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- Ocean biogeochemistry in the Canadian Earth System Model version 5.0.3: CanESM5 and
 CanESM5-CanOE
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19	Abstract. The ocean biogeochemistry components of two new versions of the Canadian Earth
20	System Model v. 5 -are presented and compared to observations and other models. CanESM5
21	employs the same biogeochemistry module ocean biology model as CanESM2 whereas
22	CanESM5-CanOE ("Canadian Ocean Ecosystem model") is a new, more complex
23	biogeochemistry modulemodel developed for CMIP6, with multiple food chains, flexible
24	phytoplankton elemental ratios, and a prognostic iron cycle. This new model is described in
25	detail and the outputs (distributions of major tracers such as oxygen, dissolved inorganic carbon,
26	and alkalinity, the iron and nitrogen cycles, plankton biomass, and historical trends in CO_2
27	uptake and export production) compared to CanESM5 and CanESM2, as well as to observations
28	and other CMIP6 models. Both CanESM5 models show gains in skill relative to CanESM2,
29	which are attributed primarily to improvements in ocean circulation. CanESM5-CanOE shows
30	improved skill relative to CanESM5 in some areas for most major tracers at most depths.
31	CanESM5-CanOE includes a prognostic iron cycle, and maintains high nutrient / low
32	chlorophyll conditions in the expected regions (in CanESM2 and CanESM5, iron limitation is
33	specified as a temporally static 'mask'). Surface nitrate concentrations are biased low in the
34	subarctic Pacific and equatorial Pacific, and high in the Southern Ocean, in both CanESM5 and
35	CanESM5-CanOE. Export production in CanESM5-CanOE is among the lowest for CMIP6
36	models; in CanESM5 it is among the highest, but shows the most rapid decline after about 1980.
37	CanESM5-CanOE shows some ability to simulate aspects of plankton community structure that a
38	single-species model can not (e.g., seasonal dominance of large cells), but is biased towards has
39	relatively low concentrations of zooplankton and detritus relative to phytoplankton., and a high
40	and relatively constant living phytoplankton fraction of total particulate organic matter. In most
41	regions, large and small phytoplankton show decoupled seasonal cycles with greater abundance

- 42 of large phytoplankton in the productive seasons. Cumulative ocean uptake of anthropogenic
- 43 carbon dioxide through 2014 is lower in both CanESM5 models than in observation-based
- 44 estimates (145 PgC) or the model ensemble mean (144 PgC), and is lower in CanESM5-CanOE
- 45 (122 PgC) than in CanESM5 (132 PgC).
- 46

47 1. Introduction

49	The Canadian Centre for Climate Modelling and Analysis has been developing coupled models
50	with an interactive carbon cycle for more than a decade (Arora et al., 2009; 2011; Christian et al.,
51	2010). The Canadian Earth System Model version 5 (CanESM5-(, Swart et al., 2019a) is an
52	updated version of CanESM2 (Arora et al., 2011), with an entirely new ocean model and an
53	atmosphere with the same T63 horizontal resolution and important improvements in atmospheric
54	physics. The CanESM5 ocean is based on the Nucleus for European Modelling of the Ocean
55	(NEMO) system version 3.4. The ocean biogeochemistry modules were developed in-house,
56	although parameterizations for some processes were adapted from the native PISCES
57	biogeochemistry model (Aumont et al., 2015). CanESM5 uses the same ocean biogeochemistry
58	biology model as CanESM1 (Christian et al., 2010) and CanESM2 (Arora et al., 2011), the
59	Canadian Model of Ocean Carbon (CMOC; Zahariev et al., 2008), adapted for use within
60	NEMO. An additional model was developed for CMIP6, called the Canadian Ocean Ecosystem
61	model (CanOE). The biological components of CanOE are of substantially greater complexity
62	than CMOC, including multiple food chains, flexible phytoplankton elemental ratios, and a
63	prognostic iron (Fe) cycle. Carbon chemistry, gas exchange and solubility of carbon dioxide
64	(CO ₂) and oxygen are identical between the two and follow the protocols specified by the Ocean
65	Model Intercomparison Project - Biogeochemistry (OMIP-BGC) (Orr et al., 2017). The two
66	coupled models are known as CanESM5 and CanESM5-CanOE, respectively. There are no
67	feedbacks between biology and the physical ocean model, so the physical climate of CanESM5
68	and CanESM5-CanOE is identical in experiments with prescribed atmospheric CO2
69	concentration.

71 The reasons for developing both models are, firstly, to evaluate the effect of changes in ocean 72 circulation between CanESM2 and CanESM5 on ocean biogeochemistry by running the new climate model with the same ocean biogeochemistry, and secondly because CanOE is 73 substantially more expensive computationally (as it has 19 tracers vs 7 the total computation cost 74 75 is 2-3 times greater). Having CMOC as an option allowed us to run many Most CMIP6 76 experiments were run with CanESM5 only, as ocean biogeochemistry is not central to their 77 purpose. Many aAdditional tracers requested by the Ocean Model Intercomparison Project -Biogeochemistry (OMIP-BGC) including abiotic and natural dissolved inorganic carbon (DIC), 78 79 DI¹⁴C, CFCs and SF₆ (see Orr et al., 2017) were run only in CanESM5, since adding these tracers on top of the larger set of biological components in CanOE would have been 80 prohibitively expensive. The CMIP6 experiments published for CanESM5-CanOE are listed in 81 82 Supplementary Table S1.

83

CMOC is a nutrient-phytoplankton-zooplankton-detritus (NPZD) model with highly 84 parameterized representations of phytoplankton Fe limitation, dinitrogen (N₂) fixation and 85 denitrification, and calcification and calcite dissolution (Zahariev et al., 2008; Supplementary 86 Figure S1). In-CanESM1 and CanESM2, CMOC did not include oxygen; In-CanESM5, CMOC 87 88 now includes oxygen as a purely 'downstream' tracer that does not affect other biogeochemical processes. , whereas in CanESM5-CanOE, denitrification is prognostic and dependent on the 89 concentration of oxygen. Among the less satisfactory aspects of CMOC biogeochemistry are, 90 91 firstly, that Fe limitation is specified as a static 'mask' that does not change with climate (being it 92 is calculated from the present-day climatological distribution of nitrate, based on the assumption

93 that regions without iron limitation will have complete drawdown of surface nitrate at some point in the year), and secondly, that denitrification is parameterized so that nitrogen (N) is conserved 94 within each vertical column, i.e., collocated with N₂ fixation in tropical and subtropical open-95 96 ocean regions (Zahariev et al., 2008; Riche and Christian, 2018). This latter simplification produced excessive accumulations of nitrate in Eastern Boundary Current (EBC) regions where 97 98 most denitrification actually occurs. CMOC also has a tendency to produce rather stark extremes of high and low primary and export production (Zahariev et al., 2008), a well-known problem of 99 NPZD models (Armstrong, 1994; Friedrichs et al., 2007). Our intent in developing CanOE was 100 101 to alleviate, or at least reduce, these biases, by including multiple food chains, a prognostic Fe cycle, and prognostic denitrification. Dinitrogen fixation is still parameterized, but the CanOE 102 103 parameterization includes Fe (but not P) limitation, whereas in CMOC N₂ fixation tends to grow without bound in a warming ocean as there is no P or Fe limitation (Riche and Christian, 2018). 104 Calcification is represented by a prognostic detrital calcite pool with its own sinking rate (distinct 105 from that of organic detritus), and calcite burial depends on the saturation state. In CMOC 106 calcification is parameterized by a temperature dependent "rain ratio" and 100% burial of calcite 107 that reaches the seafloor is assumed. 108

109

In this paper we present a detailed model description for CanOE and an evaluation of both CanESM5 and CanESM5-CanOE relative to observational data products and other available models. CMOC has been well described previously (Zahariev et al., 2008) and the details are not reiterated here. In some cases, CanESM2 results are also shown to illustrate which differences in the model solutions arise largely from the evolution of the physical ocean model, and which are specifically associated with different representations of biogeochemistry. An overall evaluation

