

Representation of phosphorus cycle in Joint UK Land Environment Simulator (vn5.5_JULES-CNP)

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Abstract

Most Land Surface Models (LSMs), the land components of Earth system models (ESMs), include representation of nitrogen (N) limitation on ecosystem productivity. However only few of these models have incorporated phosphorus (P) cycling. In tropical ecosystems, this is likely to be important as N tends to be abundant but the availability of rock-derived elements, such as P, can be very low. Thus, without a representation of P cycling, tropical forest response in areas such as Amazonia to rising atmospheric CO₂ conditions remains highly uncertain. In this study, we introduced P dynamics and its interactions with the N and carbon (C) cycles into the Joint UK Land Environment Simulator (JULES). The new model (JULES-CNP) includes the representation of P stocks in vegetation and soil pools, as well as key processes controlling fluxes between these pools. We develop and evaluate JULES-CNP using in situ data collected at a low fertility site in the Central Amazon, with a soil P content representative of 60% of soils across the Amazon basin, to parameterise, calibrate and evaluate JULES-CNP. Novel soil and plant P pool observations are used for parameterisation and calibration and the model is evaluated against C fluxes and stocks, and for those soil P pools not used for parameterisation/calibration. We then evaluate the model [at additional P limited test sites across the Amazon, in Panama and Hawaii showing a significant improvement over the C and CN only versions of the model](#). The model is then applied under elevated CO₂ (600 ppm) at our study site Central Amazon to quantify the impact of P limitation on CO₂ fertilization. We compare our results against current state of the art CNP models using the same methodology that was used in the AmazonFACE model intercomparison study. The model is able to reproduce the observed plant and soil P pools and fluxes used for evaluation under ambient CO₂. We estimate P to limit net primary productivity (NPP) by 24% under current CO₂ and by 46% under elevated CO₂. Under elevated CO₂, biomass in simulations accounting for CNP increase by 10% relative [to contemporary CO₂ conditions](#), although it is 5% lower compared with CN and C-only simulations. Our results highlight the potential for high P limitation and therefore lower CO₂ fertilization capacity in the Amazon forest with low fertility soils.

1. Introduction

Land ecosystems currently take up about 30% of anthropogenic CO₂ emissions (Friedlingstein *et al.*, 2020), thus buffering the anthropogenic increase in atmospheric CO₂. Tropical forests play a major role in the land C cycle, account for about half of global net primary production (NPP)(Schimel *et al.*, 2015), and store the highest above ground carbon among all biomes (Pan *et al.*, 2011; Mitchard, 2018).

The C sink capacity of tropical forests may be constrained by nutrient availability for plant photosynthesis and growth (Vitousek and Howarth, 1991; Elser *et al.*, 2007; LeBauer and Treseder, 2008) via P (Nordin, Högberg and Näsholm, 2001; Shen *et al.*, 2011) and/or N related processes (DeLuca, Keeney and McCarty, 1992; Perakis and Hedin, 2002). Global process-based models of vegetation dynamics and function suggest a continued land C sink in the tropical forests, largely attributed to the CO₂ fertilization effect (Sitch *et al.*, 2008; Schimel, Stephens and Fisher, 2015; Koch, Hubau and Lewis, 2021). However, many of these models typically do not consider P constraints on plant growth (Fleischer *et al.*, 2019), which is likely to be an important limiting nutrient in tropical ecosystems, characterised by old and heavily weathered soils. The importance of nutrient cycling representation in Earth System Models (ESMs), and the lack thereof, was highlighted by Hungate *et al.* (2003) and Zaehle and Dalmonech (2011), showing the significance of nitrogen inclusion in ESMs for generating more realistic estimations of the future evolution of the terrestrial C sink. However, in the Coupled Climate C Cycle Model Inter-comparison Project (C4MIP), none of the participating ESMs included N dynamics (Friedlingstein *et al.*, 2006). Seven years later, for the update in CMIP5 (Anav *et al.*, 2013), three models out of eighteen with N dynamics were included (Bentsen *et al.*, 2013; Long *et al.*, 2013; Ji *et al.*, 2014). Although much progress has been made in the inclusion of an N cycle in ESMs so far, none of the CMIP5 models included P cycling and in the most recent CMIP6, only one model includes P (ACCESS-ESM1.5 model) (Arora *et al.*, 2020).

The long history of soil development in tropical regions which involves the loss of rock-derived nutrients through weathering and leaching on geologic timescales (Vitousek *et al.*, 1997, 2010) results in highly weathered soils. Soil P is hypothesized to be among the key limiting nutrients to plant growth in tropical forests (Vitousek *et al.*, 1997, 2010; Hou *et al.*, 2020), unlike temperate forest where N is hypothesised to be the main constraint (Aerts and Chapin, 1999; Luo *et al.*, 2004). Low P availability in tropical soils is related to the limited un-weathered parent material or organic compounds as source of P (Walker and Syers, 1976), active sorption (Sanchez, 1977) and high occlusion (Yang and Post, 2011) which further reduce plant available P. Although N limitation can impact the terrestrial C sink response to increasing atmospheric CO₂ by changing plant C fixation capacity (Luo *et al.*, 2004), this can be partially ameliorated over time by input of N into the biosphere via the continuous inputs of N into ecosystems from atmospheric deposition and biological N fixation (Vitousek *et al.*, 2010). P-limitation is pervasive in natural ecosystems (Hou *et al.*, 2020) and the lack of large P inputs into ecosystems, especially those growing on highly weathered soil, may make P limitation a stronger constraint on ecosystem response to elevated CO₂ (eCO₂) than N (Gentile *et al.*, 2012; Sardans, Rivas-Ubach and Peñuelas, 2012). This causes considerable uncertainty in predicting the future of the Amazon forest C sink (Yang *et al.*, 2014).

There is evidence to suggest P limitation on plant productivity in the Amazon forest (Malhi, 2012) where it has been shown that the younger, more fertile west and south-west Amazon soils have higher tree turnover (Phillips *et al.*, 2004; Stephenson and Van Mantgem, 2005) and stem growth rates (Malhi *et al.*, 2004) and lower above ground biomass (Baker *et al.*, 2004; Malhi *et al.*, 2006) compared to their central and eastern counterparts. Total soil P has been found as the best predictor of stem growth (Quesada *et al.*, 2010) and of total NPP (Aragão *et al.*, 2009) across this fertility gradient, and foliar P is positively related to plant photosynthetic capacity (V_{cmax} and J_{cmax}) in these forests (Mercado *et al.*, 2011).

However, modelling studies are unable to reproduce observed spatial patterns of NPP and biomass in the Amazon, [one possible reason being](#) the lack of inclusion of soil P constraints on plant productivity and function (Wang, Law and Pak, 2010; Vicca *et al.*, 2012a; Yang *et al.*, 2014). Nevertheless, some modelling [studies](#) have focused on improving process and parameter representation using the observational data of spatial variation in woody biomass residence time (Johnson *et al.*, 2016), soil texture and soil P to parameterise the maximum carboxylation capacity (V_{cmax}) (Castanho *et al.*, 2013). Results from these studies successfully represent observed patterns of Amazon forest biomass growth increases with increasing soil fertility. However, the full representation of these interactions and the impact of the soil nutrient availability on biomass productivity is still missing in most of ESMs.

So far, several dynamic global vegetation models have been developed to represent P cycling within the soil (Yang *et al.*, 2013; Haverd *et al.*, 2018) and between plant and soils for tropical forests particularly (Yang *et al.*,

2014; Zhu *et al.*, 2016; Goll *et al.*, 2017). Furthermore, a comprehensive study included several models with C-N-P cycling and their feedbacks on the atmospheric C fixation and biomass growth in Amazon forests under ambient and elevated CO₂ conditions (eCO₂) (Fleischer *et al.*, 2019). Despite these developments, data to underpin them and their projections, particularly for the tropics, is sparse and remains challenging particularly for the Amazon forest (Reed *et al.*, 2015; Jiang *et al.*, 2019). Moreover, due to the lack of detailed measurements, the P-related processes such as ad/desorption and uptake represented in these models are under-constrained and likely oversimplified, thus the future predictions of Amazon forest responses to eCO₂ and climate change are uncertain. To fill this gap, in this study, we use data collected as part of the Amazon Fertilization Experiment (AFEX), the first project that focuses on experimental soil nutrient manipulation in the Amazon, with a comprehensive data collection program covering plant ecophysiology, C stocks and fluxes, soil processes including P stocks. Thus, our model parameterization compared to prior P modelling studies includes detailed P processes representation using the site measurements.

[Here, we describe the development and implementation of the terrestrial P cycle in the Joint UK Land Environment Simulator \(JULES\) \(Clark *et al.*, 2011\), the land component of the UK Earth System Model \(UKESM\), following the structure of the prior N cycle development \(Wiltshire *et al.*, 2021\) and utilising state of the art already tested and implemented descriptions of P cycling in other land surface models \(Wang, Houlton and Field, 2007; Zhu *et al.*, 2016; Goll *et al.*, 2017\).](#)

The model (JULES-CNP) is parameterized and calibrated using novel in situ P soil and plant data from a well-studied forest site in Central Amazon near to Manaus, Brazil with soil P content representative of 60% of soils across the Amazon basin. [The new developed P component estimates the sorption of the soil organic and inorganic P based on the saturation status of the adsorbed P pools, which is unique compared to the other existing P models and enable more realistic estimation of P ad/desorption processes. We first evaluate the model at our study site but also at additional five test sites across the Amazon, in Panama and Hawaii. We then apply the model under ambient and eCO₂ following the protocol of Fleischer *et al.*, \(2019\) to predict nutrient limitations on land biogeochemistry under these conditions. Predictions of the CO₂ fertilization effect in JULES-CNP are compared to those in current versions of the model with coupled C and N cycles \(JULES-CN\) and with C cycle only \(JULES-C\).](#)

2. Material and methods

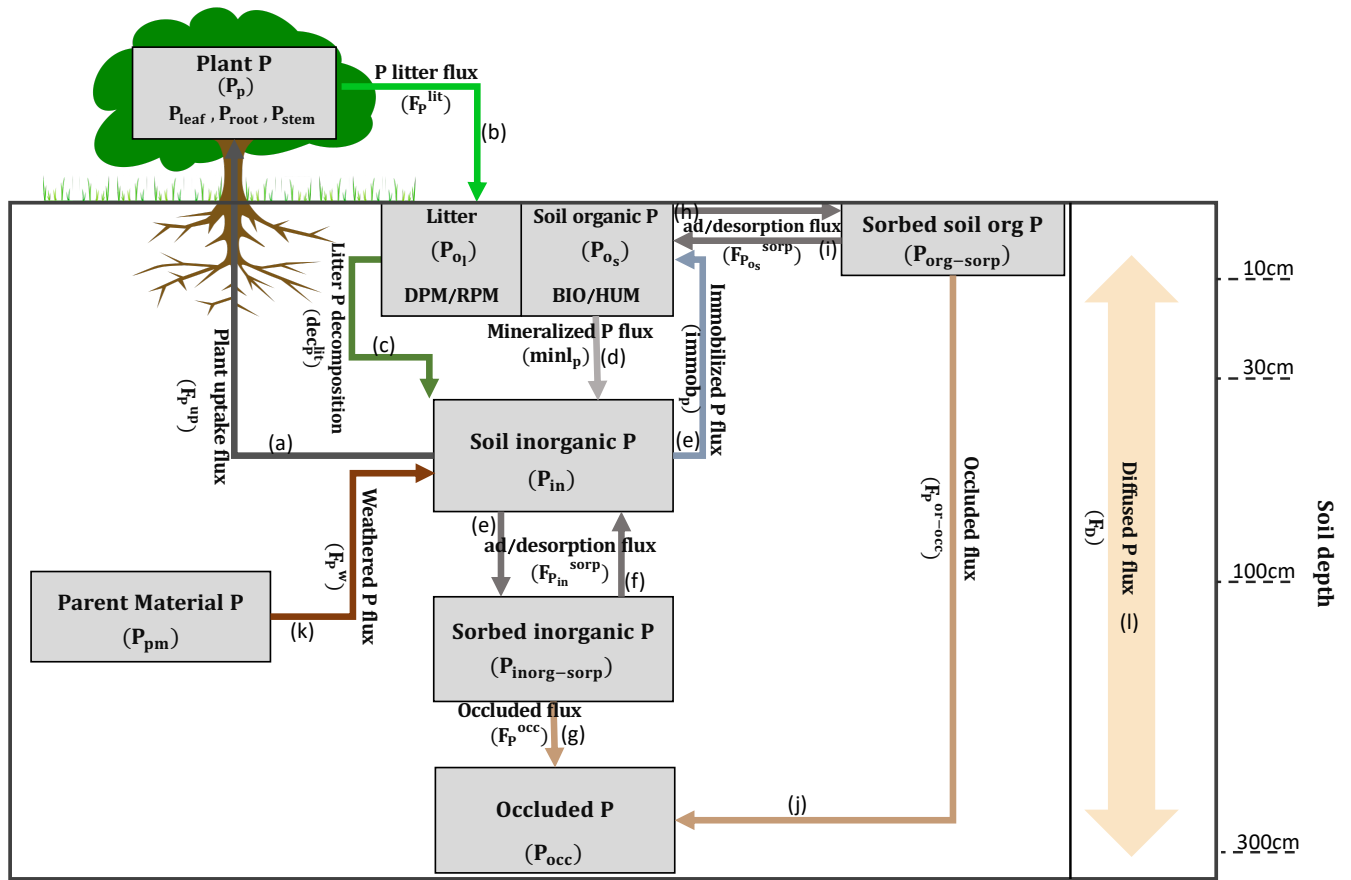
2.1 JULES

JULES is a process-based model that integrates water, energy, C cycling (JULES-C) (Clark *et al.*, 2011) and N cycling (JULES-CN) (Wiltshire *et al.*, 2021) between the atmosphere, vegetation and soil (Best *et al.*, 2011; Clark *et al.*, 2011). Vegetation dynamics are represented in JULES using the TRIFFID model, using nine distinct plant functional types (PFTs) (tropical and temperate broadleaf evergreen trees, broadleaf deciduous trees, needle-leaf evergreen and deciduous trees, C3 and C4 grasses, and evergreen and deciduous shrubs), as well as height competition (Harper *et al.*, 2016). [Leaf-level photosynthesis \(Collatz *et al.*, 1991; Collatz, Ribas-Carbo and Berry, 1992\) is scaled to estimate canopy level Gross Primary Productivity \(GPP\) using a multilayer approach that accounts for vertical variation of radiation interception and partition of sunlit and shaded leaves and associated vertical variation of leaf N and P -exponential decrease through the canopy \(Clark *et al.*, 2011; Mercado *et al.* 2007, Mercado *et al.* 2009\) - while the C:P and N:P ratios remain the same. NPP is estimated as the difference between GPP and autotrophic respiration for each living tissue \(leaf, wood, root\). NPP is then allocated to increase tissue C stocks and to spread, i.e., expand the fractional coverage of the PFT. The resultant PFT fractional coverages also depend on competition across PFTs for resources, e.g., light. Tissue turnover and vegetation mortality add C into the litter pools. Representation of soil organic C \(SOC\) follows the \[Rothamsted Carbon model\]\(#\) RothC equations \(Jenkinson *et al.*, 1990; Jenkinson and Coleman, 2008\) defining four C pools: decomposable plant material \(DPM\) and resistant plant material \(RPM\), which receive direct input from litterfall, and microbial biomass \(BIO\) and humified material \(HUM\) which receive a fraction of decomposed C from DPM and RPM which is not released to the atmosphere. The limitation of N on SOC is applied to the vegetation and soil components using a dynamic C:N ratio to modify the mineralization and immobilization processes as described in Wiltshire *et al.*, \(2021\). Note that the soil component of JULES-CN can be run either as a single box model or vertically resolved over soil depth \(JULES-CN layered\), and in this paper we build upon the vertically resolved version described in Wiltshire *et al.* \(2021\).](#)

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2.2 JULES-CNP

JULES-CNP includes the representation of the P cycle in JULES version (vn5.5) [and it is built on existing and well tested representations of P cycling in other global land surface models \(Wang, Houlton and Field, 2007; Yang et al., 2014; Goll et al., 2017; Sun et al., 2021\)](#). It includes P fluxes within the vegetation and soil components, and the specification of P pools and processes related to P cycling within the soil column (Fig. 1). A parent material pool is introduced to consider the input of weathered P. The adsorbed, desorbed and occluded fractions of P for both organic and inorganic P are also represented. However, except for parent material and occluded P pools, all other pools are estimated at each soil layer. The description of changes in pools and associated relative fluxes are explained in detail in the next sections. [Although](#) JULES-CN includes N leaching and deposition, P leaching and deposition are [not included](#) in the current version of JULES-CNP.



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Figure.1 – JULES-CNP model scheme [including P pools \(grey boxes\) and fluxes \(arrows\)](#)

2.2.1 P pools

JULES represents eight P pools comprising organic and inorganic P: plant P (P_p) and soil P pools (in each soil layer (n)), litter P (P_{0l}), soil organic P (P_{0s}), soil inorganic P (P_{in}), organic sorbed ($P_{org-sorp}$), inorganic sorbed ($P_{inorg-sorp}$), parent material (P_{pm}) and occluded (P_{occ}) P comprised of both organic and inorganic P. All pools are in units of kg P m^{-2} (Fig 1, Tables 1 and 2).

