated based on oxygen exposure time (τ_{OET}):

$$f_{aero} = \left(1 - f_{deni}\right)\left(1 - e^{-k\tau_{OET}}\right),$$ (46)

where f_{deni} denotes the fraction of denitrification and k is an empirical constant. τ_{OET} is given by

$$\tau_{OET} = \frac{OPD}{SR},$$ (47)

where OPD is the oxygen penetration depth (cm) and SR denotes a linear sedimentation rate (cm yr^{-1}). In the CANOPS-GRB model OPD is calculated by a simplified parametric law obtained from a 1-D early-diagenetic model of C and O$_2$. We performed a series of experiments (n = 5,652) in order to parameterize OPD as a polynomial function with the following variables: sedimentation rate SR (cm yr^{-1}), bottom water O$_2$ concentration [O$_2$]$_{bw}$ (μM), depositional flux of POC j_{org}^{dep} (mmol C cm^{-2} yr^{-1}) and bottom water temperature T_{bw} (°C). The variables are allowed to vary over a parameter space spanning 10^{-4} cm yr^{-1} < SR < 10^1 cm yr^{-1}, 10^0 μM < [O$_2$]$_{bw}$ < 10^3 μM, 10^{-4} mmol C cm^{-2} yr^{-1} < j_{org}^{dep} < 10^1 mmol C cm^{-2} yr^{-1}, and 0 °C < T_{bw} < 30 °C.

$$
\begin{aligned}
\log OPD \; = \; & a_0 + a_1 \log SR + a_2 \log[O_2]_{bw} + a_3 \log j_{org}^{dep} + a_4 (\log SR)^2 \\
& + a_5 (\log[O_2]_{bw})^2 + a_6 (\log j_{org}^{dep})^2 + a_7 (\log SR)(\log[O_2]_{bw}), \\
& + a_8 (\log[O_2]_{bw})(\log j_{org}^{dep}) + a_9 (\log SR)(\log j_{org}^{dep}) + a_{10} T_{bw}
\end{aligned}
$$ (48)

where a_0 = -2.24869, a_1 = 0.110645, a_2 = 1.12569, a_3 = -0.281005, a_4 = 0.014827, a_5 = -0.124721, a_6 = 0.0894604, a_7 = 0.00279531, a_8 = -0.127797, a_9 = 0.0017995, and a_{10} = 0.0085171. This parametric fit provides a rapid means of obtaining OPD from a 1-D early diagenetic model of C and O$_2$ (Fig. 8). Note that Eq. (48) is verified for [O$_2$]$_{bw}$ >1 μM. When bottom water O$_2$ concentration is lower than 1 μM, OPD is set at zero.

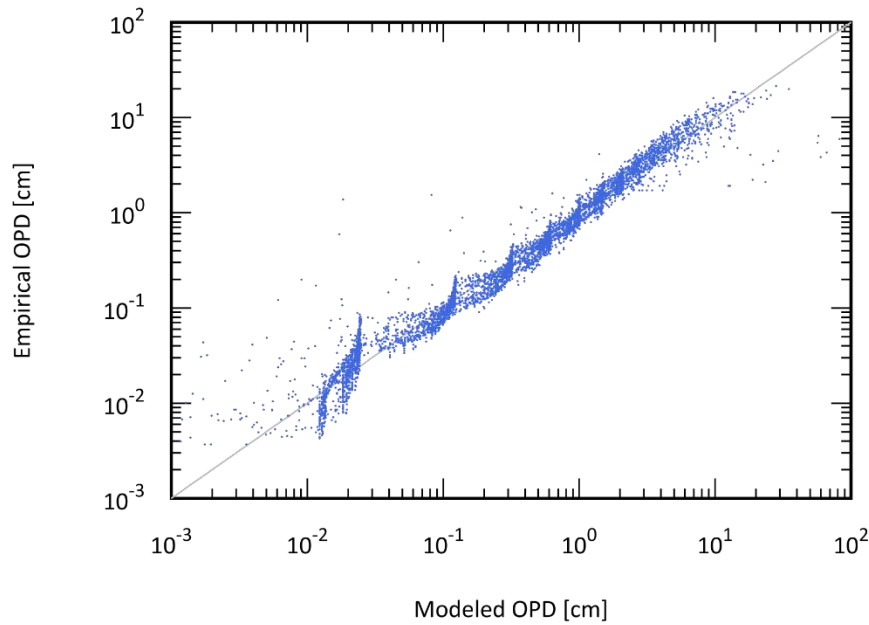



Figure 8. The correlation between the simulated OPD and the OPD obtained from an empirical relationship of equation (48) ($n = 5652$). Gray line denotes the 1:1 line. $r^2 = 0.9595$.

Phosphorus cycling

Marine P inventory is controlled not only by the riverine P input flux from land but also by the efficiency of P recycling in marine sediments (Van Cappellen and Ingall, 1994). Because the estimated P diffusive flux from seafloor sediments is much greater than the riverine P flux (Delaney, 1998; Hensen et al., 1998; Ruttenberg, 2003; Mcmanus et al., 1997; Wallmann, 2003; Wallmann, 2010), changes in diagenetic processes affecting P recycling and burial in marine sediments could have a significant impact on global oceanic biogeochemical cycles.

1000

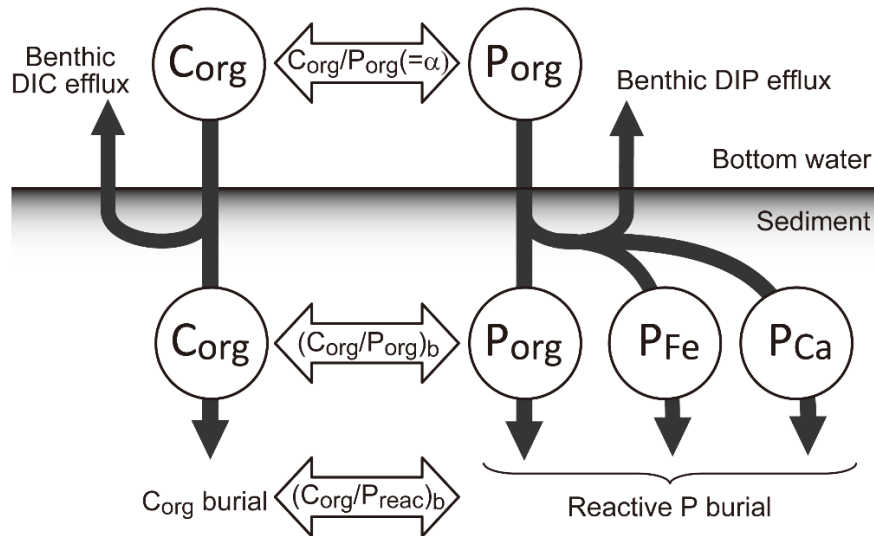



Figure 9. Schematic of P burial in marine sediments. The primary source of P to the sediment is the deposition of organic matter, which represents the C:P ratio of primary producers, α. Most of the deposited organic P is decomposed before burial and the DIP released to pore waters diffuses to the bottom water. A fraction of the liberated P is trapped by iron hydroxides or buried as authigenic minerals (e.g., carbonate fluorapatite). Phosphorus deposited in sediments is a subject of decomposition and sink-switching. Three reactive phases, organic P (P_{org}), Fe-bound P (P_{Fe}), and Ca-bound P (P_{Ca}), are considered in the CANOPS-GRB model. The burial of these species are redox-dependent: burial efficiency is affected by bottom water O_2 concentration. Because of the sink-switching, sedimentary C_{org}/P_{reac}, rather than C_{org}/P_{org} ratios, provides a correct measure of the retention versus diffusive loss of remineralized P.

A schematic of benthic P cycling is shown in Fig. 9. The majority of organic matter delivered to the sediment-water interface is regenerated (Jahnke, 1996), but a fraction of DIP released via respiration to pore waters is redistributed to other phases such as iron-hydroxide or carbonate fluorapatite within the sediments. This mechanism is known as 'sink-switching' (e.g., (Anderson et al., 2001; Filippelli, 2001)), and results in P burial other than organic P playing a more important role in the total P sink (Ruttenberg, 1993, 2003; Compton et al., 2000). Three different P pools are considered in the CANOPS-GRB model: organic P (P_{org}), Fe-hydroxide-bound P (P_{Fe}), and authigenic Ca-bound P (P_{Ca}). The sum of these pools is defined as biologically reactive P (P_{reac}) (bioavailable in the ocean to fuel primary productivity). The marine C and P cycles are coupled not only through the C_{org}/P_{org} ratio of POM (α) but also through the C_{org}/P_{reac} ratio of marine sediments. It is important to note that as argued by Anderson et al. (2001), the fundamental measure

of the retention versus diffusive loss of remineralized P is not the sedimentary C_{org}/P_{org} ratio, but the ratio of C_{org}/P_{reac}.

Field observations of marine and lacustrine sediments have revealed that the burial efficiency of P depends on the redox conditions of bottom waters (Ingall and Jahnke, 1994): Phosphorus retention potential is suppressed under anoxic bottom water conditions. Elevated C_{org}/P_{reac} ratios observed in permanently anoxic environments suggest preferential regeneration of P relative to C under these conditions (Algeo and Ingall, 2007; Anderson et al., 2001; Colman et al., 2000; Filippelli, 2001; Ingall and Jahnke, 1997). In the CANOPS-GRB model, P benthic regeneration rate is calculated at each sediment segment based on the POP depositional flux density j_P^{dep} ($=j_{org}^{dep}/\alpha$) and P burial efficiency which is a function of both $[O_2]_{bw}$ and SR. We assume the following formulation for the C_{org}/P_{org} ratio of the buried organic phase, on the basis of previous studies (Slomp and Van Cappellen, 2007; Van Cappellen and Ingall, 1994, 1996):

$$(C_{org}/P_{org})_b = \begin{cases} \dfrac{(C_{org}/P_{org})_b^{oxic}(C_{org}/P_{org})_b^{anox}}{(C_{org}/P_{org})_b^{anox}\dfrac{[O_2]_{bw}}{oxic}+\left(1-\dfrac{[O_2]_{bw}}{oxic}\right)(C_{org}/P_{org})_b^{oxic}} f_\tau & \text{for } [O_2]_{bw} < oxic \\[2em] (C_{org}/P_{org})_b^{oxic} f_\tau & \text{for } [O_2]_{bw} \geq oxic \end{cases}, \qquad (49)$$

where $(C_{org}/P_{org})_b^{oxic}$ and $(C_{org}/P_{org})_b^{anox}$ denote organic C/P ratios for fully oxic and anoxic conditions, and *oxic* ($=250\ \mu M$) is a threshold value of $[O_2]_{bw}$ below which preferential P regeneration occurs. $(C_{org}/P_{org})_b^{oxic}$ is set to twice the value of the Redfield ratio, and $(C_{org}/P_{org})_b^{anox}$ is an upper limit assumed for organic matter buried under fully anoxic overlying waters, estimated as 40 times larger than the Redfield value on the basis of previous studies on various ancient anoxic basin sediments (Slomp and Van Cappellen, 2007; Ingall et al., 1993). In Eq. (49), we also include the dependence of buried C_{org}/P_{org} ratio on SR, expressed as f_τ. Modern observations suggest that SR is a one of the major factors influencing the preservation versus remineralization of sedimentary organic C and P. Organic C preservation in marine sediments tends to be enhanced at higher SR. In contrast, the C_{org}/P_{org} ratio of sedimentary organic matter shows a non-linear relationship with respect to SR (Ingall and Cappellen, 1990) (Sect. 3.2.3), suggesting more complex behavior of benthic P cycling. Specifically, in the pelagic deep ocean, preferential P regeneration is not

1044 observed, likely due to the long timescale of diagenesis prior to burial (Ingall and Cappellen, 1990). In the

1045 CANOPS model, f_τ is formulated as follows:

$$f_\tau = 0.5 + 0.5 \exp\left(-\frac{0.001\,\text{cm}}{SR}\right). \tag{50}$$

1046 Specifically, the C_{org}/P_{org} ratio approaches the Redfield value for oxygenated pelagic sediments.

1047

1048 The burial flux density of P_{org} can be calculated as the C_{org} burial flux density divided by $(C_{org}/P_{org})_b$:

$$j_{Porg}^b = \frac{j_{org}^{b,ocn}}{\left(C_{org}/P_{org}\right)_b}. \tag{51}$$

1049 The burial efficiency of P_{org} can be written as follows:

$$BE_{Porg} \equiv \frac{j_{Porg}^b}{j_P^{dep}} = \frac{j_{Porg}^b}{j_{org}^{dep}/\alpha} = \frac{\alpha BE_{org}}{j_{org}^b/j_{Porg}^b} = \frac{\alpha BE_{org}}{\left(C_{org}/P_{org}\right)_b}, \tag{52}$$

1050 where j_P^{dep} denotes the POP settling flux density to sediments, which is coupled to the C/P stoichiometry of

1051 POM ($= j_{org}^{dep}/\alpha$).

1052

1053 Under oxic bottom water conditions, remineralized organic P can be trapped efficiently at the sediment-

1054 water interface by ferric iron phases. In contrast, under anoxic bottom water conditions, a lack of ferric iron

1055 phases allows most mineralized P to diffuse out of the sediment. This redox-dependent P burial is assumed

1056 to be linearly proportional to the $[O_2]_{bw}$ (Slomp and Van Cappellen, 2007):

$$j_{P\text{-}Fe}^b = \begin{cases} BE_{Porg}^{oxic}\left(\dfrac{[O_2]_{bw}}{oxic}\right) j_{Porg}^{dep} & \text{for } [O_2]_{bw} < oxic \\[2mm] BE_{Porg}^{oxic} j_{Porg}^{dep} & \text{for } [O_2]_{bw} \geq oxic \end{cases}, \tag{53}$$

1057 where BE_{Porg}^{oxic} denotes the burial efficiency of P_{org} under well-oxygenated bottom water conditions ($[O_2]_{bw}$

1058 $> oxic$). We assume that the retention potential of P_{Fe} in sediments overlain by oxic bottom waters is

1059 comparable to that of P_{org} (Ruttenberg, 1993).

1060

1061 Some authors have also proposed that authigenic P burial, the dominant process for P burial today

1062 (Ruttenberg, 1993), depends on the redox conditions of the bottom water (Slomp and Van Cappellen, 2007;

1063 Slomp et al., 2002). In the CANOPS-GRB model, we adopt the following redox dependence used by Slomp

1064 and Van Cappellen (2007):

$$
j_{\text{P-Ca}}^{\text{b}} = \begin{cases} 2BE_{\text{Porg}}^{\text{oxic}}\left(a_{\text{auth}} + (1-a_{\text{auth}})\dfrac{[O_2]_{\text{bw}}}{\text{oxic}}\right) j_{\text{Porg}}^{\text{dep}} & \text{for } [O_2]_{\text{bw}} < oxic \\ 2BE_{\text{Porg}}^{\text{oxic}} j_{\text{Porg}}^{\text{dep}} & \text{for } [O_2]_{\text{bw}} \geq oxic \end{cases} \tag{54}
$$

1065 We assume that P_{org}, P_{Fe} and P_{Ca} account roughly for 25%, 25%, and 50%, respectively, of the total reactive

1066 P buried in oxygenated sediments (Ruttenberg, 1993). Therefore, the burial efficiency of authigenic P phases

1067 is larger than that of P_{org} and P_{Fe} by a factor of 2. The redox-dependency of authigenic P burial is controlled

1068 by a parameter, a_{auth}. There is still great uncertainty as to the sensitivity of P retention efficiency of

1069 authigenic P phases to changing redox conditions. For instance, recent modeling study suggests that the

1070 burial of authigenic P is influenced not only by the redox state of bottom water, but also by seawater

1071 chemistry (especially Ca^{2+}) (Zhao et al., 2020), temperature, and pH (Papadomanolaki et al. 2022). In our

1072 reference run, we set a_{auth} at 1, i.e. no redox dependency for authigenic P burial.

1073

1074 When above formulations are adopted, the ratio

$$
\overline{C_{\text{org}}/P_{\text{reac}}} = \frac{\text{Marine } C_{\text{org}} \text{ burial rate}}{\text{Marine } P_{\text{reac}} \text{ burial rate}} \tag{55}
$$

1075 varies between 63 and 370 as a function of ocean redox state. This is in the range of an estimation derived

1076 from various observations of modern and ancient sediments (Papadomanolaki et al., 2022; Algeo and Ingall,

1077 2007). Given that the continental shelves are a main locus of reactive P burial, the separate treatment of

1078 continental shelves and margin sediments from the pelagic ocean could affect the non-linearity of the redox-

1079 dependent P cycle. However, this was left as one of the subjects of future work.

Nitrogen cycling

1081 The benthic denitrification rate is estimated with a semi-empirical relationship (Middelburg et al., 1996).

1082 Middelburg and colleagues performed a series of experiments ($n = 2,000$) with a 1-D early diagenetic model

1083 of C-N-O$_2$ to parameterize benthic denitrification $j_{\text{deni}}^{\text{sed}}$ (μmol C cm^{-2} d^{-1}) as a polynomial function using

1084 $j_{\text{org}}^{\text{dep}}$ (μmol C cm^{-2} d^{-1}), z (m), and bottom water concentrations of dissolved O$_2$ and NO$_3^-$ (μM):

$$\log j_{\text{deni}}^{\text{sed}} = c_0 + c_1 \log j_{\text{org}}^{\text{dep}} + c_2 \left(\log j_{\text{org}}^{\text{dep}} \right)^2 + c_3 \log[\text{NO}_3^-]_{\text{bw}} \log[\text{O}_2]_{\text{bw}}$$
$$+ c_4 \log[\text{NO}_3^-]_{\text{bw}} + c_5 \log[\text{O}_2]_{\text{bw}} + c_6 \log z + c_7 \log j_{\text{org}}^{\text{dep}} \log[\text{O}_2]_{\text{bw}} \tag{56}$$

1085   where c_0 = -2.2567, c_1 = -0.1850, c_2 = -0.2210, c_3 = -0.3995, c_4 = 1.2500, c_5 = 0.4721, c_6 = -0.0996, c_7 =

1086   0.4256. This polynomial function was obtained by examining a parameter space spanning 50 m $< z <$ 6,000

1087   m, 10 μM $<$ [O_2]$_{\text{bw}}$ $<$ 350 μM, and 1 μM $<$ [NO_3^-]$_{\text{bw}}$ $<$ 60 μM. $j_{\text{org}}^{\text{dep}}$ was allowed to vary within 2 orders of

1088   magnitude at each water depth (Middelburg et al., 1996). As pointed out by (Romaniello and Derry, 2010),

1089   the predicted contribution of denitrification to total decomposition f_{deni} (= $j_{\text{denitr}}^{\text{sed}}/j_{\text{recy}}^{\text{sed}}$) can sometimes

1090   exceed 100% for [O_2]$_{\text{bw}}$ $<$10 μM. When the fraction of benthic denitrification to total decomposition exceeds

1091   90%, benthic denitrification is limited in order to avoid unphysical values (Ozaki and Tajika, 2013;

1092   Romaniello and Derry, 2010).

1093

1094   The burial flux density of N_{org} is calculated by molar ratio of C to N of buried sediments, $(\text{C}_{\text{org}}/\text{N}_{\text{org}})_{\text{b}}$, and

1095   the burial flux of C_{org}:

$$j_{\text{Norg}}^{\text{b}} = \frac{j_{\text{org}}^{\text{b,ocn}}}{\left(\text{C}_{\text{org}} / \text{N}_{\text{org}} \right)_{\text{b}}} \tag{57}$$

1096   We assumed an average ratio of 10, which is observed in the Washington and Mexico margin (Hedges et al.,

1097   1999; Hartnett and Devol, 2003).

1098   **Sulfur cycling**

1099   The fractions of MSR and methanogenesis to total decomposition of organic matter in marine sediment are

1100   given by

$$f_{\text{MSR}} = \left(1 - f_{\text{aerobic}} - f_{\text{denitr}} \right) \frac{[\text{SO}_4^{2-}]_{\text{bw}}}{[\text{SO}_4^{2-}]_{\text{bw}} + K_{\text{MSR}}}, \tag{58}$$

$$f_{\text{meth}} = 1 - f_{\text{aero}} - f_{\text{deni}} - f_{\text{MSR}} . \tag{59}$$

1101   The production rate of hydrogen sulfide in sediment, $j_{\text{H2S}}^{\text{sed}}$ (mol S m^{-2} yr^{-1}), is given by

$$j_{\text{H}_2\text{S}}^{\text{sed}} = \frac{1}{2} f_{\text{MSR}} j_{\text{recy}}^{\text{sed}} + j_{\text{AOM}}, \tag{60}$$

1102 where j_{AOM} denotes the production rate of sulfide via AOM:

$$j_{AOM} = \frac{1}{2} f_{meth} \frac{[SO_4^{2-}]_{bw}}{[SO_4^{2-}]_{bw} + K_{MSR}} j_{recy}^{sed}. \tag{61}$$

1103 Here we assume that AOM is proportional to the CH$_4$ production rate with a sulfate-dependent term.

1104

1105 The rate of pyrite precipitation in sediments would be proportional to the sulfide production rate at the

1106 sediment-water interface:

$$j_{pyr}^{b,sed} = e_{pyr} j_{H_2S}^{sed}, \tag{62}$$

1107 where the proportional coefficient, e_{pyr}, is the pyrite burial efficiency. The rate of MSR is a function of the

1108 marine redox state, [SO$_4^{2-}$], and the availability of degradable organic matter. In the well-oxygenated

1109 modern oceans most sulfide produced in sediments is reoxidized and only a few per cent of total sulfide is

1110 buried as pyrite (Canfield, 1991; Lin and Morse, 1991; Turchyn and Schrag, 2004; Bowles et al., 2014;

1111 Jørgensen, 1982). It has been pointed out that efficient oxidation of sulfide is promoted by animal

1112 bioturbation (Berner and Westrich, 1985; Canfield and Farquhar, 2009). In contrast, the value of e_{pyr} for

1113 anoxic sediments is much greater due to the absence of bioturbation and enhanced sulfide production. We

1114 assume that e_{pyr} asymptotes toward unity with decreasing the bottom water [O$_2$] (Tarhan et al., 2015):

$$e_{pyr} = e_{pyr}^{max} - \left(e_{pyr}^{max} - e_{pyr}^{*} \right) \tanh[O_2]_{bw}, \tag{63}$$

1115 where e_{pyr}^{max} (= 1 in our reference run) denotes the maximum pyrite precipitation efficiency in anoxic

1116 sediments. The reference value, e_{pyr}^{*}, was calibrated using a present-day control simulation such that the

1117 present-day seawater [SO$_4^{2-}$] is ~29 mM. The obtained value of 0.117 is generally consistent with modern

1118 observations (Bottrell and Newton, 2006; Tarhan et al., 2015; Turchyn and Schrag, 2006) (see Sect. 3).