116	of the CanESM5 physical ocean model is given in Swart et al. (2019a). Here, we focus on
117	biogeochemical variables, and have evaluated model performance in three main areas: (1) the
118	distribution of major tracers like oxygen, DIC and alkalinity, and the resulting saturation state for
119	CaCO ₃ minerals, (2) the iron cycle and its interaction with the nitrogen cycle, and (3) plankton
120	community structure and the concentration and export of particulates. We first address the major
121	chemical species that are common to both models (and almost all other Earth System Models) to
122	determine whether a more complex biology model measurably improves skill, and whether the
123	updated circulation model improves skill relative to CanESM2. Then we examine the areas
124	where our two models differ: the presence of a prognostic iron cycle and multiple food chains in
125	CanOE. More specifically, does CanESM5-CanOE reproduce the geographic distribution of
126	High-Nutrient, Low-Chlorophyll (HNLC) regions? Does the large phytoplankton / large
127	zooplankton food chain become dominant under nutrient-rich conditions, and how does having
128	multiple detrital size classes affect particle flux and remineralization length scale? Following this
129	model evaluation, we present historical trends in ocean anthropogenic CO ₂ uptake, export
130	production, and total volume of low-oxygen waters over the historical (1850-2014) experiment.
131	Possible future changes under Shared Socioeconomic Pathway experiments will be addressed in
132	subsequent publications.
133	
134	2. Model Description
135	
136	CanESM5 (Swart et al., 2019a) is an updated version of CanESM2 (Arora et al., 2011), with an
137	entirely new ocean. The atmosphere model has the same T63 horizontal resolution, and contains
138	some important improvements in atmospheric physics (Swart et al., 2019a). The land surface

139	(Canadian Land Surface Scheme) and terrestrial carbon cycle (Canadian Terrestrial Ecosystem
140	Model) models are substantially the same as in CanESM2 with minor modifications as described
141	by Arora et al. (2020). The CanESM5 ocean is based on the NEMO modelling system version
142	3.4, with a horizontal resolution of 1°, telescoping to 1/3° in the tropics, and 45 vertical levels
143	ranging in thickness from ~6 m near the surface to ~250 m in the deep ocean (Swart et al.,
144	2019a). All physical climate model components are the same in CanESM5 and CanESM5-
145	CanOE. There are no feedbacks between biology and the physical ocean model, so the physical
146	climate of CanESM5 and CanESM5-CanOE is identical in experiments with prescribed
147	atmospheric CO ₂ concentration.
148	
149	The NEMO system is a publicly available archive of codes based on the OPA (Océan
150	PArallelisé) ocean model (Madec and Imbard, 1996; Guilyardi and Madec, 1997) and the
151	Tracers in Ocean Paradigm (TOP) module for tracer advection and mixing. It comes with two
152	options for biogeochemistry: PISCES (Pelagic Interactions Scheme for Carbon and Ecosystem
153	Studies) and LOBSTER (LODyC Ocean Biogeochemical System for Ecosystem and Resources).
154	Our ocean biogeochemistry modules CanOE and the NEMO implementation of CMOC are built
155	around the basic code structure of PISCES-within the Tracers in Ocean Paradigm (TOP) module,
156	using NEMO v3.4.1, but have also been implemented in NEMO 3.6 for regional downscaling
157	applications (Holdsworth et al., 2021).
158	
159	
	The biology, carbon chemistry, gas exchange and light attenuation components have all been
160	modified to various degrees. In a few cases PISCES parameterizations, or slightly modified

161 versions thereof, were adopted. CanOE uses PISCES three-band light attenuation while NEMO-

162	CMOC uses broadband attenuation of photosynthetically active radiation (PAR) for consistency
163	with the published version of CMOC. Carbon chemistry was modified to be consistent with is
164	based on the Best Practices Guide (Dickson et al., 2007) and the OMIP-BGC data request (Orr et
165	al., 2017) and are identical in CanESM5 and CanESM5-CanOE. All calculations are done on the
166	total scale and the recommended formulae for the equilibrium constants are employed. The
167	PISCES conventions for convergence of cThe carbon chemistry solver calculations were
168	retained, the greater was run for a fixed number of iterations (ten in the surface layer, and five in
169	the subsurface layers in CanESM5-CanOE) offering greater accuracy in calculating pCO2 and gas
170	exchange. In subsurface layers (where the only function of the carbon chemistry is to calculate
171	burial of calcite in the sediments) the number of iterations is fixed at five. CanESM5 uses the
172	same carbon chemistry but does not solve the carbon chemistry equations in the subsurface
173	layers. <u>OMIP-BGC formulations for CO₂ and O₂ solubility and gas exchange are employed. It is</u>
174	important to note here that the carbon chemistry and gas exchange formulations used in
175	CanESM2 (and other CMIP5 models) are slightly different than those used in CMIP6. However,
176	this difference is of little functional significance, i.e., it will have a negligible impact on the
177	distribution of [CO3] compared to the differences in DIC and alkalinity distribution. The
178	initialization fields for nitrate, DIC and alkalinity were also different in CanESM2. This will
179	affect the total ocean inventory of DIC but not the spatial distribution if the model is well
180	equilibrated.

The CanOE biology model is a substantially new model based on the cellular regulation model
of Geider et al. (1998). There are two phytoplankton <u>functional groupssize classes</u>, and each
group has four state variables: C, N, Fe and chlorophyll. Photosynthesis is decoupled from cell

185	production and photosynthetic rate is a function of the cell's internal N and Fe quotas. Each
186	functional group has a specified minimum and maximum N quota and Fe quota, and nutrient
187	uptake ceases when the maximal cell quota is reached. Chlorophyll synthesis is a function of N
188	uptake and increases at low irradiance. There are also two size classes each of zooplankton and
189	detritus. Small zooplankton graze on small phytoplankton, while large zooplankton graze on both
190	large phytoplankton and small zooplankton. Small detritus sinks at 2 m d ⁻¹ and large detritus at
191	<u>30 m d⁻¹ (in CanESM5 there is a single detrital pool with a sinking rate of 8 m d⁻¹).</u> Model
192	parameters and their values are listed in Table 1. A schematic of the model is shown in Figure 1.
193	
194	2.1 Photosynthesis and Phytoplankton Growth
195	
196	For simplicity and clarity, the equations are shown here for a single phytoplankton species, and
197	do not differ structurally for small and large phytoplankton. Some parameter values differ for the
198	two phytoplankton groups; all parameter values are listed in Table 1.
199	
200	Temperature dependence of photosynthetic activity is expressed by the Arrhenius equation
201	$T_f = \exp\left(-\frac{E_{ap}}{R}\left(\frac{1}{T} - \frac{1}{T_{\text{ref}}}\right)\right) \tag{1}$
202	where E_{ap} is an enzyme activation energy that corresponds approximately to that of RuBisCo (cf.
203	Raven and Geider 1988), R is the gas constant (8.314 J mol ⁻¹ K ⁻¹), and temperature T and

reference temperature T_{ref} are in Kelvin. Maximal rates of nutrient (either N or Fe, but

205 generically referred to here with the superscript X) uptake are given by

206
$$V_{max}^{X} = V_{ref}^{X} T_f \left(\frac{Q_{max}^{X} - Q^{X}}{Q_{max}^{X} - Q_{min}^{X}} \right)^{0.05}$$
(2)

where V_{max}^{X} is the maximal uptake rate in mg of nutrient X per mg of cell C, X can represent N or Fe, Q is the nutrient cell quota and Q_{min} and Q_{max} its minimum and maximum values, and V_{ref}^{X} is a (specified) basal rate at $T=T_{ref}$ and $Q=Q_{min}$. These maximum rates are then reduced according to the ambient nutrient concentration, i.e.