Plant P pool is composed of leaf (P_{leaf}), fine root (P_{root}) and stem together with coarse root (P_{stem}), which are related to their associated C pools (C_{leaf} , C_{root} , C_{stem}) in (kg C m^{-2}) and fixed C to P ratios ($C:P_{leaf}$, $C:P_{root}$, $C:P_{stem}$) as follows:

$$P_{leaf} = \frac{C_{leaf}}{C:P_{leaf}} \quad (\text{eq.1})$$

195 $P_{root} = \frac{C_{root}}{C:P_{root}}$ (eq.2)

196
197 $P_{stem} = \frac{C_{stem}}{C:P_{stem}}$ (eq.3)

198
199 Therefore, the plant P pool (P_p) is the sum of all vegetation P pools as follows:

200
201 $P_p = P_{leaf} + P_{root} + P_{stem}$ (eq.4)

202
203 Description of the plant P pool (P_p) follows Zhu *et al.*, (2016) and is estimated as the difference between the
204 input, plant uptake F_p^{up} (eq.26) and output of this pool, plant litter flux F_p^{lit} (eq.28), with both fluxes
205 expressed in kg P m⁻² yr⁻¹ as follows:

206
207 $\frac{dP_p}{dt} = F_p^{up} - F_p^{lit}$ (eq.5)

208
209 The litter P pool (P_{ol}) is estimated as a sum of P_{DPM} and P_{RPM} pools [over soil layers \(n\)](#). Each pool is formed by
210 the fluxes of plant litter input (F_p^{lit}) and the outgoing decomposed P (dec_p^{lit}) both expressed in kg P m⁻² yr⁻¹
211 (eq.28-29). Furthermore, the plant litter input is modified based on the plant type material ratio α (in order to
212 distribute the litter input based on the DPM/RPM fraction) as follows:

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214 $\frac{dP_{DPM}}{dt} = F_{P_n}^{lit} \times \alpha - dec_{P_{DPM},n}$ (eq.6)

215
216 $\frac{dP_{RPM}}{dt} = F_{P_n}^{lit} \times (1 - \alpha) - dec_{P_{RPM},n}$ (eq.7)

217
218 $P_{ol} = \sum_{n=1}^N P_{DPM,n} + \sum_{n=1}^N P_{RPM,n}$ (eq.8)

219
220 The soil organic pool (P_{os}) is represented as the sum of P_{BIO} and P_{HUM} . These pools are estimated from the
221 difference between P inputs from total immobilized (F_{immob_p}) distributed between BIO and HUM based on
222 fixed fraction (0.46 for BIO, 0.54 for HUM) (Jenkinson *et al.*, 1990; Jenkinson and Coleman, 2008) and
223 desorbed P, $F_{P_{os}}^{desorp}$ and P outputs from mineralized (F_{minl_p}), and adsorbed P fluxes ($F_{P_{os}}^{sorp}$) (adsorption:
224 eq. 40 and desorption: eq.41) with all fluxes expressed in kg P m⁻² yr⁻¹ as follows:

225
226 $\frac{dP_{BIO}}{dt} = 0.46 \times F_{immob_{P_n}} + F_{P_{os_{BIO},n}}^{desorp} - F_{minl_{P_{BIO},n}} - F_{P_{os_{BIO},n}}^{sorp}$ (eq.9)

227
228 $\frac{dP_{HUM}}{dt} = 0.54 \times F_{immob_{P_n}} + F_{P_{os_{HUM},n}}^{desorp} - F_{minl_{P_{BIO},n}} - F_{P_{os_{HUM},n}}^{sorp}$ (eq.10)

229
230 $P_{os} = \sum_{n=1}^N P_{BIO,n} + \sum_{n=1}^N P_{HUM,n}$ (eq.11)

231
232
233 Description of the inorganic sorbed P pool ($P_{inorg-sorp}$) follows Wang *et al.*, (2007) [and](#) is represented as the
234 difference between the input flux of inorganic sorption ($F_{P_{in}}^{sorp}$) (eq. 37) and output fluxes of inorganic
235 desorption ($F_{P_{in}}^{desorp}$) (eq. 38) and occluded P (F_p^{occ}) (eq. 39), with all fluxes expressed in kg P m⁻² yr⁻¹ as
236 follows:

237
238 $\frac{dP_{inorg-sorp}}{dt} = \sum_{n=1}^N F_{P_{in},n}^{sorp} - \sum_{n=1}^N F_{P_{in},n}^{desorp} - \sum_{n=1}^N F_{P_n}^{occ}$ (eq.12)

239
240 [The description](#) of the occluded (P_{occ}) P pool follows Wang *et al.*, (2007) and Hou *et al.*, (2019) [and](#) is
241 represented as the sum of input fluxes of occluded P from both organic (F_p^{or-occ}) (eq. 42) and inorganic P
242 pools (F_p^{occ}) expressed in kg P m⁻² yr⁻¹, as follows:

243
244 $\frac{dP_{occ}}{dt} = \sum_{n=1}^N F_{P_n}^{occ} + \sum_{n=1}^N F_{P_n}^{or-occ}$ (eq.13)

245

246 [The description](#) of the organic sorbed P pool ($P_{org-sorp}$) follows Wang *et al.*, (2007) [and](#) is represented as the
 247 difference between the input flux of organic sorption ($F_{P_{OS_n}^{sorp}}$) and output fluxes of organic desorption
 248 ($F_{P_{OS_n}^{desorp}}$) and occluded P ($F_{P_n^{occ}}$), with all fluxes expressed in kg P m⁻² yr⁻¹ as follows:

$$249 \frac{dP_{org-sorp}}{dt} = \sum_{n=1}^N F_{P_{OS_n}^{sorp}} - \sum_{n=1}^N F_{P_{OS_n}^{desorp}} - \sum_{n=1}^N F_{P_n^{or-occ}} \quad (\text{eq.14})$$

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 254 P from parent material (P_{pm}) pool follows Wang *et al.*, (2007) [and](#) depends on the weathering flux (F_p^w) (eq.
 255 43) in kg P m⁻² yr⁻¹ as follows:

$$256 \frac{dP_{pm}}{dt} = - \sum_{n=1}^N F_{P_n^w} \quad (\text{eq.15})$$

260 2.2.2. C and P fluxes

261
 262 NPP in JULES is calculated as the difference between GPP and autotrophic respiration. In JULES-CNP,
 263 potential NPP represents [the amount of C](#), available for tissue growth (C density increase) on a unit area, and
 264 spreading (vegetation cover increase as a result of reproduction and recruitment), [i.e.](#), to increase the area
 265 covered by the vegetation type, assuming no nutrient limitation. The reported NPP in the literature often
 266 includes other C fluxes related to the exudates, [production of](#) volatiles and non-structural carbohydrates (Malhi
 267 *et al.*, 2009; Chapin *et al.*, 2011; Walker *et al.*, 2021) which are challenging to measure (Malhi, Doughty and
 268 Galbraith, 2011). Therefore, actual NPP is for our purposes equal to Biomass Production (BP), and is calculated
 269 as potential NPP minus excess C (lost to the plant through autotrophic respiration), with the latter the C that
 270 cannot be used to grow new plant tissue due to insufficient plant nutrient supply. Hence, if the system is limited
 271 by the availability of N and/or P, NPP will be adjusted to match the growth that can be supported with the
 272 limited N or P supply, with any excess carbohydrate lost through excess C.
 273 The total excess C term (ψ_t) (kg C m⁻² yr⁻¹) is calculated as:

$$274 \psi_t = \psi_g + \psi_s \quad (\text{eq.16})$$

275
 276 where ψ_g and ψ_s are the excess C fluxes due to growth (g) and spread (s) and are assumed to be rapidly respired
 277 by plants.

278
 279 Therefore, BP is calculated as the difference between potential NPP (Π_c) and total excess C:

$$280 BP = \Pi_c - \psi_t \quad (\text{eq.17})$$

281
 282 The litter production in JULES before limitation is estimated as follows:

$$283 F_{C_n}^{lit} = \gamma_{leaf} C_{leaf} + \gamma_{root} C_{root} + \gamma_{wood} C_{wood} \quad (\text{eq.18})$$

284
 285 where γ is a temperature dependent turnover rate representing the phenological state (Clark *et al.*, 2011). P
 286 limitation is applied on the C litter production similar to the N scheme of JULES (JULES-CN) (Wiltshire *et al.*,
 287 2021). In JULES-CN the N limitation effect on the litter production is captured by estimating the available C for
 288 litter production as a difference between the NPP and excess C (Wiltshire *et al.*, 2021).
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293 Similar to other P-enabled models (Yang *et al.*, 2014; Goll *et al.*, 2017), JULES-CNP follows the same structure
 294 as its N model component. Description of the plant P and N demand follow Wang *et al.*, (2007) and are
 295 represented by the sum of demand (ϕ_t) to sustain growth (P-related: (ϕ_{gP}), N-related: (ϕ_{gN})) and to sustain
 296 vegetation spreading (to increment PFT fractional coverage) (P-related: (ϕ_{sP}), N-related: (ϕ_{sN})) and is
 297 expressed in (P-related in kg P m⁻² yr⁻¹; N-related in kg N m⁻² yr⁻¹). The total demand for growth (ϕ_g) and
 298 spreading (ϕ_s) is controlled by the dominant demand between P (ϕ_{gP}) and N (ϕ_{gN}) as follows:

$$299 \phi_t = \phi_g + \phi_s \quad (\text{eq.19})$$

$$301 \quad \phi_{gP} = \frac{P_p}{C_V} \left(\Pi_c - \frac{dC_V}{dt} - \psi_g \right) \quad (\text{eq.20})$$

$$302 \quad \phi_{sP} = \frac{P_p}{C_V} \left(\Pi_c - \frac{dC_V}{dt} - \psi_s \right) \quad (\text{eq.21})$$

$$303 \quad \phi_{gN} = \frac{N_v}{C_V} \left(\Pi_c - \frac{dC_V}{dt} - \psi_g \right) \quad (\text{eq.22})$$

$$304 \quad \phi_{sN} = \frac{N_v}{C_V} \left(\Pi_c - \frac{dC_V}{dt} - \psi_s \right) \quad (\text{eq.23})$$

$$305 \quad \phi_g = \begin{cases} \phi_{gP} & \phi_{gP} \times \frac{C_V}{P_p} > \phi_{gN} \times \frac{C_V}{N_v} \\ \phi_{gN} & \phi_{gN} \times \frac{C_V}{N_v} > \phi_{gP} \times \frac{C_V}{P_p} \end{cases} \quad (\text{eq.24})$$

$$306 \quad \phi_s = \begin{cases} \phi_{sP} & \phi_{sP} \times \frac{C_V}{P_p} > \phi_{sN} \times \frac{C_V}{N_v} \\ \phi_{sN} & \phi_{sN} \times \frac{C_V}{N_v} > \phi_{sP} \times \frac{C_V}{P_p} \end{cases} \quad (\text{eq.25})$$

307

308

309 where $\frac{P_p}{C_V}$ is the inverse of whole plant C:P ratio, $\frac{N_v}{C_V}$ is inverse plant C:N ratio, $\frac{dC_V}{dt}$ is rate of change in plant C
 310 (see Clark *et al.*, (2011) for more detail), Π_c is nutrient-unlimited, or potential, NPP ($\text{kg C m}^{-2} \text{ yr}^{-1}$), ψ_g is excess
 311 C due to either P or N limitation for plant growth ($\text{kg C m}^{-2} \text{ yr}^{-1}$) and ψ_s is excess C due to either P or N
 312 limitation for vegetation spreading ($\text{kg C m}^{-2} \text{ yr}^{-1}$).

313

314 Equations 20 and 22 are solved by first setting $\psi_g = 0.0$ to find the total plant P (eq. 20) and N demand (eq.22).

315 If the P and N demand for growth are less than the available P and N and fractional coverage (λ) (NPP fraction

316 used for fractional cover increment; for detail see Wiltshire *et al.*, (2021)) at the considered timestep Δt then

317 there is no limitation to growth (*i. e.* $\phi_{gP} < \frac{(1-\lambda)P_{avail}}{\Delta t}$; $\phi_{gN} < \frac{(1-\lambda)N_{avail}}{\Delta t}$). Where there is limited P and/or N

318 availability, the uptake equals the available P and N ($\phi_{gP} = \frac{(1-\lambda)P_{avail}}{\Delta t}$; $\phi_{gN} = \frac{(1-\lambda)N_{avail}}{\Delta t}$), and the plant

319 growth which cannot be achieved due to nutrient constraints will be deducted from potential NPP, here termed

320 excess C term (ψ_g), to give an actual NPP. Following Wiltshire *et al.*, 2021, we assume excess C is respired by

321 the plant.

322

323 Similarly, in order to estimate the P and N demand for spreading (eq. 21 and 23), initially the excess C from
 324 spreading is set to 0.0 ($\psi_s = 0.0$), *i.e.* under the assumption that there is no nutrient limitation. If the P and N

325 demand for spreading are lower than the available P and N and fractional coverage (λ) ($\phi_{sP} <$

326 $\frac{(1-\lambda)P_{avail}}{\Delta t}$; $\phi_{sN} < \frac{(1-\lambda)N_{avail}}{\Delta t}$), then there is no limitation on spreading and in case of limited P and N

327 availability, the uptake equals the available P and N ($\phi_{sP} = \frac{(1-\lambda)P_{avail}}{\Delta t}$; $\phi_{sN} = \frac{(1-\lambda)N_{avail}}{\Delta t}$), and the excess C

328 for spread (ψ_s) is subtracted from potential NPP.

329

330 Plant P uptake (F_p^{up}) (arrow a in Fig 1) is estimated based on the P demand for growth and spreading (ϕ_t) and

331 the root uptake capacity (u^{max}) ($\text{kg P kg}^{-1} \text{ C yr}^{-1}$), as follows:

332

$$333 \quad F_p^{up} = \begin{cases} \phi_t & \phi_t \leq u^{max} \\ u^{max} & \phi_t > u^{max} \end{cases} \quad (\text{eq.26})$$

334

335 **Plant P uptake** (F_p^{up}) varies spatially depending on the root uptake capacity (u^{max}) followed by Goll *et al.*,
 336 (2017). Therefore, in regions with limited P supply, the plant P uptake is limited to the u^{max} and consequently
 337 impacts the excess C and BP.

338 The root uptake capacity depends on the maximum root uptake capacity (v_{max}) ($\text{kg P kg}^{-1} \text{ C yr}^{-1}$), root depth

339 (d_{root}), the concentration of inorganic P at different soil depths (P_{in}), and a half saturation term at which half of

340 the maximum uptake capacity is reached using inorganic P at different soil depths (P_{in}), a scaling uptake ratio

341 (K_p) ($\mu\text{mol P l}^{-1}$), unit conversion (C_f) (1 kg P^{-1}), and soil moisture (θ) (1 m^{-2}), as follows:

342

$$343 \quad u^{max} = v_{max} \times d_{root} \times \sum_{n=1}^N P_{in_n} \times \left(\frac{1}{\sum_{n=1}^N P_{in_n} + c_f \times K_p \times \theta_n} \right) \quad (\text{eq.27})$$

344

345 Description of the litter production of P ($F_{P_n}^{lit}$) (arrow b in Fig 1) follows JULES-CN as in Wiltshire *et al.*,
 346 (2021) and is calculated based on the litter flux of C ($\text{kg C m}^{-2} \text{ yr}^{-1}$) using leaf, root and wood turnovers (yr^{-1}),
 347 and through the vegetation dynamics due to large-scale disturbance and litter production density, as follows:
 348

$$349 \quad F_{P_n}^{lit} = \left(\frac{(1-k_{leaf})\gamma_{leaf}C_{leaf}}{C:P_{leaf}} \right) + \left(\frac{(1-k_{root})\gamma_{root}C_{root}}{C:P_{root}} \right) + \left(\frac{\gamma_{wood}C_{wood}}{C:P_{stem}} \right) \quad (\text{eq.28})$$

350
 351 where λ is the leaf, root and stem re-translocation (at daily timestep) coefficient (Zaehle and Friend, 2010; Clark
 352 *et al.*, 2011) and the related $C:P$ ratios for P fraction and γ is a temperature dependent turnover rate representing
 353 the phenological state (Clark *et al.*, 2011).
 354
 355

356 The decomposition of litter (dec^{lit}) (arrow c in Fig 1) depends on soil respiration (R) ($\text{kg C m}^{-2} \text{ yr}^{-1}$), the litter
 357 $C:P$ ratio ($C:P_{lit}$) at each soil layer (n) as follows:
 358

$$359 \quad dec_p^{lit} = \frac{\sum_{n=1}^N R_n}{C:P_{lit}} \quad (\text{eq.29})$$

360
 361 where the $C:P_{lit}$ is calculated based on litter C pool (DPM and RPM) (lit^C) ($\text{kg C m}^{-2} \text{ yr}^{-1}$) and litter P pool
 362 (P_{O_l}) as follows:
 363

$$364 \quad C:P_{lit} = \frac{\sum_{n=1}^N lit_n^C}{P_{O_{ln}}} \quad (\text{eq.30})$$

365
 366 The mineralized (F_{minl_p}) (arrow d in Fig 1) and immobilized (F_{immob_p}) (arrow e in Fig 1) P fluxes are
 367 calculated based on C mineralization and immobilization, $C:P$ ratios of plant (i) (DPM/RPM) ($C:P_{plant}$) and
 368 soil (HUM/BIO) ($C:P_{soil}$), soil pool potential respiration (R_{POT_i}) ($\text{kg C m}^{-2} \text{ yr}^{-1}$) and the respiration partitioning
 369 fraction ($resp_frac$) as follows:
 370

$$371 \quad F_{minl_p} = \frac{\sum_{n=1}^N R_{POT_{i,n}}}{C:P_{plant}} \quad (\text{eq.31})$$

$$373 \quad F_{immob_p} = \frac{\sum_{n=1}^N R_{i,n} \times resp_frac}{C:P_{soil}} \quad (\text{eq.32})$$

374
 375 The soil respiration from each soil layer ($R_{i,n}$) is estimated from potential soil respiration ($R_{POT_{i,n}}$) for the
 376 DPM, RPM pools and the litter decomposition rate modifier (F_{P_n}) as follows:
 377

$$378 \quad R_{i,n} = R_{POT_{i,n}} \times F_{P_n} \quad (\text{eq.33})$$

379
 380 where the description of F_{P_n} for P pools (F_{P_n}) follows Wang *et al.*,(2007) and is estimated based on the soil
 381 pool (BIO/HUM) mineralization ($minl_{p-BIO_n}$, $minl_{p-HUM_n}$) and immobilization ($immob_{p-BIO_n}$,
 382 $immob_{p-HUM_n}$) (in $\text{kg P m}^{-2} \text{ yr}^{-1}$), soil inorganic P (P_{inorg_n}) (in kg P m^{-2}), and litter pools (DPM/RPM) demand
 383 (in $\text{kg P m}^{-2} \text{ yr}^{-1}$) as follows:
 384

$$385 \quad F_{P_n} = \frac{(minl_{p-BIO_n} + minl_{p-HUM_n} - immob_{p-BIO_n} - immob_{p-HUM_n}) + P_{inorg_n}}{DEM_{DPM_n} + DEM_{RPM_n}} \quad (\text{eq.34})$$

386
 387 The net demand associated with decomposition of litter pools ($DEM_{k,n}$) represents the P required by microbes
 388 which convert DPM and RPM into BIO and HUM. The limitation due to insufficient P availability is estimated
 389 based on the potential mineralization ($minl_{p-pot}$) and immobilization ($immob_{p-pot}$) (in $\text{kg P m}^{-2} \text{ yr}^{-1}$) of pools
 390 (k) as follows:
 391

$$392 \quad DEM_{k,n} = immob_{p-pot,k} - minl_{p-pot,k} \quad (\text{eq.35})$$

393
 394 The F_{P_n} estimated for N pools (F_{P_n}) follows the same formulation as P (see Wiltshire *et al.*, 2021 for [further](#)
 395 [details](#)) and the F_{P_n} is estimated based on a higher rate modifier between N and P as follows:
 396

$$F_{P_n} = \begin{cases} F_{PP_n} & F_{PP_n} > F_{PN_n} \\ F_{PN_n} & F_{PN_n} > F_{PP_n} \end{cases} \quad (\text{eq.36})$$

398

399 Description of the fluxes of adsorption ($F_{P_{in_n}}^{sorp}$) (arrow e in Fig 1) and desorption ($F_{P_{in_n}}^{desorp}$) (arrow f in Fig
400 1) of inorganic P in kg P m⁻² yr⁻¹ follow Wang *et al.*, (2010) and are calculated based on soil inorganic (P_{in_n}) and
401 sorbed inorganic ($P_{inorg-sorbed_n}$) P pools and inorganic adsorption ($K_{sorp-in}$), desorption ($K_{desorp-in}$)
402 coefficients (kg P m⁻² yr⁻¹) and maximum sorbed inorganic (P_{in-max}) (kg P m⁻²) as follows:
403

$$F_{P_{in_n}}^{sorp} = P_{in_n} \times K_{sorp-in} \times \frac{(P_{in-max_n} - P_{inorg-sorbed_n})}{P_{in-max_n}} \quad (\text{eq.37})$$

405

$$F_{P_{in_n}}^{desorp} = P_{inorg-sorbed_n} \times K_{desorp-in} \quad (\text{eq.38})$$

407

408 Description of the occluded inorganic P flux ($F_{P_n}^{occ}$) (arrow g in Fig 1) follows Wang *et al.*, (2007) and Hou *et al.*, (2019) and is calculated based on sorbed inorganic P pool and P occlusion rate (K_{occ}) (kg P m⁻² yr⁻¹) as
409 follows:
410