1119 Although our approach does not provide a mechanistic description of the complex process of pyrite

1120 precipitation, it is suitable for many purposes.

Early diagenetic modeling for quantifying the OPD

1122 A simple 1-D early diagenetic model of C and O$_2$ is employed to obtain the parameterization of OPD (Eq.

1123 (48)). The 100 cm thick sediment is vertically divided into 50 layers with an uneven grid. The grid size

increases from the sediment-water interface ($\Delta z = 0.25$ mm) to the maximum simulated sediment depth ($\Delta z = 1.6$ cm). The diagenetic model calculates transport and biogeochemical transformation processes at each grid point within these sediment columns as well as the sedimentary burial and recycling fluxes at the model boundaries. The one-dimensional mass conservation equation for POC (wt. %) and dissolved O_2 is given by

$$\frac{\partial \text{POC}}{\partial t} = D_{\text{bio}} \frac{\partial^2 \text{POC}}{\partial z^2} - SR \frac{\partial \text{POC}}{\partial z} - k\text{POC},\tag{64}$$

$$\frac{\partial [O_2]}{\partial t} = D_{O_2} \frac{\partial^2 [O_2]}{\partial z^2} - r_{O_2} k\text{POC}\left(\frac{\rho(1-\phi)}{1.2\phi}\right)\frac{[O_2]}{[O_2]+K_{O_2}},\tag{65}$$

where D_{O2} is the diffusion coefficient of O_2, SR is the sedimentation rate, and ϕ is porosity, which is assumed to be constant over the entire sediment column for simplicity. Bioturbation is formulated as a diffusive process with a coefficient D_{bio}. The effective diffusion coefficient of O_2 is then given by

$$D_{O_2} = \frac{D_{O_2}^{T=0} \times \left(1 + v_{O_2} T_{\text{bw}}\right)}{\theta^2} + D_{\text{bio}},\tag{66}$$

where $D_{O2}^{T=0}$ denotes a tracer diffusion coefficient in seawater of 0°C, v_{O2} is a coefficient for temperature dependence of molecular diffusion coefficient. The in situ diffusion coefficient is further corrected for tortuosity θ, which is related to pore water resistivity and porosity via the following expressions (Colman and Holland, 2000; Tromp et al., 1995; Berner, 1980):

$$\theta^2 = \phi F\tag{67}$$

$$F = \phi^{-m}\tag{68}$$

where F is the formation factor—defined as the ratio of bulk sediment resistivity to interstitial water resistivity—and m is an empirical constant, varying with sediment type. We assumed the average value for unconsolidated muds ($m = 2.7$) in this work (Tromp et al., 1995). The particle mixing coefficient for bioturbation D_{bio} is formulated as a function of both sediment accumulation rate and bottom water O_2 concentration (Tromp et al., 1995; Wallmann, 2003):

$$D_{\text{bio}} = 10^{1.63+0.85\log SR} \frac{[O_2]_{\text{bw}}}{[O_2]_{\text{bw}} + K_{O_2}}.\tag{69}$$

1140 At the bottom of the sediment column, a no-flux condition was applied. The parameters used in the 1-D

1141 early diagenetic model are tabulated in Table 6.

1142

1143 **Table 6.** Parameters used in the 1-D early diagenetic model.

Parameters	Label	Value	Unit	Ref.
Porosity	ϕ	0.8	–	
Dry bulk density	ρ	2.6	g cm^{-3}	
O$_2$:C ratio for aerobic respiration	r_{O2}	1.4	mol mol^{-1}	
Half-saturation constant for aerobic respiration	K_{O2}	1	μM	
O$_2$ diffusion coefficient at 0°C	$D_{O2}^{T=0}$	281	cm^2 yr^{-1}	
Coefficient for a temperature dependence of molecular diffusion coefficient	v_{O2}	0.06	°C^{-1}	
Exponent for the formation factor	m	2.7	–	

1144

2.4.5 Air-sea exchange

1146 To calculate the gas exchange of O$_2$, H$_2$S, NH$_3$, and CH$_4$ across the air–sea interface, we employed a

1147 stagnant film model (Liss and Slater, 1974). The flux of a gas X across the air–sea interface is controlled by

1148 the difference in partial pressure between the atmosphere and surface waters, which can be described by the

1149 following formula:

$$J_X^{\text{air-sea}} = v_X^{\text{pis}} \left([X]_{\text{aq}} - [X]_{\text{sat}} \right), \tag{70}$$

1150 where v_X^{pis}, $[X]_{\text{aq}}$, and $[X]_{\text{sat}}$ denote piston velocity, the dissolved concentration of species X, and the

1151 saturation concentration of species X, respectively. For O$_2$, the saturation concentration is calculated based

1152 on solubility (Garcia and Gordon, 1992; Sarmiento and Gruber, 2006) and partial pressure:

$$[O_2]_{\text{sat}} = \left(\frac{1000}{22.3916} e^l \right) \left(\frac{pO_2}{pO_2^*} \right), \tag{71}$$

1153 where

$$l = A_0 + A_1 T_s + A_2 T_s^2 + A_3 T_s^3 + A_4 T_s^4 + A_5 T_s^5 + S \times (B_0 + B_1 T_s + B_2 T_s^2 + B_3 T_s^3) + C_0 S^2, \tag{72}$$

$$T_s = \ln\left(\frac{298.15 - T}{273.15 + T}\right), \tag{73}$$

with T in °C. The constants are $A_0 = 2.00907$, $A_1 = 3.22014$, $A_2 = 4.0501$, $A_3 = 4.94457$, $A_4 = -0.256847$, $A_5 = 3.88767$, $B_0 = -6.24523 \times 10^{-3}$, $B_1 = -7.3761 \times 10^{-3}$, $B_2 = -1.0341 \times 10^{-2}$, $B_3 = -8.17083 \times 10^{-3}$, and $C_0 = -4.88682 \times 10^{-7}$. The erroneous $A_3 \times T_s^2$ term in the original equation (Garcia and Gordon, 1992) was left out (Sarmiento and Gruber, 2006).

For CH_4, H_2S and NH_3, $[X]_{sat}$ is given by (Kharecha et al., 2005)

$$[X]_{sat} = K_X^{Henry} pX, \tag{74}$$

where K_X^{Henry}, and pX denote Henry's law coefficient and the partial pressure of species X, respectively. The temperature dependence of X's solubility is expressed as:

$$K_X^{Henry} = K_X^{Henry\circ} \exp\left[K_X^T\left(\frac{1}{T} - \frac{1}{298.15}\right)\right], \tag{75}$$

where $K_X^{Henry\circ}$ denotes the Henry's law coefficient of species X at 25°C, and K_X^T is the temperature dependence constant.

$[X]_{aq}$ is the dissolved concentration of X. $[H_2S]_{aq}$ and $[NH_3]_{aq}$ can be written as follows:

$$[H_2S]_{aq} = \frac{[\Sigma H_2S]}{1 + K_{H_2S}^{dis}/[H^+]} \tag{76}$$

$$[NH_3]_{aq} = \frac{[\Sigma NH_3]}{1 + [H^+]/K_{NH_3}^{dis}} \tag{77}$$

where $[\Sigma H_2S] = [H_2S] + [HS^-]$ and $[\Sigma NH_3] = [NH_4^+] + [NH_3]$. K_{H2S}^{dis} and K_{NH3}^{dis} are the dissociation constant, defined as follows:

$$K_{H_2S}^{dis} = \frac{[HS^-][H^+]}{[H_2S]_{aq}}, \tag{78}$$

$$K_{H_2S}^{dis} = \frac{[NH_3]_{aq}[H^+]}{[NH_4^+]}. \tag{79}$$

1168 Given values of K_{H2S}^{dis}, K_{NH3}^{dis} and pH (Millero et al., 1988; Yao and Millero, 1995), $[H_2S]_{aq}$ and $[NH_3]_{aq}$

1169 can be calculated.

1170

1171 Atmospheric concentrations of H_2S and NH_3 are set at 0. H_2S and NH_3 flow past the surface layer of the

1172 ocean to the atmosphere are converted to an equal influx of SO_4^{2-} and NO_3^- to the surface ocean. The

1173 parameters used in the stagnant film model are tabulated in Table 7.

1174

1175 If atmospheric O_2 levels are lower than ~1% PAL spatial heterogeneity of the gas exchange flux is expected

1176 (Olson et al., 2016); for example primary productivity (and O_2 generation) would be more active in coastal

1177 regions than open-ocean gyres. Because our ocean model resolves only two regions for the surface oceans

1178 (low-mid latitude region L and high latitude region H), it tends to overestimate the oxidation of reductants in

1179 surface mixing layers. To mitigate this model limitation for the CH_4 degassing flux, the aerobic oxidation

1180 rate of CH_4 is decreased to 1×10^{-7} of the standard value in surface layers (Ozaki et al., 2019a).

1181

1182 **Table 7.** Parameters used in the air-sea exchange module of CANOPS-GRB.

Parameters	Label	Value	Unit	Ref.
Piston velocity of O_2	v_{O2}^{pis}	1,000	m yr^{-1}	This study
Piston velocity of NH_3	v_{NH3}^{pis}	300	m yr^{-1}	(Webbook, 2022)
Piston velocity of H_2S	v_{H2S}^{pis}	1072	m yr^{-1}	(Webbook, 2022)
Piston velocity of CH_4	v_{CH4}^{pis}	1419	m yr^{-1}	(Webbook, 2022)
Solubility of NH_3 at 25°C	$K_{NH3}^{Henry°}$	5.6×10^4	mol m^{-3} bar^{-1}	
Solubility of H_2S at 25°C	$K_{H2S}^{Henry°}$	100	mol m^{-3} bar^{-1}	
Solubility of CH_4 at 25°C	$K_{CH4}^{Henry°}$	1.4	mol m^{-3} bar^{-1}	
Temperature dependence of solubility of NH_3	K_{NH3}^{T}	4,100	K	
Temperature dependence of solubility of H_2S	K_{H2S}^{T}	2,100	K	

Temperature dependence of solubility of CH_4	K_{CH4}^T	1,600	K	
Sea surface pH at low-mid latitude region	pH^l	8.17	–	This study
Sea surface pH at high latitude region	pH^h	8.16	–	This study
Partial pressure of atmospheric NH_3	pNH_3	0	atm	This study
Partial pressure of atmospheric H_2S	pH_2S	0	atm	This study

1183

2.5 Land model

2.5.1 Net primary productivity

Terrestrial NPP is scaled by global land biomass V normalized to the modern value:

$$J_{NPP}^{lnd} = V \times J_{NPP}^{lnd,*}, \tag{80}$$

where the present value of terrestrial NPP is set at 60 Gt C yr^{-1} (Prentice et al., 2001). The global land biomass is a function of atmospheric O_2 levels:

$$V = f_{UV} f_{fire} f_{O_2}, \tag{81}$$

where f_{O2} represents the direct effect of atmospheric O_2 concentration on the C$_3$ plant growth, and f_{fire} denotes the effect of fires on land biota (Bergman et al., 2004; Lenton and Watson, 2000b):

$$f_{O_2} = \max\left\{1.5 - 0.5 pO_2^{PAL}, 0\right\}, \tag{82}$$

$$f_{fire} = \frac{k_{fire}}{k_{fire} - 1 + ignit}. \tag{83}$$

Here k_{fire} (= 3; (Lenton, 2013)) is the fire frequency constant, and *ignit* is an ignition factor representing the fire frequency as a function of oxygen (Lenton, 2013; Lenton et al., 2018; Lenton and Watson, 2000b):

$$ignit = \min\left\{\max\left\{c_1 pO_2 - c_2, 0\right\}, c_3\right\}, \tag{84}$$

with $c_1 = 48$, $c_2 = 9.08$ and $c_3 = 5$ (Lenton, 2013). CANOPS-GRB also includes an additional factor f_{UV} representing the effect of UV on the terrestrial biosphere as a function of atmospheric O_2 levels (Ozaki and Reinhard, 2021):

$$f_{\text{UV}} = \tanh\left(\frac{p\text{O}_2^{\text{PAL}}}{c_{\text{UV}}}\right), \tag{85}$$

where c_{UV} is a model parameter, which, in our standard model is set at 1% PAL, meaning that terrestrial plant activity is suppressed when atmospheric O_2 is lower than a few % PAL.

2.5.2 Terrestrial biogeochemical cycles

Phosphorus weathering flux, J_P^{w} (Eq. (2)), is treated as a boundary condition. A fraction of weathered P is ultimately buried as terrigenous organic matter (Eq. (3)), whereas the remaining fraction is delivered to the ocean via rivers (Eq. (4)). In the CANOPS-GRB model, the reference value of J_P^{r} (= 0.155 Tmol P yr^{-1}) is tuned so that the oceanic P level of the reference state is consistent with modern observations. The burial rate of terrigenous organic matter (in terms of C) can be written as follows:

$$J_{\text{org}}^{\text{b,lnd}} = \left(\text{C}_{\text{org}}/\text{P}_{\text{org}}\right)^{\text{lnd}} J_P^{\text{b,lnd}}, \tag{86}$$

where $(\text{C}_{\text{org}}/\text{P}_{\text{org}})^{\text{lnd}}$ (= 1000) is the average C/P burial ratio of terrigenous organic matter (Bergman et al., 2004). In this study, the reference value of $J_{\text{org}}^{\text{b,lnd}}$ was set at 3 Tmol C yr^{-1}. By combining Eqs. (3), (4), and (85) for the reference state, the proportional coefficient k_{11} of Eq. (3) is determined by the reference state, as follows:

$$k_{11} = \frac{J_{\text{org}}^{\text{b,lnd,*}}}{J_{\text{org}}^{\text{b,lnd,*}} + \left(\text{C}_{\text{org}}/\text{P}_{\text{org}}\right)^{\text{lnd}} J_P^{\text{r,*}}} = 0.0189. \tag{87}$$

The value of k_{11} is treated as a constant in this study.

Almost all organic matter produced by terrestrial NPP is decomposed before burial. The total decomposition rate is given by:

$$J_{\text{org}}^{\text{r,lnd}} = J_{\text{NPP}}^{\text{lnd}} - J_{\text{org}}^{\text{b,lnd}}. \tag{88}$$

CANOPS-GRB includes aerobic respiration and methanogensis as respiration pathways for terrigenous matter, and the CH$_4$ flux from the terrestrial ecosystem to the atmosphere is evaluated with the assumption that it is proportional to the burial rate of terrigenous organic matter:

$$J_{CH_4}^{lnd} = \left(\frac{J_{org}^{b,lnd}}{J_{org}^{b,lnd,*}} \right) J_{CH_4}^{lnd,*}, \tag{89}$$

where the reference value was set at 1 Tmol CH_4 yr^{-1}. The net flux of CO_2, O_2 and CH_4 from the terrestrial ecosystem to the atmosphere can be written, as follows:

$$J_{CO_2}^{lnd} = \left(g_{O_2} + \frac{1}{2}(1+\delta) g_{CH_4} \right) J_{org}^{r,lnd}, \tag{90}$$

$$J_{O_2}^{lnd} = J_{NPP}^{lnd} - \left(g_{O_2} + \delta g_{CH_4} \right) J_{org}^{r,lnd}, \tag{91}$$

$$J_{CH_4}^{lnd} = \frac{1}{2}(1-\delta) g_{CH_4} J_{org}^{r,lnd}, \tag{92}$$

where g_{O2} and g_{CH4} denote the fraction of organic matter decomposed by aerobic respiration and methanogenesis, respectively. δ represents the fraction of methane that is consumed by aerobic methanotrophy that is a function of O_2:

$$\delta = \frac{M_{O_2}^{atm}}{M_{O_2}^{atm} + K_{O_2}'}, \tag{93}$$

with K'_{O2} = 0.273×10^{18} mol (Goldblatt et al., 2006). A fraction of organic matter decomposed by methanogenesis, g_{CH4}, can be calculated based on Eqs. (89) and (91). Then, g_{O2} is determined from 1-g_{CH4}.

2.5.3 Weathering

The oxidative weathering of continental crust is a major oxygen sink on geologic timescales, providing a fundamental control on atmospheric O_2 levels. The weathering rate in the model is assumed to be proportional to sedimentary reservoir size and a global erosion factor, f_R, expressing the effect of continental denudation/erosion on terrestrial weathering:

$$J_{org}^{w} = f_R f_{orgw}^{O_2} \left(\frac{ORG}{ORG^*} \right) J_{org}^{w,*}, \tag{94}$$

$$J_{pyr}^{w} = f_R f_{pyrw}^{O_2} \left(\frac{PYR}{PYR^*} \right) J_{pyr}^{w*}, \tag{95}$$

1227 where J_{org}^w and J_{pyr}^w denotes the oxidative weathering of organic carbon and pyrite, respectively, and f_{orgw}^{O2}

1228 and f_{pyrw}^{O2} represent the O_2 dependency. For the oxidative weathering of organic matter, previous

1229 biogeochemical models have adapted a $(pO_2^{PAL})^{0.5}$ relationship (Bergman et al., 2004; Lasaga and Ohmoto,

1230 2002). In this study, we employ alternative empirical relationships based on results obtained from a 1-D

1231 weathering model (Bolton et al., 2006; Daines et al., 2017):

$$f_{orgw}^{O_2} = c_{orgw} \frac{pO_2^{PAL}}{pO_2^{PAL} + K_{orgw}} , \tag{96}$$

$$f_{pyrw}^{O_2} = c_{pyrw} \frac{pO_2^{PAL}}{pO_2^{PAL} + K_{pyrw}} , \tag{97}$$

1232 where K_{orgw} and K_{pyrw} denote half-saturation constants ($K_{orgw} = 0.334$ and $K_{pyrw} = 0.017$) and c_{orgw} and c_{pyrw}

1233 are normalized constants ($c_{orgw} = 1.334$ and $c_{pyrw} = 1.017$), respectively. The Monod-type relationship

1234 captures the fact that the rate of oxidative weathering reaches its maximum as determined by the erosion rate

1235 under highly oxygenated conditions (i.e., transport-limited regime). For example, due to the fast dissolution

1236 kinetics of pyrite, oxidative weathering can be regarded as transport-limited under modern conditions

1237 (Bolton et al., 2006) (Fig. 10). In the CANOPS-GRB model, J_{org}^{w*} is calibrated based on the global redox

1238 budget of the reference run (see Sect. 2.2.5).

1239

1240 It is important to note that above equations ignore the possible importance of microbial activity and

1241 temperature on the rate of oxidative weathering (Petsch et al., 2001; Soulet et al., 2021). Both represent

1242 important topics for future research.

1243

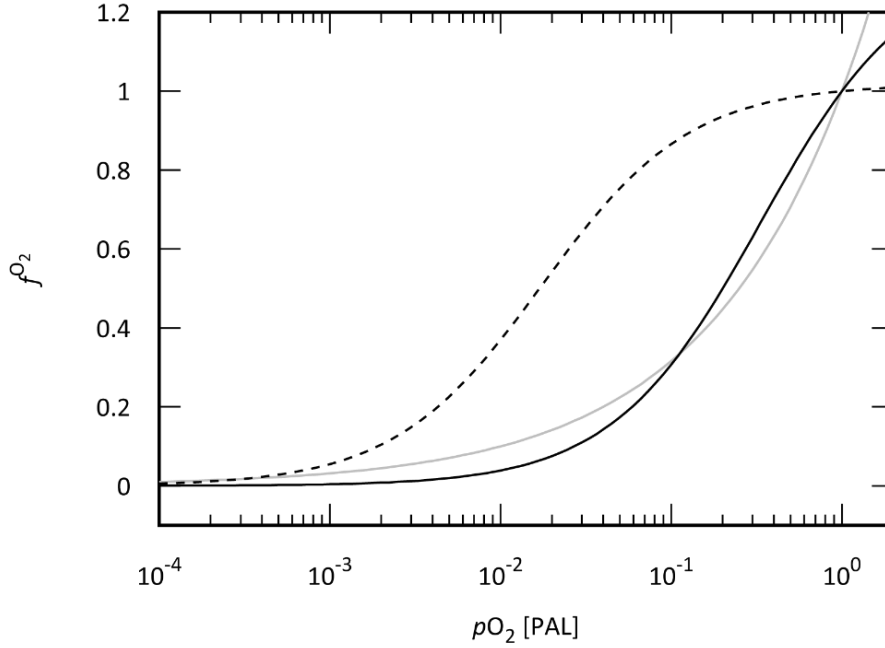



Figure 10. O_2 dependency of the oxidative weathering rate of organic matter and pyrite sulfur. Gray line denotes the $(pO_2^{PAL})^{0.5}$ relationship assumed in previous biogeochemical models (Lasaga and Ohmoto, 2002; Daines et al., 2017). Solid and dashed black lines represent the empirical Monod-type relationships for oxidative weathering of organic matter (solid) and pyrite sulfur (dashed) based on the results obtained from a 1-D weathering model (Bolton et al., 2006; Daines et al., 2017), which are adopted in the standard model of the CANOPS-GRB model. PAL = present atmospheric level.

The present riverine flux of sulfur, J_S^{r*}, is estimated at 2.6 Tmol S yr^{-1} (Raiswell and Canfield, 2012), representing the dominant source to the oceans. Riverine flux is written as the sum of the gypsum weathering flux J_{gyp}^w and the oxidative weathering of pyrite J_{pyr}^w and depends directly or indirectly on the oxidation state of the atmosphere:

$$J_S^r = J_{gyp}^w + J_{pyr}^w. \tag{98}$$

Based on previous studies (Berner, 2009; Wortmann and Paytan, 2012; Bergman et al., 2004; Markovic et al., 2015), a 3:1 ratio in modern rivers of SO_4^{2-} from gypsum versus pyrite weathering is assumed. Gypsum weathering flux is assumed to be proportional to its sedimentary reservoir size, *GYP*, and f_R:

$$J_{gyp}^w = f_R \left(\frac{GYP}{GYP^*} \right) J_{gyp}^{w*}, \tag{99}$$

1258   where * represents the present value.