211
$$V^N = V_{max}^N (L_{NH4} + (1 - L_{NH4})L_{NO3})$$
 (3a)

212 where $L_{\text{NH4}} = \frac{N_a}{K_{\text{NaX}} + N_a}$ and $L_{\text{NO3}} = \frac{N_i}{K_{\text{NiX}} + N_i}$, with N_i and N_a indicating nitrate and ammonium

213 respectively, and

214
$$V^{Fe} = V_{max}^{Fe} \left(\frac{Fe}{K_{FeX} + Fe}\right)$$
 (3b)

where X indicates large or small phytoplankton (Table 1). The maximal carbon-based growthrate is given by

217
$$P_{max}^{C} = P_{ref}^{C} T_{f} \min\{\frac{Q^{N} - Q_{min}^{N}}{Q_{max}^{N} - Q_{min}^{N}}, \frac{Q^{Fe} - Q_{min}^{Fe}}{Q_{max}^{Fe} - Q_{min}^{Fe}}\}$$
(4)

218 where P^{C}_{ref} is the rate at the reference temperature T_{ref} under nutrient-replete conditions

219 $(Q=Q_{max})$. The light-limited growth rate is then given by

220
$$P_{phot}^{C} = P_{max}^{C} \left(1 - e^{-\alpha_{chl} E\theta_{C}} / P_{max}^{C} \right)$$
(5)

221 where <u>E is irradiance and</u> θ_C is the chlorophyll-to-carbon ratio. The rate of chlorophyll synthesis 222 is

223
$$\rho_{chl} = \theta_{max}^{N} \frac{P_{phot}^{C}}{E\alpha_{chl}\theta}$$
(6)

224 These rates are then used to define a set of state equations for phytoplankton carbon (C_p) ,

225 nitrogen (N_p) , iron (Fe_p) , and chlorophyll (M).

226
$$\frac{dC_p}{dt} = (P_{phot}^C - \zeta V_N)C_p - (G + C_{XS}) - m_1C_p - m_2C_p^2 - k_{XU}C_{INTR}$$
(7)

227 where ζ is the respiratory cost of biosynthesis, G is the grazing rate (equation 12), C_{XS} is the 228 excess (above the ratio in grazer biomass) carbon in grazing losses (see below equation 16a), m_1 and m_2 are coefficients for linear and quadratic nonspecific mortality terms, C_{INTR} is the 229 230 concentration of intracellular carbohydrate carbon in excess of biosynthetic requirements, and k_{XU} is a rate coefficient for its exudation to the environment. The nonspecific mortality terms are 231 set to 0 below 0.01 mmol C m⁻³, to prevent biomass from being driven to excessively low levels 232 233 in the high latitudes in winter; linear mortality terms can result in biomass declining to levels from which recovery would take much longer than the brief Arctic summer (Hayashida, 2018). 234

235 The full equation for phytoplankton N, Fe and chlorophyll are

236
$$\frac{dN_p}{dt} = \frac{V^N}{Q_N} - (G + m_1 C_p + m_2 C_p^2) R_{\rm NC} - N_{\rm XS}$$
(8)

237
$$\frac{dFe_p}{dt} = \frac{V^{Fe}}{Q_{Fe}} - (G + m_1 C_P + m_2 C_p^2) R_{FeC} - Fe_{XS}$$
(9)

238
$$\frac{\mathrm{dM}}{\mathrm{dt}} = \frac{\rho_{\mathrm{chl}} V^N}{\theta_C} M - (G + m_1 C_p + m_2 C_p^2) \theta_C - k_{\mathrm{dgr}} M \tag{10}$$

where k_{dgr} is a rate coefficient for nonspecific losses of chlorophyll e.g., by photooxidation, in addition to losses to grazing and other processes that also affect C_p , N_p , and Fe_p . N_{XS} and Fe_{XS} are remineralization of "excess" (relative to grazer or detritus ratios) N or Fe and are defined below (equation 16).

243

244 **2.2 Grazing and Food Web Interactions**

Grazing rate depends on the phytoplankton carbon concentration, which most closely represents the food concentration available to the grazer (Elser and Urabe 1999; Loladze et al. 2000).

248 Zooplankton biomass is also in carbon units. State equations for small and large zooplankton are

249
$$\frac{dZ_s}{dt} = \lambda G_s - (R + G_Z + m_{1s}Z_s + m_{2s}Z_s^2)$$
(11a)

250
$$\frac{dZ_L}{dt} = \lambda G_L - (R + m_{1L}Z_L + m_{2L}Z_L^2)$$
(11b)

251 where

252
$$G_s = G_{so}(1 - e^{-a_s C_{ps}})Z_s$$
 (12a)

253
$$G_L = G_{L0}(1 - e^{-a_l(C_{pl} + Z_s)})Z_L$$
 (12b)

for small and large zooplankton respectively, G_Z is grazing of small zooplankton by large zooplankton, *R* is respiration, and m_1 and m_2 are nongrazing mortality rates. Large zooplankton grazing is divided into grazing on large phytoplankton and small zooplankton in proportion to the relative abundance of each

$$258 \qquad G_P = G_L \frac{P_l}{P_l + Z_s} \tag{13a}$$

259
$$G_Z = G_{\downarrow L} \frac{Z_S}{P_l + Z_S}$$
 (13b)

260 Zooplankton biomass loss to respiration is given by

261
$$R = max\{r_z T_f Z - C_{\rm XS}, 0\}$$
(14)

and uses the same activation energy as photosynthesis. Respiration (*R*) is assumed to consume only carbon and not result in catabolism of existing biomass when "excess" carbon is available in the prey. In addition, conservation of mass must be maintained by recycling to the dissolved pool grazer consumption of elements in excess of biosynthetic requirements when grazer and prey elemental ratios differ. In the case where the nutrient quota (relative to carbon) exceeds the grazer fixed ratio, the excess nutrient is remineralized to the dissolved inorganic pool. In the case where the nutrient quota is less than the grazer ratio, the grazer intake is reduced to what can be supported by the least abundant nutrient (relative to the grazer biomass ratio) and excess carbon is remineralized. For the case of two nutrients (in this case N and Fe) it is necessary to define

271
$$G' = G\min\left\{\frac{N_P}{C_P}R_{\rm CN}, \frac{Fe_P}{C_P}R_{\rm CFe}, 1\right\}$$
(15)

where *G* is equal to G_S (equation 12a) for small zooplankton and G_P (equation 13a) for large zooplankton, and R_{XY} indicates the fixed ratio of element X to element Y in grazer biomass. The 'excess' carbon available for respiration is

275
$$C_{\rm XS} = G' \left\{ \frac{C_P}{N_P} R_{\rm NC} - 1, \frac{C_P}{Fe_P} R_{\rm FeC} - 1, 0 \right\}$$
 (16a)

and the excess nutrients remineralized to their inorganic pools are

277
$$N_{\rm XS} = G'\max\left\{\frac{N_P}{C_P} - R_{\rm NC}.0\right\}\varepsilon + G'\max\left\{R_{\rm NC}\left(\frac{N_P}{Fe_P}R_{\rm FeN} - 1\right).0\right\}(1-\varepsilon)$$
(16b)

278
$$\operatorname{Fe}_{\mathrm{XS}} = G'\max\left\{\frac{\operatorname{Fe}_{P}}{C_{P}} - R_{\mathrm{FeC}}.0\right\}\varepsilon + G'\max\left\{R_{\mathrm{FeC}}(\frac{\operatorname{Fe}_{P}}{N_{P}}R_{\mathrm{NFe}} - 1).0\right\}(1 - \varepsilon)$$
(16c)

279 where

$$280 \qquad \varepsilon = \frac{\max\{C_{xs}, 0\}}{C_{xs} + \Delta}$$

is a switch to prevent double-counting in cases where one of the terms is redundant (the excess relative to the least abundant element is included in the other term), but would otherwise be nonzero (Δ is a constant equal to 10⁻¹⁵, to prevent divide-by-zero). For three elements, there are 3! = 6 possible cases: for N greater or less than C_PR_{NC}, Fe may be either in excess relative to both C and N, deficient relative to both, or in excess relative to one but not the other (Table 2).

287 2.3 Organic and Inorganic Pools

288

289 There are two pools of detritus with different sinking rates but the same fixed elemental ratios.

290 Detrital C/N/Fe ratios are the same as zooplankton, so zooplankton mortality or grazing of small

291 zooplankton by large zooplankton produce no 'excess'. Phytoplankton mortality, and defecation

by zooplankton grazing on phytoplankton, produces excess nutrient or excess C that needs to be

recycled into the inorganic pool in a similar fashion as outlined above for the assimilated fraction

294 of grazing on phytoplankton.