411

$$F_{P_n}^{occ} = P_{inorg-sorbed_n} \times K_{occ} \quad (\text{eq.39})$$

413

414 Description of the fluxes of adsorption ($F_{P_{OS_n}}^{sorp}$) (arrow h in Fig 1) and desorption ($F_{P_{OS_n}}^{desorp}$) (arrow i in Fig
415 1) of organic P follow Wang *et al.*, (2010) are calculated based on soil organic and sorbed organic P pools and
416 organic adsorption ($K_{sorp-or}$) (kg P m⁻² yr⁻¹), desorption ($K_{desorp-or}$) coefficients (kg P m⁻² yr⁻¹) and maximum
417 sorbed organic ($P_{org-max}$) (which corresponds to the sorbed soil P saturation, thus modifying the sorption rate
418 respectively) (kg P m⁻²) as follows:
419

$$F_{P_{OS_n}}^{sorp} = P_{OS_n} \times K_{sorp-or} \times \frac{(P_{or-max_n} - P_{org-sorbed_n})}{P_{or-max_n}} \quad (\text{eq.40})$$

421

$$F_{P_{OS_n}}^{desorp} = P_{org-sorbed_n} \times K_{desorp-or} \quad (\text{eq.41})$$

423

424 Description of the occluded organic P flux ($F_{P_n}^{or-occ}$) (kg P m⁻² yr⁻¹) (arrow j in Fig 1) follows Wang *et al.*,
425 (2007) and Hou *et al.*, (2019) is calculated based on sorbed organic P pool ($P_{org-sorbed_n}$) and P occlude rate
426 (K_{occ}) (kg P m⁻² yr⁻¹) as follows:
427

428

$$F_{P_n}^{or-occ} = P_{org-sorbed_n} \times K_{occ} \quad (\text{eq.42})$$

429

430 Description of the P flux from weathered parent material ($F_{P_n}^w$) (arrow k in Fig 1) follows Wang *et al.*, (2007)
431 and is calculated based on amount of P in the parent material (P_{pm}) and P weathering rate (K_w) (kg P m⁻² yr⁻¹) as
432 follows:
433

434

$$F_{P_n}^w = P_{pm_n} \times K_w \quad (\text{eq.43})$$

435

436 Description of P diffusion between soil layers (F_{D_n}) expressed in (kg P m⁻² yr⁻¹) (arrow l in Fig 1) follows Goll
437 *et al.*, (2017) and is calculated following Fick's second law and it is a function of the diffusion coefficient (Dz)
438 in m² s⁻¹, the concentration of inorganic P at different soil depths (P_{in}) in kg P m⁻², the distance (z) between the
439 midpoints of soil layers in metres and seconds to year unit conversion (Yr):
440

$$F_{D_n} = \frac{\partial}{\partial z} (D_{zn} \frac{\partial P_{zn}}{\partial z}) \times Yr \quad (\text{eq.44})$$

442

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Table 1. Model variables

Variable	Unit	Definition
ψ	kg C m ⁻² yr ⁻¹	Excess C flux
\emptyset	kg P m ⁻² yr ⁻¹	Plant demand for uptake
Π_c	kg C m ⁻² yr ⁻¹	Potential NPP
u^{max}	kg P kg ⁻¹ C yr ⁻¹	Root uptake capacity
DEM	kg P m ⁻² yr ⁻¹	Plant pool P associated decomposition demand
dec_P^{lit}	kg P m ⁻² yr ⁻¹	Litter decomposition
F_D	kg P m ⁻² yr ⁻¹	Plant diffusion flux
F_P	-	Plant litter decomposition rate modifier
F_P^{lit}	kg P m ⁻² yr ⁻¹	Plant litter flux
F_P^{up}	kg P m ⁻² yr ⁻¹	Plant uptake
$F_{PO_S}^{sorp}$	kg P m ⁻² yr ⁻¹	Sorbed organic P flux
$F_{P_{in}}^{sorp}$	kg P m ⁻² yr ⁻¹	Sorbed inorganic P flux
$F_{PO_S}^{desorp}$	kg P m ⁻² yr ⁻¹	Desorbed organic P flux
$F_{P_{in}}^{desorp}$	kg P m ⁻² yr ⁻¹	Desorbed inorganic P flux
F_P^{occ}	kg P m ⁻² yr ⁻¹	Occluded inorganic P flux
F_P^{or-occ}	kg P m ⁻² yr ⁻¹	Occluded organic P flux
F_P^w	kg P m ⁻² yr ⁻¹	Weathered P flux
F_{immob_P}	kg P m ⁻² yr ⁻¹	Immobilized P flux
lit_C	kg C m ⁻² yr ⁻¹	C litter flux
lit_{frac}	-	Litter fraction
lit_{leaf}	kg C m ⁻² yr ⁻¹	Leaf litter flux
lit_{root}	kg C m ⁻² yr ⁻¹	Root litter flux
lit_{wood}	kg C m ⁻² yr ⁻¹	Woody litter flux
F_{minl_P}	kg P m ⁻² yr ⁻¹	Mineralized P flux
P_p	kg P m ⁻²	Plant P pool
P_{O_l}	kg P m ⁻²	Litter organic pool
P_{O_s}	kg P m ⁻²	Soil organic pool
P_{in}	kg P m ⁻²	Soil inorganic pool
$P_{inorg-sorp}$	kg P m ⁻²	Soil inorganic sorbed pool
$P_{org-sorp}$	kg P m ⁻²	Soil organic sorbed pool
P_{occ}	kg P m ⁻²	Soil occluded pool
P_{pm}	kg P m ⁻²	Parent material pool
R	kg C m ⁻² yr ⁻¹	Total respiration
R_{POT}	kg C m ⁻² yr ⁻¹	Total potential respiration
R^s	kg C m ⁻² yr ⁻¹	Soil respiration
R_d	kg C m ⁻² yr ⁻¹	Leaf dark respiration
T_{ref}	K	Soil reference temperature
T_s	K	Soil temperature
Veg_c	kg C m ⁻²	Sum of biomass
z	m	Soil depth

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453

Table 2. P Model parameters

Parameter	Value	Unit	Eq.	Description	Source
C and N related					
α	0.25	-	6	Plant type material ratio	(Clark <i>et al.</i> , 2011)
a_{wl}	1.204	kg C m ⁻²	50	Allometric coefficient	calibrated
σ_l	0.0375	kg C m ⁻² per unit LAI	48	Specific leaf density	Clark <i>et al.</i> , 2011
b_{wl}	1.667	-	50	Allometric exponent.	Clark <i>et al.</i> , 2011
f_{dr}	0.005	-	47	Respiration scale factor	Calibrated
$resp_frac$	0.25	-	32	Respiration fraction	(Clark <i>et al.</i> , 2011)
k_{leaf}	0.5	-	28	Leaf N re-translocation coefficient	(Zachle and Friend, 2010)
k_{root}	0.2	-	28	Root N re-translocation coefficient	(Zachle and Friend, 2010)
d_{root}	3.0	-	27	Root fraction in each soil layer	(Clark <i>et al.</i> , 2011)
v_{int}	7.21	$\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$	45	Intercept in the linear regression between V_{cmax} and N_{area}	Calibrated (Clark <i>et al.</i> , 2011)
v_{sl}	19.22	$\mu\text{mol CO}_2 \text{ gN}^{-1} \text{ s}^{-1}$	45	Slope in the linear regression between V_{cmax} and N_{area}	Calibrated (Clark <i>et al.</i> , 2011)
LMA	131.571852	g m ⁻²	45	Observed Leaf Mass per Area	Study site
$Leaf\ N$	1.79007596	g g ⁻¹	45, 46	Foliar N concentrations	Study site
P related					
$C:P_{soil}$	1299.6	-	32	Soil C:P ratio	(Fleischer <i>et al.</i> , 2019)
v_{max}	0.0007	kg P kg ⁻¹ C yr ⁻¹	27	Maximum root uptake capacity	Calibrated (Goll <i>et al.</i> , 2017)
P	0.7083062	g kg ⁻¹	46	Foliar P concentrations	Study site
c_f	3.1×10^{-5}	l kg P ⁻¹	27	Conversion factor	(Goll <i>et al.</i> , 2017)
D_z	0.001	m ² s ⁻¹	44	Diffusion coefficient	(Burke <i>et al.</i> , 2017)
K_{occ}	1.2×10^{-5}	yr ⁻¹	39, 42	P occlusion rate	(Yang <i>et al.</i> , 2014)
K_p	3.0	kg P l ⁻¹	27	Scaling uptake ratio	Calibrated
$K_{sorp-in}$	0.0054	kg P m ⁻² yr ⁻¹	37	Inorganic P adsorption coefficient	Calibrated (Hou <i>et al.</i> , 2019)
$K_{sorp-or}$	0.00054	kg P m ⁻² yr ⁻¹	40	Organic P adsorption coefficient	Calibrated
K_{in-max}	0.0075	kg P m ⁻² yr ⁻¹	37	Maximum sorbed inorganic P	Study site
K_{or-max}	0.0042	kg P m ⁻² yr ⁻¹	40	Maximum sorbed organic P	Study site
K_w	3×10^{-6}	kg P m ⁻² yr ⁻¹	43	P weathering rate	(Wang <i>et al.</i> , 2010)

2.3 Study sites

This study [primarily](#) uses data from two nearby sites in Central Amazon in Manaus, Brazil. The main site from here on termed *study site* (2°35′21.08″ S, 60°06′53.63″ W) (Lugli *et al.*, 2020) is for model development and evaluation. The second site is the Manaus K34 flux site (2°36′32.67″ S, 60°12′33.48″ W) which provides meteorological station data for running the model but also provides data for model evaluation. Our *study site* is the main lowland tropical forest site maintained by the National Institute for Amazon Research (INPA). [Research at this site focuses on projects, combining experimental approaches \(Keller *et al.*, 2004; Malhi *et al.*, 2009\) with modelling \(Lapola and Norby, 2014\).](#) We use detailed novel soil and plant P pool data from the *study site* (Lugli *et al.*, 2020, 2021) for model parameterisation and calibration and carbon stock data for model validation. The *study site* has a very similar forest, geomorphology, soil chemistry and species composition to the well-known and studied K34 flux site (Araújo *et al.*, 2002). The average reported annual precipitation is 2431 (mm yr⁻¹), with a monthly range of 95 to 304 (mm month⁻¹), and averaged temperature is 26°C (Araújo *et al.*, 2002). [Soil type](#) at this site is Geric Ferrosol with a high clay content and weathering activities (Malhi *et al.*, 2004).

473 [In addition to the *study site* we use data from other P limited locations from the Amazon, Panama and Hawaii](#)
474 [\(Table 3\) for model evaluation. Old-growth forest sites in the Amazon are located across a fertility gradient](#)
475 [from west to east \(AGP-01, SA3, CAX,\) with detailed C cycle measurements available \(Aragão *et al.*, \(2009\)\).](#)
476 [The site in Panama is located in the Gigante Peninsula in the Barro Colorado Nature Reserve and is a 200 year](#)
477 [old semi-deciduous rainforest \(Wright *et al.*, 2011\) growing on Oxisols developed on Miocene basalt \(Dieter,](#)
478 [Elsenbeer and Turner, 2010\) with the topsoil a dominant clay texture \(Turner and Condron, 2013\). It is the](#)
479 [location of a long term running nutrient fertilization experiment since 1998 \(Mirabello *et al.*, 2013\). The site in](#)
480 [Hawaii \(Hawaii Kokee\) is a P limited chronosequence that developed on the 4 million year old oxisols soil](#)
481 [\(Vitousek, 2004\) and has a long term fertilization experiment. Site information is provided in Table 3.](#)

482 **Table 3.** Test sites name, location and climate characterises.
483

Site	Name	Location		Climate	
		Lat.	Lon.	Rainfall (mm yr ⁻¹)	Temperature(°C)
Study site	AFEX project	-2.58	-60.11	2431	26
AGP-01	Agua pudre plot E	-3.72	-70.3	2723	25.5
CAX	Caxiuanã flux tower site	-1.72	-51.5	2314	26.9
SA3	Tapajós flux tower site	-2.5	-55	1968	26.1
Gig. Pen.	Gigante peninsula (control data)	-9.1	-79.84	2600	26
Hawaii K.	Hawaii Kokee (control data)	22.13	-159.62	2500	16

484

485

486 **2.4 Model parameterisation, calibration and evaluation [at study site](#)**

487

488 We use observations from the four control plots of the study site to parameterise, calibrate and evaluate different
489 processes in JULES (Table 4). The observations were collected at 4 soil depths and processed using the Hedley
490 sequential fractionation (Hedley, Stewart and Chauhan, 1982; Quesada *et al.*, 2010). Observed Leaf Mass per
491 Area (LMA), leaf N and leaf P estimated from fresh leaves were used as input parameters to JULES to estimate
492 photosynthetic capacity and respiration parameters. JULES vn5.5 (JULES CN in this study) estimates V_{cmax}
493 ($\mu\text{mol m}^{-2} \text{s}^{-2}$) based on Kattge *et al.* (2009) using foliar N concentrations in area basis (n_{leaf}), as follows:

494

$$495 V_{cmax} = v_{int} + v_{sl} * n_{leaf} \quad (\text{eq.45})$$

496

497 where v_{int} is the estimated intercept and v_{sl} is the slope of the linear regression derived for the V_{cmax} estimation.

498

499 We incorporated an additional P dependency on the estimation of V_{cmax} following Walker *et al.* (2014) as

500

$$501 \ln(V_{cmax}) = 3.946 + 0.921 \ln(N) + 0.121 \ln(P) + 0.282 \ln(N) \ln(P) \quad (\text{eq.46})$$

502

503 Where N and P are foliar concentrations in area basis.

504

505 Implementation of eq. 46 resulted in higher V_{cmax} than in the original version of JULES. A higher V_{cmax} predicted
506 higher leaf and plant respiration (eq.47). Constrained by observations of NPP and plant respiration at the study
507 site, we modified one of the most uncertain parameters in the description of plant respiration (f_{dr}) (eq.47) which
508 is the scale factor for leaf dark respiration (R_d) as follows:

509

$$510 R_d = f_{dr} V_{cmax} \quad (\text{eq.47})$$

511

512 The default value is 0.01 (Clark *et al.*, 2011), and for JULES-CNP simulations at our study site it was modified
513 to 0.005.

514

515 Observations of aboveground biomass were used to calibrate the non PFT dependent allometric relationships in
516 JULES (Clark *et al.* 2011) (eq 48-50) for leaf, root and wood C. Specifically, the a_{wl} parameter (eq 50) was
517 modified from 0.65 to 1.204 to match better tropical forest allometry:

518

$$518 C_{leaf} = \sigma_l L_b \quad (\text{eq.48})$$

519

$$519 C_{root} = C_{leaf} \quad (\text{eq.49})$$

$$C_{stem} = a_{wl} L_b^{b_{wl}} \quad (\text{eq.50})$$

Where σ_l is specific leaf density (kg C m⁻² per unit LAI), L_b is balanced (or seasonal maximum) leaf area index (m² m⁻²), a_{wl} is allometric coefficient (kg C m⁻²) and b_{wl} is the allometric exponent.

Note that JULES-CNP uses the C3 and C4 photosynthesis model from Collatz et al., 1991; Collatz, Ribas-Carbo and Berry, 1992, which does not include estimation of J_{max} .

JULES-CNP has fixed stoichiometry and C:P ratios of leaf and root (measured), and wood (estimated from fresh coarse wood (Lugli, 2013)) which were taken from the *study site* and prescribed in JULES to simulate P dynamics in the plant. The following belowground data were used to represent various soil P pools: Resin and bicarbonate inorganic P (inorganic P: P_{in}), organic bicarbonate P (organic P: P_{O_S}), NaOH organic P (sorbed organic P: $P_{org-sorp}$), NaOH inorganic P (sorbed inorganic P: $P_{inorg-sorp}$), residual P (occluded P: P_{occ}) and HCL P (parent material P: P_{pm}) (Table 4). The measurements were collected between 2017 and 2018 in control plots. All measurements were conducted in four soil layers (0-5, 5-10, 10-20, 20-30 cm). However, to be consistent with the JULES model soil layer discretization scheme, we defined 4 soil layers (0-10 cm, 10-30 cm, 30-100 cm and 100-300 cm) and we used the average between 0 and 30 cm to compare against the measurement from the same depth for model evaluation.

Vegetation C stocks were derived based on tree diameter measurements at breast height, that are linked to allometric equations and wood density databases to estimate the C stored in each individual tree, and then scaled to the plot (Chave et al., 2014).

The organic and inorganic soil P was assumed to be always at equilibrium with the relative sorbed pools (Wang, Law and Pak, 2010). Thus, in order to cap P sorption and uptake capacity, the maximum sorption capacities (P_{in-max_n} , P_{or-max_n} , eq.37 and 39) (adopted from (Wang, Houlton and Field, 2007)) were prescribed using maximum observed sorbed inorganic and organic P. Hence, the maximum sorption capacity defines the equilibrium state of sorbed and free-soil P. Moreover, despite the initial representation of the parent material pool in JULES and its depletion through weathering (eq. 43), as the magnitude of changes in the occluded and parent material pools are insignificant over a short-term (20 years) simulation period (Vitousek et al., 1997), these two pools were prescribed using observations. Remaining parameters used to describe soil P fluxes (eqs. 27-44) were prescribed using values from the literature (Table 4).

We used a combination of data from the *study site* and the nearby K34 site for model evaluation of C fluxes (GPP, NPP) and C pools (soil and vegetation C, leaf, root and wood C) with no calibration of plant and soil organic and soil inorganic P pools included (Table 4).

Table 4. Observations from study site (taken during 2017-2018) and from Manaus site K34 used for model parameterisation and evaluation

Process	Variables	Purpose of use	Reference and site
C associated	GPP	Evaluation	Fleischer et al., 2019, K34
	NPP	Evaluation	Fleischer et al., 2019, K34
	Soil C	Evaluation	Malhi et al., 2009, K34
	CUE	Evaluation	Malhi et al., 2009, K34
	Veg C	Evaluation	Study site
	Leaf C	Evaluation	Study site
	Wood C	Evaluation	Study site
	Root C	Evaluation	Study site
	LAI	Initialisation	Study site
LMA	Parameterisation	Study site	
P associated	Resin	Evaluation	Study site
	Pi Bic	Evaluation	Study site
	Po Bic	Evaluation	Study site
	Po NaOH	Calibration	Study site
	Pi NaOH	Calibration	Study site
	P residual	Parameterisation	Study site
	P HCL	Parameterisation	Study site
	Leaf N	Parameterisation	Study site
	Leaf P	Parameterisation	Study site
	Root P	Parameterisation	Study site
Plant C:P ratio	Parameterisation	Study site	

2.4.1 Model parameterisation and evaluation at test sites

JULES-CNP was parameterised using reported C:P ratios and maximum sorbed organic and inorganic P for each test site (Table 5) as follows:

Table 5. Additional test sites data used for model parameterisation

	<u>AGP-01^{a,b}</u>	<u>CAX^{a,b}</u>	<u>SA3^{a,b}</u>	<u>Gig. Pen.^c</u>	<u>Hawaii K.^{b,d}</u>
<u>LeafC:P</u>	<u>600</u>	<u>600</u>	<u>600</u>	<u>700</u>	<u>691.5</u>
<u>RootC:P</u>	<u>1000</u>	<u>1000</u>	<u>1000</u>	<u>1750</u>	<u>1100</u>
<u>WoodC:P</u>	<u>3000</u>	<u>3000</u>	<u>3000</u>	<u>5500</u>	<u>5937.5</u>
<u>SoilC:P</u>	<u>2000</u>	<u>2000</u>	<u>2000</u>	<u>800</u>	<u>2000</u>
<u>K_{or-max}</u>	<u>0.001</u>	<u>0.001</u>	<u>0.001</u>	<u>0.0033</u>	<u>0.001</u>
<u>K_{in-max}</u>	<u>0.001</u>	<u>0.001</u>	<u>0.001</u>	<u>0.0185</u>	<u>0.001</u>

^aC:P ratios from Wang, Law and Pak, 2010 and ^bmaximum sorbed P capacities from Yang *et al.*, 2014.