1259

1260   In the previous version of the CANOPS (Ozaki et al., 2019a), oxidative weathering of pyrite was divided to

1261   biogenic and abiotic weathering fluxes. In this study, we simplify this (Eq. (94)). Also, oxidative weathering

1262   of Fe(II)-bearing minerals is ignored in this study, which simplifies the framework of the global O_2 budget

1263   (Sect. 2.2.5).

1264 2.5.4 Volcanic degassing

1265   Volcanic outgassing fluxes of carbon and sulfur are assumed to be proportional to their respective crustal

1266   reservoir sizes:

$$J_{org}^{m} = \left(\frac{ORG}{ORG^*} \right) J_{org}^{m,*}, \tag{100}$$

$$J_{pyr}^{m} = \left(\frac{PYR}{PYR^*} \right) J_{pyr}^{m,*}, \tag{101}$$

$$J_{gyp}^{m} = \left(\frac{GYP}{GYP^*} \right) J_{gyp}^{m,*}. \tag{102}$$

1267   We set the reference value of the volcanic outgassing flux of organic carbon, $J_{org}^{m,*}$, at 1.25 Tmol C yr^{-1}

1268   (Bergman et al., 2004). The estimates of modern volcanic fluxes of sulfur fall within the range of ~0.3–3

1269   Tmol S yr^{-1} (Kagoshima et al., 2015; Catling and Kasting, 2017; Raiswell and Canfield, 2012; Walker and

1270   Brimblecombe, 1985). We adopted a recent estimate of 0.8 Tmol S yr^{-1} (Kagoshima et al., 2015).

1271 2.5.5 Sedimentary reservoirs

1272   We extend the original model framework to the explicit calculation of the secular evolution of the

1273   sedimentary reservoirs, linking the biogeochemical cycles in the ocean-atmosphere system to the rock cycle.

1274   The mass balance equation for sedimentary organic carbon (ORG) can be written as follows:

$$\frac{dORG}{dt} = J_{org}^{b} - J_{org}^{w} - J_{org}^{m}, \tag{103}$$

1275   where J_{org}^{b} denotes the sum of the burial rate of marine and terrigenous organic matter ($J_{org}^{b,ocn} + J_{org}^{b,lnd}$),

1276   the primary source of sedimentary organic carbon. Primary outputs are oxidative weathering, volcanic

1277 outgassing and metamorphism. Previous estimates of the present reservoir size of *ORG* fall in the range of
1278 1000–1300 Emol (1 E $=10^{18}$) (Berner, 1989; Garrels and Perry, 1974; Mackenzie et al., 1993). We assumed
1279 1250 Emol for the reference value of *ORG*.

1280

1281 The sedimentary reservoir sizes of pyrite sulfur (*PYR*) and gypsum sulfur (*GYP*) are also written as the
1282 balance between the input (burial) and outputs (weathering and outgassing):

$$\frac{dPYR}{dt} = J_{pyr}^{b} - J_{pyr}^{w} - J_{pyr}^{m}, \tag{104}$$

$$\frac{dGYP}{dt} = J_{gyp}^{b} - J_{gyp}^{w} - J_{gyp}^{m}, \tag{105}$$

1283 where J_{pyr}^{b} represents the sum of pyrite precipitation rates in the water column and sediments, $J_{pyr}^{b,wc}$ +
1284 $J_{pyr}^{b,sed}$. Previous estimates of present reservoir sizes of *GYP* and *PYR* fall in the range of 77−300 Emol and
1285 155−300 Emol (Berner, 2006; Bottrell and Newton, 2006; Yaroshevsky, 2006; Kump, 1989; Lasaga, 1989;
1286 Holser et al., 1989; Sleep, 2005; Schlesinger and Bernhardt, 2013), respectively. We adopted 200 Emol and
1287 200 Emol for GYP^{*} and PYR^{*}.

1288 2.6 Atmosphere model

1289 2.6.1 Hydrogen escape

1290 The rate of hydrogen escape is assumed to be diffusion-limited as it is today. Thus, the total concentration of
1291 all H-bearing compounds in the lower stratosphere determines the rate of hydrogen escape (Walker, 1977).
1292 For Proterozoic-Phanerozoic atmospheres, CH_4 appears to have been the dominant hydrogen-bearing species
1293 in the stratosphere, and the flux, J_{Hesc} (mol yr^{-1}), is calculated as

$$J_{Hesc} = sM_{CH_4}^{atm}, \tag{106}$$

1294 where M_{CH4}^{atm} denotes the abundance of CH_4 in the atmosphere (mol) and s (= 3.7×10^{-5} yr^{-1}) is a
1295 proportional coefficient (Goldblatt et al., 2006).

2.6.2 Photochemistry

CANOPS-GRB includes parameterized O_2-O_3-CH_4 photochemistry that allows quantification of the abundances of atmospheric O_2 and CH_4. The rate of oxidation of CH_4 is calculated by the following empirical parameterization that was obtained from a 1-D photochemistry model (Claire et al., 2006):

$$J_{CH_4ox} = k_{CH_4ox} M_{O_2}^{atm} M_{CH_4}^{atm},$$
(107)

where $M_{O_2}^{atm}$ and $M_{CH_4}^{atm}$ denote the abundance of O_2 and CH_4 in the atmosphere (mol). The reaction rate k_{CH_4ox} (mol^{-1} yr^{-1}) is expressed as a polynomial function of the reservoir sizes of O_2 and CH_4 (Ozaki and Reinhard, 2021):

$$\log k_{CH_4ox} = \alpha_0^j + \alpha_1^j \cdot \varphi_{O_2} + \alpha_2^j \cdot \varphi_{O_2}^2 + \alpha_3^j \cdot \varphi_{O_2}^3 + \alpha_4^j \cdot \varphi_{O_2}^4 + \alpha_5^j \cdot \varphi_{O_2}^5 + \alpha_6^j \cdot \varphi_{O_2}^6,$$
(108)

where a_i^j are fitting coefficients for given atmospheric CH_4 levels and φ_{O_2} is $\log pO_2$ (in bar) (Supplementary Table 4 of Ozaki and Reinhard, 2021). The oxidation rate was evaluated using Fig. 3 of Claire et al. (2006), showing the oxidation rate as a function of pO_2 and pCH_4. We took the relationship between k_{CH_4ox} and pO_2 for pCH_4 of 10^{-6}, 10^{-5}, 10^{-4}, 10^{-3}, 2×10^{-3} bar, and k_{CH_4ox} is calculated as a function of pO_2 and pCH_4 with a log-linear interpolation method. Note that the default photochemical parameterization presented above limits the applicability of CANOPS-GRB v1.0 to Earth-like planets around the Sun-like host stars. Current work is focused on elaborating parameterized photochemistry across a wider range of spectral energy distributions.

2.6.3 Mass balance

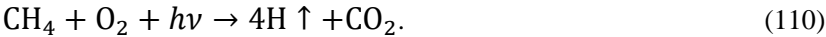
CANOPS-GRB accounts for the atmospheric concentrations of O_2 and CH_4. The atmospheric concentration of O_2 is determined by the biogenic source (from the ocean and terrestrial ecosystems) and the consumption through the series of oxidation reaction (the continental weathering of kerogen and pyrite, volcanic outgassing, and photochemical oxidation of methane):

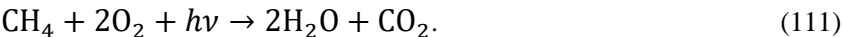
$$\frac{dM_{O_2}^{atm}}{dt} = J_{O_2\uparrow}^{air-sea} + J_{O_2\uparrow}^{air-lnd} - \left(J_{Hesc} + 2J_{CH_4ox}\right) - \left(J_{org}^w + J_{org}^m\right) - 2\left(J_{pyr}^w + J_{pyr}^m\right),$$
(109)

where $M_{O_2}^{atm}$ denotes the mass of O_2 in the atmosphere (moles), and the first and second term on the right hand side represents the biogenic flux of O_2 from marine and terrestrial ecosystems. The third term denotes O_2 consumption via photochemistry, and the fourth and fifth terms are the O_2 consumption via organic C

1318 and pyrite S sub-cycles. Note that the hydrogen escape to space is represented as the O_2 sink, because the

1319 hydrogen escape via CH_4 followed by the oxidation of carbon to CO_2 is represented as:

$$CH_4 + O_2 + h\nu \rightarrow 4H \uparrow + CO_2. \tag{110}$$

1320 On the other hand, the photochemical oxidation of CH_4 can be written, as follows:

$$CH_4 + 2O_2 + h\nu \rightarrow 2H_2O + CO_2. \tag{111}$$

1321 Thus, the hydrogen escape to space represents the net gain of oxidizing power to the system (see Eq. (13)).

1322

1323 The abundance of CH_4 in the atmosphere, M_{CH4}^{atm}, is determined by input from the ecosystems and the

1324 consumption of CH_4 via photolysis, as well as by the hydrogen escape:

$$\frac{dM_{CH_4}^{atm}}{dt} = J_{CH_4\uparrow}^{air\text{-}sea} + J_{CH_4\uparrow}^{air\text{-}lnd} - \left(J_{Hesc} + J_{CH_4ox} \right). \tag{112}$$

1325 No abiotic CH_4 input via hydrothermal systems is included.

3 Validation against the modern global ocean

Here, a steady-state simulation mimicking the present-day condition was run to evaluate the overall performance of CANOPS-GRB. To do this, the ocean model was run until reaching steady state, assuming the present atmospheric O_2 level and reference values of boundary fluxes (weathering and volcanic fluxes). The simulated circulation and biogeochemistry for the modern global ocean was compared with modern oceanographic observations from the Global Ocean Data Analysis Project (Key et al., 2015; Olsen et al., 2016).

3.1 Distribution of circulation tracers

Comparisons of model output with circulation tracers, such as potential temperature (θ) and radiocarbon ($\Delta^{14}C$), permit a test of the physical exchange scheme. Figure 11 depicts the simulated patterns of physical tracers with observational data. The physical circulation in the model generally agrees well with oceanic observations, although we note that model temperatures for low-mid latitudes above 1,000 m water depth tend to be higher than observed because temperature distribution in the real ocean is strongly controlled by vertical structure and advective processes that are not captured in our simple circulation scheme. Despite this model limitation, the modelled temperature distribution generally reproduces the observed distribution. The $\Delta^{14}C$ minimum in the model for the low-mid latitude region corresponds well with observations. The modelled background radiocarbon for young deep waters (about -150±25‰) is closer to the value for the Southern Ocean (approximately -150‰) than for North Atlantic deep waters (approximately -80‰), and old deep waters (-200±15‰) correspond to the South Pacific. We conclude that the simulated circulation tracers generally match well with ocean data.

1347

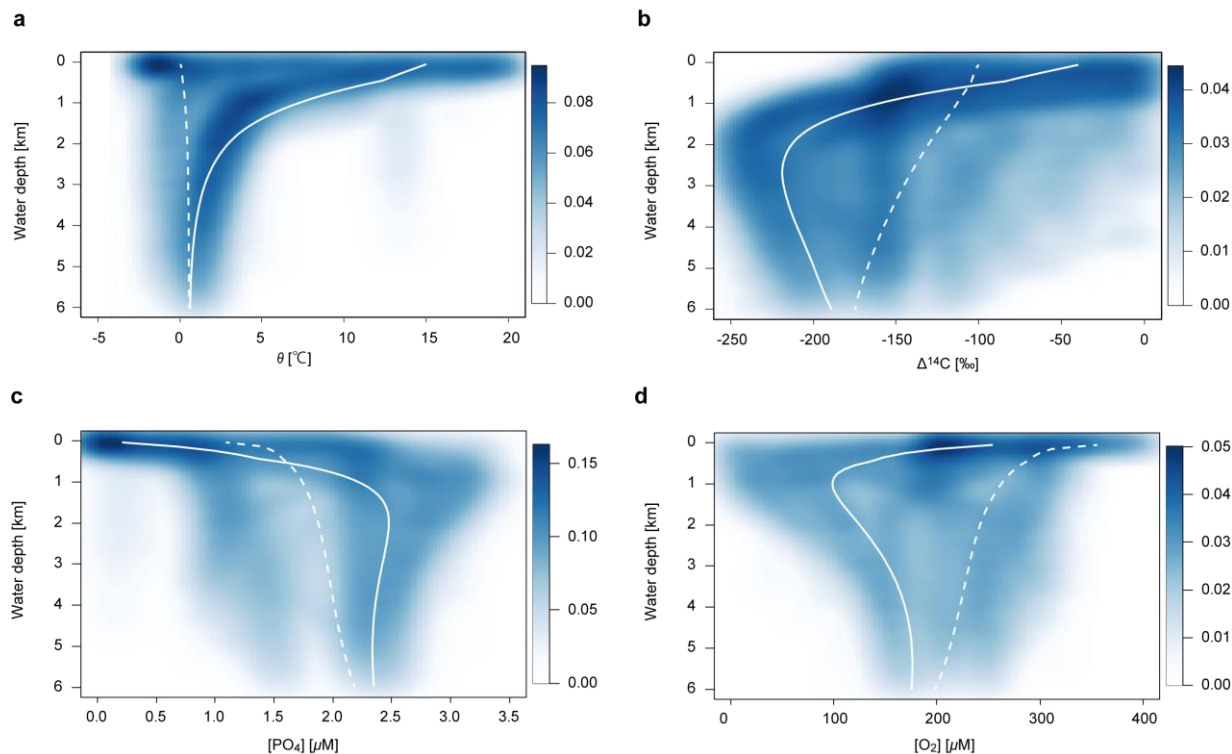



Figure 11. Simulated steady state depth-profiles of (**a**) potential temperature, θ, (**b**) radio carbon, $\Delta^{14}C$, (**c**) DIP (dissolved inorganic phosphorus, PO_4^{3-}), and (**d**) dissolved oxygen, O_2. Solid and dashed white lines denote the simulated profiles for LD and HD regions, respectively. Simulation results are compared with the dataset from the Global Ocean Data Analysis Project (GLODAP) data base (GLODAPv2_2019; (Key et al., 2015; Olsen et al., 2016)). The color represents the density of observational points.

3.2 Ocean biogeochemistry

Having demonstrated that CANOPS-GRB's ocean circulation model does a reasonable job of representing water mass exchange, we next assess the performance of the oceanic biogeochemistry model by comparing its output to ocean biogeochemical data. Model-generated global fluxes and inventories of C, P, N and S cycles are summarized in Fig. 12. Those compare well with independent observational estimates. Below, we provide a brief discussion of globally-integrated biogeochemical flux estimates.

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1360

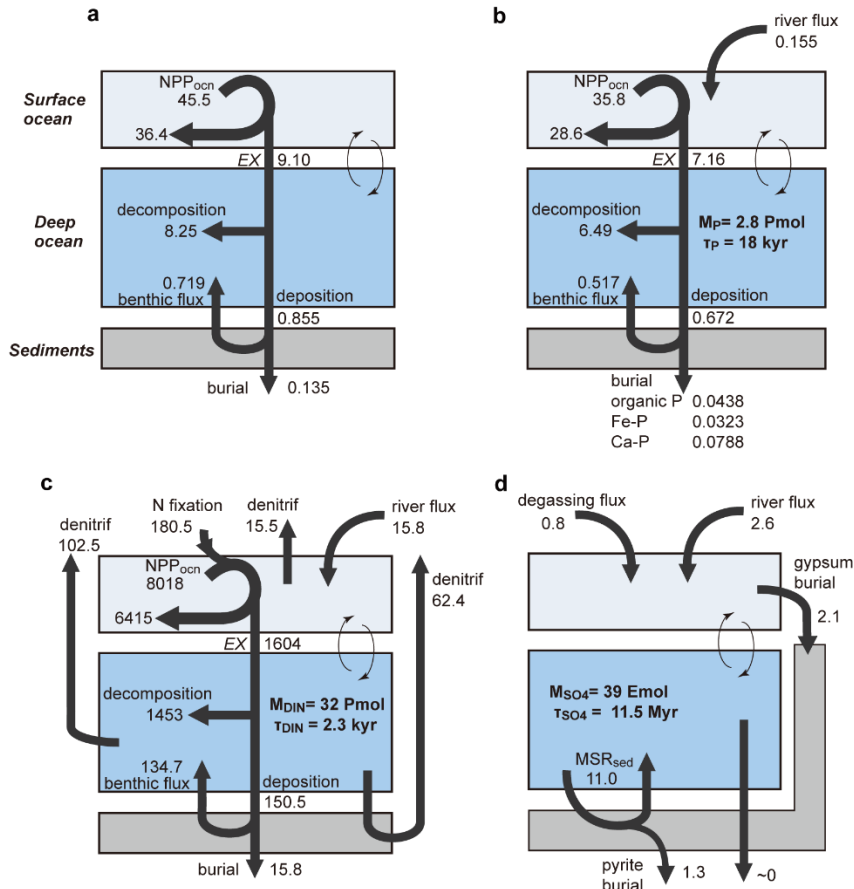



Figure 12. Schematics of the simulated material flow in the ocean for the reference run. (**a**) Organic carbon (in Gt C yr^{-1}), (**b**) phosphorus (in Tmol P yr^{-1}), (**c**) nitrogen (in Tg N yr^{-1}), and (**d**) sulfur (in Tmol S yr^{-1}). NPP$_{ocn}$ = oceanic net primary production. EX = export production. M_X = mass of X in the ocean. τ_X = residence time of X in the ocean. Pmol = 10^{15} mol.

3.2.1 Distribution of nutrients and oxygen

The simulated vertical profile of phosphate captures the characteristic features and values of observational data (Fig. 11c). More specifically, the distribution in the low-mid latitude region is more similar to that in the Pacific and Indian Ocean, and distribution of high-mid latitude region is similar to that in the Southern Ocean. This is a consequence of limiting high-latitude productivity (preformed DIP is 1.1 μM) which results in higher concentrations in the ocean interior. The model dissolved O$_2$ profile for low-mid latitude shows a minimum of approximately 100 μM at water depth of 1,000 m, corresponding to the oxygen minimum zone

1372 (Fig. 11d). In contrast, dissolved O_2 for high-mid latitude sector (HD) shows a monotonically decreasing

1373 trend. This is because of oxygen consumption via POM decomposition during downwelling.

3.2.2 Carbon cycling

1375 The marine export/new production in our model is 9.1 Gt C yr^{-1} (8.36 Gt C yr^{-1} at L and 0.74 Gt C yr^{-1} at H).

1376 This is consistent with previously estimated global values of 8.5–12 Gt C yr^{-1} (Dunne et al., 2007; Laws et

1377 al., 2000; Sarmiento and Gruber, 2006; Heinze et al., 2009). In particular, our estimate is close to the mid-

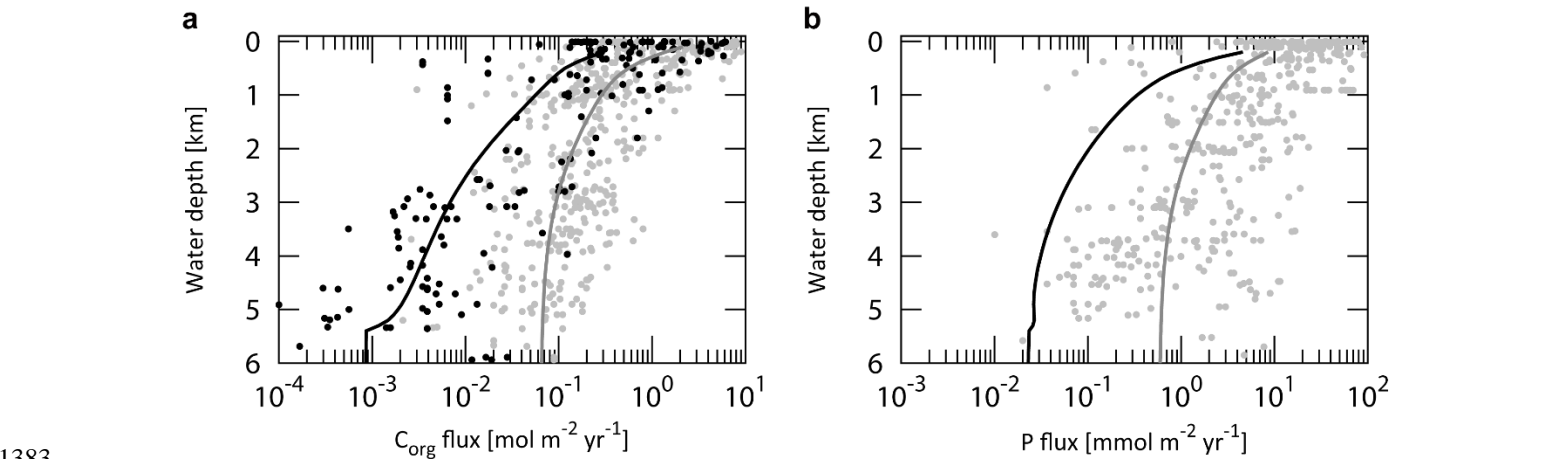
1378 point of the previously estimated range of 9.6±3.6 Gt C yr^{-1} (Dunne et al., 2007). This is a marked

1379 improvement from earlier studies with box models which have underestimated marine new production by a

1380 factor of 2 or more (Archer et al., 2000; Shaffer et al., 2008). Simulated global oceanic NPP is 45.5 Gt C yr^{-1}.

1381 This is also consistent with the previous estimated range of 44–65 Gt C yr^{-1} (Prentice et al., 2001;

1382 Woodward, 2007; Carr et al., 2006; Berelson et al., 2007).

1383

1384 **Figure 13.** Simulated steady-state depth-profiles of organic C and reactive P flux density for the LD region.
1385 In (**a**), gray dots denote observations of depositional/settling flux density, whereas black dots represent
1386 observations of burial flux density compiled from literature survey (Baturin, 2007; Betts and Holland, 1991;
1387 Colman et al., 2000; Lutz et al., 2002). Gray and black solid lines denote the simulated POC depositional
1388 and burial flux densities obtained from the reference run. (**b**) Gray dots denote the benthic P efflux density
1389 obtained from literature survey (Hartnett and Devol, 2003; Hensen et al., 1998; Ingall and Jahnke, 1994,
1390 1997; Mcmanus et al., 1997; Colman et al., 2000; Schenau and De Lange, 2001; Zabel et al., 1998), whereas
1391 gray and black solid lines represent the simulated benthic P efflux density and burial flux density of reactive
1392 P obtained from the reference run. The burial flux density of reactive P is not shown due to the sparseness of
1393 such observations.