295 The conservation equations for detrital C are

296
$$\frac{dD_s}{dt} = m_1(C_{ps} + Z_s) + m_2(C_{ps}^2 + Z_s^2) - r_1 D_s T_g - w_s \frac{dD_s}{dz}$$
(17a)

297
$$\frac{\mathrm{d}D_l}{\mathrm{d}t} = m_1(C_{\mathrm{pl}} + Z_L) + m_2(C_{pl}^2 + Z_L^2) - r_2 D_l T_g - w_l \frac{\mathrm{d}D_l}{\mathrm{d}z}$$
(17b)

where T_g is an Arrhenius function for temperature dependence of remineralization and *w* is the sinking speed. The conservation equations for inorganic C, N, and Fe are

300
$$\frac{dC_i}{dt} = (\zeta V^N - P_{\text{phot}}^C)C_p + R + C_{\text{XS}} + (r_1 D_s + r_2 D_l)T_g$$
(18a)

301
$$\frac{dN_i}{dt} = -\frac{V^N}{Q^N} N_p(\frac{L_{NO3}}{L_{NO3} + L_{NH4}}) + N_{ox} - N_{dentr}(1 - A_f)$$
(18b)

302
$$\frac{dN_a}{dt} = -\frac{V^N}{Q^N} N_p \left(\frac{L_{\text{NH4}}}{L_{\text{NO3}} + L_{\text{NH4}}}\right) + \frac{R}{R_{\text{CN}}} + N_{\text{XS}} + (r_1 D_s + r_2 D_l) R_{\text{NC}} T_g - N_{\text{ox}} + N_{\text{dnf}} - N_{\text{dentr}} A_f \quad (18c)$$

303
$$\frac{\mathrm{dFe}}{\mathrm{dt}} = \frac{V^{\mathrm{Fe}}}{Q^{\mathrm{Fe}}} \mathrm{Fe}_p + \frac{R}{R_{\mathrm{CFe}}} + \mathrm{Fe}_{\mathrm{XS}} + (r_1 D_s + r_2 D_l) R_{\mathrm{FeC}} T_g$$
(18d)

where N_{ox} is microbial oxidation of ammonium to nitrate (nitrification), N_{dnf} and N_{dentr} are sources and sinks associated with dinitrogen fixation and denitrification, and A_f is the ammonium fraction of denitrification losses, associated with anaerobic ammonium oxidation ("anammox"). The oxygen equation is essentially the inverse of equation 18a, with additional terms foroxidation and reduction of N, i.e.,

$$309 \quad \frac{dO_2}{dt} = -\frac{dC_i}{dt} + 2\frac{V^N}{Q^N}N_p(\frac{L_{NO3}}{L_{NO3}+L_{NH4}}) - 2N_{ox}$$
(19)

310 Nitrification is given by

311
$$N_{\rm ox} = k_{\rm NH4ox} N_a \frac{\kappa_E}{\kappa_E + E(z)}$$
(20)

where E(z) is the layer mean irradiance at depth *z*. Dinitrogen fixation is parameterized as an external input of ammonium dependent on light, temperature and Fe availability, and inhibited by high ambient concentrations of inorganic N,

315
$$N_{\rm dnf} = k_{\rm dnf} T_{\rm dnf} (1 - e^{-aE}) \left(\frac{Fe}{K_{\rm Fe} + Fe} \right) \left(\frac{K_{\rm NO3}}{K_{\rm NO3} + N_i + N_a} \right)$$
 (21)

where T_{dnf} =max(0, 1.962(T_f - 0.773)), i.e., a linear multiple of equation (1) that is 0 at T<20°C and unity at T=30°C.

318

319 Denitrification is parameterized as a fraction of total remineralization that increases as a linear 320 function of oxygen concentration for concentrations less than a threshold concentration O_{mxd}

321
$$N_{\text{frxn}} = 1 - \frac{\min(O_2, O_{\text{mxd}})}{O_{\text{mxd}}}$$
 (22)

Remineralization is then divided among oxygen $(1-N_{frxn})$, nitrate $(0.875N_{frxn})$, and ammonium ($0.125N_{frxn}$) assuming an average anammox contribution of 25% (Babbin et al., 2014). We use this average ratio of anammox to classical denitrification to partition fixed N losses between NO₃⁻ and NH₄⁺; the DIC sink and organic matter source associated with anammox are small and are neglected here.

328 **2.4** Calcification, Calcite Dissolution, and Alkalinity

329

- 330 In CanOE, calcification is represented by a prognostic detrital calcite pool with its own sinking
- rate (distinct from that of organic detritus), and calcite burial or dissolution in the sediments
- <u>depends on the saturation state (100% burial when $\Omega_{C} \ge 1$, 100% dissolution when $\Omega_{C} \le 1$).</u>
- 333 Calcification is represented by a detrital calcium carbonate (CaCO₃) state variable, but no
- explicit calcifier groups. Detrital CaCO₃ sinks in the same fashion as detrital particulate organic
- carbon (POC), with a sinking rate independent of those for large and small organic detritus.
- 336 Calcite production is represented as a fixed fraction of detritus production from small
- 337 phytoplankton and small zooplankton mortality:

338
$$\frac{dCa}{dt} = m_1 (C_{ps} + Z_s) P_{Ca} + m_2 (C_{ps}^2 + Z_s^2) P_{Ca} - k_{Ca} Ca - w_{Ca} \frac{dCa}{dz}$$
(23)

339 Calcite dissolution occurs throughout the water column as a first order process (i.e., no dependence on temperature or saturation state). Approximately 80% of calcite produced is 340 exported from the euphotic zone. Burial in the sediments is represented as a simple 'on/off' 341 342 switch dependent on the calcite saturation state (zero when $\Omega_C < 1$ and 1 when $\Omega_C \geq 1$). In 343 CanESM5, calcification is parameterized by a temperature dependent "rain ratio" (Zahariev et al., 2008) and 100% burial of calcite that reaches the seafloor is assumed. Calcite burial in both 344 models is balanced by an equivalent source of DIC and alkalinity at the ocean surface (in the 345 346 same vertical column) as a crude parameterization of fluvial sources.

347

348 For each mole of calcite production two moles of alkalinity equivalent are lost from the

- 349 dissolved phase; the reverse occurs during calcite dissolution. There are additional sources and
- sinks for alkalinity associated with phytoplankton nutrient (NH_4^+, NO_3^-) uptake, organic matter

351 remineralization, nitrification, denitrification and dinitrogen fixation (Wolf-Gladrow et al., 2007, see Supplementary Table S2). The anammox reaction does not in itself contribute to alkalinity 352 (Jetten at al., 2001), but there is a sink associated with ammonium oxidation to nitrite (the model 353 does not distinguish between nitrite and nitrate). Autotrophic production of organic matter by 354 anammox bacteria is a net source of alkalinity (Strous et al., 1998) but this source is extremely 355 356 small (~0.03 mol/molN) and is neglected here. Globally, the sources and sinks of alkalinity from the N cycle offset each other such that there is no net gain or loss as long as the global fixed N 357 pool is conserved (see below Sect. 2.5). If dinitrogen fixation and denitrification are allowed to 358 359 vary freely, there will generally be a net gain or loss of fixed N and, therefore, of alkalinity.