^cMirabello *et al.*, 2013 ^d C:P ratios from Vitousek, 2004

Model evaluation at test sites, was performed using observed NPP, litterfall, autotrophic respiration, biomass and soil C pools taken from different sources. We used NPP and litterfall for the Amazon sites from Aragão *et al.*, (2009) and for Gigante Peninsula from Chave *et al.*, (2003), Hawaii Kokee NPP as reported in Goll *et al.*, (2017) and litterfall as reported in (Yang *et al.*, 2014). Plant respiration was only available at two of Amazon sites (AGP and CAX) (Malhi *et al.*, 2009). The biomass and soil C pools for Amazon sites (CAX and SA3) are taken from Malhi *et al.*, (2009) and biomass from AGP is taken from Jiménez *et al.*, (2009). The Gigante Peninsula biomass is taken from Chave *et al.*, (2003), soil C from Turner *et al.*, (2015), and the Hawaii Kokee C pools are taken as reported in Yang *et al.*, (2014).

2.5 JULES simulations

JULES was first applied at the K34 flux tower site using observed meteorological forcing data from 1999-2019 (Fleisher *et al.* 2019) at half hourly resolution. The following meteorological variables are needed to drive JULES (model inputs) (Best *et al.*, 2011): atmospheric specific humidity (kg kg^{-1}), atmospheric temperature (K), air pressure at the surface (Pa), short and longwave radiation at the surface (W m^{-2}), wind speed (m s^{-1}) and total precipitation ($\text{kg m}^{-2} \text{s}^{-1}$). Furthermore, the averaged measured LAI from study site was used to initialise the vegetation phenology module, but was allowed to vary in subsequent prognostic calculations. Soil organic and inorganic sorbed P pools were initialised with study site observations. The JULES-CNP simulations were initialized following the same methodology as in Fleischer *et al.*, (2019), by the spin-up from 1850 resulted in equilibrium state (Figure S1). The spin up was performed separately for three versions of JULES (C/CN/CNP) following the same procedure. Furthermore, the transient run was performed for the period 1851-1998 using time-varying CO_2 and N deposition fields. Finally, for the extended simulation period (1999-2019) two runs were performed, the first with ambient the second elevated CO_2 concentrations.

We evaluate the impact of including a P cycle in JULES using three model configurations (JULES C, CN and CNP). We apply JULES in all three configurations using present day climate under both ambient CO_2 and $e\text{CO}_2$. Ambient and $e\text{CO}_2$ were prescribed following Fleischer *et al.*, (2019), with present-day CO_2 based on global monitoring stations, and an abrupt (step) increase in atmospheric CO_2 of +200 ppm on the onset of the transient period (i.e., 1999). However, the comparison period is limited to 2017-18 for which the P measurements are available.

We compare simulated C fluxes (GPP, NPP, litterfall C), C stocks (total vegetation, fine root, leaf, wood, soil) and the CO_2 fertilization effect across model configurations. The CO_2 fertilization effect ($\text{CO}_{2\text{fert-eff}}$) (eq.51) is calculated based on simulated vegetation C under ambient ($\text{VegC}(a\text{CO}_2)$) and $e\text{CO}_2$ ($\text{VegC}(e\text{CO}_2)$) as follows:

$$\text{CO}_{2\text{fert-eff}} = \frac{(\text{VegC}(e\text{CO}_2) - \text{VegC}(a\text{CO}_2)) \times 100}{\text{VegC}(a\text{CO}_2)} \quad (\text{eq.51})$$

Furthermore, the net biomass increases due to CO_2 fertilization effect (ΔCveg) is estimated as follows:

607
608 $\Delta C_{veg} = \Delta BP - \Delta litterfall C$ (eq.52)
609

610 We studied the Water Use Efficiency (WUE) (eq. 53) at half-hourly timestep, then aggregated per month as one
611 of the main indicators of GPP changes (Xiao *et al.*, 2013), and soil moisture content (SMCL), as one of the
612 main controllers of maximum uptake capacity (eq. 27), in order to better understanding the changes in GPP, P
613 demand and uptake as well as excess C fluxes.

614
615 $WUE = GPP/Transpiration$ (eq.53)
616

617 Moreover, we also estimated the Carbon Use Efficiency (CUE) as an indicator of the required C for the growth
618 (Bradford and Crowther, 2013) as follows:

619
620 $CUE = BP/GPP$ (eq.54)
621

622 We use JULES-CNP to evaluate the extent of P limitation under ambient and eCO₂ at this rainforest site in
623 Central Amazon. P limitation is represented by the amount of C that is not used to grow new plant tissue due to
624 insufficient P in the system (excess C) (eq. 27). The excess C flux is highly dependent on the plant P and the
625 overall P availability to satisfy demand. We also explore the distribution of the inorganic and organic soil P and
626 their sorbed fraction within the soil layers and under ambient and eCO₂.

627 628 2.5.1 Model sensitivity

629
630 To test the sensitivity of the P and C related processes to [individual](#) model P parameters, six sets of simulations
631 were conducted [independently](#) with modified plant C:P stoichiometry (Plant C:P: *SENS1*), P uptake scaling
632 factor (K_P) (K_P : *SENS2*), inorganic ($K_{P_sorb_in}$: *SENS3*) and organic ($K_{P_sorb_or}$: *SENS4*) P adsorption
633 coefficients ($K_{sorp-or}$, $K_{sorp-in}$), and maximum inorganic ($K_{P_sorb_in_max}$: *SENS5*) and organic
634 ($K_{P_sorb_or_max}$: *SENS6*) sorbed P (K_{or-max} , K_{in-max}). These values were prescribed to vary between $\pm 50\%$
635 of the observed values and their effect on C pools (plant and soil C) and fluxes (NPP and excess C), and P pools
636 (plant, soil, and soil sorbed P) was assessed. As the derived model parameters from measurements have their
637 own level of uncertainty, we took 50% change to test these parameters at reasonable degree. However, the
638 occluded and weathered P pools are prescribed for this model application, the occluded and weather P
639 coefficients (other two P-related model parameters) were not part of sensitivity tests.

640
641 Our model evaluation period is limited to years 2017-18 due to the P measurement availability. However, in
642 order to [compare](#) with 15 models studied by Fleischer *et al.*, (2019) we also studied the response of GPP, NPP
643 and BP to eCO₂ for both initial (1999) and 15 years periods (between 1999-2013).

644 645 646 2.5.1 Simulations at test sites

647
648 [To perform JULES \(C, CN, CNP\) simulations at test sites we extracted the meteorological input data to drive](#)
649 [the model from a global dataset \(CRU-NCEP\)\(Harris *et al.*, 2014\) by selecting the closest grid cell to each site](#)
650 [when data were not available for a given site \(Table 3\). Soil texture ancillaries for each site were extracted from](#)
651 [a global soil data \(HWSD\)\(Nachtergaele *et al.*, 2010\). All simulations were initialised from a global JULES-CN](#)
652 [run \(Wiltshire *et al.*, 2020\) extracted for each site and further spun-up for 2000 years over the 1980-2000 period](#)
653 [for the three versions of JULES \(C/CN/CNP\). Finally, the transient \(2000-2013\) run was performed using the](#)
654 [output of the spin-up for each site.](#)

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664 3. Results

665

666 3.1 Model application under ambient CO₂

667

668 3.1.1 Calibration of simulated soil P pools [at study site](#)

669

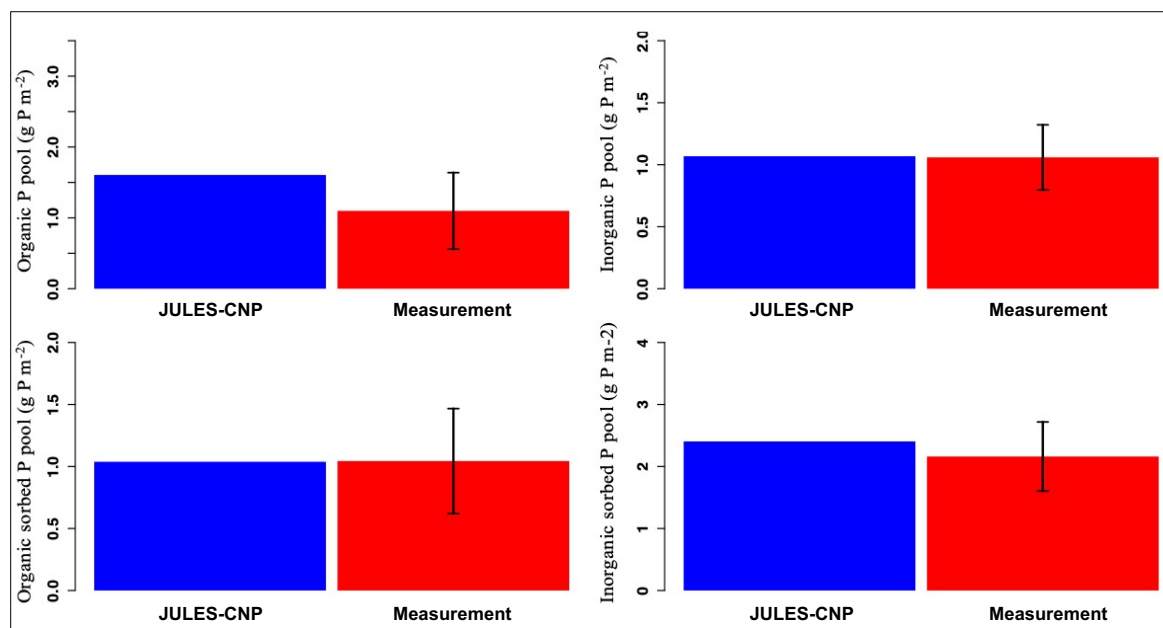
670 The maximum sorption capacities (P_{in-max_n} , P_{or-max_n} , eq.37 and 40) were calibrated to the observed P pools.

671 As a result, JULES-CNP could reproduce the measured soil P pools (Fig. 2 and Table 6). Simulated inorganic

672 soil P and sorbed organic and inorganic soil P closely matched the observations (Table 7 and Fig. 2). However,

673 simulated organic soil P overestimates the observations by 60 %.

674



675

676 **Figure. 2-** Modelled vs measured soil phosphorus pools under ambient CO₂ (for the soil depth of 0-30cm). Black line
677 represents standard deviation

678

679

680 **Table 6.** Observed and simulated phosphorus pools and fluxes. Occluded and weathered P pools were prescribed using the
681 observed values (between period 2017-18).

	Phosphorus pools and fluxes		
	Measured	Modelled Ambient CO ₂	Modelled Elevated CO ₂
Organic P (g P m ⁻²)	1.09±0.53	1.6	1.57
Inorganic P (g P m ⁻²)	1.05±0.33	1.07	0.96
Sorbed organic P (g P m ⁻²)	1.04±0.42	1.04	1.03
Sorbed inorganic P (g P m ⁻²)	2.1±0.55	2.4	2.4
Occluded P (g P m ⁻²)	7.98±2.38	prescribed	prescribed
Weathered P (g P m ⁻²)	0.59±12	prescribed	prescribed
Total vegetation P (g P m ⁻²)	4.15	4.66	5.11
Soil P – 30cm (g P m ⁻²)	13.85	14.7	14.56
Total ecosystem P (g P m ⁻²)	-	35.97	35.97
P litter flux (g P m ⁻² yr ⁻¹)	0.3	0.28	0.29

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686

687 3.1.2 Model evaluation

688
 689 JULES_CNP could reproduce the plant and soil C (Figure 2 and Table 7) and N pools and fluxes (Figure S6 and
 690 Table 8) under ambient CO₂. Our results show that simulated GPP, is within the range of measurement (3.02 kg
 691 C m⁻² yr⁻¹ model vs 3-3.5 kg C m⁻² yr⁻¹ observed, respectively, Table 7).

692
 693 Simulated NPP, is close to the measured values (NPP: 1.14 - 1.31 observed vs 1.26 modelled kg C m⁻² yr⁻¹) with
 694 autotrophic respiration (RESP) also closely following the observations (1.98 observed vs 1.81 modelled kg C m⁻²
 695 yr⁻¹). Biomass production is estimated as a difference between NPP and the amount of C which is not fixed by
 696 plants due to the insufficient P in the system (excess C) (eq. 27). The excess C flux [depends](#) on the plant P and
 697 the overall P availability to satisfy demand (Table 7). [The](#) simulated flux of excess C is 0.3 kg C m⁻² yr⁻¹ under
 698 ambient CO₂. In JULES-CNP this flux is subtracted from NPP in order to give the BP (eq. 17) (Table 7). Our
 699 simulated litterfall overestimates the observations by 32%, however simulated vegetation and its components
 700 (fine root, leaf and wood) and soil C stocks match well the observations (Table 7).

701
 702 **Table 7.** Observed and simulated carbon pools and fluxes with JULES-CNP (between period 2017-18)

	Measured	Modelled Ambient CO ₂	Modelled Elevated CO ₂
GPP (kg C m ⁻² yr ⁻¹)	3.0 – 3.5	3.06	3.9
NPP_{pot} (kg C m ⁻² yr ⁻¹)	-	1.27	1.77
Plant respiration (kg C m ⁻² yr ⁻¹)	1.98	1.78	2.12
Excess C flux (kg C m ⁻² yr ⁻¹)	-	0.30	0.81
Biomass Production (kg C m ⁻² yr ⁻¹)	1.14±0.12	0.96	0.94
Litter C flux (kg C m ⁻² yr ⁻¹)	0.69±0.15	0.91	0.83
Leaf C (kg C m ⁻²)	0.37±0.2	0.38	0.40
Wood C (kg C m ⁻²)	22.01	22.4	24.71
Root C (kg C m ⁻²)	0.37±0.2	0.38	0.40
Vegetation C (kg C m ⁻²)	22.75±0.3	23.16	25.52
Soil C stock (kg C m ⁻²)	12.7	13.2	12.71
LAI (m ² m ⁻²)	5.6±0.36	5.77	6.12

703 704 3.1.3 Comparison of JULES C, CN and CNP under ambient CO₂ [at study site](#)

705
 706 We compare simulated C pools and fluxes from JULES-C, JULES-CN and JULES-CNP (Figure 3). There is no
 707 difference between C stocks and fluxes in simulations from JULES C and CN indicating that there is no N
 708 limitation at this tropical site in the CN simulations. However, simulated BP and litter flux of C by JULES
 709 C/CN are higher than in JULES-CNP but also overestimate the observations (litter flux of JULES C/CN: 1.18,
 710 JULES-CNP: 0.91 and obs 0.69 (kg C m⁻² yr⁻¹) and BP of JULES C/CN: 1.24, JULES-CNP: 0.96 and obs 1.14-
 711 1.31 (kg C m⁻² yr⁻¹), respectively). By including P cycling in JULES an excess C flux of 0.3 (kg C m⁻² yr⁻¹) is
 712 simulated, indicating a 24% P limitation to BP at this site according to JULES-CNP, which represents a 29%
 713 decrease in BP compared to JULES-C/CN. Consequently, the total vegetation C stock for models without P
 714 inclusion is higher than the CNP version (+3% difference) due to the lack of representation of P limitation. The
 715 simulated soil C stock in JULES C and JULES CN is also higher than in the CNP version (JULES C/CN: 13.93
 716 vs. JULES-CNP: 13.18 (kg C m⁻² yr⁻¹)) and higher than the observations. Moreover, CUE in JULES C/CN
 717 (eq.54) is higher than observations and JULES_CNP (JULES C/CN: 0.38 vs. JULES-CNP: 0.31, obs: 0.34
 718 ±0.1(dimensionless)).

719
 720

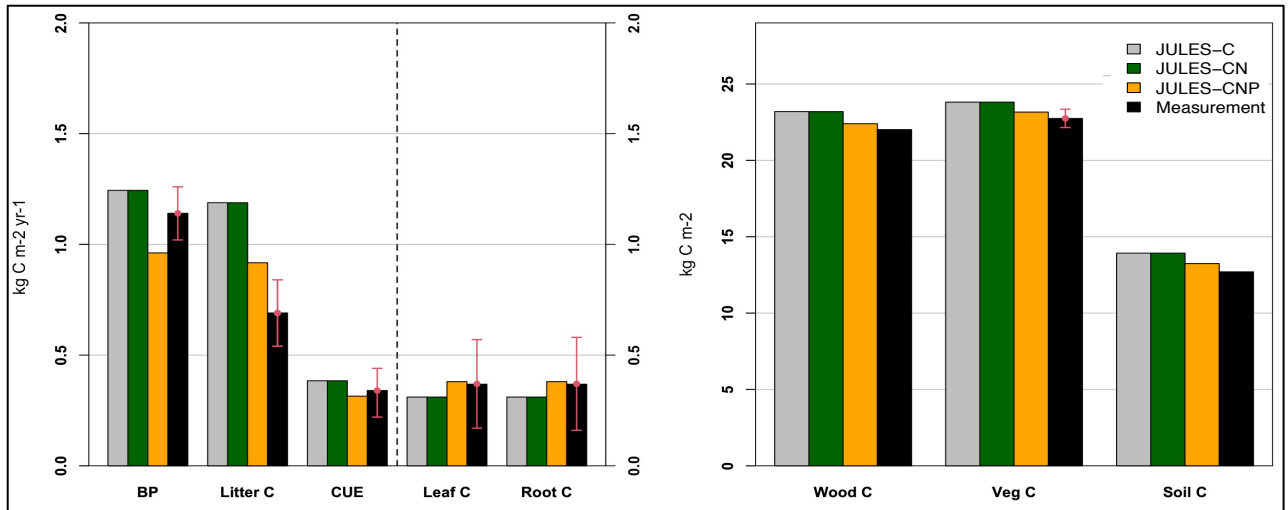


Figure 3- JULES C, CN, CNP modelled vs measured C pools (Leaf, root, wood, Veg and Soil C) (in kg C m⁻²) and fluxes (BP and Litter C) (in kg C m⁻² yr⁻¹) and CUE under ambient CO₂. Note that CUE is unitless.