1394

1395 The global marine POC flux depends largely on water depth. Model-generated fluxes compare well with

1396 independent estimates of deposition, burial, and regeneration. The gray line in Fig. 13a shows the simulated

1397 sinking flux density of POC in the water column for LD region, compared with observations (Archer et al.,

1398 2002; Betts and Holland, 1991; Lutz et al., 2002; Baturin, 2007). The preferential consumption of labile

1399 compounds (G_1 and G_2) during the settling process leads to a continuous decrease in reactivity and therefore,

1400 remineralization rates from the surface ocean down to the deep. Our estimate lies well within the range of

1401 observations. The model tends to give lower fluxes than observed above 2,000 m water depth, and higher

1402 below 5,000 m water depth. This is probably because of the assumption of homogeneous productivity in the

1403 surface ocean. In the real ocean, oceanic productivity is generally greater at the continental margins than in

1404 the pelagic gyre regions (Lutz et al., 2002). This is a model limitation, but the simulated biological pump is

1405 sufficient to describe the general characteristics of global ocean biogeochemistry.

1406

1407 Of total exported POC, 91% (8.25 Gt C yr^{-1}) is decomposed in the water column and the rest (0.85 Gt C yr^{-1})

1408 sinks to the sediment surface (Fig. 12a). The simulated global POC depositional flux is comparable not only

1409 with observational estimates of 0.93 Gt C yr^{-1} (Muller-Karger et al., 2005) and 0.67±0.48 Gt C yr^{-1} for off-

1410 shore regions (Dunne et al., 2007), but also with an estimate using EMIC (0.87 Gt C yr^{-1}) (Ridgwell and

1411 Hargreaves, 2007). The depositional fluxes of C_{org} in marginal (<2,000 m) and deep-sea sediments (>2,000

1412 m) are estimated at 0.58 Gt C yr^{-1} and 0.27 Gt C yr^{-1}, respectively. These estimates are slightly lower than

1413 previous estimates of 0.62–1.98 Gt C yr^{-1} and 0.31–0.62 Gt C yr^{-1} (Bohlen et al., 2012; Dunne et al., 2007;

1414 Muller-Karger et al., 2005; Burdige, 2007).

1415

1416 In our standard run, benthic remineralization removes 7.9% of the exported POC (0.72 Gt C yr^{-1}), equivalent

1417 of 84% of the global POC sedimentation rate. As a result, only 1.5% (0.135 Gt C yr^{-1} or 11.3 Tmol C yr^{-1}) of

1418 the global POC export production is ultimately buried in marine sediments. Our model demonstrates that

1419 much (91%) of the total burial occurs on the continental margins (<2,000 m water depth), where the settling

1420 flux and burial efficiency are relatively high. Previous studies (Dunne et al., 2007; Muller-Karger et al.,

1421 2005) estimated a C_{org} burial rate of 0.29±0.15 Gt C yr^{-1} and >0.06±0.06 Gt C yr^{-1} at the margin. Our

1422   estimate of 0.123 Gt C yr^{-1} lies between these values, whereas our estimate for the deep sea, 0.012 Gt C yr^{-1},

1423   is on the lower end of previous estimates of 0.012±0.02 Gt C yr^{-1} (Dunne et al., 2007) and 0.017±0.005 Gt C

1424   yr^{-1} (Hayes et al., 2021). Also, (Sarmiento and Gruber, 2006; Hayes et al., 2021) estimated the burial rate

1425   below 1,000 m as 0.02±0.006 Gt C yr^{-1}; our estimate of 0.019 Gt C yr^{-1} is consistent with this. Combined

1426   with the prescribed burial rate of terrigenous C_{org} 0.036 Gt C yr^{-1} (3 Tmol C yr^{-1}), the total burial rate is

1427   calculated to be 0.17 Gt C yr^{-1} (14.3 Tmol C yr^{-1}). This is somewhat higher than previous estimates (Berner,

1428   1982; Burdige, 2005; Muller-Karger et al., 2005), but given the rather large uncertainty we consider it

1429   defensible.

1430

1431   Figure 14 shows OPD as a function of water depth. Although the benthic data could be biased towards

1432   highly specific environments, such as sediments underlying upwelling areas and continental margins, our

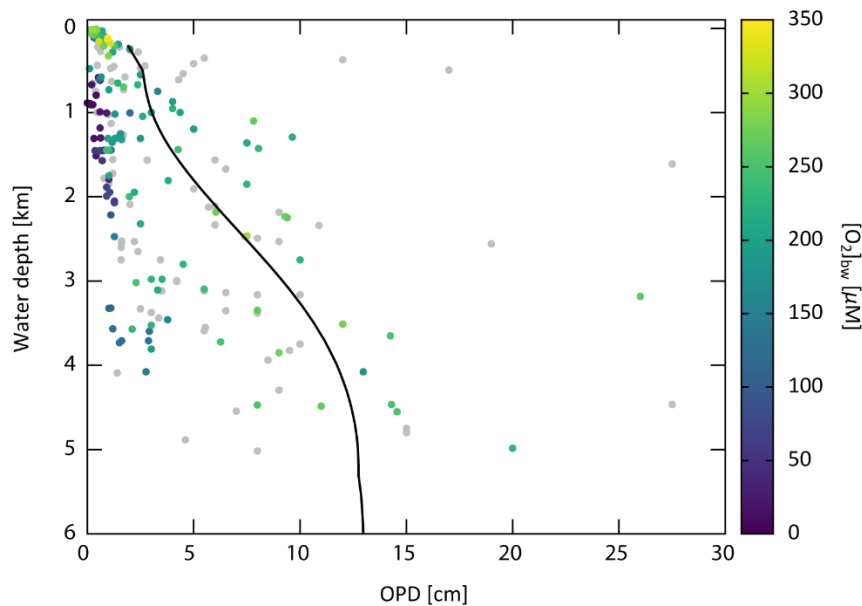
1433   estimates capture the general features of modern observations.

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Figure 14. Oxygen penetration depth (OPD) as a function of water depth. Color dots denote the observational data obtained from literature survey (Bradley et al., 2020; Donis et al., 2016; Nierop et al., 2017; Rowe et al., 2008; Mcmanus et al., 2005; Martin and Sayles, 2014; Pfeifer et al., 2002; Hyacinthe et al., 2001; Hartnett et al., 1998; Hedges et al., 1999; Morford and Emerson, 1999; Devol and Christensen, 1993; Gundersen and Jorgensen, 1990; Sachs et al., 2009). The color represents the O_2 concentration of bottom water, $[O_2]_{bw}$, with grey dots for the unknown $[O_2]_{bw}$ value. The simulated OPD obtained from the reference run is shown as a black line.

3.2.3 Phosphorus cycling

The removal of phosphate from surface waters occurs through photosynthetic fixation by primary producers and subsequent export in the form of POP into deeper waters, where it is largely remineralized back into DIP. Through this process there is a vertical partitioning of DIP within the ocean with reduced surface concentrations. Phosphorus export production is 7.16 Tmol P yr^{-1}, which is coupled with carbon according to the POM compositional ratio (C:P = 106:1 for our standard model). The remineralization in the water column (6.49 Tmol P yr^{-1}) and total sedimentation rate (0.672 Tmol P yr^{-1}) are also proportional to those of POC. In contrast, the benthic DIP flux is decoupled from the carbon flux. Figure 13b shows modelled DIP benthic efflux and burial flux together with observed fluxes. Some observational data showing a relatively large abyssal (4−6 km) benthic flux are from upwelling regions in the South Atlantic (Hensen et al., 1998). The deviation is therefore not critical for our globally averaged model. Our model gives the total benthic efflux of DIP as 0.517 Tmol P yr^{-1}, which is roughly three times the riverine reactive P input rate. This is within the range of previous estimates of 0.05−1.25 Tmol P yr^{-1} (Wallmann, 2003; Wallmann, 2010; Compton et al., 2000; Colman and Holland, 2000).

The preservation efficiency (here defined as burial flux divided by the export flux) of P is 2.1%. This is higher than that of organic carbon (1.5%), indicating that more P is trapped in marine sediments than might be expected from Redfield stoichiometry. In marine sediments overlain by oxic bottom waters, a fraction of the DIP released to pore waters from POM decomposition can be absorbed by iron-oxyhydroxide or precipitated as authigenic fluorapatite (Fig. 9). Therefore, the global averaged C_{org}/P_{reac} ratio of buried sediments is generally less than the Redfield of 106 (approximately 65±25 based on observations; (Algeo and Ingall, 2007)). The modelled global average C_{org}/P_{reac} ratio of buried sediment is 73, consistent with this. The P burial fluxes of organic P, Fe-bound P and authigenic P are estimated at 0.044 Tmol P yr^{-1}, 0.032 Tmol P yr^{-1}, and 0.079 Tmol P yr^{-1}, respectively.

The C_{org}/P_{org} ratio of burying organic matter shows a non-linear relationship with respect to sedimentation rate. The observed C_{org}/P_{org} ratios are generally greater than the Redfield value of 106, especially for sediments in oxygen minimum zones (OMZs), which are characterized by a high depositional flux of

1470 organic matter (C_{org}/P_{org} ratios up to 600 for the present open ocean) (Ingall and Cappellen, 1990). For

1471 example, the averaged C_{org}/P_{org} molar ratio at the Peru-Chile OMZ and Black Sea are 600, and the estimated

1472 burial ratio of sapropel S1 of Mediterranean Sea is in the range of 400–800 (Slomp et al., 2002).This reflects

1473 the preferential regeneration of P relative to C during microbial remineralization of marine organic matter

1474 and reflect the more labile nature of P-biochemicals relative to most non-phosphorus containing organic

1475 carbon compounds. Additional rationale for this observation is that P is preferentially targeted for

1476 remineralization to support subsequent biological productivity as an essential and potentially limiting

1477 nutrient. Our model demonstrates that we can reproduce the first-order relationship between C_{org}/P_{org} and

1478 sediment accumulation rate (Fig. 15).

1479

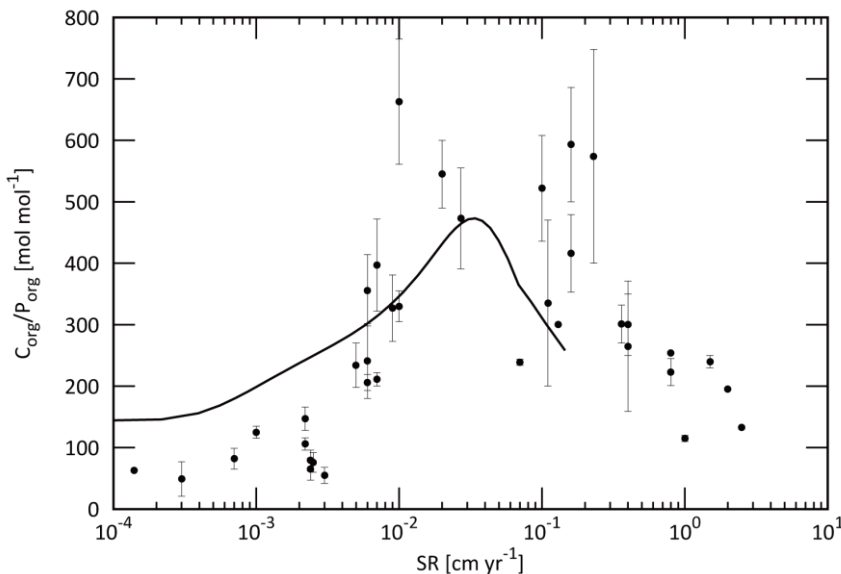
1480 The modelled marine DIP inventory is 2.75×10^{15} mol, consistent with the observational estimate of around

1481 3×10^{15} mol (e.g., (Delaney, 1998; Guidry et al., 2000)). Given the riverine reactive P input flux of 0.155

1482 Tmol P yr^{-1}, the phosphorus residence time is estimated at 18 kyr, which is also consistent with previous

1483 estimates of 20 kyr or shorter (Benitez-Nelson, 2000; Ruttenberg, 2003).

1484

1485

1486 **Figure 15.** C_{org}/P_{org} ratios of buried sediments as a function of sedimentation rate. Black dots represent the
1487 observational data (Ingall and Cappellen, 1990). The simulated C_{org}/P_{org} ratios for the LD region obtained
1488 from our reference run is shown as a black line.

3.2.4 Nitrogen cycling

Nitrogen export production is 1603 Tg N yr^{-1}, which is coupled with carbon according to the C:N stoichiometry of organic matter. Simulated N fixation required for the N balance in the ocean is 180 Tg N yr^{-1}, which is higher than the range of previous estimates of 110–150 Tg N yr^{-1} (Luo et al., 2012; Gruber and Sarmiento, 1997; Galloway et al., 2004; Karl et al., 2002; Fowler et al., 2013; Duce et al., 2008; Deutsch et al., 2007), while a recent study (Großkopf et al., 2012) suggests a higher value of ~180 Tg N yr^{-1}. This discrepancy is partly because atmospheric deposition is ignored in the CANOPS-GRB model, which contributes 25.8 Tg N yr^{-1} (Wang et al., 2019). Gruber and Sarmiento (2002) (Gruber and Sarmiento, 2002) estimated the pre-industrial value of the total source of N as 188±44 Tg N yr^{-1}. Our estimate of 196 Tg N yr^{-1} is within this range.

Nitrogen fluxes in an oxic water column are tightly coupled with the C_{org} fluxes, whereas decoupling appears in suboxic environments. Simulated denitrification in the water column is 102 Tg N yr^{-1}, within the range of the observational estimates (50–150 Tg N yr^{-1}) (Devries et al., 2012; Devries et al., 2013; Brandes and Devol, 2002; Gruber, 2008; Gruber and Sarmiento, 2002; Oschlies et al., 2008; Wang et al., 2019). Modelled benthic denitrification is 62 Tg N yr^{-1}, which is lower than the estimated range of 90–300 Tg N yr^{-1} (Devries et al., 2012; Devries et al., 2013; Brandes and Devol, 2002; Eugster and Gruber, 2012; Devol, 2015; Wang et al., 2019) by a factor of 1.5–5, suggesting that further efforts are required to improve representation of this process. One possible explanation for this discrepancy is that our model is not sufficient to express benthic N cycling because we ignore localized upwelling regions (such as the eastern Tropical Pacific and the Arabian Sea) and coastal regions where benthic denitrification is significant POM decomposition pathway in favor of globally averaged parameterizations. The separate treatment of continental shelves and margin sediments from the pelagic ocean could improve this issue. We also ignore another denitrification mechanism: anaerobic ammonium oxidation (anammox), which will often play an important role in the loss of fixed nitrogen in marine sediments and pelagic anoxic zones (Karthäuser et al., 2021; Kuypers et al., 2005).

1516 The modelled DIN inventory is $4.5×10^5$ Tg N. Given the total source flux of 196 Tg N yr^{-1}, the residence

1517 time of DIN is estimated at 2.3 kyr.

3.2.5 Sulfur cycling

1519 MSR is a major early diagenetic pathway of carbon oxidation in organic-rich sediments deposited below

1520 oxygenated waters. For the standard run, aerobic oxidation is a dominant process in the water column, but

1521 MSR contributes 37% of benthic degradation. CANOPS-GRB estimates a global rate of benthic sulfate

1522 reduction at 11 Tmol S yr^{-1}. This is lower than the previously reported value of gross MSR (40–75 Tmol S

1523 yr^{-1}; (Canfield and Farquhar, 2009; Jørgensen and Kasten, 2006) but agrees better with net MSR (Bowles et

1524 al., 2014). Bowles et al. (Bowles et al., 2014) have estimated global net MSR at 6.2 Tmol S yr^{-1} and 11.3

1525 Tmol S yr^{-1} for $z > 200$ m depth and $z > 0$ m depth, respectively. Our estimate is within this range. MSR is

1526 most pronounced on the shelf where high fluxes of organic matter to the seafloor lead to shallow OPD, high

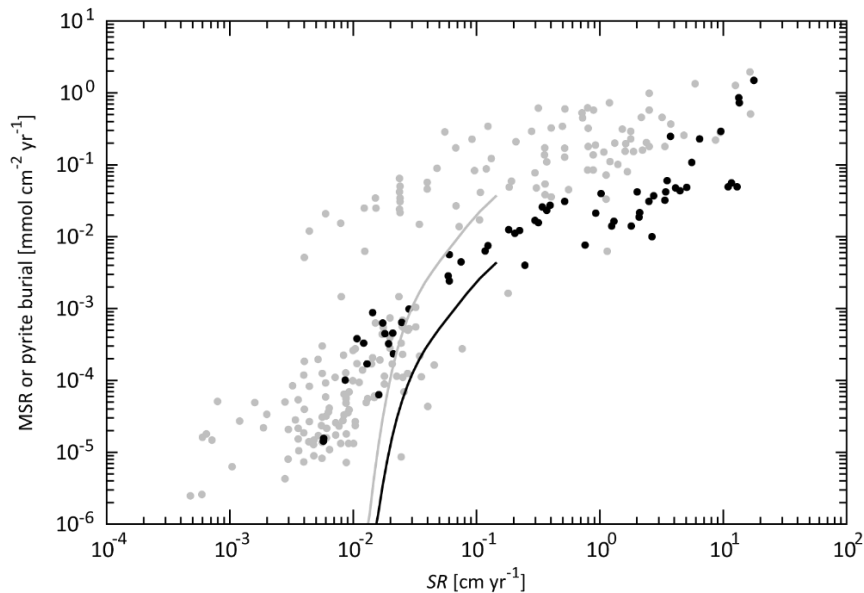
1527 sulfide production, and consequently high pyrite precipitation (Fig. 16).

1528

Figure 16. MSR and pyrite burial flux density as a function of sedimentation rate. Gray and black dots depict observational data compilation of depth-integrated MSR flux density and pyrite burial flux density for normal (oxic) marine sediments (Berner and Canfield, 1989; Canfield, 1989; Raiswell and Canfield, 2012). The unit of sedimentation rate was converted from g cm^{-2} yr^{-1} to cm yr^{-1} with assuming the dry bulk density of 2.5 g cm^{-3} and porosity of 0.9. Solid lines are the results obtained from the reference run of the CANOPS-GRB model.

1535

1536 In the CANOPS-GRB model, pyrite burial efficiency e_{pyr} (Sect. 2.3.4) for sediments underlying oxic bottom

1537 waters is set such that simulated seawater [SO_4^{2-}] of the reference run is consistent with the modern value of

1538 28.9 mM. The tuned value of 11.7% agrees well with observations suggesting that pyrite precipitation rate is

1539 about 10–20% of the rate of MSR (Fig. 16). Our reference value is also consistent with other estimates of

1540 11–20% (ref.(Bottrell and Newton, 2006; Tarhan et al., 2015; Turchyn and Schrag, 2006)).

1541

1542 The sulfate inventory of our reference state is 39×10^{18} mol. Given the total source flux of 3.4 Tmol S yr^{-1},

1543 the residence time of sulfate is 11.5 Myr.

3.3 Global oxygen cycling

1544

1545 The global O_2 budget for our reference state is shown in Fig. 17. The simulated O_2 inventory in the ocean-

1546 atmosphere system is 38×10^{18} mol (atmosphere = 38×10^{18} mol, ocean = 0.23×10^{18} mol). Organic carbon

1547 burial represents a major O_2 source flux (marine = 11.3 Tmol O_2 equiv. yr^{-1} and terrigenous = 3 Tmol O_2

1548 equiv. yr^{-1}). Pyrite burial and hydrogen escape to space contribute 2.6 Tmol O_2 equiv. yr^{-1} and 0.001 Tmol

1549 O_2 equiv. yr^{-1}, respectively. Given the total source/sink flux of 16.9 Tmol O_2 yr^{-1}, the residence time of O_2 in

1550 the ocean-atmosphere system of our reference state is estimated as 2.26 Myr, which is consistent with

1551 previous estimates of 2–4 Myr (Berner, 1989; Berner, 2004a; Garrels and Perry, 1974).

1552

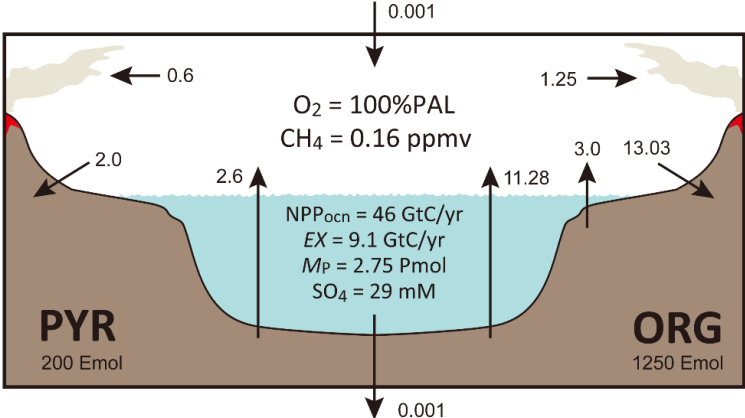



Figure 17. Schematics of global redox (O_2) budget for the reference run. Arrows represent the O_2 flux in terms of 10^{12} mol O_2 equiv. yr^{-1}. PAL = present atmospheric level. Pmol = 10^{15} mol. Emol = 10^{18} mol. ORG = sedimentary organic carbon. PYR = sedimentary pyrite sulfur.

4 Sensitivity experiment

Based on the results obtained above, we conclude that the CANOPS-GRB model is sufficient to describe basic biogeochemical characteristics in the modern ocean-atmosphere system. As a next step, we assess the dynamic response of the full model by performing sensitivity experiments with respect to P availability in surface environments.

4.1 Dynamic response to changes in P weathering

Here, we conduct a sensitivity experiment with respect to the P weathering rate in order to see how the atmospheric and oceanic O_2 levels respond to changes in P availability in the exogenic system over a wide range of timescales. Specifically, we performed four simulations, varying the values of f_P in Eq. (2) over two orders of magnitude. The reference state presented in the previous section is assumed for the initial condition, and the full model is allowed to evolve freely for three billion model years. These experiments demonstrate how P availability in surface environments affects global biogeochemical cycles and redox states of the atmosphere and oceans over a diverse range of timescales.