360

- 361 **2.5 External Nutrient Sources and Sinks**
- 362

External sources and sinks consist of river inputs, aeolian deposition, biological N₂ fixation, 363 364 denitrification, mobilization of Fe from reducing sediments, loss of Fe to scavenging, and burial of calcium carbonate in the sediments. Aeolian deposition of Fe is calculated from a climatology 365 366 of mineral dust deposition generated from offline (atmosphere-only) simulations with CanAM4 367 (von Salzen et al., 2013), with an Fe mass fraction of 5% and a fractional solubility of 1.4% in 368 the surface layer. Subsurface dissolution is parameterized based on PISCESv2 (Aumont et al., 369 2015); the total dissolution is 6.35%, with 22% of soluble Fe input into the first vertical layer 370 (see Supplementary material). Iron from reducing sediments is also based on PISCES, with a constant areal flux of 1000 nmol m⁻² d⁻¹ in the first model level, declining exponentially with 371 372 increasing seafloor depth (i.e., assuming that shelf sediments are the strongest source and the 373 sediments become progressively more oxygenated with increasing seafloor depth) with an efolding length scale of about 200 m. Scavenging of dissolved iron is first-order with a high rate (2.5 d⁻¹) for concentrations in excess of 0.6 nM (Johnson et al., 1997). For concentrations below this threshold, the rate is much lower (0.001 d⁻¹) and is weighted by the concentration of organic detritus (Christian et al., 2002b), i.e.,

378
$$\frac{dFe}{dt} = -FeS_{Fe1}min\{(D_S + D_L)P_{Fe}, 1\}$$
(24)

379 where Fe is the dissolved iron concentration, D_S and D_L are the small and large detritus 380 concentrations, S_{Fe1} is the first-order scavenging rate in surface waters with abundant particulates, and P_{Fe} is an empirical parameter to determine the dependence on particle 381 382 concentration (Table 1). The basis for this parameterization is that the rate of scavenging must depend not only on the concentration of iron but on the concentration of particles available for it 383 384 to precipitate onto, and assumes that POC is strongly positively correlated with total particulate matter. Scavenging is treated as irreversible, i.e., scavenged Fe is not tracked and does not 385 reenter the dissolved phase. 386

387

388 Unlike in CMOC, N₂ fixation and denitrification vary independently in CanOE, so the global total N pool can change. Conservation is imposed by adjusting the global total N pool according 389 to the difference between the gain from N₂ fixation and the loss to denitrification. A slight 390 adjustment is applied to the nitrate concentration at every grid point, while preserving the overall 391 spatial structure of the nitrate field. Adjustments are multiplicative rather than additive to avoid 392 producing negative concentrations. This adjustment does not maintain (to machine precision) a 393 constant global N inventory but is intended to minimize long term drift, keeping it much smaller 394 than the free surface error (see below). This adjustment is applied every 10 days and has a 395 magnitude of approximately $7x10^{-8}$ of the total N. 396

398	When the total fixed N adjustment is applied, One one mole of alkalinity is removed per mole of
399	N added or removed, to account for the since there are alkalinity sources of 1 mol/molN
400	associated with both N_2 fixation (creation of new NH_4^+) and denitrification (removal of NO_3^-)
401	(Wolf-Gladrow et al., 2007, see Supplementary Table S2). As there is, offset by a 2 mol/molN
402	sink associated with nitrification, this formulation is globally conservative. As noted above, in
403	CanOE CaCO ₃ can dissolve or be buried in the sediments depending on the calcite saturation
404	state. DIC and alkalinity lost to burial are reintroduced at the ocean surface, at the same grid
405	point as burial occurs, providing a crude parameterization of river inputs so that global
406	conservation is maintained (fresh water runoff contains no DIC or alkalinity). However, the OPA
407	free surface formulation is inherently imperfect with regard to tracer conservation. Drift in total
408	ocean alkalinity and nitrogen over time is on the order of 0.01% and 0.03% per thousand years,
409	respectively (losses due to the free surface are generally larger for tracers with less homogeneous
410	distributions).

2.6 Ancillary data

For first-order model validation we have relied largely on global gridded data products rather
than individual profile data. Global gridded data from World Ocean Atlas 20132018
(WOA20132018) (Locarnini et al., 20132018; Zweng et al., 20132018; Garcia et al.,
2014a2018a; 2014b2018b) were used for temperature, salinity, and oxygen and nitrate
concentration. DIC and alkalinity were taken from the GLODAPv2.2016b gridded data product
(Key et al., 2015; Lauvset et al., 2016). Offline carbon chemistry calculations were done

following the Best Practices Guide (Dickson et al., 2007) and the OMIP-BGC protocols (Orr et al., 2017), which and are identical to those used in the models except that constant reference concentrations were used for phosphate (1 μ M) and silicate (10 μ M).

423

There is no global gridded data product for Fe, but we have made use of the GEOTRACES 424 Intermediate Data Product 2017 (Schlitzer et al., 2018), and the data compilations from MBARI 425 (Johnson et al., 1997; 2003) and PICES Working Group 22 (Takeda et al., 2013). The latter two 426 are concentrated in the Pacific, while GEOTRACES is more global. The combined data sets 427 provide more than 10000 bottle samples from more than 1000 different locations (Supplementary 428 429 Figure <u>S4aS9a</u>) (excluding some surface transect data that involve frequent sampling of closely spaced locations along the ship track). More detail about model comparison to these data 430 compilations and the list of original references are given in the Supplementary information. 431

432

433 Satellite ocean colour estimates of surface chlorophyll were taken from the combined SeaWiFS/MODIS climatology described by Tesdal et al. (2016). Climatological satellite POC 434 435 was downloaded from the NASA ocean colour web site and is based on the algorithm of 436 Stramski et al. (2008) using MODIS-Aqua data. This climatology differs slightly from the 437 chlorophyll one in terms of years included and sensors utilized, but as only climatological 438 concentrations are considered and each climatology covers ~15 years, these differences will have 439 negligible effect on the results presented. Satellite chlorophyll concentrations greater than 1 mg m⁻³ were excluded as these are mostly associated with coastal regions not resolved by coarse-440 441 resolution global ocean models.

443	CMIP6 model data were regridded by distance-weighted averaging using the Climate Data
444	Operators (https://code.mpimet.mpg.de/projects/cdo/) to a common grid (2x2°, 33 levels) to
445	facilitate ensemble averaging. The vertical levels used are those used in GLODAP and in earlier
446	(through 2009) versions of the World Ocean Atlas (e.g., Locarnini et al., 2010). following the
447	GLODAP levels) to facilitate ensemble averaging. For large scale tracer distributions, using a 1°
448	or 2° grid makes little difference (for example, the spatial pattern correlation between CanESM5
449	and observed oxygen concentration at specific depths differs by an average of 0.0011). The years
450	1986-2005 of the Historical experiment were averaged into climatologies or annual means, for
451	meaningful comparison with observation-based data products. The CMIP6 Historical experiment
452	runs from 1850-2014 with atmospheric CO ₂ concentration (and other atmospheric forcings)
453	based on historical observed values. A single realization was used in each case (see Table S3); as
454	20 year averages are used, internal variability is assumed to have little effect (e.g., Arguez and
455	Vose, 2011, see Table S4). Where time series are shown, 5-year means are used.
456	
457	Sampling among CMIP6 models was somewhat opportunistic and the exact suite of models
458	varies among the analyses presented. When we conducted a search for a particular data field, we
459	included in the search parameters all models that published that field, and repeated the search at
460	least once for models that were unavailable the first time the search was executed. In some cases,
461	model ensemble means excluded all but one model from a particular 'family' (e.g., there are
462	three different MPI-ESM models for which ocean biogeochemistry fields were published), as the
463	solutions were found to be similar and would bias the ensemble mean towards their particular
464	climate. The models used are ACCESS-ESM1-5, CESM2, CESM2-WACCM, CNRM-ESM2-1,
465	GFDL-CM4, GFDL-ESM4, IPSL-CM6A-LR, MIROC-ES2L, MPI-ESM-1-2-HAM, MPI-

466	ESM1-2-LR, MPI-ESM1-2-HR, MRI-ESM2-0, NorESM2-LM, NorESM2-MM, and UKESM1-
467	0-LL. More dDetails of which variables and realizations are used for which models are is given in
468	Supplementary Table S3.
469	
470	3. Results
471	
472	We first describe here the large-scale distribution of oxygen, DIC, alkalinity, and the saturation
473	state with respect to CaCO ₃ that derives from these large-scale tracer distributions. Tracer
474	distributions result partly from ocean circulation and partly from biogeochemical processes. An
475	overall evaluation of the ocean circulation model is given in Swart et al. (2019a). Analyzing
476	CanESM5 and CanESM5-CanOE (with identical circulation) as well as CanESM2 where
477	possible (same biogeochemistry as CanESM5 but different circulation) allows us to separate the
478	effects of physical circulation and biogeochemistry on evolving model skill with respect to large-
479	scale tracer distributions. In subsequent sections we address the main areas where CanESM5 and
480	CanESM5-CanOE differ, such as the interaction of the iron and nitrogen cycles and plankton
481	community structure. Finally, we present some temporal trends over the course of the historical
482	experiment (1850-2014).
483	
484	3.1 Distribution of oxygen
485	
486	The spatial distribution of oxygen concentration ([O ₂]) at selected intermediate depths (400, 900,
487	and 1400 m) is shown in Figure 2 for gridded data from WOA2018 and differences of
488	CanESM5, CanESM5-CanOE, a model ensemble mean (MEM) of CMIP6 models (excluding
	23