3.1.4 Model evaluation at test sites under ambient CO₂

Evaluation of JULES C, CN and CNP at five test sites against the observed C pools and fluxes demonstrate that the inclusion of P processes improved the simulation of C pools and fluxes across all test sites (Figure 4). At all Amazon sites JULES C and CN overestimated BP compared to JULES-CNP which estimated lower BP values which were closer to the measurements for AGP (JULES-C: +35%, JULES-CN: +33%; JULES-CNP: +21%), CAX (JULES-C: +45%, JULES-CN: +44%; JULES-CNP: +7%) and SA3 (JULES-C: +27%, JULES-CN: +26%; JULES-CNP: -23%). Moreover, at Gigante Peninsula the C and CN versions overestimated BP (+42% and +40%, respectively), and CNP slightly underestimated BP (-15%). Furthermore, at the Hawaii Kokee site, all three versions of JULES underestimated the BP (C:-8%, CN:-8%, CNP: -32%). The litterfall and respiration fluxes in JULES-CNP have decreased compared to the JULES C and CN versions which overestimated both fluxes at all the test sites compared to the measurements. The litterfall flux comparisons show a significant overestimation using JULES C and CN versions across all the tested sites. Along the Amazon sites inclusion of P limitation reduced the litterfall flux but still overestimated (AGP: +50%, CAX: +24% and SA3: +16%) and at Gigante Peninsula and Hawaii Kokee slightly underestimated (Gigante Peninsula: -9% and Hawaii Kokee -19%). The respiration measurements were only available at two Amazon sites (CAX and SA3) at which inclusion of P limitation resulted in a well estimated flux at both sites compared to the JULES C/CN versions (CAX site: C-only: +38%, CN: +38%, CNP: -1%; SA3 site: C-only: +38%, CN: +38%, CNP: -2%). The total vegetation biomass also reduced using JULES-CNP compared to the other versions and yield closer values to the measurements across all the sites. However, except at the AGP site in which all three versions of JULES slightly underestimated the biomass (C: -1%, CN: -1%, CNP: -6%), at the other test sites JULES-CNP estimated lower biomass pools compared to the other versions which overestimated total vegetation biomass. Similarly, the soil C pool was overestimated prior to P limitation inclusion in JULES at the test sites, and the JULES-CNP estimated a closer value compared to the measurements (slight underestimation at CAX and SA3 sites: -5% and -18% respectively, and close values at Gigante Peninsula and Hawaii Kokee: +3% and +4%, respectively).

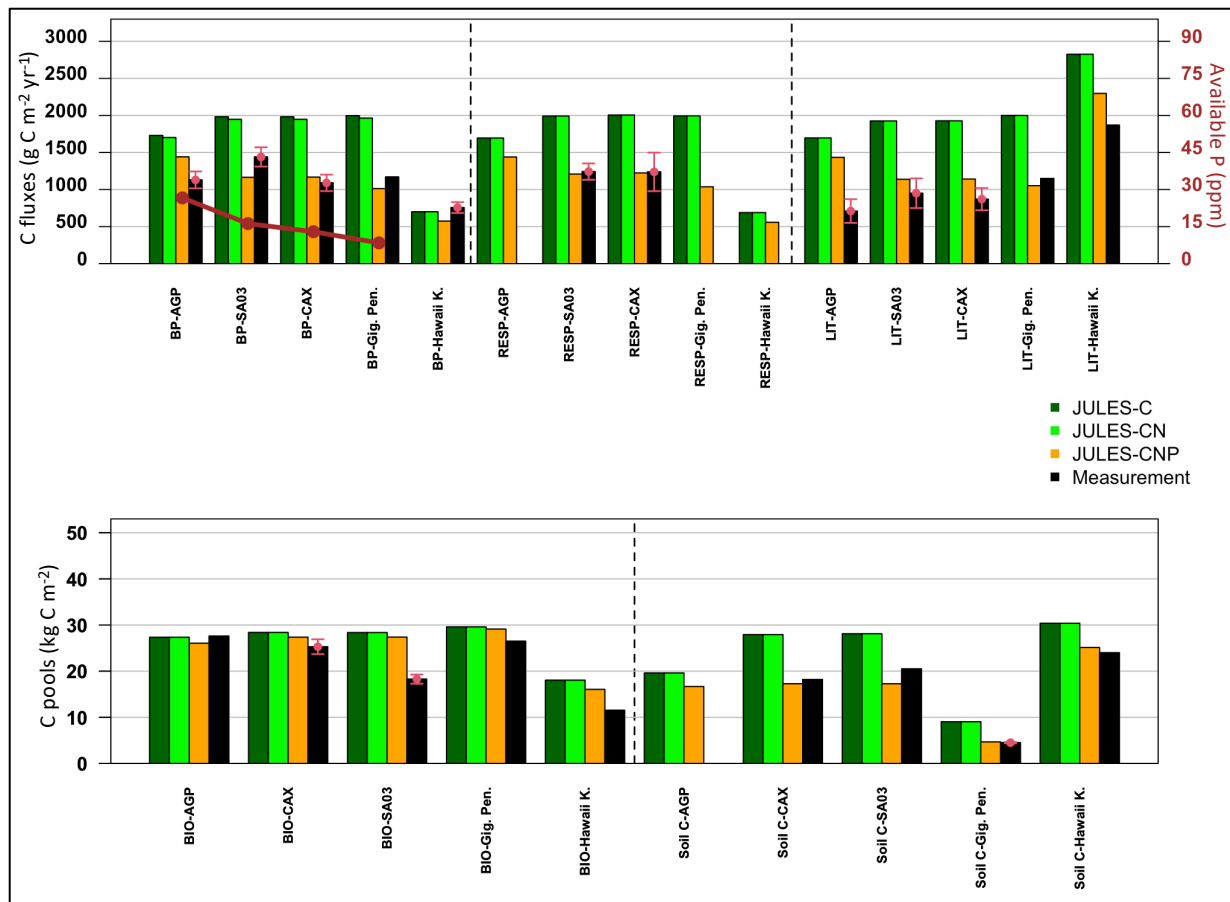
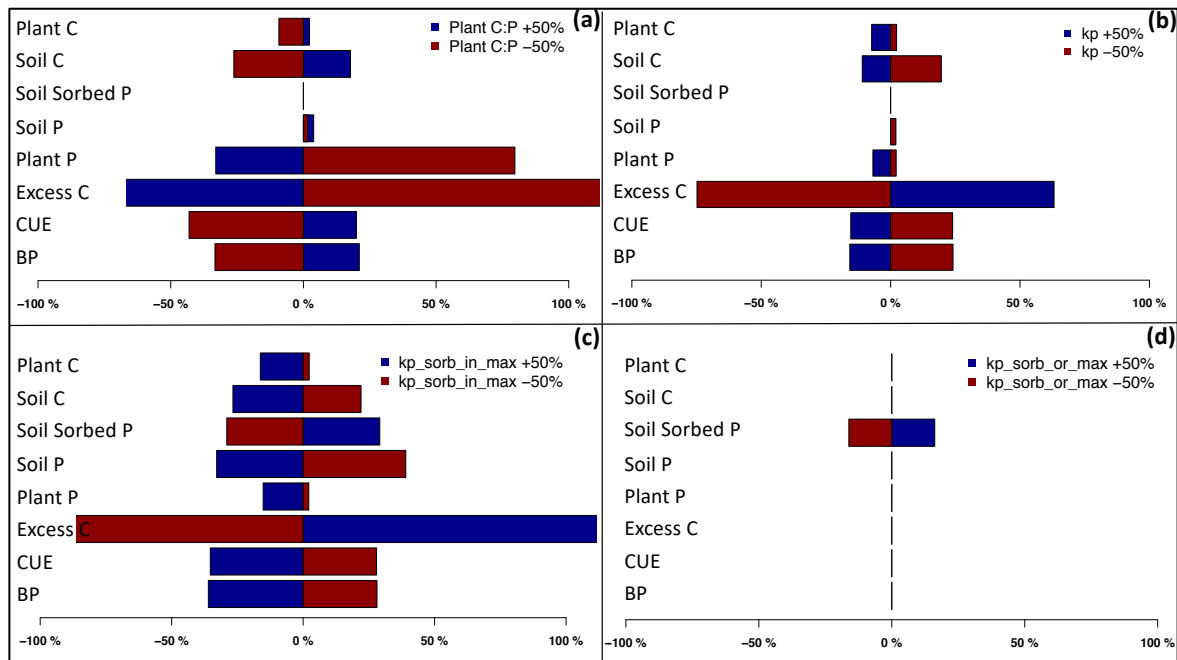


Figure 4- Observed and simulated (JULES C, CN, CNP) C fluxes and pools (averaged measurements: red points, sd: red arrows) and available observed P (dark red points and lines (reported in ppm)) at test sites across the Amazon (AGP, SA03, CAX), Gigante Peninsula (Gig. Pen.) and Hawaii Kokee (Hawaii K.).

3.1.5 Model sensitivity

The results indicate that among all the corresponding C and P pools and fluxes, the excess C flux – which demonstrates P limitation to growth – shows the highest sensitivity to changes in C:P ratios (Figure 5-a), K_p (Figure 5-b), and K_{or-max} (Figure 5-c) and K_{in-max} (Figure 5-d). A decrease in plant C:P results in a large increase in excess C. This is due to the higher plant P demand as a result of lower plant C:P ratios. An increase in the uptake factor and maximum sorbed organic and inorganic P also results in an increase in excess C. This is due to the higher uptake demand through higher uptake capacity (due to higher K_p) and lower available P for uptake due to higher organic and inorganic sorbed P (due to higher K_{or-max} , K_{in-max}). Since the total P in the system is lower than the plant demand, the uptake capacity and sorbed P, higher P limitation is placed on growth (decreasing BP) which results in an increase in excess C and decrease in plant C, but also soil C which is a result of lower litter input (Figure 5). Total soil P shows low sensitivity to changes in plant C:P and uptake factor but high sensitivity to maximum inorganic sorbed P. Moreover, sorbed P shows middle to high sensitivity to maximum organic and inorganic sorbed P respectively (Figure. S5). Nevertheless, organic and inorganic P adsorption coefficients ($K_{sorp-or}$, $K_{sorp-in}$) show no sensitivity to C and P pools and fluxes. This is due to limiting the organic and inorganic P sorption terms to be controlled only by maximum sorption capacity, hence no effect applied by organic and inorganic adsorption coefficients.



776
777 **Figure 5.** Sensitivity of C and P pools to variation in key model parameters: prescribed tissue C:P (a), K_p (b),
778 $K_{p_sorb_in}$ (c), $K_{p_sorb_or}$ (d) under ambient CO_2 .
779

780
781 **3.2 Model application under elevated CO_2**

782
783 **3.2.1 Simulated plant and soil C and P pools and fluxes -JULES-CNP: eCO_2 vs ambient CO_2**

784
785 The eCO_2 simulation using JULES-CNP yields a higher GPP compared to the ambient CO_2 ($0.83 \text{ (kg C m}^{-2} \text{ yr}^{-1})$
786 increase), as a result of CO_2 fertilization. Moreover, due to the GPP increase, NPP and RESP also increased
787 compared to ambient CO_2 (NPP: 0.49 and RESP: $0.3 \text{ (kg C m}^{-2} \text{ yr}^{-1})$ increase) (Table 7). The total simulated
788 vegetation C pool increases under eCO_2 compared to ambient CO_2 (0.41 kg C m^{-2}), hence the estimated plant P
789 (estimated as a fraction of C:P ratios) increases as well ($+0.45 \text{ (g P m}^{-2})$) (Fig. 6, Table 6). Thus, the simulated
790 plant P demand is higher, and as the total available soil P for uptake is limited, the simulated excess C flux
791 increases to $0.51 \text{ (kg C m}^{-2} \text{ yr}^{-1})$. Moreover, despite the higher NPP under eCO_2 compared to simulated NPP
792 under ambient CO_2 , due to the substantial increase in simulated excess C, the BP is similar to the ambient CO_2
793 (2% difference).
794

795 The simulated organic soil P under eCO_2 were close to those under ambient CO_2 (1.6 g P m^{-2}) (Table 7). This is
796 due to the same parameterization of the output fluxes from this pool for eCO_2 and ambient CO_2 . The simulated
797 pool of inorganic P under eCO_2 decreases compared to the ambient CO_2 by $0.11 \text{ (g P m}^{-2})$ due to the increased
798 plant P pools and slight increase in uptake ($+0.13 \%$).

799 However, the simulated sorbed organic and inorganic soil P from eCO_2 are similar to those simulated under the
800 ambient CO_2 which is due to the same parameterization of sorption function (maximum sorption capacity) from
801 the ambient CO_2 run as explained in calibration section. Moreover, the modelled occluded and weathered soil P
802 were similar to those in the ambient CO_2 simulation (Table 7) which is due to the same prescribed observational
803 data that was used for this simulation.
804

805 **3.2.2 Comparison of JULES C, CN and CNP under elevated CO_2**

806
807 JULES C/CN show higher vegetation and soil C pools, BP and litter flux compared to JULES-CNP: (Table 8,
808 Figure. S2). Under eCO_2 , simulated NPP using JULES C-CN is 4.5% higher than JULES-CNP and the BP with
809 JULES- C/CN is 96.8% higher than in JULES-CNP which simulates an excess C flux of $0.81 \text{ (kg C m}^{-2} \text{ yr}^{-1})$
810 equivalent to 46% P limitation under eCO_2 . As a result of P limitation and eCO_2 , the simulated CO_2 fertilization
811 effect estimated based on changes in biomass under ambient and eCO_2 was reduced from 13% with JULES-
812 C/CN to 10% JULES-CNP. Moreover, the CUE from JULES C/CN is 87.5% higher than the JULES-CNP as a
813 result of high P limitation over biomass production.
814

815 **Table 8.** Simulated C pools and fluxes with JULES C/CN and difference in percentage with JULES-CNP model under
 816 eCO₂. A positive % means larger respective values simulated with JULES C and JULES CN than with JULES-CNP
 817 (between period 2017-18).

	GPP	NPP	BP	CUE	Litter C	Leaf C	Root C	Wood C	Soil C
JULES C/CN	4.1	1.85	1.85	45%	1.77	0.42	0.42	26.1	19.2
JULES-CNP	3.9	1.77	0.94	24%	0.83	0.4	0.4	24.71	12.71
ΔC/CN: CNP	5.1%	4.5%	96.8%	87.5%	113.3%	5%	5%	5%	51.1%

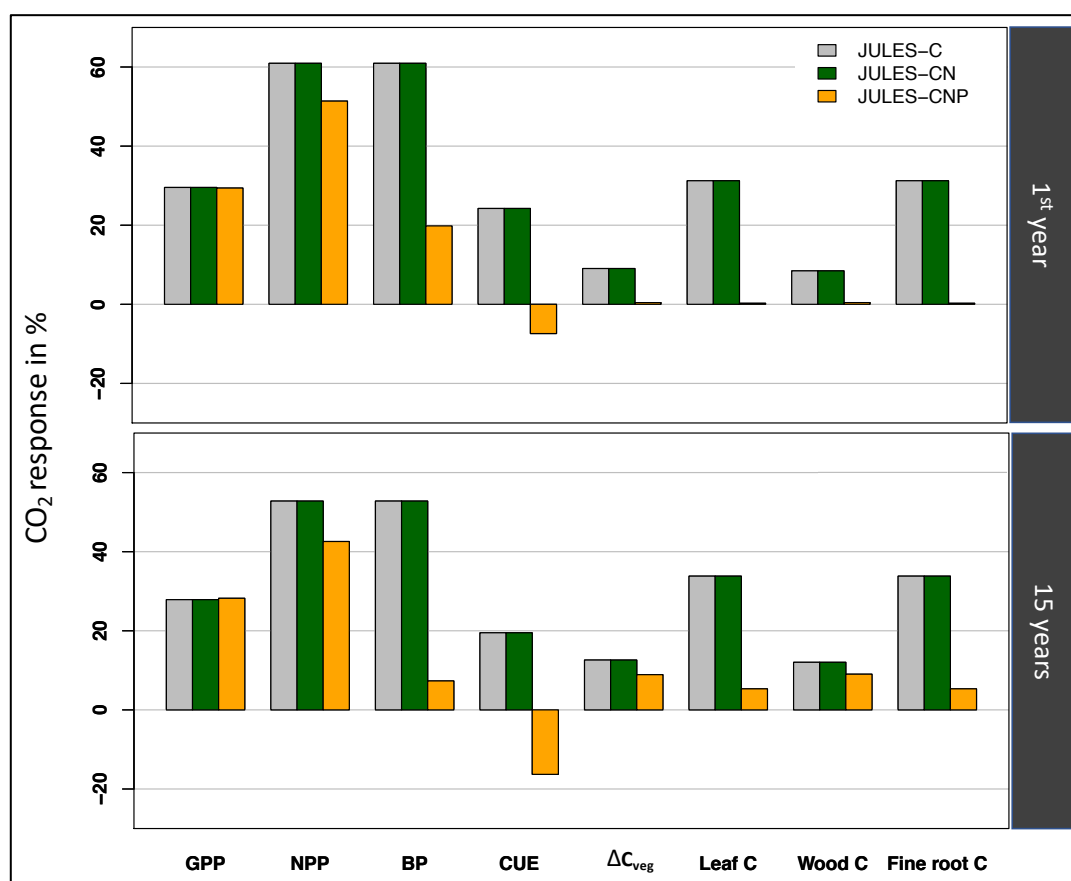
818
 819 **3.2.2.1 Inter-models under elevated CO₂**
 820

821 Following Fleischer *et al.*, (2019), we report the simulated response to eCO₂ for year 1999 (initial: CO₂ effect)
 822 and 1999-2013 (15 years: final effect) which are different than our evaluation period (2017-18). Using JULES-C
 823 and JULES-CN under eCO₂, simulated GPP and NPP during the 1st year increase by 30% and 61% respectively
 824 and by 28% and 52% after 15 years (Figure. 6). However, using JULES-CNP, eCO₂ increases simulated GPP,
 825 NPP and BP responses during the 1st year by 29%, 51% and 20% and by 28%, 43% and 7%, after 15 years,
 826 respectively.

827
 828 Corresponding simulated CUE during the 1st year and 15 years shows an increase of 24% and 20% in response
 829 to eCO₂ using JULES C/CN, respectively. However, using JULES-CNP, simulated CUE for the 1st and after 15
 830 years is reduced by 7% and 17% in response to eCO₂.

831
 832 Simulated total biomass (leaf, fine root and wood C) (ΔC_{veg}) using JULES-C/CN for the 1st and 15 years of
 833 eCO₂ increased by 9% and 13% respectively. However, using JULES-CNP ΔC_{veg} only increases by 0.5% and
 834 9% for 1st and 15 years of eCO₂, respectively.