The simulated transient response is shown in Fig. 18. As expected, lower P availability leads to a lower oceanic P inventory (Fig. 18a), resulting in suppressed biological productivity in the ocean (Fig. 18b). Given the residence time of P in the ocean (20 kyr, see Sect. 3.2.4), these responses occur within 10^5 yr. The suppressed biological productivity leads a decline of burial rate of organic matter in sediments (Fig. 18c). Specifically, 10% and 1% of f_P give rise to the burial rate of marine C_{org} of 1 Tmol C yr^{-1} and 0.13 Tmol C yr^{-1} at 10^5 yr, respectively (cf. the reference value of 11.3 Tmol C yr^{-1}).

On the timescales of 10^5–10^6 yr, the system reaches a quasi-steady state, but there is still a large redox imbalance due mainly to the suppression of C_{org} burial (Fig. 18i). This gives rise to deoxygenation of the atmosphere on a time scale of millions of years (Fig. 18d). Note that once the ocean interior becomes anoxic, the enhanced P recycling and preservation of organic matter in anoxic marine sediments tend to buffer the atmospheric deoxygenation (Figs. 18a and c). However, these passive responses do not alter the fundamental behavior: lower P availability results in lower atmospheric O_2 levels. After atmospheric deoxygenation (>~4

Myr), the system again reaches its quasi-steady state. Specifically, f_P values of 10% and 1% result in atmospheric O_2 levels of 9% PAL and 0.6% PAL, respectively.

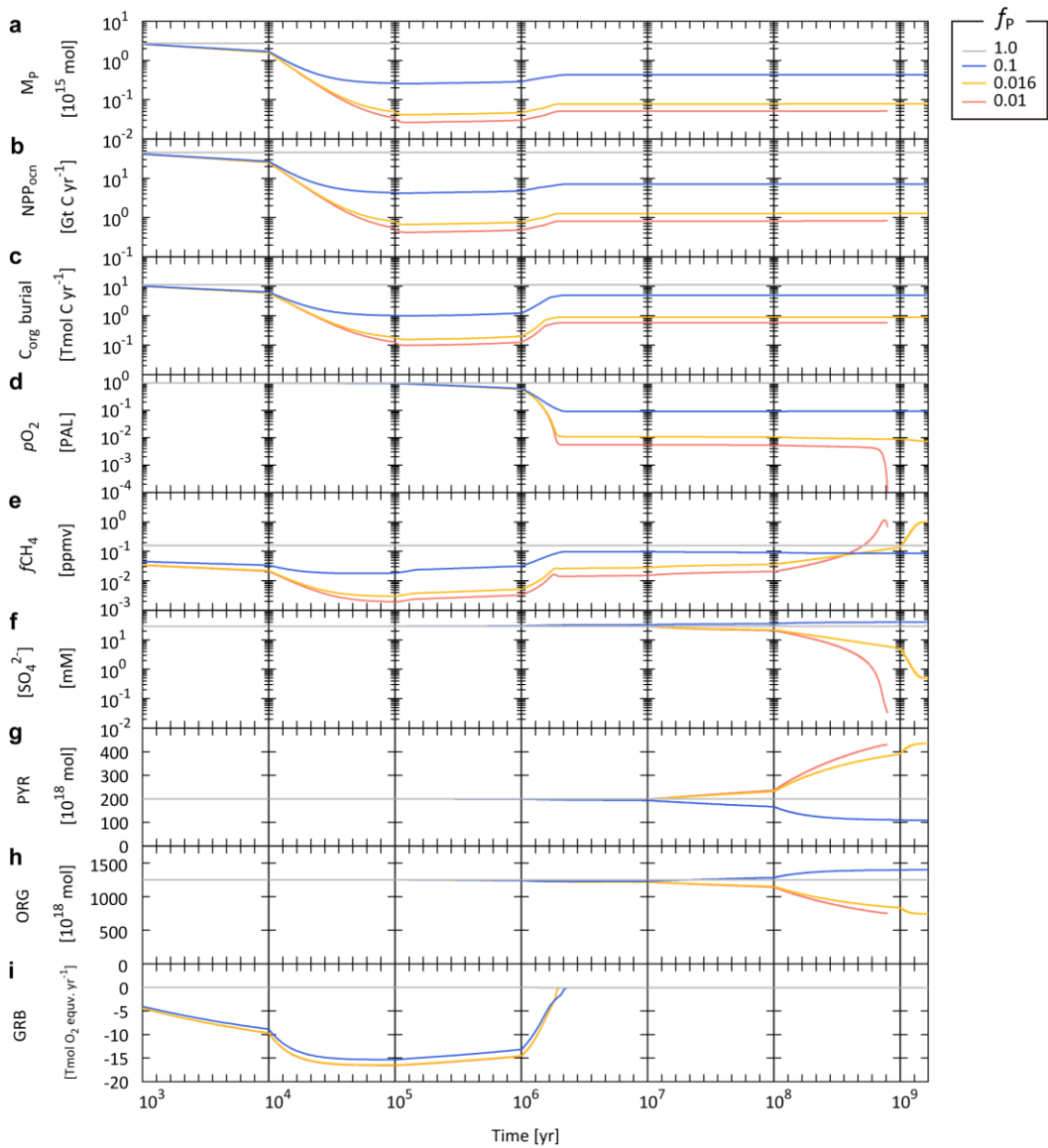



Figure 18. Biogeochemical responses obtained from the CANOPS-GRB model with different values of P availability, f_P. (**a**) Oceanic phosphate inventory, M_P. (**b**) Oceanic net primary production (NPP$_{ocn}$). (**c**) Burial rate of organic carbon (C$_{org}$) in marine sediments. (**d**) Atmospheric partial pressure of O_2. PAL = present atmospheric level. (**e**) Atmospheric CH$_4$ mixing ratio. (**f**) Sulfate concentration in the surface ocean layer. (**g**) Sedimentary reservoir size of pyrite sulfur, *PYR*. (**h**) Sedimentary reservoir size of organic carbon, *ORG*. (**i**) Global redox budget, *GRB*. For the f_P = 1% run (red line), the calculation stopped when the atmospheric O_2 level decreased to ~10^{-5} PAL due to the numerical instability.

1593

1594 The following change is driven by the response of oceanic S cycle, which is characterized by the long

1595 residence time of 11.5 Myr (see Sect. 3.2.6). Ocean anoxia promotes the MSR and subsequent precipitation

1596 of pyrite in the ocean interior. However, our model demonstrates that the decline of seawater SO_4^{2-} on a

1597 timescale of tens of millions of years is small (Fig. 18f), because the rate of MSR depends not only on the

1598 oceanic redox state but on the availability of organic matter for the MSR. The significant reduction of

1599 seawater SO_4^{2-} occurs on the longer timescales (>100 Myr) for extremely low f_P scenarios (0.016 and 0.01),

1600 in which atmospheric O_2 levels decrease to <1% PAL. These scenarios also accompany with a growth of

1601 sedimentary S from gypsum to pyrite (Fig. 18g).

1602

1603 On longer timescales, sedimentary reservoirs affect the redox state of the atmosphere and oceans. The

1604 present result demonstrates that f_P of 1% finally leads to the catastrophic decrease in atmospheric O_2 level at

1605 around 0.9 billion years (Fig. 18d). The simulation was stopped at this point due to the numerical instability.

1606 For other scenarios, the system reaches a new steady state after roughly three billion model years.

1607

1608 Biogenic CH_4 production tends to be enhanced in anoxic oceans. However, the present result demonstrates

1609 that CH_4 degassing to the atmosphere is inhibited by both limited availability of organic matter for

1610 methanogenesis and the anaerobic oxidation of CH_4 by SO_4^{2-}. Once seawater $[SO_4^{2-}]$ decreases below 1 mM,

1611 CH_4 can escape from oceans to the atmosphere, promoting the buildup of CH_4 in the atmosphere.

1612 Nevertheless, because of limited biological activity, atmospheric CH_4 levels remain comparable to the

1613 modern value (~1 ppmv) (Fig. 18e).

1614 **4.2 O_2 budget for the less oxygenated scenario**

1615 Figure 19 shows the O_2 budget of the less oxygenated state (f_P = 1.6% scenario). Because P availability

1616 exerts a primary control on biospheric O_2 production, the strongly suppressed P delivery to the ocean leads

1617 to low oceanic P levels and commensurately low biological productivity (0.08 Pmol and 1.3 Gt C yr^{-1},

1618 respectively). As a consequence, the atmospheric O_2 level is low (0.75% PAL). In this scenario the ocean

1619 interior is globally anoxic, and the preservation of organic C in marine sediments is enhanced. However, the

1620   suppressed biological productivity results in a low overall burial rate of organic C (0.9 Tmol O_2 equiv. yr^{-1};

1621   ~9% of the reference value). When combined with the burial rate of terrigenous organic C, total O_2

1622   production by the organic C sub-cycle is 0.97 Tmol O_2 equiv. yr^{-1}. This O_2 source is balanced by the sum of

1623   oxidative weathering and metamorphism. The role of the pyrite S sub-cycle in the global redox budget is

1624   also shown in Fig. 19. Most of the SO_4^{2-} entering the anoxic ocean is buried as pyrite, representing a major

1625 O_2 source (2.64 Tmol O_2 equiv. yr^{-1}). This O_2 source is balanced by oxidation of sedimentary pyrite S

1626   through weathering (1.33 Tmol O_2 equiv. yr^{-1}) and metamorphism (1.31 Tmol O_2 equiv. yr^{-1}). In other words,

1627   the O_2 budget for the weakly-oxygenated Earth system is largely affected by the crustal S sub-cycle. This is

1628   in marked contrast to the well-oxygenated Earth system, on which the O_2 budget is mainly controlled by

1629   organic C sub-cycle.

1630

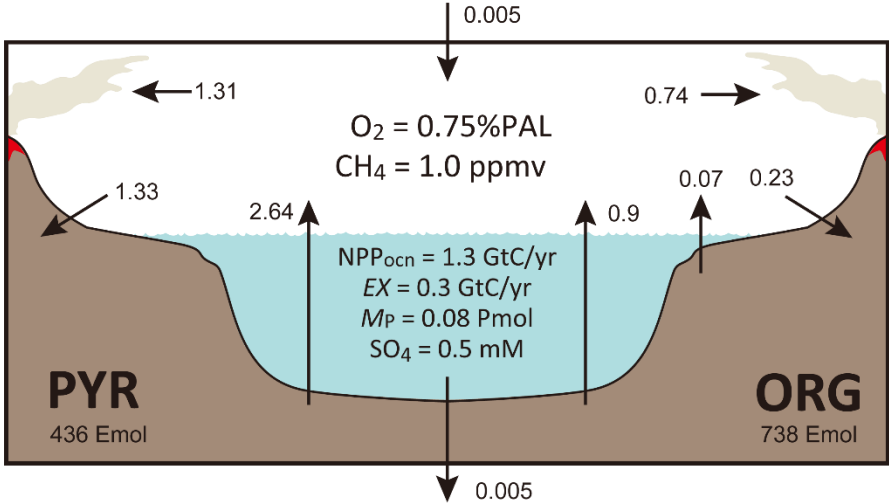
1631   The present result demonstrates that low atmospheric O_2 states (~1% PAL) can be achieved in scenarios

1632   where the availability of P is strongly limited. However, a slight decrease of f_P to 1% leads to the

1633   destabilization of global O_2 budget, providing implications for the stability and evolution of atmospheric O_2

1634   levels during the Proterozoic. This point will be further systematically examined in future work.

1635

1636

1637   **Figure 19.** Schematics of global redox (O_2) budget for the $f_P = 1.6\%$ ($=10^{-1.8}$) run. Arrows represent the O_2
1638   flux in terms of 10^{12} mol O_2 equiv. yr^{-1}. PAL = present atmospheric level. Pmol = 10^{15} mol. Emol = 10^{18} mol.
1639   Gt C = 10^{15} g C. ORG = sedimentary organic C. PYR = sedimentary pyrite S.

5 Discussion

The reference run under the present condition demonstrates generally good agreement with modern observations (Sect. 3). The water circulation scheme provides an adequate representation of general ocean circulation, resulting in robust and reliable tracer distributions that are comparable to the modern observations. This provides a mechanistic foundation for simulating generalized ocean biogeochemical cycles. The ocean biogeochemistry module includes a series of biogeochemical processes in oxic-anoxic-sulfidic environments. The reference run gives rise to the distributions of nutrients and dissolved O_2 that capture fundamental properties observed in the modern ocean. Integrated biogeochemical fluxes of the global ocean, such as biological productivity, material flow in the water column, and burial into sediments are also consistent with observational data. In our reference run the estimated organic carbon burial and oxidative weathering fluxes are relatively high compared to some previous estimates, though there remains significant uncertainty in globally integrated organic carbon weathering and burial fluxes. Further work will also be needed to better quantify the biogeochemical cycling in the continental shelf, which is a major locus of organic matter burial. In addition, some future developments to the N cycle may be needed, especially with regard to denitrification (e.g., anammox, coastal benthic denitrification). Despite of these remaining challenges, our biogeochemical model is adequate for representing the general property of the coupled C-N-P-O_2-S cycles.

A new scheme for oxidative weathering of organic matter and pyrite sulfur, mass balance calculation of O_2 in the atmosphere, and time evolution of sedimentary reservoirs are explicitly included in the CANOPS-GRB model. These are a significant improvement from the previous versions of CANOPS (Lenton, 2020). The simplified framework for the global O_2 budget is also useful to understand the response of complex biogeochemical systems. The computational efficiency of our CANOPS-GRB model allows us to conduct simulations over billions of model years with reasonable wall times (on the order of weeks), providing a useful tool for exploring the wide range of topics about the oxygenation history of Earth's atmosphere.

Sensitivity experiments with respect to the terrestrial weathering rate of P were conducted in order to see how the redox state of the ocean-atmosphere system responds to varying P availability in the surface system

1668 (Sect. 4). The CANOPS-GRB model appears to adequately simulate the biogeochemical dynamics over a
1669 wide range of timescales and is applicable for quantitative assessment of the evolution and stability of
1670 Earth's O_2 cycling. Perhaps even more importantly, our results encourage us to perform further systematic
1671 examinations with Earth system models which have different complexities. Such an 'Earth system model
1672 intercomparison' would be a critical step towards better mechanistic understanding of the stability and
1673 dynamics of atmospheric O_2 levels over Earth's history.

1674

1675 Due to a lack of explicit Fe cycling and anaerobic metabolisms (such as anoxygenic photosynthesis), the
1676 current version of the model cannot be applicable for the simulation under the Archean-like weakly-
1677 oxygenated (pO_2 <10^{-5} PAL) conditions. These topics are left to future studies, but it would be an achievable
1678 goal (Ozaki et al., 2018; Ozaki et al., 2019b; Van De Velde et al., 2021). The model design presented here
1679 also ignores the interaction between the surface system and the mantle (e.g., subduction) except for the
1680 degassing of reducing gases from the mantle. We note, however, that the surface-mantle interaction would
1681 have exerted a primary control on the long-term redox budget of Earth's surface system through the Earth's
1682 history (Canfield, 2004; Eguchi et al., 2020; Hayes and Waldbauer, 2006) and may be important for the
1683 discussion about the distant future (Ozaki and Reinhard, 2021). The importance of mantle and solid Earth
1684 controls on surficial environments is a crucially important topic for future research.

1685

1686 The CANOPS-GRB model has the basic capability to simulate the time evolution of the abundance of
1687 atmospheric biosignature gases (O_2 and CH_4) on a wide range of timescales. While the biogeochemical
1688 model is based on process studies to the extent possible, many processes are derived from empirical
1689 calibrations to Earth-like planets around sun-like stars. Clearly, some these parameterizations, such as the
1690 photochemical parameterization among O_2-O_3-CH_4 (Eq. 107), must be modified when applying the model to
1691 a range of habitable Earth-like exoplanets.

6 Conclusions

A new Earth system box model was developed — CANOPS-GRB v1.0. The new code release provides an improved description of the coupled C-N-P-O_2-S biogeochemical cycles in the ocean-atmosphere-crust system, which can be utilized to examine the dynamics and stability of Earth's O_2 cycle over a wide range of timescales. The computational efficiency and simple model design of CANOPS-GRB make it relatively easy to modify existing processes or add entirely new processes and components. CANOPS-GRB is thus a new and uniquely flexible tool capable of providing a coherent mechanistic framework for quantifying the biogeochemical cycles regulating Earth's O_2 cycle. CANOPS-GRB is also a useful tool for the development of more comprehensive, low- to intermediate-complexity Earth system box models of biogeochemistry.

CANOPS-GRB provides an important step forward when coupled to new and existing geochemical proxy data. The accumulating geological/geochemical records have led to new hypothesis for the evolution of atmospheric O_2 levels on Earth. CANOPS-GRB was designed to facilitate simulation of a wide range of past conditions so as to permit more explicit testing of hypothesis about the function of biogeochemical cycles and its effect on the redox budget through Earth history. Through the model-data synergy, CANOPS-GRB has great potential to provide an integrated, quantitative, and statistically informative picture of biogeochemical states, opening new perspectives on a wide range of scientific questions in research seeking to understand the Earth's chemical evolution, and in particular the cause-and-effect relationships with evolving biosphere.

CANOPS-GRB also provides significant steps forward in our predictive understanding of the links between geology, biogeochemistry, and the evolution of Earth's biosphere. It will allow for a fundamentally new and more precise quantitative understanding of evolving atmospheric biosignatures (O_2, O_3, CH_4) on Earth, and will broaden the interpretive power of Earth system evolution in the search for life beyond our planet. Additional elaboration of the CANOPS-GRB code could represent an important avenue for developing a more robust tool for diagnosing atmospheric biosignatures for future analysis of extrasolar worlds. In sum, it is anticipated that CANOPS-GRB will have many applications for problems linking the coupled evolution of life and the atmosphere on Earth and habitable rocky exoplanets.

1720

1721

Code availability. The bulk of CANOPS-GRB is written in Fortran as a stand-alone model. The model code can be found at GitHub (doi:10.5281/zenodo.5893804). This model is still undergoing regular development and it is recommended that potential users contact the corresponding author (Kazumi Ozaki; ozaki.k.ai@m.titech.ac.jp) to obtain the latest version.

Author contribution. KO designed the study, wrote the code, and ran model simulations. DBC and CTR contributed to code debugging. KO wrote the manuscript, with inputs from DBC, CTR, and ET.

Competing interests. The authors declare that they have no conflict of interest.

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References

Alcott, L. J., Mills, B. J. W., and Poulton, S. W.: Stepwise Earth oxygenation is an inherent property of global biogeochemical cycling, Science, 366, 1333-1337, doi:10.1126/science.aax6459, 2019.

Algeo, T. J. and Ingall, E.: Sedimentary C_{org}:P ratios, paleocean ventilation, and Phanerozoic atmospheric pO_2, Palaeogeogr. Palaeoclimatol. Palaeoecol., 256, 130-155, http://dx.doi.org/10.1016/j.palaeo.2007.02.029, 2007.

Anderson, L. D., Delaney, M. L., and Faul, K. L.: Carbon to phosphorus ratios in sediments: Implications for nutrient cycling, Glob. Biogeochem. Cycles, 15, 65-79, https://doi.org/10.1029/2000GB001270, 2001.

Archer, D., Kheshgi, H., and Maier-Reimer, E.: Dynamics of fossil fuel CO_2 neutralization by marine $CaCO_3$, Glob. Biogeochem. Cycles, 12, 259-276, https://doi.org/10.1029/98GB00744, 1998.

Archer, D. E., Morford, J. L., and Emerson, S. R.: A model of suboxic sedimentary diagenesis suitable for automatic tuning and gridded global domains, Glob. Biogeochem. Cycles, 16, 17-11-17-21, 10.1029/2000gb001288, 2002.

Archer, D. E., Eshel, G., Winguth, A., Broecker, W., Pierrehumbert, R., Tobis, M., and Jacob, R.: Atmospheric pCO_2 sensitivity to the biological pump in the ocean, Glob. Biogeochem. Cycles, 14, 1219-1230, https://doi.org/10.1029/1999GB001216, 2000.

Armstrong, R. A., Lee, C., Hedges, J. I., Honjo, S., and Wakeham, S. G.: A new, mechanistic model for organic carbon fluxes in the ocean based on the quantitative association of POC with ballast minerals, Deep Sea Research Part II: Topical Studies in Oceanography, 49, 219-236, https://doi.org/10.1016/S0967-0645(01)00101-1, 2001.

Arndt, S., Regnier, P., Godderis, Y., and Donnadieu, Y.: GEOCLIM *reloaded* (v 1.0): a new coupled earth system model for past climate change, Geoscientific Model Development, 4, 451-481, 10.5194/gmd-4-451-2011, 2011.

Baturin, G. N.: Issue of the relationship between primary productivity of organic carbon in ocean and phosphate accumulation (Holocene-Late Jurassic), Lithology and Mineral Resources, 42, 318-348, 10.1134/s0024490207040025, 2007.

Beal, E. J., Claire, M. W., and House, C. H.: High rates of anaerobic methanotrophy at low sulfate concentrations with implications for past and present methane levels, Geobiology, 9, 131-139, 10.1111/j.1472-4669.2010.00267.x, 2011.

Belcher, C. M. and McElwain, J. C.: Limits for combustion in low O_2 redefine paleoatmospheric predictions for the Mesozoic, Science, 321, 1197-1200, 10.1126/science.1160978, 2008.

Bellefroid, E. J., Hood, A. v. S., Hoffman, P. F., Thomas, M. D., Reinhard, C. T., and Planavsky, N. J.: Constraints on Paleoproterozoic atmospheric oxygen levels, Proc. Natl Acad. Sci. USA, 115, 8104-8109, 10.1073/pnas.1806216115, 2018.

Benitez-Nelson, C. R.: The biogeochemical cycling of phosphorus in marine systems, Earth-Science Reviews, 51, 109-135, https://doi.org/10.1016/S0012-8252(00)00018-0, 2000.

Berelson, W. M.: Particle settling rates increase with depth in the ocean, Deep Sea Research Part II: Topical Studies in Oceanography, 49, 237-251, https://doi.org/10.1016/S0967-0645(01)00102-3, 2001a.

Berelson, W. M.: The Flux of Particulate Organic Carbon Into the Ocean Interior: A Comparison of Four U.S. JGOFS Regional Studies, Oceanography, 14, 2001b.