489	CanESM5 and CanESM5-CanOE) from the observational data product., and gridded data from
490	WOA2013 is shown in Figure 2. The depths were chosen to span the depth range where low
491	oxygen concentrations exist; these low-oxygen environments are of substantial scientific and
492	societal interest and are sensitive to model formulation. The major features are consistent across
493	the models. Both CanESM models as well as the MEM All three cases show elevated oxygen
494	concentrations relative to observations, particularly in the North Pacific, the North Atlantic and
495	the Southern Ocean. In the Indian Ocean, both CanESM models show high oxygen
496	concentrations in the Arabian Sea and deeper layers of the Bay of Bengal relative to observations
497	and other CMIP6 models the MEM; these biases are somewhat smaller in CanESM5-CanOE than
498	in CanESM5 (Figure 2).
499	
500	The ocean's oxygen minimum zones (OMZs) are mostly located in the eastern Pacific Ocean, the
501	northern North Pacific, and the northern Indian Ocean; the spatial pattern changes with
502	increasing depth (Figure 2), but the OMZs are mostly located between 200 and 2000 m depth.
503	Biases in the eastern boundary current EBC regions are depth and model specific. CanESM5
504	shows particularly strong oxygen depletion at 1400 m in the eastern tropical Pacific. In the
505	southeastern Atlantic, models tend to be biased low at the shallower depths, and show somewhat
506	more variation at greater depths (Figure 2). Overall, [O ₂] biases tend to be positive over large
507	areas of ocean with the exception of some eastern boundary current EBC regions, implying that
508	models exaggerate the extent to which remineralization is concentrated in these regions. An
509	alternate version of Figure 2 that shows the modelled concentrations model errors relative to the
510	observational data product is given in Supplementary Figure <u>\$352</u> .
1	

512	The zonal mean oxygen concentration, saturation concentration, and apparent oxygen utilization
513	(AOU) are shown in Figure 3 for the same four cases. Again, the models generally show a
514	positive bias in [O ₂], particularly in high-latitude deep waters. The major ocean circulation
515	features are reproduced fairly well in all cases (e.g., weaker ventilation of low-latitude
516	subsurface waters, greater vertical extent of well-ventilated surface waters in the subtropics). The
517	saturation concentration (a function of temperature and salinity) generally shows relatively little
518	bias, implying that the bias in [O ₂] arises mainly from remineralization and/or ventilation. AOU
519	is lower than observed over much of the subsurface ocean. Regional biases are quite consistent
520	across models, but are slightly greater in CanESM5 than in CanESM5-CanOE or the ensemble
521	mean <u>MEM</u> , except in the Arctic Ocean. Again, Supplementary Figure S23 includes a version of
522	this plot that shows the modelled differences from the observations concentration fields.

524 The skill of each model with respect to the distribution of O_2 at different depths is represented by 525 Taylor diagrams (Taylor, 2001) in Figure 4. These diagrams allow us to assess how well the 526 model reproduces the spatial distribution at a range of depths, because different physical and 527 biogeochemical processes determine the distribution in different depth ranges, in which, all All of the CMIP6 models that were shown as an ensemble mean in Figures 2 and 3 are shown 528 529 individually. The large blue dots represent CanESM5, red CanESM5-CanOE, and grey the ensemble mean of all CMIP6 models except CanESM5 and CanESM5-CanOEMEM; the smaller 530 531 grey dots represent the individual models. CanESM5-CanOE shows slightly higher pattern 532 correlation than CanESM5 at all depths. Both models compare favourably with the full suite of CMIP6 models, with r>0.85 for CanESM5 and r>0.9 for CanESM5-CanOE at all depths 533 534 examined, and a normalized standard deviation within $\pm 25\%$ of unity.

536	The total volume of ocean with $[O_2]$ less than 6 mmol m ⁻³ (the threshold for denitrification
537	(Devol, 2008)) and 60 mmol m ⁻³ (a commonly used index of hypoxia) is shown in Figure 5. The
538	total volume is highly variable among models (note, however, that there are several clusters of
539	related models with quite similar totals). CanESM5 and CanESM5-CanOE have among the
540	lowest total volumes (i.e., the interior ocean is relatively well ventilated) and are among the
541	nearest to the observed total. For $[O_2] \leq 60 \text{ mmol m}^{-3}$ the bias is, nonetheless, quite large (i.e., the
542	observed volume is underestimated by almost 50% in both models). The volume of water with
543	[O ₂] below the denitrification threshold is overestimated in both CanESM5 and CanESM5-
544	CanOE; CanESM5-CanOE has a much smaller total that is closer to the observed value. The bias
545	in the spatial pattern of hypoxia (not shown) is generally similar to the bias in dissolved oxygen
546	distribution (Figure 2). The low-oxygen regions are generally more concentrated in the eastern
547	tropical Pacific in the models than in observations, and the low-oxygen region in the northwest
548	Pacific is not well reproduced in CanESM models.
549	

550 **3.2 Distribution of DIC, alkalinity, and CaCO3 saturation**

551

The spatial distribution of aragonite saturation state (Ω_A) at selected depths is shown in Figure 6. (<u>#The first two depths are the same as in Figure 2, but a much greater depth is also included, as</u> the length scale for CaCO₃ dissolution is greater than for organic matter remineralization the third is much deeper). In this case the observations are a combination of GLODAPv2 (Key et al., 2015; Lauvset et al., 2016) for DIC and alkalinity, and WOA20132018 for temperature and salinity. CanESM5 and CanESM5-CanOE generally compare well with other models and

558	observations. The low saturation bias in the eastern tropical Pacific is substantially reduced in			
559	CanESM5-CanOE compared to CanESM5. On the other hand CanESM5 generally does better			
560	than CanESM5-CanOE, or the ensemble meanMEM, at reproducing the low saturation states in			
561	the northwestern Pacific and the Bering Sea. Both <u>CanESM</u> models show a high saturation state			
562	bias in the North Atlantic and the well-ventilated regions of the north Pacific subtropical gyre			
563	These these biases are reduced slightly smaller in CanESM5-CanOE, probably due to the smaller			
564	average remineralization length scale for organic detritus. Maps of the calcite and aragonite			
565	saturation horizon (Ω =1) depth are shown in Supplementary Figure S3; these generally confirm			
566	the same biases noted in Figure 6.			
567				
568	Zonal mean distributions of aragonite saturation state (Ω_A), calcite saturation state (Ω_C), and			
569	carbonate ion concentration ([CO ₃]) and the differences of the models from the observations are			
570	shown in Figure 7 (Supplementary Figure <u>S3-S2</u> includes versions of Figures 6 and 7 that			
571	explicitly show the modelled fields differences from the observations). The models generally			
572	compare well with the observations in the representation of the latitude/depth distribution of high			
573	and low saturation waters. CanESM5 has a high saturation bias in low-latitude surface waters			
574	that is somewhat reduced in CanESM5-CanOE. Both CanESM5 models show a high saturation			
575	bias in Northern Hemisphere intermediate (e.g., 200-1000 m) depth waters that is larger than in			
576	the ensemble mean <u>MEM</u> . This is primarily a result of low Ω in the North Atlantic Ocean (Figure			
577	<u>6).</u>			
l 578				
570	Taylor diagrams for a range of depths are shown for DIC in Figure 8 and for O, in Figure 9 (for			

579 Taylor diagrams for a range of depths are shown for DIC in Figure 8 and for Ω_A in Figure 9 (for 580 alkalinity, see Supplementary Figure S42). As expected, the MEM generally compares