835
 836



837 **Figure. 6-** Relative effect of eCO₂ on simulated GPP, NPP, BP, CUE, ΔC_{veg}, leaf C, wood C and fine root C, using three
 838 versions of JULES model in 1st (initial response) and 15 years periods (final response).
 839
 840
 841
 842

843 **3.3 Plant P Demand, uptake and excess C under ambient and elevated CO₂**

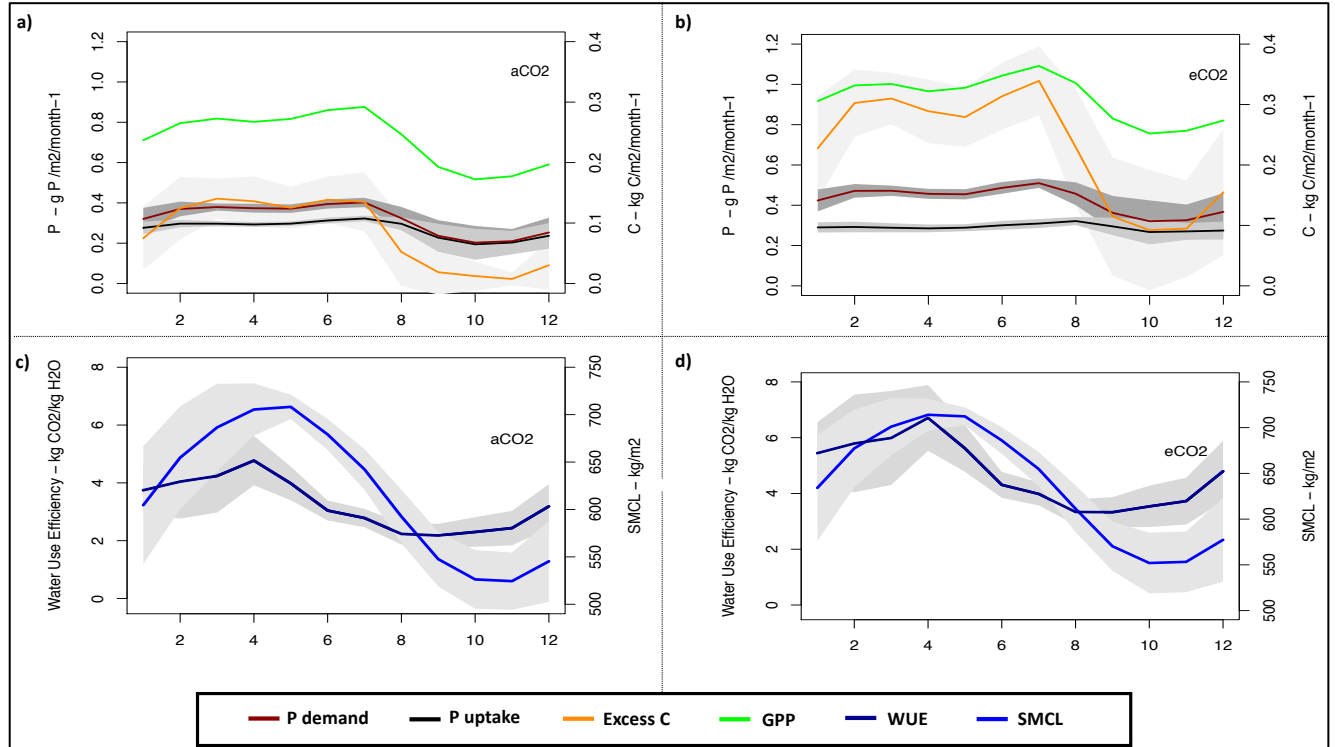
844
845 To understand further the CP-cycle dynamics, we studied the monthly averaged plant P demand and the relative
846 (limited) P uptake (eq. 26) under both ambient and elevated CO₂ conditions (Figure. 7).

847
848 Under ambient CO₂ condition the highest GPP is estimated at $0.29 \pm 0.016 \text{ kg C m}^{-2} \text{ month}^{-1}$ in July and the
849 lowest at $0.17 \pm 0.051 \text{ kg C m}^{-2} \text{ month}^{-1}$ in October (Figure. 7-a). The estimated WUE and SMCL in October is
850 among the lowest estimated monthly values at $2.3 \pm 0.51 \text{ kg CO}_2/\text{kg H}_2\text{O}$ and $526.2 \pm 31 \text{ kg m}^{-2}$ respectively
851 (Figure. 7-c). The highest P demand is estimated at $0.4 \pm 0.02 \text{ g P m}^{-2} \text{ month}^{-1}$ in July and the lowest demand at
852 $0.2 \pm 0.08 \text{ g P m}^{-2} \text{ month}^{-1}$ in October. Consequently, the highest and lowest uptake (0.32 ± 0.01 and $0.19 \pm 0.07 \text{ g P}$
853 $\text{m}^{-2} \text{ month}^{-1}$, respectively). The excess C for the highest and lowest GPP and demand periods are estimated at
854 0.4 ± 0.15 and $0.04 \pm 0.07 \text{ kg C m}^{-2} \text{ month}^{-1}$, respectively.

855
856 However, similar to ambient CO₂, under eCO₂ condition the highest estimated GPP is in July at $0.36 \pm 0.017 \text{ kg}$
857 $\text{C m}^{-2} \text{ month}^{-1}$ and lowest for October $0.25 \pm 0.062 \text{ kg C m}^{-2} \text{ month}^{-1}$ (Figure. 7-b). The estimated WUE and soil
858 moisture content (SMCL) for the lowest GPP period is among the lowest monthly estimated values at 3.5 ± 0.74
859 $\text{kg CO}_2/\text{kg H}_2\text{O}$ and $552 \pm 33 \text{ kg m}^{-2}$ for October respectively (Figure. 7-d). The highest P demand is estimated
860 for July at $0.51 \pm 0.02 \text{ g P m}^{-2} \text{ month}^{-1}$ with the uptake flux of $0.31 \pm 0.02 \text{ g P m}^{-2} \text{ month}^{-1}$ and the lowest demand
861 is estimated for October at $0.32 \pm 0.1 \text{ g P m}^{-2} \text{ month}^{-1}$ with the estimated uptake flux of $0.26 \pm 0.06 \text{ g P m}^{-2} \text{ month}^{-1}$
862 ¹. The highest excess C flux is also for July at $1.01 \pm 0.17 \text{ kg C m}^{-2} \text{ month}^{-1}$ and lowest for October $0.27 \pm 0.29 \text{ kg}$
863 $\text{C m}^{-2} \text{ month}^{-1}$, respectively.

864
865 However, despite the P limitation in both eCO₂ and ambient CO₂ conditions, the P uptake flux under eCO₂ is
866 higher than the ambient CO₂ condition. This is due to the higher WUE and increased SMCL (controlling uptake
867 capacity (eq. 27)) under eCO₂ condition, hence more water availability during the dry season to maintain
868 productivity and critically transport P to the plant (see eq. 27), compared to ambient CO₂ condition (Figure. 7-c
869 and d). Additionally, in JULES both the vertical discretisation (Burke, Chadburn and Ekici, 2017) and
870 mineralisation terms (Wiltshire *et al.*, 2021) depend on the soil moisture and temperature. Thus, higher P
871 concentration and uptake under eCO₂ condition.

872



873
874 **Figure. 7-** Simulated monthly plant P demand and uptake ($\text{g P m}^{-2} \text{ month}^{-1}$), excess C and GPP ($\text{kg C m}^{-2} \text{ month}^{-1}$) under a)
875 aCO₂ and b) eCO₂, water use efficiency ($\text{g m}^{-2} \text{ month}^{-1}$) under c) ambient CO₂ (aCO₂) and d) eCO₂ conditions. The grey area
876 represents the standard deviation.

877
878

3.4 Soil P pools profile under ambient CO₂ and elevated CO₂

We explored the distribution of the inorganic and organic soil P and their sorbed fraction within the soil layers and under different CO₂ conditions (Figure. S3). Both the ambient and eCO₂ simulations have a close inorganic soil P distribution at the topsoil layer (0-30cm) (0.85 vs. 0.9 (g P m⁻²) respectively) as well as similar organic soil P distribution (0.85 vs 0.9 (g P m⁻²) respectively).

However, the organic soil P and sorbed forms of inorganic and organic soil P profiles are not changing significantly between different sets due to the similar parameterization of the processes that control these pools (processes which are related to the physical aspects of soils, hence not changing under eCO₂ condition) and the same parameter values used for both ambient and eCO₂ runs.

Moreover, the soil P within 30cm soil depth for ambient and eCO₂ conditions is at 14.7 (g P m⁻²) and 14.56 (g P m⁻²) respectively, and the total ecosystem P for both ambient and eCO₂ conditions is at 35.97 (g P m⁻²).

However, the slightly lower soil P in the eCO₂ condition is due to the higher plant P demand compared to the ambient condition, hence the higher allocated P vegetation (10%) under eCO₂ condition.

4. Discussion

Studies show the significant role of the tropical forests, and Amazonia in particular, in C uptake and regulating atmospheric CO₂ (Brienen *et al.*, 2015; Phillips *et al.*, 2017). As soil P availability is low in the majority of Amazonia (Quesada *et al.*, 2012), the competition for nutrients by both plant and soil communities is high (Lloyd *et al.*, 2001). The responses of these communities to eCO₂ under P limited conditions remains uncertain (Fleischer *et al.*, 2019). These responses in P enabled models are represented in different ways regarding the excess C which is not used for plant growth due to P limitation. Either growth is directly downregulated taking the minimum labile plant C, N and P (Goll *et al.*, 2017), or photosynthesis is downregulated via V_{cmax} and J_{max} (Comins and McMurtrie, 1993; Yang *et al.*, 2014; Zhu *et al.*, 2016) and models like JULES-CNP downregulate NPP via respiration of excess carbon that cannot be used for growth due to plant nutrient constraints (Haverd *et al.*, 2018). The estimated CUE depends on the modelling approach. Models that down regulate the photosynthetic capacity and GPP consequently (Comins and McMurtrie, 1993; Yang *et al.*, 2014; Zhu *et al.*, 2016) simulate a positive CUE response to CO₂ fertilization while models that down regulate the NPP and respire the excess C (Haverd *et al.*, 2018) simulate a negative CUE response (Fleischer *et al.*, 2019) which is in line with field studies showing lower CUE when nutrient availability declines (Vicca *et al.*, 2012b). However, this remains a major uncertainty in understanding the implication of P limitation on terrestrial biogeochemical cycles.

[The JULES-CNP structure represent key](#) P processes in both plant and soil pools and can be applied to the Amazon region using existing soil (Quesada *et al.*, 2011) and foliar structural and nutrient (Fyllas *et al.*, 2009) data for parameterisation. [The model can be applied globally and under](#) future [climate](#) projections [using](#) global soil P data (Sun *et al.*, 2021) for model initialization and PFT-specific plant (Zechmeister-Boltenstern *et al.*, 2015) [and](#) soil stoichiometries (Zechmeister-Boltenstern *et al.* 2015; Tipping *et al.* (2016), sorption and weathering ratios (based on lithological class specific from the GLIM lithological map (Hartmann and Moosdorf, 2012) and soil shielding from Hartmann *et al.*, (2014)).

4.1. Evaluation of model performance

[At the study site](#), JULES-CNP could reproduce the magnitude of soil organic and inorganic P pools and fluxes. The relative distribution of total organic P, total inorganic P and residue P fractions of total P in soils under Brazilian Eucalyptus plantations (Costa *et al.*, 2016) shows inorganic P fraction of 28% from total soil P which is close to our estimation of 24% and organic P fraction of 30% from total soil P which is higher than our estimated fraction of 18%. Thus, we may need to improve the process representation or parameters that control the organic P concentration, such as litter flux and decomposition, soil organic P mineralization, and immobilization in the future.

Our estimated maximum P uptake, which represents the actual available P for plant uptake (Goll *et al.*, 2017), for both ambient and eCO₂, is highly correlated with the plant P demand ($R^2 = 0.96$ and 0.52 respectively). The plant P demand depends on the GPP changes which are reflected by the WUE (Hatfield and Dold, 2019). Hence, under ambient CO₂, JULES-CNP simulates lower GPP and plant P demand during the dry season than during the wet season. Sufficient P uptake during these periods results in the lowest P limitation, thus the lowest simulated excess C. Nevertheless, under eCO₂ the same pattern is simulated but a higher availability of soil P due to the stomatal closure in the dry season. Hence, due to the plant's more efficient water usage, the soil

939 moisture in the dry season is higher (Xu *et al.*, 2016) which impacts our capped P uptake flux (eq. 27) and
940 increases the uptake capacity respectively.

941
942 Overall, JULES-CNP reproduced the observed C pools and fluxes which are in the acceptable ranges compared
943 to the measurements. However, using the JULES default V_{cmax} estimation method (eq. 40), the model slightly
944 underestimates the total GPP (2.9 kg C m⁻² yr⁻¹ vs. 3-3.5 kg C m⁻² yr⁻¹). Therefore, in this version of the model,
945 we used the improved V_{cmax} estimation method based on N and P (eq. 46) which resulted [in](#) a final estimated
946 GPP closer to the measurements (3.06 kg C m⁻² yr⁻¹).

947
948 Our results show an increase in GPP (21%) in response to eCO₂ which is higher than the average increase of
949 GPP reported in mature eucalyptus forests (11%), also growing under low P soils at the free air CO₂ enrichment
950 experiment (EucFACE) facility in Australia (Jiang *et al.*, 2020). This can be related to the lower decrease of
951 biomass growth response estimated by JULES-CNP (-3%) compared to the measurements from mature
952 [eucalyptus](#) forests (-8%) (Ellsworth *et al.*, 2017), due to the P limitation which [was shown](#) to impact the above-
953 ground biomass growth response in mature forests (Körner *et al.*, 2005; Ryan, 2013; Klein *et al.*, 2016).

954
955 In order to estimate the biomass production (BP), we deducted the excess C fluxes from NPP. Using JULES
956 C/CN models, [the simulated](#) biomass productivity enhancement due to eCO₂ (49%) is in the middle range of the
957 reported [for](#) different biomes by Walker *et al.*, (2021). Moreover, our estimated difference of BP between
958 ambient and eCO₂ conditions (2%) is close to the estimated difference for mature forests (3%) (Jiang *et al.*,
959 2020).

960
961 A global estimation for tropical forests using [the](#) CASA-CNP model which includes N and P limitations on
962 terrestrial C cycling, shows that NPP is reduced by 20% on average due to the insufficient P availability (Wang,
963 Law and Pak, 2010) which is close to our estimated P limitation of 24%. This finding is in line with [a field](#) study
964 that shows a strong correlation between the total NPP and the soil available P (Aragão *et al.*, 2009).
965 The estimated decrease of NPP in response to eCO₂ as a result of P limitation is in line with the findings from
966 CLM-CNP model at five tropical forests (Yang *et al.*, 2014) which indicates the CO₂ fertilization dependency
967 on the processes that affect P availability or uptake.

968
969 Our estimated CUE (0.31) is close to [that](#) by Jiang *et al.* (2020) for mature [eucalyptus](#) forests (0.31±0.03), as
970 well as to the measurement for our study site (0.34 ±0.1). There is currently a lack of representation of stand age
971 in JULES-CNP which can significantly [affect CUE](#) (e.g. mature trees are less responsive to the nutrient
972 limitations) (De Lucia *et al.*, 2007; Norby *et al.*, 2016). However, a recent development of Robust Ecosystem
973 Demography (RED) model in JULES (Argles *et al.*, 2020) and its integration into JULES-CNP in the future can
974 [address](#) this issue.

975
976 [Under](#) low P availability, all available P is considered to be adsorbed or taken by plant and microbes for further
977 consumption, with leaching considered to be minor within the time scales of our study period (Went and Stark,
978 1968; Bruijnzeel, 1991; Neff, Hobbie and Vitousek, 2000). [Despite studies that show the possibility of P](#)
979 [fixation as a source of available P for plants \(Van Langenhove *et al.*, 2020; Gross *et al.*, 2021\), due to the strong](#)
980 [fixation of P in the soil \(Aerts & Chapin, 2000; Goodale, Lajtha, Nadelhoffer, Boyer, & Jaworski, 2002\), the P](#)
981 [deposited is unlikely to be available to plants in the short term \(de Vries *et al.*, 2014\), for this reason this](#)
982 [version of JULES-CNP did not include P deposition](#). However both P deposition and leaching are likely to have
983 a very important role on sustaining the productivity of tropical forests in the Amazon over longer time scales
984 (Van Langenhove *et al.*, 2020) and needs to be considered in future studies. Moreover, biochemical
985 mineralisation is [also](#) not included in the current version of JULES-CNP [which](#) only accounts for total
986 mineralization. However, models [that](#) include this process show no significant difference between total and
987 biochemical mineralized P which can be due to complexity of identifying the inclination of mineralization
988 versus uptake (Martins *et al.*, 2021). [Lastly](#), in order to capture plant internal nutrient impact on the C storage,
989 future work should focus on [implementing](#) recent developments [including](#) Non-Structural Carbohydrate [pools](#)
990 (NSC) (Jones *et al.*, 2020) in JULES-CNP.

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4.1.1. Evaluation of model performance at test sites

Overall, inclusion of P processes in JULES-CNP improved the previously overestimated C fluxes and pools using JULES-C and -CN versions. Generally, the biomass productivity tends to follow the observed P availability (Figure 4), where the sites with higher available P for uptake simulated higher productivity which is in line with observations across P availability in the Amazon (Aragão *et al.*, 2009). Nevertheless, this tendency could be altered if the natural conditions in these forests are perturbed. For instance, in case of the high mortality events in these P limited sites (Malhi *et al.*, 2009; Pyle *et al.*, 2009), regrowing forests developing over the highly weathered oxisols with limited available P (Davidson *et al.*, 2004), results in the shifting limitation from P to N (Herbert, Williams and Rastetter, 2003). Hence, the controlling processes under N limitation will be N-related and processes such as N leaching or outgassing (Yang *et al.*, 2014) will define the productivity. This shifting in limitation condition is not represented by JULES-CNP, therefore at few tested sites the model overestimated the P limitation, thus underestimated the productivity below the measured values. Moreover, the higher (than other sites) BP in JULES C/CN at the the Gigante Peninsula is related to the higher solar radiation in the forcing data at this site (Figure S8).

The estimated litterfall and respiration fluxes were considerably lower with JULES-CNP than JULES-C and -CN due to the lower simulated NPP with the former in closer agreement with the observations at all sites. Consequently, the total vegetation and soil C pools have lower values under the P limitation (Malhi *et al.*, 2009), which could not be captured by JULES-C and -CN and successfully represented by JULES-CNP.

As shown in Figure 5, JULES-CNP is highly sensitive to the five parameters needed to run JULES-CNP in addition to JULES-C and JULES-CN which were prescribed for simulations at test sites. The successful model performance at these sites demonstrates the importance of these parameters in JULES-CNP with implications for global scale simulations.

4.2. Inter-models Comparison of JULES C, CN and CNP

The comparison of simulated GPP enhancement across JULES versions for the 1st year is within the middle range of the 1st year CO₂ responses of the C/CN models studied by Fleischer *et al.*, (2019) evaluating simulated eCO₂ effects at a site in Manaus using the same meteorological forcing and methodology used in this study for a range of DGVM's. However, comparison for 15 years of eCO₂, shows that the simulated response with JULES-CNP is on the higher end of Fleischer *et al.*, (2019) study which is due to the higher estimated biomass growth by JULES-CNP (Table S1). Similarly, using JULES-CNP our estimated GPP enhancement is on the higher end of model estimations in Fleischer *et al.*, (2019). Moreover, comparing the GPP responses between different versions of (JULES C/CN and CNP), the JULES-CNP shows a slightly higher response to CO₂ fertilization associated with the higher WUE changes (Xiao *et al.*, 2013) (Figure. S4). This is due to the higher sensitivity of the plant to water availability than P availability in the P limited system (He and Dijkstra, 2014). Hence, under eCO₂ due to water-saving strategy of plants and stomatal closure (Medlyn *et al.*, 2016), simulated transpiration is decreased (Sampaio *et al.*, 2021) and photosynthesis is enhanced compared ambient CO₂.

To that end, the monthly changes of WUE in JULES-CNP are highly correlated to the GPP, hence the lowest and highest WUE follow the same periods as GPP similar to responses captured with models studied by Fleischer *et al.*, (2019) (Table. S1).

Our estimated NPP enhancement using JULES C/CN models for both 1st and 15 years period is within the middle range of the models in Fleischer *et al.*, (2019). Nevertheless, JULES-CNP response of BP is in the lower band of the CNP models in Fleischer *et al.*, (2019) and close to the estimations from CABLE (Haverd *et al.*, 2018) and ORCHIDEE (Goll *et al.*, 2017) models, which may be due to the similar representation of P processes and limitation between these models. However, our results show a 29% decrease in NPP using JULES-CNP compared to JULES-C/CN which is smaller than the differences between the CLM-CNP and CLM-CN versions (51% decrease) (Yang *et al.*, 2014). The lower estimated decrease in JULES highlights the need to further study the fully corresponding plant C pools and fluxes to the changes in soil and plant P. Therefore, future work should be focused on the improvement of the total P availability and the plant C feedbacks. Moreover, there are other environmental factors such as temperature which shows a possible impact on the CO₂ elevation and the changes of NPP (Baig *et al.*, 2015) which needs further improvement in our model.