Berelson, W. M., Balch, W. M., Najjar, R., Feely, R. A., Sabine, C., and Lee, K.: Relating estimates of $CaCO_3$ production, export, and dissolution in the water column to measurements of $CaCO_3$ rain into sediment traps and dissolution on the sea floor: A revised global carbonate budget, Glob. Biogeochem. Cycles, 21, https://doi.org/10.1029/2006GB002803, 2007.

Bergman, N. M., Lenton, T. M., and Watson, A. J.: COPSE: A new model of biogeochemical cycling over Phanerozoic time, Am. J. Sci., 304, 397-437, 10.2475/ajs.304.5.397, 2004.

Berner, R. A., Holland, H. D. (Ed.): Early diagenesis: A theoretical approach, Princeton University Press, Princeton, 256 pp.1980.

Berner, R. A.: Burial of organic carbon and pyrite sulfur in the modern ocean; its geochemical and environmental significance, Am. J. Sci., 282, 451-473, 10.2475/ajs.282.4.451, 1982.

Berner, R. A.: Biogeochemical cycles of carbon and sulfur and their effect on atmospheric oxygen over phanerozoic time, Palaeogeogr. Palaeoclimatol. Palaeoecol., 75, 97-122, http://dx.doi.org/10.1016/0031-0182(89)90186-7, 1989.

Berner, R. A.: The Phanerozoic Carbon Cycle: CO_2 and O_2, Oxford University Press2004a.

Berner, R. A.: A model for calcium, magnesium and sulfate in seawater over Phanerozoic time, Am. J. Sci., 304, 438-453, 10.2475/ajs.304.5.438, 2004b.

Berner, R. A.: GEOCARBSULF: A combined model for Phanerozoic atmospheric O_2 and CO_2, Geochim. Cosmochim. Acta, 70, 5653-5664, http://dx.doi.org/10.1016/j.gca.2005.11.032, 2006.

Berner, R. A.: Phanerozoic atmospheric oxygen: New results using the GEOCARBSULF model, Am. J. Sci., 309, 603-606, 10.2475/07.2009.03, 2009.

Berner, R. A. and Canfield, D. E.: A new model for atmospheric oxygen over Phanerozoic time, Am. J. Sci., 289, 333-361, 10.2475/ajs.289.4.333, 1989.

Berner, R. A. and Westrich, J. T.: Bioturbation and the early diagenesis of carbon and sulfur, Am. J. Sci., 285, 193-206, 10.2475/ajs.285.3.193, 1985.

Betts, J. N. and Holland, H. D.: The oxygen content of ocean bottom waters, the burial efficiency of organic carbon, and the regulation of atmospheric oxygen, Palaeogeogr. Palaeoclimatol. Palaeoecol., 97, 5-18, http://dx.doi.org/10.1016/0031-0182(91)90178-T, 1991.

Bohlen, L., Dale, A. W., and Wallmann, K.: Simple transfer functions for calculating benthic fixed nitrogen losses and C:N:P regeneration ratios in global biogeochemical models, Glob. Biogeochem. Cycles, 26, GB3029, 10.1029/2011gb004198, 2012.

Bolton, E. W., Berner, R. A., and Petsch, S. T.: The Weathering of Sedimentary Organic Matter as a Control on Atmospheric O_2: II. Theoretical Modeling, Am. J. Sci., 306, 575-615, 10.2475/08.2006.01, 2006.

Bottrell, S. H. and Newton, R. J.: Reconstruction of changes in global sulfur cycling from marine sulfate isotopes, Earth-Science Reviews, 75, 59-83, https://doi.org/10.1016/j.earscirev.2005.10.004, 2006.

Boudreau, B. P.: A method-of-lines code for carbon and nutrient diagenesis in aquatic sediments, Computers & Geosciences, 22, 479-496, http://dx.doi.org/10.1016/0098-3004(95)00115-8, 1996.

Bowles, M. W., Mogollón, J. M., Kasten, S., Zabel, M., and Hinrichs, K.-U.: Global rates of marine sulfate reduction and implications for sub–sea-floor metabolic activities, Science, 344, 889-891, 10.1126/science.1249213, 2014.

Bradley, J. A., Arndt, S., Amend, J. P., Burwicz, E., Dale, A. W., Egger, M., and LaRowe, D. E.: Widespread energy limitation to life in global subseafloor sediments, Science Advances, 6, eaba0697, doi:10.1126/sciadv.aba0697, 2020.

Brandes, J. A. and Devol, A. H.: A global marine-fixed nitrogen isotopic budget: Implications for Holocene nitrogen cycling, Glob. Biogeochem. Cycles, 16, GB001856, 10.1029/2001gb001856, 2002.

Broecker, W. S. and Peng, T.-H.: Tracers in the sea, Eldigio Pr, New York1982.

Burdige, D. J.: Burial of terrestrial organic matter in marine sediments: A re-assessment, Glob. Biogeochem. Cycles, 19, 10.1029/2004gb002368, 2005.

Burdige, D. J.: Preservation of Organic Matter in Marine Sediments: Controls, Mechanisms, and an Imbalance in Sediment Organic Carbon Budgets?, Chemical Reviews, 107, 467-485, 10.1021/cr050347q, 2007.

Canfield, D. E.: Sulfate reduction and oxic respiration in marine sediments: implications for organic carbon preservation in euxinic environments, Deep Sea Research Part A. Oceanographic Research Papers, 36, 121-138, https://doi.org/10.1016/0198-0149(89)90022-8, 1989.

Canfield, D. E.: Sulfate reduction in deep-sea sediments, Am. J. Sci., 291, 177-188, 10.2475/ajs.291.2.177, 1991.

Canfield, D. E.: Organic Matter Oxidation in Marine Sediments, in: Interactions of C, N, P and S Biogeochemical Cycles and Global Change, edited by: Wollast, R., Mackenzie, F. T., and Chou, L., Springer Berlin Heidelberg, Berlin, 333-363, 1993.

Canfield, D. E.: The evolution of the Earth surface sulfur reservoir, Am. J. Sci., 304, 839-861, 10.2475/ajs.304.10.839, 2004.

Canfield, D. E. and Farquhar, J.: Animal evolution, bioturbation, and the sulfate concentration of the oceans, Proc. Natl Acad. Sci. USA, 106, 8123-8127, 10.1073/pnas.0902037106, 2009.

Canfield, D. E., Zhang, S., Frank, A. B., Wang, X., Wang, H., Su, J., Ye, Y., and Frei, R.: Highly fractionated chromium isotopes in Mesoproterozoic-aged shales and atmospheric oxygen, Nature Commun., 9, 2871, 10.1038/s41467-018-05263-9, 2018.

Carr, M.-E., Friedrichs, M. A. M., Schmeltz, M., Noguchi Aita, M., Antoine, D., Arrigo, K. R., Asanuma, I., Aumont, O., Barber, R., Behrenfeld, M., Bidigare, R., Buitenhuis, E. T., Campbell, J., Ciotti, A., Dierssen, H., Dowell, M., Dunne, J., Esaias, W., Gentili, B., Gregg, W., Groom, S., Hoepffner, N., Ishizaka, J., Kameda, T., Le Quéré, C., Lohrenz, S., Marra, J., Mélin, F., Moore, K., Morel, A., Reddy, T. E., Ryan, J., Scardi, M., Smyth, T., Turpie, K., Tilstone, G., Waters, K., and Yamanaka, Y.: A comparison of global estimates of marine primary production from ocean color, Deep Sea Research Part II: Topical Studies in Oceanography, 53, 741-770, https://doi.org/10.1016/j.dsr2.2006.01.028, 2006.

Catling, D. C. and Kasting, J. F.: Atmospheric Evolution on Inhabited and Lifeless Worlds, Cambridge University Press2017.

Catling, D. C. and Zahnle, K. J.: The Archean atmosphere, Science Advances, 6, eaax1420, 10.1126/sciadv.aax1420, 2020.

Cha, H. J., Lee, C. B., Kim, B. S., Choi, M. S., and Ruttenberg, K. C.: Early diagenetic redistribution and burial of phosphorus in the sediments of the southwestern East Sea (Japan Sea), Marine Geology, 216, 127-143, 10.1016/j.margeo.2005.02.001, 2005.

Claire, M. W., Catling, D. C., and Zahnle, K. J.: Biogeochemical modelling of the rise in atmospheric oxygen, Geobiology, 4, 239-269, 10.1111/j.1472-4669.2006.00084.x, 2006.

Cole, D. B., Ozaki, K., and Reinhard, C. T.: Atmospheric Oxygen Abundance, Marine Nutrient Availability, and Organic Carbon Fluxes to the Seafloor, Global Biogeochemical Cycles, 36, e2021GB007052, https://doi.org/10.1029/2021GB007052, 2022.

Cole, D. B., Reinhard, C. T., Wang, X., Gueguen, B., Halverson, G. P., Gibson, T., Hodgskiss, M. S. W., McKenzie, N. R., Lyons, T. W., and Planavsky, N. J.: A shale-hosted Cr isotope record of low atmospheric oxygen during the Proterozoic, Geology, 44, 555-558, 10.1130/g37787.1, 2016.

Colman, A. S. and Holland, H. D.: The global diagenetic flux of phosphorus from marine sediments to the oceans: redox sensitivity and the control of atmosphreic oxygen levels, in: Marine authigenesis: from global to microbial, edited by: Glenn, C. R., Prevot-Lucas, L., and Lucas, J., SEPM (Society for Sedimentary Geology), 2000.

Colman, A. S., Holland, H. D., Glenn, C. R., Prévôt-Lucas, L., and Lucas, J.: The Global Diagenetic Flux of Phosphorus from Marine Sediments to the Oceans: Redox Sensitivity and the Control of Atmospheric Oxygen Levels, in: Marine Authigenesis: From Global to Microbial, SEPM Society for Sedimentary Geology, 0, 10.2110/pec.00.66.0053, 2000.

Compton, J., Mallinson, D., Glenn, C. R., Filippelli, G., Follmi, K., Shields, G. A., and Zanin, Y.: Variations in the global phosphorus cycle, in: Marine authigenesis: from global to microbial, edited by: Glenn, C. R., Prevot-Lucas, L., and Lucas, J., SEPM (Society for Sedimentary Geology), 21-33, 2000.

Crichton, K. A., Wilson, J. D., Ridgwell, A., and Pearson, P. N.: Calibration of temperature-dependent ocean microbial processes in the cGENIE.muffin (v0.9.13) Earth system model, Geosci. Model Dev., 14, 125-149, 10.5194/gmd-14-125-2021, 2021.

Crockford, P. W., Hayles, J. A., Bao, H., Planavsky, N. J., Bekker, A., Fralick, P. W., Halverson, G. P., Bui, T. H., Peng, Y., and Wing, B. A.: Triple oxygen isotope evidence for limited mid-Proterozoic primary productivity, Nature, 559, 613-616, 10.1038/s41586-018-0349-y, 2018.

Daines, S. J., Mills, B. J. W., and Lenton, T. M.: Atmospheric oxygen regulation at low Proterozoic levels by incomplete oxidative weathering of sedimentary organic carbon, Nature Commun., 8, 14379, 10.1038/ncomms14379

https://www.nature.com/articles/ncomms14379#supplementary-information, 2017.

Dale, A. W., Meyers, S. R., Aguilera, D. R., Arndt, S., and Wallmann, K.: Controls on organic carbon and molybdenum accumulation in Cretaceous marine sediments from the Cenomanian–Turonian interval including Oceanic Anoxic Event 2, Chem. Geol., 324–325, 28-45, http://dx.doi.org/10.1016/j.chemgeo.2011.10.004, 2012.

Delaney, M. L.: Phosphorus accumulation in marine sediments and the oceanic phosphorus cycle, Glob. Biogeochem. Cycles, 12, 563-572, 10.1029/98gb02263, 1998.

Dellwig, O., Leipe, T., März, C., Glockzin, M., Pollehne, F., Schnetger, B., Yakushev, E. V., Böttcher, M. E., and Brumsack, H.-J.: A new particulate Mn–Fe–P-shuttle at the redoxcline of anoxic basins, Geochim. Cosmochim. Acta, 74, 7100-7115, http://dx.doi.org/10.1016/j.gca.2010.09.017, 2010.

Derry, L. A.: Causes and consequences of mid-Proterozoic anoxia, Geophys. Res. Lett., 42, 2015GL065333, 10.1002/2015gl065333, 2015.

Des Marais, D. J., Harwit, M. O., Jucks, K. W., Kasting, J. F., Lin, D. N., Lunine, J. I., Schneider, J., Seager, S., Traub, W. A., and Woolf, N. J.: Remote Sensing of Planetary Properties and Biosignatures on Extrasolar Terrestrial Planets, Astrobiology, 2, 153-181, 10.1089/15311070260192246, 2002.

Deutsch, C., Sarmiento, J. L., Sigman, D. M., Gruber, N., and Dunne, J. P.: Spatial coupling of nitrogen inputs and losses in the ocean, Nature, 445, 163, 10.1038/nature05392 https://www.nature.com/articles/nature05392#supplementary-information, 2007.

Devol, A. and Christensen, J. P.: Benthic fluxes and nitrogen cycling in sediments of the continental margin of the eastern North Pacific, Journal of Marine Research, 51, 345-372, 1993.

Devol, A. H.: Denitrification, Anammox, and N2 Production in Marine Sediments, Ann. Rev. Mar. Sci., 7, 403-423, 10.1146/annurev-marine-010213-135040, 2015.

DeVries, T., Deutsch, C., Rafter, P. A., and Primeau, F.: Marine denitrification rates determined from a global 3-D inverse model, Biogeosciences, 10, 2481-2496, 10.5194/bg-10-2481-2013, 2013.

DeVries, T., Deutsch, C., Primeau, F., Chang, B., and Devol, A.: Global rates of water-column denitrification derived from nitrogen gas measurements, Nat. Geosci., 5, 547, 10.1038/ngeo1515 https://www.nature.com/articles/ngeo1515#supplementary-information, 2012.

Doney, S. C., Lindsay, K., Caldeira, K., Campin, J. M., Drange, H., Dutay, J. C., Follows, M., Gao, Y., Gnanadesikan, A., Gruber, N., Ishida, A., Joos, F., Madec, G., Maier-Reimer, E., Marshall, J. C., Matear, R. J., Monfray, P., Mouchet, A., Najjar, R., Orr, J. C., Plattner, G. K., Sarmiento, J., Schlitzer, R., Slater, R., Totterdell, I. J., Weirig, M. F., Yamanaka, Y., and Yool, A.: Evaluating global ocean carbon models: The importance of realistic physics, Glob. Biogeochem. Cycles, 18, GB3017, 10.1029/2003gb002150, 2004.

Donis, D., McGinnis, D. F., Holtappels, M., Felden, J., and Wenzhofer, F.: Assessing benthic oxygen fluxes in oligotrophic deep sea sediments (HAUSGARTEN observatory), Deep-Sea Research Part I-Oceanographic Research Papers, 111, 1-10, doi:10.1016/j.dsr.2015.11.007, 2016.

Duce, R. A., LaRoche, J., Altieri, K., Arrigo, K. R., Baker, A. R., Capone, D. G., Cornell, S., Dentener, F., Galloway, J., Ganeshram, R. S., Geider, R. J., Jickells, T., Kuypers, M. M., Langlois, R., Liss, P. S., Liu, S. M., Middelburg, J. J., Moore, C. M., Nickovic, S., Oschlies, A., Pedersen, T., Prospero, J., Schlitzer, R., Seitzinger, S., Sorensen, L. L., Uematsu, M., Ulloa, O., Voss, M., Ward, B., and Zamora, L.: Impacts of Atmospheric Anthropogenic Nitrogen on the Open Ocean, Science, 320, 893-897, 10.1126/science.1150369, 2008.

Dunne, J. P., Sarmiento, J. L., and Gnanadesikan, A.: A synthesis of global particle export from the surface ocean and cycling through the ocean interior and on the seafloor, Glob. Biogeochem. Cycles, 21, n/a-n/a, 10.1029/2006gb002907, 2007.

Eguchi, J., Seales, J., and Dasgupta, R.: Great Oxidation and Lomagundi events linked by deep cycling and enhanced degassing of carbon, Nat. Geosci., 13, 71-76, 10.1038/s41561-019-0492-6, 2020.

Etheridge, D. M., Steele, L. P., Francey, R. J., and Langenfelds, R. L.: Atmospheric methane between 1000 A.D. and present: Evidence of anthropogenic emissions and climatic variability, J. Geophys. Res., 103, 15979-15993, https://doi.org/10.1029/98JD00923, 1998.

Eugster, O. and Gruber, N.: A probabilistic estimate of global marine N-fixation and denitrification, Glob. Biogeochem. Cycles, 26, 10.1029/2012gb004300, 2012.

Fakhraee, M., Planavsky, N. J., and Reinhard, C. T.: The role of environmental factors in the long-term evolution of the marine biological pump, Nat. Geosci., 13, 812-816, 10.1038/s41561-020-00660-6, 2020.

Fiebig, J., Woodland, A. B., D'Alessandro, W., and Püttmann, W.: Excess methane in continental hydrothermal emissions is abiogenic, Geology, 37, 495-498, 10.1130/g25598a.1, 2009.

Filippelli, G. M.: Carbon and phosphorus cycling in anoxic sediments of the Saanich Inlet, British Columbia, Marine Geology, 174, 307-321, https://doi.org/10.1016/S0025-3227(00)00157-2, 2001.

Fowler, D., Coyle, M., Skiba, U., Sutton, M. A., Cape, J. N., Reis, S., Sheppard, L. J., Jenkins, A., Grizzetti, B., Galloway, J. N., Vitousek, P., Leach, A., Bouwman, A. F., Butterbach-Bahl, K., Dentener, F., Stevenson, D., Amann, M., and Voss, M.: The global nitrogen cycle in the twenty-first century, Phil. Trans. R. Soc. B, 368, 10.1098/rstb.2013.0164, 2013.

Francois, R., Honjo, S., Krishfield, R., and Manganini, S.: Factors controlling the flux of organic carbon to the bathypelagic zone of the ocean, Glob. Biogeochem. Cycles, 16, 1087, 10.1029/2001gb001722, 2002.

Froelich, P. N., Klinkhammer, G. P., Bender, M. L., Luedtke, N. A., Heath, G. R., Cullen, D., Dauphin, P., Hammond, D., Hartman, B., and Maynard, V.: Early oxidation of organic matter in pelagic sediments of the eastern equatorial Atlantic: suboxic diagenesis, Geochim. Cosmochim. Acta, 43, 1075-1090, http://dx.doi.org/10.1016/0016-7037(79)90095-4, 1979.

Föllmi, K. B.: The phosphorus cycle, phosphogenesis and marine phosphate-rich deposits, Earth-Science Reviews, 40, 55-124, https://doi.org/10.1016/0012-8252(95)00049-6, 1996.

Galbraith, E. D. and Martiny, A. C.: A simple nutrient-dependence mechanism for predicting the stoichiometry of marine ecosystems, Proc. Natl Acad. Sci. USA, 112, 8199-8204, 10.1073/pnas.1423917112, 2015.

Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S. P., Asner, G. P., Cleveland, C. C., Green, P. A., Holland, E. A., Karl, D. M., Michaels, A. F., Porter, J. H., Townsend, A. R., and Vöosmarty, C. J.: Nitrogen Cycles: Past, Present, and Future, Biogeochemistry, 70, 153-226, 10.1007/s10533-004-0370-0, 2004.

Garcia, H. E. and Gordon, L. I.: Oxygen solubility in seawater: Better fitting equations, Limnology and Oceanography, 37, 1307-1312, 10.4319/lo.1992.37.6.1307, 1992.

Garrels, R. M. and Lerman, A.: Phanerozoic cycles of sedimentary carbon and sulfur, Proc. Natl Acad. Sci. USA, 78, 4652-4656, 1981.

Garrels, R. M. and Perry, J. E. A.: Cycling of carbon, sulfur, and oxygen through geologic time, The Sea, Wiley-Interscience, New York1974.

Goldblatt, C., Lenton, T. M., and Watson, A. J.: Bistability of atmospheric oxygen and the Great Oxidation, Nature, 443, 683-686, http://www.nature.com/nature/journal/v443/n7112/suppinfo/nature05169_S1.html, 2006.

Graham, W. F. and Duce, R. A.: Atmospheric pathways of the phosphorus cycle, Geochim. Cosmochim. Acta, 43, 1195-1208, https://doi.org/10.1016/0016-7037(79)90112-1, 1979.

Großkopf, T., Mohr, W., Baustian, T., Schunck, H., Gill, D., Kuypers, M. M. M., Lavik, G., Schmitz, R. A., Wallace, D. W. R., and LaRoche, J.: Doubling of marine dinitrogen-fixation rates based on direct measurements, Nature, 488, 361, 10.1038/nature11338 https://www.nature.com/articles/nature11338#supplementary-information, 2012.

Gruber, N.: Chapter 1 - The Marine Nitrogen Cycle: Overview and Challenges, in: Nitrogen in the Marine Environment (2nd Edition), Academic Press, San Diego, 1-50, https://doi.org/10.1016/B978-0-12-372522-6.00001-3, 2008.

Gruber, N. and Sarmiento, J. L.: Global patterns of marine nitrogen fixation and denitrification, Glob. Biogeochem. Cycles, 11, 235-266, 10.1029/97gb00077, 1997.

Gruber, N. and Sarmiento, J. L.: Biogeochemical/physical interactions in elemental cycles, in: THE SEA: Biological-Physical Interactions in the Oceans, edited by: Robinson, A. R., McCarthy, J. J., and Rothschild, B. J., John Wiley and Sons, New York, 337-399, 2002.

Guidry, M. W., Mackenzie, F. T., and Arvidson, R. S.: Role of tectonics in phosphorus distribution and cycling, in: Marine Authigenesis: From Global to Microbial, edited by: Glenn, C. R., Prevot-Lucas, L., and Lucas, J., SEPM, 35-51, 2000.

Gundersen, J. K. and Jorgensen, B. B.: Microstructure of diffusive boundary layers and the oxygen uptake of the sea floor, Nature, 345, 604, 10.1038/345604a0, 1990.

Halevy, I., Peters, S. E., and Fischer, W. W.: Sulfate Burial Constraints on the Phanerozoic Sulfur Cycle, Science, 337, 331-334, 10.1126/science.1220224, 2012.