581	favourably with the individual models (e.g., Lambert and Boer, 2001). CanESM5 and
582	CanESM5-CanOE compare favourably with the full suite of CMIP6 models. CanESM5-CanOE
583	shows a gain in skill relative to CanESM5, and both show improvement relative to CanESM2. At
584	400 m, CanESM2 stands out as having extremely high variance, which is mostly due to
585	extremely high DIC concentrations occurring over a limited area in the eastern equatorial Pacific
586	(not shown). This bias is present in CanESM5 and in CMIP6 models generally (Figure 6) but
587	involves much lower concentrations spread over a larger area.
588	

589 **3.3** N and Fe cycles

590

An important difference between CMOC-CanESM5 and CanESM5-CanOE is the inclusion of a 591 prognostic Fe cycle. The CMOC iron mask (Zahariev et al., 2008) was a pragmatic solution in 592 the face of resource limitations but is inherently compromised as it can not evolve with a 593 594 changing climate. Other centres that introduced a prognostic Fe cycle between CMIP5 and 595 CMIP6 include JAMSTEC (MIROC-ESM, MIROC-ES2L) and the UK Met Office (HadGEM2-596 ES, UKESM1-0-LL). The first order test of a model with prognostic, interacting Fe and N cycles 597 is whether it can reproduce the distribution of High-Nutrient, Low-Chlorophyll (HNLC) regions 598 and the approximate surface macronutrient concentrations within these. CanESM5-CanOE 599 succeeded by this standard, although the surface nitrate concentrations are biased low in the 600 subarctic Pacific and equatorial Pacific and high in the Southern Ocean and in the global mean (Figure 10). 601

602

603 The seasonal cycle of the zonal mean surface nitrate concentration for a selection of CMIP6

604	models is shown in Figure 11. CanESM5, CanESM5-CanOE, and CNRM-ESM2-1 reproduce the
605	equatorial enrichment and the low concentrations in the tropical-subtropical latitudes fairly well.
606	Some models either have very weak equatorial enrichment (MPI-ESM1-2-LR) or too high a
607	concentration in the off-equatorial regions (UKESM1-0-LL, NorESM2-LM). UKESM1-0-LL
608	has very high concentrations throughout the low-latitude Pacific, which biases the ensemble
609	mean (Figure 11a). <u>Supplementary</u> Figure <u>S611b</u> shows the same data as Figure 11a but for a
610	more limited latitude range to better illustrate model behaviour in the tropics. CanESM5,
611	CanESM5-CanOE, and CNRM-ESM2-1 reproduce the seasonal cycle of tropical upwelling (e.g.,
612	Philander and Chao, 1991), with highest concentrations in summer.

The surface distribution of dissolved iron (dFe) in various CMIP6 models is shown in Figure 12. 614 For Fe there is no observation-based global climatology with which to compare the model 615 616 solutions (some comparisons to available profile data are shown in Supplementary Figures 617 <u>S4bS9b-gh</u>). CanESM5-CanOE shows a similar overall spatial pattern to other models, and generally falls in the middle of the spread, particularly regarding concentrations in the Southern 618 Ocean. Several models show extremely high concentrations in the tropical-subtropical North 619 620 Atlantic (Sahara outflow region). CanESM5-CanOE, along with CNRM-ESM2-1 and CESM2, 621 has much less elevated concentrations in this region, due to lower deposition or greater scavenging or both. CanESM5-CanOE has its lowest concentration in the eastern subtropical 622 623 South Pacific, which is common to many models (Figure 12). The area of strong surface 624 depletion is generally more spatially restricted in CanESM5-CanOE than in other models, and 625 surface dFe concentrations are greater over large areas of the Pacific. Both the north-south and 626 east-west asymmetry of distribution in the Pacific is greater in CanESM5-CanOE than in most

other models, some of which show the South Pacific minimum extending westward across the
entire basin, and others into the Northern Hemisphere. Only in CESM2 is this minimum
similarly limited to the southeast Pacific.

630

The mean depth profiles of dFe are shown in Figure 13. Some models show more of a "nutrient-631 type" (increasing with depth due to strong near-surface biological uptake and subsequent 632 remineralization) profile, some a more "scavenged-type" (maximal at the surface, declining with 633 depth) profile (cf. Li, 1991; Nozaki, 2001), and others a hybrid profile (increasing downward but 634 with a surface enrichment). CanESM5-CanOE is at the "nutrient-type" end of spectrum with a 635 636 generally monotonic increase with depth to a near-constant deep-water concentration of 0.6 nM and a very slight near-surface enrichment (see also Supplementary Figures S9S4b,c). In 637 CanESM5-CanOE the scavenging model is very simple, with distinct regimes for concentrations 638 greater or less than 0.6 nM; scavenging rates are very high above this threshold which causes 639 deep-water concentrations to converge on this value. The generally nutrient-like profile suggest 640 that in CanOE the scavenging rate is quite low for concentrations below 0.6 nM. CanOE 641 considers particulate organic matter (POM) as an index of all particulate matter available for 642 scavenging onto, and model POM concentrations fall off rapidly below the euphotic zone (not 643 shown). 644

645

Mean surface nitrate and dFe concentrations for selected ocean regions are shown in Figure 14.
CanESM5-CanOE shows concentrations that are within the range of CMIP6 models, although in
some cases at the higher or lower end. Surface nitrate concentrations generally compare
favourably with the observation-based climatology, but are biased low in HNLC regions other

650 than the Southern Ocean. These biases are not necessarily a consequence of having too much or too little iron. For example, in the Southern Ocean CanESM5-CanOE has among the highest 651 surface nitrate concentrations, but it also has some of the highest dFe concentrations, and the 652 high nitrate bias is present in CanESM5 as well. Comparisons with the limited GEOTRACES 653 data available suggest that near surface dFe concentrations in the Southern Ocean are biased high 654 rather than low in CanESM5-CanOE (not shown). One region where there does seem to be a 655 strong correlation between surface nitrate and dFe concentrations is the western subarctic 656 657 Pacific. All but two models (CNRM-ESM2-1, NorESM2-LM) fall along a spectrum from high 658 Fe / low nitrate to low Fe / high nitrate. CanESM5-CanOE falls near the high Fe / low nitrate end of the range. 659

660

Surface nitrate concentrations along the Pacific equator during the upwelling season (June-661 662 October) for CanESM5 and CanESM5-CanOE are shown in Figure 15. The range of other CMIP6 models is not shown here because it is large and therefore adds little information (see 663 Figure 11 and Supplementary Figure S6). CanESM5-CanOE better represents the east-west 664 gradient, while CanESM5 has slightly higher concentrations in the core upwelling region. Both 665 666 models underestimate the highest concentrations around 100°W. Some Although some localized 667 maxima in this data product are due to undersampling, equatorial upwelling is strong at this location (e.g., Lukas, 2001) and the spatial coherence of the data strongly suggests that this 668 669 maximum accurately reflects reality; however, examination of ancillary data sets such as satellite 670 sea surface temperature suggests that the enrichment at 100°W accurately reflects ocean upwelling (not shown). Although It should be noted that CanESM5 iron limitation is calculated 671 672 from an earlier version of the same data product, however, the Fe mask is based on the

673	minimum nitrate concentration over the annual cycle, whereas the data shown here are for th			
674	upwelling season. In CanESM5-CanOE, the distribution of surface nitrate is an emergent			
675	property of the model, and the fidelity to the observed distribution is generally good.			
676				

677 3.4 Plankton biomass, detritus, and particle flux

678

The relative abundance of the four living plankton groups are shown in Figure 16 for a range of ocean regions. Both CanESM models mostly compare favourably with observation-based estimates of phytoplankton biomass, except in the tropics where CanESM5-CanOE has very high biomass. Both CanESM models have low phytoplankton biomass in the North Atlantic. In the North Pacific and the Southern Ocean, CanESM5-CanOE reproduces the observation-based estimates well, and CanESM5 slightly less well. The general pattern is that large and small phytoplankton have similar abundance, and are substantially more abundant than zooplankton.