The CUE estimations of 1st year and 15 years response to CO₂ elevation from JULES C/CN are in the middle range of C/CN models in Fleischer *et al.*, (2019). However, the estimated CUE using JULES-CNP for 1st and 15 years are in the low range of CNP models reported by Fleischer *et al.*, (2019) which is due to the same reason discussed for NPP comparison.

1059 Finally, our estimated total biomass enhancement (ΔC_{veg}) using JULES C/CN for the 1st and 15 years are in the
1060 middle range of C/CN models from Fleischer *et al.*, (2019) and in lower range of CNP models from Fleischer *et al.*
1061 *et al.*, (2019) using JULES-CNP. Nevertheless, while JULES-CNP includes the trait-based parameters (Harper *et al.*
1062 *et al.*, 2016), other functions such as flexible C allocation and spatial variation of biomass turnover are still
1063 missing and future model improvement should be focused on their inclusion.

1064 1065 **5. Conclusion**

1066 Land ecosystems are a significant sink of atmospheric CO₂, ergo buffering the anthropogenic increase of this
1067 flux. While tropical forests contribute substantially to the global land C sink, observational studies show that a
1068 stalled increase in carbon gains over the recent decade (Brienen *et al.*, 2015; Hubau *et al.*, 2020). However
1069 modelling studies that lack representation of P cycling processes predict an increasing sink (Fernández-Martínez
1070 *et al.*, 2019; Fleischer *et al.*, 2019). This is particularly relevant for efforts to mitigate dangerous climate change
1071 and assumptions on the future efficacy of the land C sink. Therefore, in this study, we presented the full
1072 terrestrial P cycling and its feedback on the C cycle within the JULES framework. Our results show that the
1073 model is capable of representing plant and soil P pools and fluxes at a site in Central Amazon [and across the](#)
1074 [extended P limited test sites in Amazon, Gigante Peninsula and Hawaii chronosequence provided with site level](#)
1075 [data for model parameterisation](#). Moreover, the model estimated a significant NPP limitation under ambient
1076 CO₂, due to the high P deficiency at [these sites](#) which is representative of Central Amazon [and tropical P limited](#)
1077 [sites](#), and elevated CO₂ resulted in a further subsequent decrease in the land C sink capacity relative to the
1078 model without P limitation. While our study is a step toward the full nutrient cycling representation in ESMs, it
1079 can also help the empirical community to test different hypotheses (i.e., dynamic allocation and stoichiometry)
1080 and generate targeted experimental measurements (Medlyn *et al.*, 2015).

1081 1082 *Code availability*

1084 The modified version of JULES vn5_5 and the P extension developed for this paper are freely available on Met
1085 Office Science Repository Service:
1086 https://code.metoffice.gov.uk/svn/jules/main/branches/dev/mahdinakhavali/vn5.5_JULES_PM_NAKHAVALI/
1087 after registration (http://jules-lsm.github.io/access_req/JULES_access.html) and completion of software license
1088 form. Codes for compiling model available at: (<https://doi.org/10.5281/zenodo.5711160>). Simulations were
1089 conducted using two sets of model configurations (namelists): ambient CO₂ condition
1090 (<https://doi.org/10.5281/zenodo.5711144>) and elevated CO₂ condition
1091 (<https://doi.org/10.5281/zenodo.5711150>).

1092 *Data availability*

1093 The model outputs related to the results in this paper are provided on Zenodo repository
1094 (<https://doi.org/10.5281/zenodo.5710898>). All the R scripts used for processing the model outputs and
1095 producing results in form of table or figures are provided on Zenodo repository
1096 (<https://doi.org/10.5281/zenodo.5710896>).

1097 *Author contributions.* MAN, LMM, SS, SEC, CAQ, AJW, IAP, KMA and DBC developed the model, per-
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1099 measurements for the model parasitisation and evaluation. MAN, LMM, SS, IAP, SEC, FVC, RP, LFL, KMA
1100 and DBC contributed in writing the manuscript.

1101
1102 *Competing interests.* The authors declare no competing interests

1103
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1114 **References:**

- 1115
- 1116 Aerts, R. and Chapin, F. S. (1999) 'The Mineral Nutrition of Wild Plants Revisited: A Re-evaluation of
- 1117 Processes and Patterns', *Advances in Ecological Research*, 30(C), pp. 1–67. doi: 10.1016/S0065-
- 1118 2504(08)60016-1.
- 1119 Anav, A. *et al.* (2013) 'Evaluating the land and ocean components of the global carbon cycle in the CMIP5 earth
- 1120 system models', *Journal of Climate*, 26(18), pp. 6801–6843. doi: 10.1175/JCLI-D-12-00417.1.
- 1121 Aragão, L. E. O. C. *et al.* (2009) 'Above- and below-ground net primary productivity across ten Amazonian
- 1122 forests on contrasting soils', *Biogeosciences Discussions*, 6(1), pp. 2441–2488. doi: 10.5194/bgd-6-2441-2009.
- 1123 Araço, L. E. O. C. *et al.* (2009) 'Above- and below-ground net primary productivity across ten Amazonian
- 1124 forests on contrasting soils', *Biogeosciences*, 6(12), pp. 2759–2778. doi: 10.5194/bg-6-2759-2009.
- 1125 Araújo, A. C. *et al.* (2002) 'Comparative measurements of carbon dioxide fluxes from two nearby towers in a
- 1126 central Amazonian rainforest: The Manaus LBA site', *Journal of Geophysical Research*, 107(D20), p. 8090.
- 1127 doi: 10.1029/2001JD000676.
- 1128 Argles, A. P. K. *et al.* (2020) 'Robust Ecosystem Demography (RED version 1.0): A parsimonious approach to
- 1129 modelling vegetation dynamics in Earth system models', *Geoscientific Model Development*, 13(9), pp. 4067–
- 1130 4089. doi: 10.5194/gmd-13-4067-2020.
- 1131 Arora, V. K. *et al.* (2020) 'Carbon–concentration and carbon–climate feedbacks in CMIP6 models and their
- 1132 comparison to CMIP5 models', *Biogeosciences*, 17(16), pp. 4173–4222. doi: 10.5194/bg-17-4173-2020.
- 1133 Baig, S. *et al.* (2015) 'Does the growth response of woody plants to elevated CO₂ increase with temperature? A
- 1134 model-oriented meta-analysis', *Global Change Biology*, 21(12), pp. 4303–4319. doi: 10.1111/gcb.12962.
- 1135 Baker, T. R. *et al.* (2004) 'Variation in wood density determines spatial patterns in Amazonian forest biomass',
- 1136 *Global Change Biology*, 10(5), pp. 545–562. doi: 10.1111/j.1365-2486.2004.00751.x.
- 1137 Bentsen, M. *et al.* (2013) 'The Norwegian Earth System Model, NorESM1-M – Part 1: Description and basic
- 1138 evaluation of the physical climate', *Geoscientific Model Development*, 6(3), pp. 687–720. doi: 10.5194/gmd-6-
- 1139 687-2013.
- 1140 Best, M. J. *et al.* (2011) 'The Joint UK Land Environment Simulator (JULES), model description – Part 1:
- 1141 Carbon fluxes and vegetation dynamics', *Geoscientific Model Development*, 4(3), pp. 701–722. doi:
- 1142 10.5194/gmd-4-701-2011.
- 1143 Bradford, M. A. and Crowther, T. W. (2013) 'Carbon use efficiency and storage in terrestrial ecosystems', *New*
- 1144 *Phytologist*, 199(1), pp. 7–9. doi: 10.1111/nph.12334.
- 1145 Brienen, R. J. W. *et al.* (2015) 'Long-term decline of the Amazon carbon sink', *Nature*, 519(7543), pp. 344–
- 1146 348. doi: 10.1038/nature14283.
- 1147 Bruijnzeel, L. A. (1991) 'Nutrient input—output budgets of tropical forest ecosystems: A review', *Journal of*
- 1148 *Tropical Ecology*, 7(1), pp. 1–24. doi: 10.1017/S0266467400005010.
- 1149 Burke, E. J., Chadburn, S. E. and Ekici, A. (2017) 'A vertical representation of soil carbon in the JULES land
- 1150 surface scheme (vn4.3-permafrost) with a focus on permafrost regions', *Geoscientific Model Development*,
- 1151 10(2), pp. 959–975. doi: 10.5194/gmd-10-959-2017.
- 1152 Castanho, A. D. A. *et al.* (2013) 'Improving simulated Amazon forest biomass and productivity by including
- 1153 spatial variation in biophysical parameters', *Biogeosciences*, 10(4), pp. 2255–2272. doi: 10.5194/bg-10-2255-
- 1154 2013.
- 1155 Chapin, F. S. *et al.* (2011) *Principles of Terrestrial Ecosystem Ecology*. Springer New York (Biomedical and
- 1156 Life Sciences). Available at: <https://books.google.co.uk/books?id=68nFNpceRmIC>.
- 1157 Chave, J. *et al.* (2003) 'Spatial and temporal variation of biomass in a tropical forest: Results from a large
- 1158 census plot in Panama', *Journal of Ecology*, 91(2), pp. 240–252. doi: 10.1046/j.1365-2745.2003.00757.x.
- 1159 Chave, J. *et al.* (2014) 'Improved allometric models to estimate the aboveground biomass of tropical trees',
- 1160 *Global Change Biology*, 20(10), pp. 3177–3190. doi: 10.1111/gcb.12629.
- 1161 Clark, D. B. *et al.* (2011) 'The Joint UK Land Environment Simulator (JULES), model description – Part 2:
- 1162 Carbon fluxes and vegetation dynamics', *Geoscientific Model Development*, 4(3), pp. 701–722. doi:
- 1163 10.5194/gmd-4-701-2011.
- 1164 Collatz, G. J. *et al.* (1991) 'Physiological and environmental regulation of stomatal conductance, photosynthesis
- 1165 and transpiration: a model that includes a laminar boundary layer', *Agricultural and Forest Meteorology*, 54(2–
- 1166 4), pp. 107–136. doi: 10.1016/0168-1923(91)90002-8.
- 1167 Collatz, G., Ribas-Carbo, M. and Berry, J. (1992) 'Coupled Photosynthesis-Stomatal Conductance Model for
- 1168 Leaves of C₄ Plants', *Functional Plant Biology*, 19(5), p. 519. doi: 10.1071/pp9920519.
- 1169 Comins, H. N. and McMurtrie, R. E. (1993) 'Long-Term Response of Nutrient-Limited Forests to CO₂
- 1170 Enrichment; Equilibrium Behavior of Plant-Soil Models', *Ecological Applications*, 3(4), pp. 666–681. doi:
- 1171 10.2307/1942099.
- 1172 Costa, M. G. *et al.* (2016) 'Labile and Non-Labile Fractions of Phosphorus and Its Transformations in Soil
- 1173 under Eucalyptus', pp. 1–15. doi: 10.3390/f7010015.

1174 Davidson, E. A. *et al.* (2004) ‘Nitrogen and phosphorus limitation of biomass growth in a tropical secondary
1175 forest’, *Ecological Applications*, 14(4 SUPPL.), pp. 150–163. doi: 10.1890/01-6006.
1176 DeLuca, T. H., Keeney, D. R. and McCarty, G. W. (1992) ‘Effect of freeze-thaw-events on mineralization of
1177 soil nitrogen’, *Biol. Fertil. Soils*, 14, pp. 116–120. doi: 10.1007/BF00336260.
1178 Dieter, D., Elsenbeer, H. and Turner, B. L. (2010) ‘Phosphorus fractionation in lowland tropical rainforest soils
1179 in central Panama.’, *Catena*, 82(2), pp. 118–125. doi: 10.1016/j.catena.2010.05.010.
1180 Ellsworth, D. S. *et al.* (2017) ‘Elevated CO₂ does not increase eucalypt forest productivity on a low-phosphorus
1181 soil’, *Nature Climate Change*, 7(4), pp. 279–282. doi: 10.1038/nclimate3235.
1182 Elser, J. J. *et al.* (2007) ‘Global analysis of nitrogen and phosphorus limitation of primary producers in
1183 freshwater, marine and terrestrial ecosystems’, *Ecology Letters*, 10(12), pp. 1135–1142. doi: 10.1111/j.1461-
1184 0248.2007.01113.x.
1185 Fernández-Martínez, M. *et al.* (2019) ‘Global trends in carbon sinks and their relationships with CO₂ and
1186 temperature’, *Nature Climate Change*, 9(1), pp. 73–79. doi: 10.1038/s41558-018-0367-7.
1187 Fleischer, K. *et al.* (2019) ‘Amazon forest response to CO₂ fertilization dependent on plant phosphorus
1188 acquisition’, *Nature Geoscience*. doi: 10.1038/s41561-019-0404-9.
1189 Friedlingstein, P. *et al.* (2006) ‘Climate-carbon cycle feedback analysis: Results from the C4MIP model
1190 intercomparison’, *Journal of Climate*, 19(14), pp. 3337–3353. doi: 10.1175/JCLI3800.1.
1191 Friedlingstein, P. *et al.* (2019) ‘Comment on “The global tree restoration potential”’, *Science*. doi:
1192 10.1126/science.aay8060.
1193 Fyllas, N. M. *et al.* (2009) ‘Basin-wide variations in foliar properties of Amazonian forest: phylogeny, soils and
1194 climate’, *Biogeosciences*, 6(11), pp. 2677–2708. doi: 10.5194/bg-6-2677-2009.
1195 Gentile, R. *et al.* (2012) ‘Effects of long-term exposure to enriched CO₂ on the nutrient-supplying capacity of a
1196 grassland soil’, *Biology and Fertility of Soils*, 48(3), pp. 357–362. doi: http://dx.doi.org/10.1007/s00374-011-
1197 0616-7.
1198 Goll, D. S. *et al.* (2017) ‘A representation of the phosphorus cycle for ORCHIDEE (revision 4520)’,
1199 *Geoscientific Model Development*, 10(10), pp. 3745–3770. doi: 10.5194/gmd-10-3745-2017.
1200 Gross, A. *et al.* (2021) ‘Direct foliar uptake of phosphorus from desert dust’, *New Phytologist*, 230(6), pp.
1201 2213–2225. doi: 10.1111/nph.17344.
1202 Harper, A. B. *et al.* (2016) ‘Improved representation of plant functional types and physiology in the Joint UK
1203 Land Environment Simulator (JULES v4.2) using plant trait information’, *Geoscientific Model Development*,
1204 9(7), pp. 2415–2440. doi: 10.5194/gmd-9-2415-2016.
1205 Harris, I. *et al.* (2014) ‘Updated high-resolution grids of monthly climatic observations - the CRU TS3.10
1206 Dataset’, *International Journal of Climatology*, 34(3), pp. 623–642. doi: 10.1002/joc.3711.
1207 Hatfield, J. L. and Dold, C. (2019) ‘Water-use efficiency: Advances and challenges in a changing climate’,
1208 *Frontiers in Plant Science*, 10(February), pp. 1–14. doi: 10.3389/fpls.2019.00103.
1209 Haverd, V. *et al.* (2018) ‘A new version of the CABLE land surface model (Subversion revision r4601)
1210 incorporating land use and land cover change, woody vegetation demography, and a novel optimisation-based
1211 approach to plant coordination of photosynthesis’, *Geoscientific Model Development*, 11(7), pp. 2995–3026.
1212 doi: 10.5194/gmd-11-2995-2018.
1213 He, M. and Dijkstra, F. A. (2014) ‘Drought effect on plant nitrogen and phosphorus: A meta-analysis’, *New
1214 Phytologist*, 204(4), pp. 924–931. doi: 10.1111/nph.12952.
1215 Hedley, M. J., Stewart, J. W. B. and Chauhan, B. S. (1982) ‘Changes in Inorganic and Organic Soil Phosphorus
1216 Fractions Induced by Cultivation Practices and by Laboratory Incubations’, *Soil Science Society of America
1217 Journal*, 46(5), pp. 970–976. doi: https://doi.org/10.2136/sssaj1982.03615995004600050017x.
1218 Herbert, D. A., Williams, M. and Rastetter, E. B. (2003) ‘A model analysis of N and P limitation on carbon
1219 accumulation in Amazonian secondary forest after alternate land-use abandonment’, *Biogeochemistry*, 65(1), pp.
1220 121–150. doi: 10.1023/A:1026020210887.
1221 Hou, E. *et al.* (2019) ‘Quantifying Soil Phosphorus Dynamics: A Data Assimilation Approach’, *Journal of
1222 Geophysical Research: Biogeosciences*, 124(7), pp. 2159–2173. doi: 10.1029/2018JG004903.
1223 Hou, E. *et al.* (2020) ‘Global meta-analysis shows pervasive phosphorus limitation of aboveground plant
1224 production in natural terrestrial ecosystems’, *Nature Communications*, 11(1), pp. 1–9. doi: 10.1038/s41467-020-
1225 14492-w.
1226 Hubau, W. *et al.* (2020) ‘Asynchronous carbon sink saturation in African and Amazonian tropical forests’,
1227 *Nature*, 579(7797), pp. 80–87. doi: 10.1038/s41586-020-2035-0.
1228 Hungate, B. a *et al.* (2003) ‘Nitrogen and Climate Change’, *Science*, 302(November), pp. 1512–1513.
1229 Jenkinson, D. S. *et al.* (1990) ‘The turnover of organic carbon and nitrogen in soil’, *The Royal Society*,
1230 329(1255). doi: https://doi.org/10.1098/rstb.1990.0177.
1231 Jenkinson, D. S. and Coleman, K. (2008) ‘The turnover of organic carbon in subsoils. Part 2. Modelling carbon
1232 turnover’, *European Journal of Soil Science*, 59(2), pp. 400–413. doi: 10.1111/j.1365-2389.2008.01026.x.
1233 Ji, D. *et al.* (2014) ‘Description and basic evaluation of Beijing Normal University Earth System Model (BNU-

1234 ES(M) version 1', *Geoscientific Model Development*, 7(5), pp. 2039–2064. doi: 10.5194/gmd-7-2039-2014.

1235 Jiang, M. *et al.* (2019) 'Towards a more physiological representation of vegetation phosphorus processes in land

1236 surface models', *New Phytologist*, 222(3), pp. 1223–1229. doi: 10.1111/nph.15688.

1237 Jiang, M. *et al.* (2020) 'The fate of carbon in a mature forest under carbon dioxide enrichment', *Nature*,

1238 580(7802), pp. 227–231. doi: 10.1038/s41586-020-2128-9.

1239 Jiménez, E. M. *et al.* (2009) 'Fine root dynamics for forests on contrasting soils in the Colombian Amazon',

1240 *Biogeosciences*, 6(12), pp. 2809–2827. doi: 10.5194/bg-6-2809-2009.

1241 Johnson, M. O. *et al.* (2016) 'Variation in stem mortality rates determines patterns of above-ground biomass in

1242 Amazonian forests: implications for dynamic global vegetation models', *Global Change Biology*, 22(12), pp.

1243 3996–4013. doi: 10.1111/gcb.13315.

1244 Jones, S. *et al.* (2020) 'The impact of a simple representation of non-structural carbohydrates on the simulated

1245 response of tropical forests to drought', *Biogeosciences*, 17(13), pp. 3589–3612. doi: 10.5194/bg-17-3589-2020.