Handoh, I. C. and Lenton, T. M.: Periodic mid-Cretaceous oceanic anoxic events linked by oscillations of the phosphorus and oxygen biogeochemical cycles, Glob. Biogeochem. Cycles, 17, 1092, 10.1029/2003gb002039, 2003.

Hartnett, H. E. and Devol, A. H.: Role of a strong oxygen-deficient zone in the preservation and degradation of organic matter: a carbon budget for the continental margins of northwest Mexico and Washington State, Geochim. Cosmochim. Acta, 67, 247-264, http://dx.doi.org/10.1016/S0016-7037(02)01076-1, 2003.

Hartnett, H. E., Keil, R. G., Hedges, J. I., and Devol, A. H.: Influence of oxygen exposure time on organic carbon preservation in continental margin sediments, Nature, 391, 572-575, 1998.

Hayes, C. T., Costa, K. M., Anderson, R. F., Calvo, E., Chase, Z., Demina, L. L., Dutay, J.-C., German, C. R., Heimbürger-Boavida, L.-E., Jaccard, S. L., Jacobel, A., Kohfeld, K. E., Kravchishina, M. D., Lippold, J., Mekik, F., Missiaen, L., Pavia, F. J., Paytan, A., Pedrosa-Pamies, R., Petrova, M. V., Rahman, S., Robinson, L. F., Roy-Barman, M., Sanchez-Vidal, A., Shiller, A., Tagliabue, A., Tessin, A. C., van Hulten, M., and Zhang, J.: Global Ocean Sediment Composition and Burial Flux in the Deep Sea, Glob. Biogeochem. Cycles, 35, e2020GB006769, https://doi.org/10.1029/2020GB006769, 2021.

Hayes, J. M. and Waldbauer, J. R.: The carbon cycle and associated redox processes through time, Phil. Trans. R. Soc. B, 361, 931-950, 10.1098/rstb.2006.1840, 2006.

Hedges, J. I., Hu, F. S., Devol, A. H., Hartnett, H. E., Tsamakis, E., and Keil, R. G.: Sedimentary organic matter preservation; a test for selective degradation under oxic conditions, Am. J. Sci., 299, 529-555, 10.2475/ajs.299.7-9.529, 1999.

Heinze, C., Kriest, I., and Maier-Reimer, E.: Age offsets among different biogenic and lithogenic components of sediment cores revealed by numerical modeling, Paleoceanography, 24, n/a-n/a, 10.1029/2008pa001662, 2009.

Henrichs, S. M. and Reeburgh, W. S.: Anaerobic mineralization of marine sediment organic matter: Rates and the role of anaerobic processes in the oceanic carbon economy, Geomicrobiology Journal, 5, 191-237, 10.1080/01490458709385971, 1987.

Hensen, C., Landenberger, H., Zabel, M., and Schulz, H. D.: Quantification of diffusive benthic fluxes of nitrate, phosphate, and silicate in the southern Atlantic Ocean, Glob. Biogeochem. Cycles, 12, 193-210, 10.1029/97gb02731, 1998.

Hitchcock, D. R. and Lovelock, J. E.: Life detection by atmospheric analysis, Icarus, 7, 149-159, http://dx.doi.org/10.1016/0019-1035(67)90059-0, 1967.

Holland, H. D.: The Chemistry of the Atmosphere and Oceans, John Wiley & Sons, New York1978.

Holser, W. T., Maynard, J. B., and Cruikshank, K. M.: Modelling the natural cycle of sulphur through Phanerozoic time, in: Evolution of the Global Biogeochemical Sulphur Cycle, edited by: Brimblecombe, P., and Lein, A. Y., John Wiley & Sons Ltd, New York, 21-56, 1989.

Honjo, S.: Material fluxes and modes of sedimentation in the mesopelagic and bathypelagic zones, Journal of Marine Research, 38, 53-97, 1980.

Honjo, S. and Manganini, S. J.: Annual biogenic particle fluxes to the interior of the North Atlantic Ocean; studied at 34°N 21°W and 48°N 21°W, Deep Sea Research Part II: Topical Studies in Oceanography, 40, 587-607, https://doi.org/10.1016/0967-0645(93)90034-K, 1993.

Hotinski, R. M., Kump, L. R., and Najjar, R. G.: Opening Pandora's Box: The impact of open system modeling on interpretations of anoxia, Paleoceanography, 15, 267-279, 10.1029/1999pa000408, 2000.

Hyacinthe, C., Anschutz, P., Carbonel, P., Jouanneau, J. M., and Jorissen, F. J.: Early diagenetic processes in the muddy sediments of the Bay of Biscay, Marine Geology, 177, 111-128, https://doi.org/10.1016/S0025-3227(01)00127-X, 2001.

Ingall, E. and Jahnke, R.: Evidence for enhanced phosphorus regeneration from marine sediments overlain by oxygen depleted waters, Geochim. Cosmochim. Acta, 58, 2571-2575, http://dx.doi.org/10.1016/0016-7037(94)90033-7, 1994.

Ingall, E. and Jahnke, R.: Influence of water-column anoxia on the elemental fractionation of carbon and phosphorus during sediment diagenesis, Marine Geology, 139, 219-229, https://doi.org/10.1016/S0025-3227(96)00112-0, 1997.

Ingall, E. D. and Cappellen, P. V.: Relation between sedimentation rate and burial of organic phosphorus and organic carbon in marine sediments, Geochim. Cosmochim. Acta, 54, 373-386, http://dx.doi.org/10.1016/0016-7037(90)90326-G, 1990.

Ingall, E. D., Bustin, R. M., and Van Cappellen, P.: Influence of water column anoxia on the burial and preservation of carbon and phosphorus in marine shales, Geochim. Cosmochim. Acta, 57, 303-316, https://doi.org/10.1016/0016-7037(93)90433-W, 1993.

Ittekkot, V.: The abiotically driven biological pump in the ocean and short-term fluctuations in atmospheric CO_2 contents, Global and Planetary Change, 8, 17-25, https://doi.org/10.1016/0921-8181(93)90060-2, 1993.

Jahnke, R. A.: The global ocean flux of particulate organic carbon: Areal distribution and magnitude, Glob. Biogeochem. Cycles, 10, 71-88, https://doi.org/10.1029/95GB03525, 1996.

Joos, F., Sarmiento, J. L., and Siegenthaler, U.: Estimates of the effect of Southern Ocean iron fertilization on atmospheric CO_2 concentrations, Nature, 349, 772-775, 10.1038/349772a0, 1991.

Jørgensen, B. B.: Mineralization of organic matter in the sea bed—the role of sulphate reduction, Nature, 296, 643, 10.1038/296643a0, 1982.

Jørgensen, B. B. and Kasten, S.: Sulfur cycling and methane oxidation, 2006.

Kagoshima, T., Sano, Y., Takahata, N., Maruoka, T., Fischer, T. P., and Hattori, K.: Sulphur geodynamic cycle, Sci Rep, 5, 8330, 10.1038/srep08330, 2015.

Karl, D., Michaels, A., Bergman, B., Capone, D., Carpenter, E., Letelier, R., Lipschultz, F., Paerl, H., Sigman, D., and Stal, L.: Dinitrogen fixation in the world's oceans, in: The Nitrogen Cycle at

Regional to Global Scales, edited by: Boyer, E. W., and Howarth, R. W., Springer Netherlands, Dordrecht, 47-98, 10.1007/978-94-017-3405-9_2, 2002.

Karl, D. M., Beversdorf, L., Björkman, K. M., Church, M. J., Martinez, A., and Delong, E. F.: Aerobic production of methane in the sea, Nat. Geosci., 1, 473-478, 10.1038/ngeo234, 2008.

Karthäuser, C., Ahmerkamp, S., Marchant, H. K., Bristow, L. A., Hauss, H., Iversen, M. H., Kiko, R., Maerz, J., Lavik, G., and Kuypers, M. M. M.: Small sinking particles control anammox rates in the Peruvian oxygen minimum zone, Nature Commun., 12, 3235, 10.1038/s41467-021-23340-4, 2021.

Kashiyama, Y., Ozaki, K., and Tajika, E.: Impact of the Evolution of Carbonate Ballasts on Marine Biogeochemistry in the Mesozoic and Associated Changes in Energy Delivery to Subsurface Waters, Paleontological Research, 15, 89-99, 10.2517/1342-8144-15.2.089, 2011.

Katsev, S. and Crowe, S. A.: Organic carbon burial efficiencies in sediments: The power law of mineralization revisited, Geology, 43, 607-610, 10.1130/g36626.1, 2015.

Key, R. M., Olsen, A., van Heuven, S., Lauvset, S. K., Velo, A., Lin, X., Schirnick, C., Kozyr, A., Tanhua, T., Hoppema, M., Jutterström, S., Steinfeldt, R., Jeansson, E., Ishii, M., Perez, F. F., and Suzuki, T.: Global Ocean Data Analysis Project, Version 2 (GLODAPv2), doi:10.3334/CDIAC/OTG. NDP093_GLODAPv2, 2015.

Kharecha, P., Kasting, J., and Siefert, J.: A coupled atmosphere–ecosystem model of the early Archean Earth, Geobiology, 3, 53-76, 10.1111/j.1472-4669.2005.00049.x, 2005.

Klaas, C. and Archer, D. E.: Association of sinking organic matter with various types of mineral ballast in the deep sea: Implications for the rain ratio, Glob. Biogeochem. Cycles, 16, 63-61-63-14, 10.1029/2001gb001765, 2002.

Knox, F. and McElroy, M. B.: Changes in atmospheric CO_2: Influence of the marine biota at high latitude, J. Geophys. Res., 89, 4629-4637, https://doi.org/10.1029/JD089iD03p04629, 1984.

Krissansen-Totton, J., Garland, R., Irwin, P., and Catling, D. C.: Detectability of Biosignatures in Anoxic Atmospheres with the James Webb Space Telescope: A TRAPPIST-1e Case Study, The Astronomical Journal, 156, 114, 10.3847/1538-3881/aad564, 2018.

Kump, L. R.: Chemical stability of the atmosphere and ocean, Palaeogeogr. Palaeoclimatol. Palaeoecol., 75, 123-136, http://dx.doi.org/10.1016/0031-0182(89)90187-9, 1989.

Kump, L. R.: The rise of atmospheric oxygen, Nature, 451, 277-278, 2008.

Kuypers, M. M. M., Lavik, G., Woebken, D., Schmid, M., Fuchs, B. M., Amann, R., Jørgensen, B. B., and Jetten, M. S. M.: Massive nitrogen loss from the Benguela upwelling system through anaerobic ammonium oxidation, Proc. Natl Acad. Sci. USA, 102, 6478-6483, 10.1073/pnas.0502088102, 2005.

Kuznetsov, I., Neumann, T., and Burchard, H.: Model study on the ecosystem impact of a variable C:N:P ratio for cyanobacteria in the Baltic Proper, Ecological Modelling, 219, 107-114, https://doi.org/10.1016/j.ecolmodel.2008.08.002, 2008.

Laakso, T. A. and Schrag, D. P.: Regulation of atmospheric oxygen during the Proterozoic, Earth Planet. Sci. Lett., 388, 81-91, http://dx.doi.org/10.1016/j.epsl.2013.11.049, 2014.

Larsson, U., Hajdu, S., Walve, J., and Elmgren, R.: Baltic Sea nitrogen fixation estimated from the summer increase in upper mixed layer total nitrogen, Limnology and Oceanography, 46, 811-820, https://doi.org/10.4319/lo.2001.46.4.0811, 2001.

Lasaga, A. C.: A new approach to isotopic modeling of the variation of atmospheric oxygen through the Phanerozoic, Am. J. Sci., 289, 411-435, 10.2475/ajs.289.4.411, 1989.

Lasaga, A. C. and Ohmoto, H.: The oxygen geochemical cycle: dynamics and stability, Geochim. Cosmochim. Acta, 66, 361-381, http://dx.doi.org/10.1016/S0016-7037(01)00685-8, 2002.

Laws, E. A., Falkowski, P. G., Smith, W. O., Ducklow, H., and McCarthy, J. J.: Temperature effects on export production in the open ocean, Glob. Biogeochem. Cycles, 14, 1231-1246, 10.1029/1999gb001229, 2000.

Ledwell, J. R., Watson, A. J., and Law, C. S.: Mixing of a tracer in the pycnocline, J. Geophys. Res., 103, 21499-21529, https://doi.org/10.1029/98JC01738, 1998.

Lenton, T. M.: Fire Feedbacks on Atmospheric Oxygen, in: Fire Phenomena and the Earth System, 289-308, 10.1002/9781118529539.ch15, 2013.

Lenton, T. M.: On the use of models in understanding the rise of complex life, Interface Focus, 10, 20200018, doi:10.1098/rsfs.2020.0018, 2020.

Lenton, T. M. and Watson, A. J.: Redfield revisited: 1. Regulation of nitrate, phosphate, and oxygen in the ocean, Glob. Biogeochem. Cycles, 14, 225-248, 10.1029/1999gb900065, 2000a.

Lenton, T. M. and Watson, A. J.: Redfield revisited: 2. What regulates the oxygen content of the atmosphere?, Glob. Biogeochem. Cycles, 14, 249-268, 10.1029/1999gb900076, 2000b.

Lenton, T. M., Daines, S. J., and Mills, B. J. W.: COPSE reloaded: An improved model of biogeochemical cycling over Phanerozoic time, Earth-Science Reviews, 178, 1-28, https://doi.org/10.1016/j.earscirev.2017.12.004, 2018.

Lenton, T. M., Dahl, T. W., Daines, S. J., Mills, B. J. W., Ozaki, K., Saltzman, M. R., and Porada, P.: Earliest land plants created modern levels of atmospheric oxygen, Proc. Natl Acad. Sci. USA, 113, 9704-9709, 10.1073/pnas.1604787113, 2016.

Lin, S. and Morse, J. W.: Sulfate reduction and iron sulfide mineral formation in Gulf of Mexico anoxic sediments, Am. J. Sci., 291, 55-89, 10.2475/ajs.291.1.55, 1991.

Liss, P. S. and Slater, P. G.: Flux of Gases across the Air-Sea Interface, Nature, 247, 181-184, 1974.

Lord, N. S., Ridgwell, A., Thorne, M. C., and Lunt, D. J.: An impulse response function for the "long tail" of excess atmospheric CO_2 in an Earth system model, Glob. Biogeochem. Cycles, 30, 2-17, https://doi.org/10.1002/2014GB005074, 2016.

Lovelock, J. E.: A Physical Basis for Life Detection Experiments, Nature, 207, 568-570, 1965.

Lovelock, J. E.: Gaia as seen through the atmosphere, Atmospheric Environment, 6, 579-580, https://doi.org/10.1016/0004-6981(72)90076-5, 1972.

Lovelock, J. E.: Thermodynamics and the recognition of alien biospheres, Proceedings of the Royal Society of London. Series B. Biological Sciences, 189, 167-181, doi:10.1098/rspb.1975.0051, 1975.

Lumpkin, R. and Speer, K.: Global Ocean Meridional Overturning, Journal of Physical Oceanography, 37, 2550-2562, 10.1175/jpo3130.1, 2007.

Luo, Y. W., Doney, S. C., Anderson, L. A., Benavides, M., Berman-Frank, I., Bode, A., Bonnet, S., Boström, K. H., Böttjer, D., Capone, D. G., Carpenter, E. J., Chen, Y. L., Church, M. J., Dore, J. E., Falcón, L. I., Fernández, A., Foster, R. A., Furuya, K., Gómez, F., Gundersen, K., Hynes, A. M., Karl, D. M., Kitajima, S., Langlois, R. J., LaRoche, J., Letelier, R. M., Marañón, E., McGillicuddy Jr, D. J., Moisander, P. H., Moore, C. M., Mouriño-Carballido, B., Mulholland, M. R., Needoba, J. A., Orcutt, K. M., Poulton, A. J., Rahav, E., Raimbault, P., Rees, A. P., Riemann, L., Shiozaki, T., Subramaniam, A., Tyrrell, T., Turk-Kubo, K. A., Varela, M., Villareal, T. A., Webb, E. A., White, A. E., Wu, J.,

and Zehr, J. P.: Database of diazotrophs in global ocean: abundance, biomass and nitrogen fixation rates, Earth Syst. Sci. Data, 4, 47-73, 10.5194/essd-4-47-2012, 2012.

Lutz, M., Dunbar, R., and Caldeira, K.: Regional variability in the vertical flux of particulate organic carbon in the ocean interior, Glob. Biogeochem. Cycles, 16, 11-11-11-18, 10.1029/2000gb001383, 2002.

Lyons, T. W. and Gill, B. C.: Ancient Sulfur Cycling and Oxygenation of the Early Biosphere, Elements, 6, 93-99, 10.2113/gselements.6.2.93, 2010.

Lyons, T. W., Reinhard, C. T., and Planavsky, N. J.: The rise of oxygen in Earth's early ocean and atmosphere, Nature, 506, 307-315, 10.1038/nature13068, 2014.

Mackenzie, F. T., Ver, L. M., Sabine, C., Lane, M., and Lerman, A.: C, N, P, S Global Biogeochemical Cycles and Modeling of Global Change, in: Interactions of C, N, P and S Biogeochemical Cycles and Global Change, edited by: Wollast, R., Mackenzie, F. T., and Chou, L., Springer Berlin Heidelberg, Berlin, Heidelberg, 1-61, 1993.

Maier-Reimer, E.: Geochemical cycles in an ocean general circulation model. Preindustrial tracer distributions, Glob. Biogeochem. Cycles, 7, 645-677, 10.1029/93gb01355, 1993.

Markovic, S., Paytan, A., and Wortmann, U. G.: Pleistocene sediment offloading and the global sulfur cycle, Biogeosciences, 12, 3043-3060, 10.5194/bg-12-3043-2015, 2015.

Martin, J. H., Knauer, G. A., Karl, D. M., and Broenkow, W. W.: VERTEX: carbon cycling in the northeast Pacific, Deep Sea Research Part A. Oceanographic Research Papers, 34, 267-285, https://doi.org/10.1016/0198-0149(87)90086-0, 1987.

Martin, W. R. and Sayles, F. L.: The Recycling of Biogenic Material at the Sea Floor, in: Treatise on Geochemistry (Second Edition), edited by: Turekian, K. K., Elsevier, Oxford, 33-59, https://doi.org/10.1016/B978-0-08-095975-7.00702-6, 2014.

Mayor, M. and Queloz, D.: A Jupiter-mass companion to a solar-type star, Nature, 378, 355-359, 10.1038/378355a0, 1995.

McManus, J., Berelson, W. M., Coale, K. H., Johnson, K. S., and Kilgore, T. E.: Phosphorus regeneration in continental margin sediments, Geochim. Cosmochim. Acta, 61, 2891-2907, http://dx.doi.org/10.1016/S0016-7037(97)00138-5, 1997.

McManus, J., Berelson, W. M., Klinkhammer, G. P., Hammond, D. E., and Holm, C.: Authigenic uranium: Relationship to oxygen penetration depth and organic carbon rain, Geochim. Cosmochim. Acta, 69, 95-108, https://doi.org/10.1016/j.gca.2004.06.023, 2005.

Meadows, V. S.: Reflections on O_2 as a Biosignature in Exoplanetary Atmospheres, Astrobiology, 17, 1022-1052, 10.1089/ast.2016.1578, 2017.

Meadows, V. S., Reinhard, C. T., Arney, G. N., Parenteau, M. N., Schwieterman, E. W., Domagal-Goldman, S. D., Lincowski, A. P., Stapelfeldt, K. R., Rauer, H., DasSarma, S., Hegde, S., Narita, N., Deitrick, R., Lustig-Yaeger, J., Lyons, T. W., Siegler, N., and Grenfell, J. L.: Exoplanet Biosignatures: Understanding Oxygen as a Biosignature in the Context of Its Environment, Astrobiology, 18, 630-662, 10.1089/ast.2017.1727, 2018.

Middelburg, J. J., Soetaert, K., and Herman, P. M. J.: Empirical relationships for use in global diagenetic models, Deep Sea Research Part I: Oceanographic Research Papers, 44, 327-344, https://doi.org/10.1016/S0967-0637(96)00101-X, 1997.

Middelburg, J. J., Soetaert, K., Herman, P. M. J., and Heip, C. H. R.: Denitrification in marine sediments: A model study, Glob. Biogeochem. Cycles, 10, 661-673, 10.1029/96gb02562, 1996.

Millero, F. J.: The oxidation of H_2S in Black Sea waters, Deep Sea Research Part A. Oceanographic Research Papers, 38, S1139-S1150, https://doi.org/10.1016/S0198-0149(10)80028-7, 1991.

Millero, F. J.: Chemical Oceanography, 3rd edn., Taylor & Francis Group CRC Press, Boca Raton, 496 pp.2006.

Millero, F. J., Plese, T., and Fernandez, M.: The dissociation of hydrogen-sulfide in seawater, Limnology and Oceanography, 33, 269-274, 1988.

Morford, J. L. and Emerson, S.: The geochemistry of redox sensitive trace metals in sediments, Geochim. Cosmochim. Acta, 63, 1735-1750, https://doi.org/10.1016/S0016-7037(99)00126-X, 1999.

Muller-Karger, F. E., Varela, R., Thunell, R., Luerssen, R., Hu, C., and Walsh, J. J.: The importance of continental margins in the global carbon cycle, Geophys. Res. Lett., 32, 10.1029/2004gl021346, 2005.

National Academies of Sciences, E. and Medicine: An Astrobiology Strategy for the Search for Life in the Universe, The National Academies Press, Washington, DC, 188 pp., doi:10.17226/25252, 2019.

Nierop, K. G. J., Reichart, G.-J., Veld, H., and Sinninghe Damsté, J. S.: The influence of oxygen exposure time on the composition of macromolecular organic matter as revealed by surface sediments on the Murray Ridge (Arabian Sea), Geochim. Cosmochim. Acta, 206, 40-56, 10.1016/j.gca.2017.02.032, 2017.

Oguz, T., Ducklow, H. W., and Malanotte-Rizzoli, P.: Modeling distinct vertical biogeochemical structure of the Black Sea: Dynamical coupling of the oxic, suboxic, and anoxic layers, Glob. Biogeochem. Cycles, 14, 1331-1352, https://doi.org/10.1029/1999GB001253, 2000.

Oguz, T., Murray, J. W., and Callahan, A. E.: Modeling redox cycling across the suboxic–anoxic interface zone in the Black Sea, Deep Sea Research Part I: Oceanographic Research Papers, 48, 761-787, https://doi.org/10.1016/S0967-0637(00)00054-6, 2001.