1246 Kattge, J. *et al.* (2009) 'Quantifying photosynthetic capacity and its relationship to leaf nitrogen content for

1247 global-scale terrestrial biosphere models', *Global Change Biology*, 15(4), pp. 976–991. doi:

1248 <https://doi.org/10.1111/j.1365-2486.2008.01744.x>.

1249 Keller, M. *et al.* (2004) 'Ecological research in the Large-scale Biosphere-Atmosphere Experiment in

1250 Amazonia: Early results', *Ecological Applications*, 14(4 SUPPL.), pp. 3–16. doi: 10.1890/03-6003.

1251 Klein, T. *et al.* (2016) 'Growth and carbon relations of mature *Picea abies* trees under 5 years of free-air CO₂

1252 enrichment', *Journal of Ecology*, 104(6), pp. 1720–1733. doi: 10.1111/1365-2745.12621.

1253 Koch, A., Hubau, W. and Lewis, S. L. (2021) 'Earth System Models Are Not Capturing Present-Day Tropical

1254 Forest Carbon Dynamics', *Earth's Future*, 9(5), pp. 1–19. doi: 10.1029/2020EF001874.

1255 Körner, C. *et al.* (2005) 'Ecology: Carbon flux and growth in mature deciduous forest trees exposed to elevated

1256 CO₂', *Science*, 309(5739), pp. 1360–1362. doi: 10.1126/science.1113977.

1257 Van Langenhove, L. *et al.* (2020) 'Atmospheric deposition of elements and its relevance for nutrient budgets of

1258 tropical forests', *Biogeochemistry*, 149(2), pp. 175–193. doi: 10.1007/s10533-020-00673-8.

1259 Lapola, D. M. and Norby, R. (2014) 'Assessing the effects of increased atmospheric CO₂ on the ecology and

1260 resilience of the Amazon forest', *Science plan et implementation strategy*, AMAZON FAC.

1261 LeBauer, D. and Treseder, K. (2008) 'Nitrogen Limitation of Net Primary Productivity', *Ecology*, 89(2), pp.

1262 371–379.

1263 Lloyd, J. *et al.* (2001) 'Should Phosphorus Availability Be Constraining Moist Tropical Forest Responses to

1264 Increasing CO₂ Concentrations?', in *Global Biogeochemical Cycles in the Climate System*. Elsevier, pp. 95–

1265 114. doi: 10.1016/B978-012631260-7/50010-8.

1266 Long, M. C. *et al.* (2013) 'Twentieth-century oceanic carbon uptake and storage in CESM1(BGC)', *Journal of*

1267 *Climate*, 26(18), pp. 6775–6800. doi: 10.1175/JCLI-D-12-00184.1.

1268 De Lucia, E. H. *et al.* (2007) 'Forest carbon use efficiency: Is respiration a constant fraction of gross primary

1269 production?', *Global Change Biology*, 13(6), pp. 1157–1167. doi: 10.1111/j.1365-2486.2007.01365.x.

1270 Lugli, L. F. (2013) *Estoque de nutrientes na serrapilheira fina e grossa em função de fatores edáficos em*

1271 *florestas do Amazonas, Brasil*. Instituto Nacional de Pesquisas da Amazônia - INPA. Available at:

1272 <https://repositorio.inpa.gov.br/handle/1/5028>.

1273 Lugli, L. F. *et al.* (2020) 'Multiple phosphorus acquisition strategies adopted by fine roots in low-fertility soils

1274 in Central Amazonia', *Plant and Soil*, 450(1–2), pp. 49–63. doi: 10.1007/s11104-019-03963-9.

1275 Lugli, L. F. *et al.* (2021) 'Rapid responses of root traits and productivity to phosphorus and cation additions in a

1276 tropical lowland forest in Amazonia', *New Phytologist*, 230(1), pp. 116–128. doi: 10.1111/nph.17154.

1277 Luo, Y. *et al.* (2004) 'Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon

1278 dioxide', *BioScience*, 54(8), pp. 731–739. doi: 10.1641/0006-3568(2004)054[0731:PNLOER]2.0.CO;2.

1279 Malhi, Y. *et al.* (2004) 'The above-ground coarse wood productivity of 104 Neotropical forest plots', *Global*

1280 *Change Biology*, 10(5), pp. 563–591. doi: 10.1111/j.1529-8817.2003.00778.x.

1281 Malhi, Y. *et al.* (2006) 'The regional variation of aboveground live biomass in old-growth Amazonian forests',

1282 *Global Change Biology*, 12(7), pp. 1107–1138. doi: 10.1111/j.1365-2486.2006.01120.x.

1283 Malhi, Y. *et al.* (2009) 'Comprehensive assessment of carbon productivity, allocation and storage in three

1284 Amazonian forests', *Global Change Biology*, 15(5), pp. 1255–1274. doi: 10.1111/j.1365-2486.2008.01780.x.

1285 Malhi, Y. (2012) 'The productivity, metabolism and carbon cycle of tropical forest vegetation', *Journal of*

1286 *Ecology*, 100(1), pp. 65–75. doi: 10.1111/j.1365-2745.2011.01916.x.

1287 Malhi, Y., Doughty, C. and Galbraith, D. (2011) 'The allocation of ecosystem net primary productivity in

1288 tropical forests', *Philosophical Transactions of the Royal Society B: Biological Sciences*, 366(1582), pp. 3225–

1289 3245. doi: 10.1098/rstb.2011.0062.

1290 Martins, N. P. *et al.* (2021) 'Fine roots stimulate nutrient release during early stages of leaf litter decomposition

1291 in a Central Amazon rainforest', *Plant and Soil*, 469(1–2), pp. 287–303. doi: 10.1007/s11104-021-05148-9.

1292 Medlyn, B. E. *et al.* (2015) 'Using ecosystem experiments to improve vegetation models', *Nature Climate*

1293 *Change*, 5(6), pp. 528–534. doi: 10.1038/nclimate2621.

1294 Medlyn, B. E. *et al.* (2016) ‘Using models to guide field experiments: a priori predictions for the CO₂ response
1295 of a nutrient- and water-limited native Eucalypt woodland’, *Global Change Biology*, 22(8), pp. 2834–2851. doi:
1296 10.1111/gcb.13268.

1297 Mercado, L. M. *et al.* (2011) ‘Variations in Amazon forest productivity correlated with foliar nutrients and
1298 modelled rates of photosynthetic carbon supply’, *Philosophical Transactions of the Royal Society B: Biological
1299 Sciences*, 366(1582), pp. 3316–3329. doi: 10.1098/rstb.2011.0045.

1300 Mirabello, M. J. *et al.* (2013) ‘Soil phosphorus responses to chronic nutrient fertilisation and seasonal drought in
1301 a humid lowland forest, Panama’, *Soil Research*, 51(3), pp. 215–221. doi: 10.1071/SR12188.

1302 Mitchard, E. T. A. (2018) ‘The tropical forest carbon cycle and climate change’, *Nature*, 559(7715), pp. 527–
1303 534. doi: 10.1038/s41586-018-0300-2.

1304 Nachtergaele, F. *et al.* (2010) ‘The Harmonized World Soil Database’, *Proceedings of the 19th World Congress
1305 of Soil Science, Soil Solutions for a Changing World, Brisbane, Australia, 1-6 August 2010*, pp. 34–37. doi:
1306 3123.

1307 Neff, J. C., Hobbie, S. E. and Vitousek, P. M. (2000) ‘Nutrient and mineralogical control on dissolved organic
1308 C, N and P fluxes and stoichiometry in Hawaiian soils’, *Biogeochemistry*, 51(3), pp. 283–302. doi:
1309 10.1023/A:1006414517212.

1310 Norby, R. J. *et al.* (2016) ‘Model–data synthesis for the next generation of forest free-air <sc>CO</sc>₂
1311 enrichment (<sc>FACE</sc>) experiments’, *New Phytologist*, 209(1), pp. 17–28. doi: 10.1111/nph.13593.

1312 Nordin, A., Högborg, P. and Näsholm, T. (2001) ‘Soil nitrogen form and plant nitrogen uptake along a boreal
1313 forest productivity gradient’, *Oecologia*, 129(1), pp. 125–132. doi: 10.1007/s004420100698.

1314 Pan, Y. *et al.* (2011) ‘A Large and Persistent Carbon Sink in the World’s Forests’, *Science*, 333(6045), pp. 988–
1315 993. doi: 10.1126/science.1201609.

1316 Perakis, S. S. and Hedin, L. O. (2002) ‘Nitrogen loss from unpolluted South American forests mainly via
1317 dissolved organic compounds’, *Nature*, 415(6870), pp. 416–419. doi: 10.1038/415416a.

1318 Phillips, O. L. *et al.* (2004) ‘Pattern and process in Amazon tree turnover, 1976–2001’, *Philosophical
1319 Transactions of the Royal Society B: Biological Sciences*, 359(1443), pp. 381–407. doi: 10.1098/rstb.2003.1438.

1320 Phillips, O. L. *et al.* (2017) ‘Carbon uptake by mature Amazon forests has mitigated Amazon nations’ carbon
1321 emissions’, *Carbon Balance and Management*, 12(1), pp. 1–9. doi: 10.1186/s13021-016-0069-2.

1322 Pyle, E. H. *et al.* (2009) ‘Dynamics of carbon, biomass, and structure in two Amazonian forests’, *Journal of
1323 Geophysical Research: Biogeosciences*, 114(1), pp. 1–20. doi: 10.1029/2007JG000592.

1324 Quesada, C. A. *et al.* (2010) ‘Variations in chemical and physical properties of Amazon forest soils in relation to
1325 their genesis’, *Biogeosciences*, 7(5), pp. 1515–1541. doi: 10.5194/bg-7-1515-2010.

1326 Quesada, C. A. *et al.* (2011) ‘Soils of Amazonia with particular reference to the RAINFOR sites’,
1327 *Biogeosciences*, 8(6), pp. 1415–1440. doi: 10.5194/bg-8-1415-2011.

1328 Quesada, C. A. *et al.* (2012) ‘Basin-wide variations in Amazon forest structure and function are mediated by
1329 both soils and climate’, *Biogeosciences*, 9(6), pp. 2203–2246. doi: 10.5194/bg-9-2203-2012.

1330 Reed, S. C., Yang, X. and Thornton, P. E. (2015) ‘Incorporating phosphorus cycling into global modeling
1331 efforts: A worthwhile, tractable endeavor’, *New Phytologist*, 208(2), pp. 324–329. doi: 10.1111/nph.13521.

1332 Ryan, M. G. (2013) ‘Three decades of research at Flakaliden advancing whole-tree physiology, forest ecosystem
1333 and global change research’, *Tree Physiology*, 33(11), pp. 1123–1131. doi: 10.1093/treephys/tpt100.

1334 Sampaio, G. *et al.* (2021) ‘CO₂ physiological effect can cause rainfall decrease as strong as large-scale
1335 deforestation in the Amazon’, *Biogeosciences*, 18(8), pp. 2511–2525. doi: 10.5194/bg-18-2511-2021.

1336 Sanchez, P. A. (1977) ‘Properties and Management of Soils in the Tropics’, *Soil Science*, 124(3). Available at:
1337 [https://journals.lww.com/soilsci/Fulltext/1977/09000/Properties_and_Management_of_Soils_in_the_Tropics.12.](https://journals.lww.com/soilsci/Fulltext/1977/09000/Properties_and_Management_of_Soils_in_the_Tropics.12.aspx)
1338 [aspx](https://journals.lww.com/soilsci/Fulltext/1977/09000/Properties_and_Management_of_Soils_in_the_Tropics.12.aspx).

1339 Sardans, J., Rivas-Ubach, A. and Peñuelas, J. (2012) ‘The C:N:P stoichiometry of organisms and ecosystems in
1340 a changing world: A review and perspectives’, *Perspectives in Plant Ecology, Evolution and Systematics*, 14(1),
1341 pp. 33–47. doi: 10.1016/j.ppees.2011.08.002.

1342 Schimel, D., Stephens, B. B. and Fisher, J. B. (2015) ‘Effect of increasing CO₂ on the terrestrial carbon cycle’,
1343 *Proceedings of the National Academy of Sciences of the United States of America*, 112(2), pp. 436–441. doi:
1344 10.1073/pnas.1407302112.

1345 Shen, J. *et al.* (2011) ‘Phosphorus dynamics: From soil to plant’, *Plant Physiology*, 156(3), pp. 997–1005. doi:
1346 10.1104/pp.111.175232.

1347 Sitch, S. *et al.* (2008) ‘Evaluation of the terrestrial carbon cycle, future plant geography and climate-carbon
1348 cycle feedbacks using five Dynamic Global Vegetation Models (DGVMs)’, *Global Change Biology*, 14(9), pp.
1349 2015–2039. doi: 10.1111/j.1365-2486.2008.01626.x.

1350 Stephenson, N. L. and Van Mantgem, P. J. (2005) ‘Forest turnover rates follow global and regional patterns of
1351 productivity’, *Ecology Letters*, 8(5), pp. 524–531. doi: 10.1111/j.1461-0248.2005.00746.x.

1352 Sun, Y. *et al.* (2021) ‘Global evaluation of the nutrient-enabled version of the land surface model ORCHIDEE-
1353 CNP v1.2 (r5986)’, *Geoscientific Model Development*, 14(4), pp. 1987–2010. doi: 10.5194/gmd-14-1987-2021.

1354 Turner, B. L. *et al.* (2015) ‘Seasonal changes in soil organic matter after a decade of nutrient addition in a
1355 lowland tropical forest’, *Biogeochemistry*, 123(1–2), pp. 221–235. doi: 10.1007/s10533-014-0064-1.
1356 Turner, B. L. and Condon, L. M. (2013) ‘Pedogenesis, nutrient dynamics, and ecosystem development: the
1357 legacy of T.W. Walker and J.K. Syers’, *Plant and Soil*, 367(1–2), pp. 1–10. doi: 10.1007/s11104-013-1750-9.
1358 Vicca, S. *et al.* (2012a) ‘Fertile forests produce biomass more efficiently’, *Ecology Letters*, 15(6), pp. 520–526.
1359 doi: 10.1111/j.1461-0248.2012.01775.x.
1360 Vicca, S. *et al.* (2012b) ‘Fertile forests produce biomass more efficiently’, *Ecology Letters*, 15(6), pp. 520–526.
1361 doi: 10.1111/j.1461-0248.2012.01775.x.
1362 Vitousek, P. M. *et al.* (1997) ‘Human Domination of Earth Ecosystems’, *Science*, 278(5335), p. 21. Available
1363 at: <http://www.cheric.org/research/tech/periodicals/view.php?seq=257860>.
1364 Vitousek, P. M. (2004) *Nutrient Cycling and Limitation: Hawai‘i as a Model System*. Princeton University Press
1365 (Princeton Environmental Institute series). Available at:
1366 <https://books.google.com.fj/books?id=6EmAtAEACAAJ>.
1367 Vitousek, P. M. *et al.* (2010) ‘Terrestrial phosphorus limitation: Mechanisms, implications, and nitrogen-
1368 phosphorus interactions’, *Ecological Applications*, 20(1), pp. 5–15. doi: 10.1890/08-0127.1.
1369 Vitousek, P. M. and Howarth, R. W. (1991) ‘Nitrogen limitation on land and in the sea: How can it occur?’,
1370 *Biogeochemistry*, 13(2), pp. 87–115. doi: 10.1007/BF00002772.
1371 Walker, A. P. *et al.* (2021) ‘Integrating the evidence for a terrestrial carbon sink caused by increasing
1372 atmospheric CO₂’, *New Phytologist*, 229(5), pp. 2413–2445. doi: 10.1111/nph.16866.
1373 Walker, T. W. and Syers, J. K. (1976) ‘The fate of phosphorus during pedogenesis’, *Geoderma*, 15(1), pp. 1–19.
1374 doi: 10.1016/0016-7061(76)90066-5.
1375 Wang, Y. P., Houlton, B. Z. and Field, C. B. (2007) ‘A model of biogeochemical cycles of carbon, nitrogen, and
1376 phosphorus including symbiotic nitrogen fixation and phosphatase production’, *Global Biogeochemical Cycles*,
1377 21(1), pp. 1–15. doi: 10.1029/2006GB002797.
1378 Wang, Y. P., Law, R. M. and Pak, B. (2010) ‘A global model of carbon, nitrogen and phosphorus cycles for the
1379 terrestrial biosphere’, *Biogeosciences*, 7(7), pp. 2261–2282. doi: 10.5194/bg-7-2261-2010.
1380 Went, F. W. and Stark, N. (1968) ‘Mycorrhiza’, *BioScience*, 18(11), pp. 1035–1039. doi: 10.2307/1294552.
1381 Wiltshire, A. J. *et al.* (2020) ‘JULES-CN : a coupled terrestrial Carbon-Nitrogen Scheme (JULES vn5 . 1)’,
1382 (July), pp. 1–40.
1383 Wiltshire, A. J. *et al.* (2021) ‘Jules-cn: A coupled terrestrial carbon-nitrogen scheme (jules vn5.1)’,
1384 *Geoscientific Model Development*, 14(4), pp. 2161–2186. doi: 10.5194/gmd-14-2161-2021.
1385 Wright, S. J. *et al.* (2011) ‘Potassium, phosphorus, or nitrogen limit root allocation, tree growth, or litter
1386 production in a lowland tropical forest’, *Ecology*, 92(8), pp. 1616–1625. doi: 10.1890/10-1558.1.
1387 Xiao, J. *et al.* (2013) ‘Carbon fluxes, evapotranspiration, and water use efficiency of terrestrial ecosystems in
1388 China’, *Agricultural and Forest Meteorology*, 182–183, pp. 76–90. doi: 10.1016/j.agrformet.2013.08.007.
1389 Xu, Z. *et al.* (2016) ‘Elevated-CO₂ response of stomata and its dependence on environmental factors’, *Frontiers
1390 in Plant Science*, 7(MAY2016), pp. 1–15. doi: 10.3389/fpls.2016.00657.
1391 Yang, X. *et al.* (2013) ‘The distribution of soil phosphorus for global biogeochemical modeling’,
1392 *Biogeosciences*, 10(4), pp. 2525–2537. doi: 10.5194/bg-10-2525-2013.
1393 Yang, X. *et al.* (2014) ‘The role of phosphorus dynamics in tropical forests – a modeling study using CLM-
1394 CNP’, *Biogeosciences*, 11(6), pp. 1667–1681. doi: 10.5194/bg-11-1667-2014.
1395 Yang, X. and Post, W. M. (2011) ‘Phosphorus transformations as a function of pedogenesis: A synthesis of soil
1396 phosphorus data using Hedley fractionation method’, *Biogeosciences*, 8(10), pp. 2907–2916. doi: 10.5194/bg-8-
1397 2907-2011.
1398 Zaehle, S. and Dalmonech, D. (2011) ‘Carbon-nitrogen interactions on land at global scales: Current
1399 understanding in modelling climate biosphere feedbacks’, *Current Opinion in Environmental Sustainability*,
1400 3(5), pp. 311–320. doi: 10.1016/j.cosust.2011.08.008.
1401 Zaehle, S. and Friend, A. D. (2010) ‘Carbon and nitrogen cycle dynamics in the O-CN land surface model: 1.
1402 Model description, site-scale evaluation, and sensitivity to parameter estimates’, *Global Biogeochemical Cycles*,
1403 24(1), pp. 1–13. doi: 10.1029/2009GB003521.
1404 Zhu, Q. *et al.* (2016) ‘Multiple soil nutrient competition between plants, microbes, and mineral surfaces: model
1405 development, parameterization, and example applications in several tropical forests’, *Biogeosciences*, 13(1), pp.
1406 341–363. doi: 10.5194/bg-13-341-2016.
1407