Olsen, A., Key, R. M., van Heuven, S., Lauvset, S. K., Velo, A., Lin, X., Schirnick, C., Kozyr, A., Tanhua, T., Hoppema, M., Jutterström, S., Steinfeldt, R., Jeansson, E., Ishii, M., Pérez, F. F., and Suzuki, T.: The Global Ocean Data Analysis Project version 2 (GLODAPv2) – an internally consistent data product for the world ocean, Earth Syst. Sci. Data, 8, 297-323, 10.5194/essd-8-297-2016, 2016.

Olson, S. L., Reinhard, C. T., and Lyons, T. W.: Limited role for methane in the mid-Proterozoic greenhouse, Proc. Natl Acad. Sci. USA, 113, 11447-11452, 10.1073/pnas.1608549113, 2016.

Oschlies, A., Schulz, K. G., Riebesell, U., and Schmittner, A.: Simulated 21st century's increase in oceanic suboxia by CO_2-enhanced biotic carbon export, Glob. Biogeochem. Cycles, 22, GB4008, 10.1029/2007gb003147, 2008.

Ozaki, K. and Reinhard, C. T.: The future lifespan of Earth's oxygenated atmosphere, Nat. Geosci., 14, 138-142, 10.1038/s41561-021-00693-5, 2021.

Ozaki, K. and Tajika, E.: Biogeochemical effects of atmospheric oxygen concentration, phosphorus weathering, and sea-level stand on oceanic redox chemistry: Implications for greenhouse climates, Earth Planet. Sci. Lett., 373, 129-139, http://dx.doi.org/10.1016/j.epsl.2013.04.029, 2013.

Ozaki, K., Reinhard, C. T., and Tajika, E.: A sluggish mid-Proterozoic biosphere and its effect on Earth's redox balance, Geobiology, 17, 3-11, doi:10.1111/gbi.12317, 2019a.

Ozaki, K., Tajima, S., and Tajika, E.: Conditions required for oceanic anoxia/euxinia: Constraints from a one-dimensional ocean biogeochemical cycle model, Earth Planet. Sci. Lett., 304, 270-279, http://dx.doi.org/10.1016/j.epsl.2011.02.011, 2011.

Ozaki, K., Tajika, E., Hong, P. K., Nakagawa, Y., and Reinhard, C. T.: Effects of primitive photosynthesis on Earth's early climate system, Nat. Geosci., 11, 55-59, 10.1038/s41561-017-0031-2, 2018.

Ozaki, K., Thompson, K. J., Simister, R. L., Crowe, S. A., and Reinhard, C. T.: Anoxygenic photosynthesis and the delayed oxygenation of Earth's atmosphere, Nature Commun., 10, 3026, 10.1038/s41467-019-10872-z, 2019b.

Pallud, C. and Van Cappellen, P.: Kinetics of microbial sulfate reduction in estuarine sediments, Geochim. Cosmochim. Acta, 70, 1148-1162, https://doi.org/10.1016/j.gca.2005.11.002, 2006.

Papadomanolaki, N. M., Lenstra, W. K., Wolthers, M., and Slomp, C. P.: Enhanced phosphorus recycling during past oceanic anoxia amplified by low rates of apatite authigenesis, Science Advances, 8, eabn2370, 10.1126/sciadv.abn2370,

Petsch, S. T. and Berner, R. A.: Coupling the geochemical cycles of C, P, Fe, and S; the effect on atmospheric O_2 and the isotopic records of carbon and sulfur, Am. J. Sci., 298, 246-262, 10.2475/ajs.298.3.246, 1998.

Petsch, S. T., Eglinton, T. I., and Edwards, K. J.: ^{14}C-Dead Living Biomass: Evidence for Microbial Assimilation of Ancient Organic Carbon During Shale Weathering, Science, 292, 1127-1131, doi:10.1126/science.1058332, 2001.

Pfeifer, K., Hensen, C., Adler, M., Wenzhfer, F., Weber, B., and Schulz, H. D.: Modeling of subsurface calcite dissolution, including the respiration and reoxidation processes of marine sediments in the region of equatorial upwelling off Gabon, Geochim. Cosmochim. Acta, 66, 4247-4259, https://doi.org/10.1016/S0016-7037(02)01073-6, 2002.

Planavsky, N. J., Cole, D. B., Isson, T. T., Reinhard, C. T., Crockford, P. W., Sheldon, N. D., and Lyons, T. W.: A case for low atmospheric oxygen levels during Earth's middle history, Emerging Topics in Life Sciences, 2, 149-159, 10.1042/etls20170161, 2018.

Planavsky, N. J., Cole, D. B., Reinhard, C. T., Diamond, C., Love, G. D., Luo, G., Zhang, S., Konhauser, K. O., and Lyons, T. W.: No evidence for high atmospheric oxygen levels 1,400 million years ago, Proc. Natl Acad. Sci. USA, 113, E2550-E2551, 10.1073/pnas.1601925113, 2016.

Prentice, I. C., Farquhar, G. D., Fasham, M. J. R., Goulden, M. L., Heimann, M., Jaramillo, V. J., Kheshgi, H. S., Le Quere, C., Scholes, R. J., and Wallace, D. W. R.: The carbon cycle and atmospheric carbon dioxide, in: Climate Change 2001: the Scientific Basis, edited by: Houghton, J. T., Ding, Y., Griggs, D. J., Noguer, N., van der Linden, P. J., Xiaosu, D., Maskell, K., and Johnson, C. A., Cambridge University Press, New York, 2001.

Quigg, A., Finkel, Z. V., Irwin, A. J., Rosenthal, Y., Ho, T.-Y., Reinfelder, J. R., Schofield, O., Morel, F. M. M., and Falkowski, P. G.: The evolutionary inheritance of elemental stoichiometry in marine phytoplankton, Nature, 425, 291-294, 10.1038/nature01953, 2003.

Raiswell, R. and Canfield, D. E.: The Iron Biogeochemical Cycle Past and Present, Geochemical Perspectives, 1, 1-2, 2012.

Raynaud, D., Jouzel, J., Barnola, J. M., Chappellaz, J., Delmas, R. J., and Lorius, C.: The Ice Record of Greenhouse Gases, Science, 259, 926-934, doi:10.1126/science.259.5097.926, 1993.

Redfield, A. C., Ketchum, B. H., and Richards, F. A.: The influence of organisms on the composition of sea-water, in: The Sea, edited by: Hill, M. N., Interscience Publishers, New York, 26-77, 1963.

Reimers, C. E., Jahnke, R. A., and McCorkle, D. C.: Carbon fluxes and burial rates over the continental slope and rise off central California with implications for the global carbon cycle, Glob. Biogeochem. Cycles, 6, 199-224, 10.1029/92gb00105, 1992.

Reinhard, C. T., Olson, S. L., Schwieterman, E. W., and Lyons, T. W.: False Negatives for Remote Life Detection on Ocean-Bearing Planets: Lessons from the Early Earth, Astrobiology, 17, 287-297, 10.1089/ast.2016.1598, 2017a.

Reinhard, C. T., Olson, S. L., Kirtland Turner, S., Pälike, C., Kanzaki, Y., and Ridgwell, A.: Oceanic and atmospheric methane cycling in the cGENIE Earth system model – release v0.9.14, Geosci. Model Dev., 13, 5687-5706, 10.5194/gmd-13-5687-2020, 2020.

Reinhard, C. T., Planavsky, N. J., Gill, B. C., Ozaki, K., Robbins, L. J., Lyons, T. W., Fischer, W. W., Wang, C., Cole, D. B., and Konhauser, K. O.: Evolution of the global phosphorus cycle, Nature, 541, 386-389, 10.1038/nature20772 http://www.nature.com/nature/journal/v541/n7637/abs/nature20772.html#supplementary-information, 2017b.

Ridgwell, A. and Hargreaves, J. C.: Regulation of atmospheric CO_2 by deep-sea sediments in an Earth system model, Glob. Biogeochem. Cycles, 21, GB2008, 10.1029/2006gb002764, 2007.

Romaniello, S. J. and Derry, L. A.: An intermediate-complexity model for simulating marine biogeochemistry in deep time: Validation against the modern global ocean, Geochem. Geophys. Geosyst., 11, Q08001, 10.1029/2009gc002711, 2010.

Rowe, G. T., Morse, J., Nunnally, C., and Boland, G. S.: Sediment community oxygen consumption in the deep Gulf of Mexico, Deep Sea Research Part II: Topical Studies in Oceanography, 55, 2686-2691, https://doi.org/10.1016/j.dsr2.2008.07.018, 2008.

Ruttenberg, K. C.: Reassessment of the oceanic residence time of phosphorus, Chem. Geol., 107, 405-409, http://dx.doi.org/10.1016/0009-2541(93)90220-D, 1993.

Ruttenberg, K. C.: The Global Phosphorus Cycle, in: Treatise on Geochemistry, edited by: Turekian, K. K., Pergamon, Oxford, 585-643, https://doi.org/10.1016/B0-08-043751-6/08153-6, 2003.

Sachs, O., Sauter, E. J., Schlüter, M., Rutgers van der Loeff, M. M., Jerosch, K., and Holby, O.: Benthic organic carbon flux and oxygen penetration reflect different plankton provinces in the Southern Ocean, Deep Sea Research Part I: Oceanographic Research Papers, 56, 1319-1335, 10.1016/j.dsr.2009.02.003, 2009.

Sagan, C., Thompson, W. R., Carlson, R., Gurnett, D., and Hord, C.: A search for life on Earth from the Galileo spacecraft, Nature, 365, 715-721, 1993.

Sarmiento, J. L. and Gruber, N.: Ocean biogeochemical dynamics, Princeton University Press2006.

Sarmiento, J. L. and Toggweiler, J. R.: A new model for the role of the oceans in determining atmospheric P_{CO2}, Nature, 308, 621-624, 10.1038/308621a0, 1984.

Schenau, S. J. and De Lange, G. J.: Phosphorus regeneration vs. burial in sediments of the Arabian Sea, Marine Chemistry, 75, 201-217, http://dx.doi.org/10.1016/S0304-4203(01)00037-8, 2001.

Schlesinger, W. H. and Bernhardt, E. S.: The Global Cycles of Sulfur and Mercury, in: Biogeochemistry (Third Edition), Academic Press, Boston, 469-486, https://doi.org/10.1016/B978-0-12-385874-0.00013-3, 2013.

Schwieterman, E. W., Kiang, N. Y., Parenteau, M. N., Harman, C. E., DasSarma, S., Fischer, T. M., Arney, G. N., Hartnett, H. E., Reinhard, C. T., Olson, S. L., Meadows, V. S., Cockell, C. S., Walker, S. I.,

Grenfell, J. L., Hegde, S., Rugheimer, S., Hu, R., and Lyons, T. W.: Exoplanet Biosignatures: A Review of Remotely Detectable Signs of Life, Astrobiology, 18, 663-708, 10.1089/ast.2017.1729, 2018.

Shaffer, G.: Phosphate pumps and shuttles in the Black Sea, Nature, 321, 515-517, 10.1038/321515a0, 1986.

Shaffer, G. and Sarmiento, J. L.: Biogeochemical cycling in the global ocean: 1. A new, analytical model with continuous vertical resolution and high-latitude dynamics, J. Geophys. Res., 100, 2659-2672, https://doi.org/10.1029/94JC01167, 1995.

Shaffer, G., Malskær Olsen, S., and Pepke Pedersen, J. O.: Presentation, calibration and validation of the low-order, DCESS Earth System Model (Version 1), Geosci. Model Dev., 1, 17-51, 10.5194/gmd-1-17-2008, 2008.

Sharoni, S. and Halevy, I.: Geologic controls on phytoplankton elemental composition, Proc. Natl Acad. Sci. USA, 119, e2113263118, 10.1073/pnas.2113263118, 2022.

Siegenthaler, U. and Wenk, T.: Rapid atmospheric CO_2 variations and ocean circulation, Nature, 308, 624-626, 10.1038/308624a0, 1984.

Sleep, N. H.: Dioxygen over geological time, in: Metal ions in biological systems, edited by: Sigel, A., Sigel, H., and Sigel, R. K. O., Taylor & Francis Group, Boca Raton, 49-73, 2005.

Slomp, C. P. and Van Cappellen, P.: The global marine phosphorus cycle: sensitivity to oceanic circulation, Biogeosciences, 4, 155-171, 10.5194/bg-4-155-2007, 2007.

Slomp, C. P., Thomson, J., and de Lange, G. J.: Enhanced regeneration of phosphorus during formation of the most recent eastern Mediterranean sapropel (S1), Geochim. Cosmochim. Acta, 66, 1171-1184, http://dx.doi.org/10.1016/S0016-7037(01)00848-1, 2002.

Sloyan, B. M.: Spatial variability of mixing in the Southern Ocean, Geophys. Res. Lett., 32, L18603, 10.1029/2005gl023568, 2005.

Soulet, G., Hilton, R. G., Garnett, M. H., Roylands, T., Klotz, S., Croissant, T., Dellinger, M., and Le Bouteiller, C.: Temperature control on CO_2 emissions from the weathering of sedimentary rocks, Nat. Geosci., 14, 665-671, 10.1038/s41561-021-00805-1, 2021.

Southam, J. R., Peterson, W. H., and Brass, G. W.: Dynamics of anoxia, Palaeogeogr. Palaeoclimatol. Palaeoecol., 40, 183-198, https://doi.org/10.1016/0031-0182(82)90089-X, 1982.

Steefel, C. I. and MacQuarrie, K. T. B.: Approaches to modeling of reactive transport in porous media, Reviews in Mineralogy and Geochemistry, 34, 85-129, 1996.

Suess, E.: Particulate organic carbon flux in the oceans—surface productivity and oxygen utilization, Nature, 288, 260-263, 10.1038/288260a0, 1980.

Tang, D., Shi, X., Wang, X., and Jiang, G.: Extremely low oxygen concentration in mid-Proterozoic shallow seawaters, Precambrian Res., 276, 145-157, http://dx.doi.org/10.1016/j.precamres.2016.02.005, 2016.

Tarhan, L. G., Droser, M. L., Planavsky, N. J., and Johnston, D. T.: Protracted development of bioturbation through the early Palaeozoic Era, Nat. Geosci., 8, 865, 10.1038/ngeo2537 https://www.nature.com/articles/ngeo2537#supplementary-information, 2015.

Tarpgaard, I. H., Røy, H., and Jørgensen, B. B.: Concurrent low- and high-affinity sulfate reduction kinetics in marine sediment, Geochim. Cosmochim. Acta, 75, 2997-3010, http://dx.doi.org/10.1016/j.gca.2011.03.028, 2011.

The LUVOIR Team: Mission Concept Study Final Report. In: arXiv e-prints, 2019.

Tostevin, R., Turchyn, A. V., Farquhar, J., Johnston, D. T., Eldridge, D. L., Bishop, J. K. B., and McIlvin, M.: Multiple sulfur isotope constraints on the modern sulfur cycle, Earth Planet. Sci. Lett., 396, 14-21, https://doi.org/10.1016/j.epsl.2014.03.057, 2014.

Tromp, T. K., Van Cappellen, P., and Key, R. M.: A global model for the early diagenesis of organic carbon and organic phosphorus in marine sediments, Geochim. Cosmochim. Acta, 59, 1259-1284, http://dx.doi.org/10.1016/0016-7037(95)00042-X, 1995.

Tsunogai, S. and Noriki, S.: Particulate fluxes of carbonate and organic carbon in the ocean. Is the marine biological activity working as a sink of the atmospheric carbon?, Tellus B: Chemical and Physical Meteorology, 43, 265-266, 10.3402/tellusb.v43i2.15272, 1991.

Turchyn, A. V. and Schrag, D. P.: Oxygen Isotope Constraints on the Sulfur Cycle over the Past 10 Million Years, Science, 303, 2004-2007, 10.1126/science.1092296, 2004.

Turchyn, A. V. and Schrag, D. P.: Cenozoic evolution of the sulfur cycle: Insight from oxygen isotopes in marine sulfate, Earth Planet. Sci. Lett., 241, 763-779, https://doi.org/10.1016/j.epsl.2005.11.007, 2006.

Turnewitsch, R. and Pohl, C.: An estimate of the efficiency of the iron- and manganese-driven dissolved inorganic phosphorus trap at an oxic/euxinic water column redoxcline, Glob. Biogeochem. Cycles, 24, GB4025, 10.1029/2010gb003820, 2010.

Tyrrell, T.: The relative influences of nitrogen and phosphorus on oceanic primary production, Nature, 400, 525-531, 10.1038/22941, 1999.

Van Cappellen, P. and Ingall, E. D.: Benthic phosphorus regeneration, net primary production, and ocean anoxia: A model of the coupled marine biogeochemical cycles of carbon and phosphorus, Paleoceanography, 9, 677-692, https://doi.org/10.1029/94PA01455, 1994.

Van Cappellen, P. and Ingall, E. D.: Redox Stabilization of the Atmosphere and Oceans by Phosphorus-Limited Marine Productivity, Science, 271, 493-496, 10.1126/science.271.5248.493, 1996.

Van Cappellen, P. and Wang, Y.: Cycling of iron and manganese in surface sediments; a general theory for the coupled transport and reaction of carbon, oxygen, nitrogen, sulfur, iron, and manganese, Am. J. Sci., 296, 197-243, 10.2475/ajs.296.3.197, 1996.

van de Velde, S. J., Hülse, D., Reinhard, C. T., and Ridgwell, A.: Iron and sulfur cycling in the cGENIE.muffin Earth system model (v0.9.21), Geosci. Model Dev., 14, 2713-2745, 10.5194/gmd-14-2713-2021, 2021.

Volk, T. and Hoffert, M. I.: Ocean carbon pumps: Analysis of relative strengths and efficiencies in ocean-driven atmospheric CO_2 changes, in: The Carbon Cycle and Atmospheric CO_2: Natural Variations Archean to Present, 99-110, https://doi.org/10.1029/GM032p0099, 1985.

Walker, J. C. G.: Evolution of the atmosphere, Macmillan, New York, 318 pp.1977.

Walker, J. C. G. and Brimblecombe, P.: Iron and sulfur in the pre-biologic ocean, Precambrian Res., 28, 205-222, https://doi.org/10.1016/0301-9268(85)90031-2, 1985.

Wallmann, K.: Feedbacks between oceanic redox states and marine productivity: A model perspective focused on benthic phosphorus cycling, Glob. Biogeochem. Cycles, 17, 1084, 10.1029/2002gb001968, 2003.

Wallmann, K.: Phosphorus imbalance in the global ocean?, Glob. Biogeochem. Cycles, 24, GB4030, 10.1029/2009gb003643, 2010.

Wang, W.-L., Moore, J. K., Martiny, A. C., and Primeau, F. W.: Convergent estimates of marine nitrogen fixation, Nature, 566, 205-211, 10.1038/s41586-019-0911-2, 2019.

WebBook, N. C.: NIST Chemistry WebBook, https://doi.org/10.18434/T4D303, 2022.

Westrich, J. T. and Berner, R. A.: The role of sedimentary organic matter in bacterial sulfate reduction: The G model tested, Limnology and Oceanography, 29, 236-249, 10.4319/lo.1984.29.2.0236, 1984.

Wheat, C. G., Feely, R. A., and Mottl, M. J.: Phosphate removal by oceanic hydrothermal processes: An update of the phosphorus budget in the oceans, Geochim. Cosmochim. Acta, 60, 3593-3608, https://doi.org/10.1016/0016-7037(96)00189-5, 1996.

Wheat, C. G., McManus, J., Mottl, M. J., and Giambalvo, E.: Oceanic phosphorus imbalance: Magnitude of the mid-ocean ridge flank hydrothermal sink, Geophys. Res. Lett., 30, https://doi.org/10.1029/2003GL017318, 2003.

Woodward, F. I.: Global primary production, Current Biology, 17, R269-R273, 10.1016/j.cub.2007.01.054, 2007.

Wortmann, U. G. and Paytan, A.: Rapid Variability of Seawater Chemistry Over the Past 130 Million Years, Science, 337, 334-336, 10.1126/science.1220656, 2012.

Yakushev, E. V. and Neretin, L. N.: One-dimensional modeling of nitrogen and sulfur cycles in the aphotic zones of the Black and Arabian Seas, Glob. Biogeochem. Cycles, 11, 401-414, https://doi.org/10.1029/97GB00782, 1997.

Yakushev, E. V., Pollehne, F., Jost, G., Kuznetsov, I., Schneider, B., and Umlauf, L.: Analysis of the water column oxic/anoxic interface in the Black and Baltic seas with a numerical model, Marine Chemistry, 107, 388-410, 10.1016/j.marchem.2007.06.003, 2007.

Yamanaka, Y. and Tajika, E.: The role of the vertical fluxes of particulate organic matter and calcite in the oceanic carbon cycle: Studies using an ocean biogeochemical general circulation model, Glob. Biogeochem. Cycles, 10, 361-382, 10.1029/96gb00634, 1996.

Yao, W. and Millero, F.: The chemistry of the anoxic waters in the Framvaren Fjord, Norway, Aquatic Geochemistry, 1, 53-88, 10.1007/bf01025231, 1995.

Yaroshevsky, A. A.: Abundances of chemical elements in the Earth's crust, Geochem. Int., 44, 48-55, 10.1134/s001670290601006x, 2006.

Zabel, M., Dahmke, A., and Schulz, H. D.: Regional distribution of diffusive phosphate and silicate fluxes through the sediment–water interface: the eastern South Atlantic, Deep Sea Research Part I: Oceanographic Research Papers, 45, 277-300, http://dx.doi.org/10.1016/S0967-0637(97)00073-3, 1998.

Zhang, S., Wang, X., Wang, H., Bjerrum, C. J., Hammarlund, E. U., Dahl, T. W., and Canfield, D. E.: Reply to Planavsky et al.: Strong evidence for high atmospheric oxygen levels 1,400 million years ago, Proc. Natl Acad. Sci. USA, 113, E2552-E2553, 10.1073/pnas.1603982113, 2016.

Zhao, M., Zhang, S., Tarhan, L. G., Reinhard, C. T., and Planavsky, N.: The role of calcium in regulating marine phosphorus burial and atmospheric oxygenation, Nature Commun., 11, 2232, 10.1038/s41467-020-15673-3, 2020.