687 Part of the rationale for multiple food chains is that they better represent the way that actual plankton communities adapt to different physical ocean regimes and therefore are better able to 688 simulate distinct ocean regions with a single parameter set (e.g., Chisholm, 1992; Armstrong, 689 690 1994; Landry et al., 1997; Friedrichs et al., 2007). The expectation is that small phytoplankton will be more temporally stable and large phytoplankton will fluctuate more strongly between 691 high and low abundances. The mean annual cycles of surface chlorophyll largely conform to 692 693 thise expected pattern, e.g., in the North Atlantic and the western subarctic Pacific large phytoplankton are dominant in summer and much more variable over the seasons (Figure 17). 694 Compared to observations, CanESM5 models underestimate the amplitude of the seasonal cycle 695

696 in the North Atlantic and overestimate it in the North Pacific. CanESM5 shows a stronger and earlier North Atlantic spring bloom compared to CanESM5-CanOE; the observations are in 697 between the two in terms of timing, and both models underestimate the amplitude (Figure 17). In 698 699 the tropics, the seasonal cycle is weak. CanESM5-CanOE in Tthe tropical Atlantic shows the expected seasonal cycle but not the expected dominance of large phytoplankton in summer. 700 These size-fractionation patterns are difficult to validate against observations. CanESM5-CanOE 701 generally overestimates the total near surface chlorophyll in both the tropical Pacific and the 702 tropical Atlantic. 703

704

705 Zooplankton biomass (especially microzooplankton) is also somewhat difficult to test against observations, but our model concentrations appear to be biased low. White et al. (1995), for 706 example, show a transect of vertically resolved mesozooplankton abundance along 140°W in the 707 tropical Pacific, with biomass ranging from about 0.1-0.7 mmolC m³; CanESM5-CanOE 708 concentrations in this region are much lower. Stock et al. (2014) estimated depth-integrated 709 biomass of phytoplankton, mesozooplankton, and microzooplankton for a range of oceanic 710 locations in which intensive field campaigns have occurred (estimates of microzooplankton 711 biomass are relatively sparse). They found that in most locations phytoplankton and (combined) 712 zooplankton biomass are of comparable magnitude, whereas in CanESM5-CanOE zooplankton 713 biomass is consistently lower (Figure 16). The global integral biomass of mesozooplankton is 714 715 about an order of magnitude less than the 0.19 PgC estimated by Moriarty and O'Brien (2013). 716 The CanESM5 total of 0.14 Pg is relatively close to the Moriarty estimate but implicitly includes 717 microzooplankton.

718

719 Surface chlorophyll and POC for CanESM5-CanOE and for ocean colour observational data are shown in Figure 18 (POC in the model is the sum of phytoplankton, microzooplankton, and 720 detrital carbon). The observations have a lower limit for POC that is not present in the model 721 722 (~17 mgC m⁻³), which is unsurprising given the processes neglected in the model, i.e., in regions of very low chlorophyll there is still substantial dissolved organic carbon, bacteria that consume 723 724 it, and microzooplankton that consume the bacteria and produce particulate detritus. The observational data show a fairly linear relationship at low concentrations, but with a curvature 725 that implies a greater phytoplankton fraction in more eutrophic environments (cf. Chisholm, 726 727 1992). The model, by contrast, shows a fairly linear relationship over the whole range of concentrations. In other words, the phytoplankton share of POC is higher and more constant in 728 the model than in the observations. The living biomass (phytoplankton + microzooplankton) 729 fraction of total POC in CanOE is generally in excess of 50% (not shown), which is implausible 730 for a real-world oceanic microbial community (e.g., Christian and Karl, 1994) but consistent 731 with the relatively low rates of export from the euphotic zone. 732

733

Export production for a range of CMIP6 models is shown in Figure 19a. CanESM5-CanOE is at 734 the low end of the range. Observations are not shown because the range of observational 735 estimates covers almost the entire range of the plotmodel estimates (e.g., Siegel et al., 2016). 736 Note also that CanESM5 export is quite a bit lower than in CanESM2, which is relatively high 737 738 for CMIP5 models (not shown). The difference between CanESM2 and CanESM5 is attributable 739 primarily to different circulation, although the different initialization fields for nitrate might also play a small role. The lower rate in CanESM5-CanOE is consistent with the above results 740 741 regarding plankton community structure (e.g., the concentration of detritus is generally low

compared to living biomass), as well as the lower sinking rate for small detritus. The latitudinal
distribution of export is shown in Figure 19b. CanESM5 shows very high export in the midlatitudes of the Southern Ocean, similar to CanESM2 (not shown). Both CanESM5 and
<u>CanESM5-CanOE models</u> show latitudinal patterns consistent with the range of other <u>CMIP6</u>
models. CanESM5 has slightly greater export in the equatorial zone; in both CanESM<u>5 and</u>
<u>CanESM5-CanOE models</u> the equatorial enrichment attenuates very rapidly with latitude and the
rates are low in the subtropics.

749

750 **3.5 Historical trends**

751

752 Cumulative ocean uptake of CO_2 is shown in Figure 20 for the historical experiment (1850-2014). CanESM models are biased low relative to observation based estimates (~145 PgC, see 753 754 Friedlingstein et al., 2020) and the ensemble mean of other modelsMEM (144 PgC, Figure 20), 755 but fall well within the spread of CMIP6 models. CanESM5-CanOE has lower cumulative 756 uptake than CanESM5 by ~ 10 PgC. As the models were not fully equilibrated when the 757 historical run was launched, this difference does not necessarily arise from the biogeochemical 758 model structure; part of the difference can be attributed to differences in the spinup protocol (cf. 759 Séférian et al., 2016). The drift in the piControl experiment over the 165 years from the 760 branching off of the historical experiment is -10.5-0 PgC in CanESM5-CanOE and -5.9-1 PgC in 761 CanESM5 (see Supplementary Table S6), so drift accounts for about half (484%) of the 762 difference in net ocean CO₂ uptake. The vertical spatial distribution of anthropogenic DIC is 763 very similar between CanESM5 and CanESM5-CanOE (not shownSupplementary Figure S7).

765 The long-term trend in global total export production is shown in Figure 21. The model values must be normalized in order to compare trends, since the differences among means are large 766 compared to the changes over the historical period (Figure 19). Such trends are difficult or 767 impossible to meaningfully constrain with observations, but the general expectation has been that 768 export will decline somewhat due to increasing stratification (e.g., Steinacher et al., 2010). 769 CanESM5 shows a greater decline than most other CMIP6 models, while CanESM5-CanOE is 770 771 more similar to non-CanESM models. Such trends are difficult or impossible to meaningfully constrain with observations, but the general expectation has been that export will decline 772 somewhat due to increasing stratification (e.g., Steinacher et al., 2010). The change in CanESM5 773 is geographically widespread and not concentrated in a specific region or regions: export is 774 maximal in the tropics and the northern and southern mid-latitudes (Figure 19b) and declines 775 776 over the historical period in all of these regions (Supplementary Figure S84). In CanESM5-CanOE, export declines in the same regions, but the magnitude of the change is smaller, and in 777 the Southern Ocean increases and decreases in different latitude bands largely offset each other. 778

779

The trend in the volume of ocean water with O₂ concentration less than 6 or 60 mmol m⁻³ is 780 shown in Figure 22. Again, the totals are normalized to a value close to the preindustrial, as the 781 differences among models are large (Figure 5). For the volume with <60 mmol m⁻³, CanESM 782 models show relatively little change; in CanESM5 the volume actually declines slightly, while in 783 784 CanESM5-CanOE it increases, but the total change is <1% in each case. As with the baseline 785 volumes, the range among models is large, with one model showing an increase approaching 10% of the total volume estimated for WOA20132018 (Figures 5b and 22b). For the volume 786 with <6 mmol m⁻³ (Figure 22a), CanESM models are among the most stable over time. In 787

788	CanESM5, the volume again declines, although this is within the range of internal variability.				
789	Again some models show fairly large excursions, but in this case none shows a strong secular				
790	trend over the last half-century.				
791					
792	4. Discussion				
793					
794	CanESM5 and CanESM5-CanOE are new coupled ocean-atmosphere climate models with				
795	prognostic ocean biogeochemistry. The two have the same physical climate (in experiments with				
796	specified atmospheric CO ₂) and differ only in their ocean biogeochemistry components.				
797	CanESM5-CanOE has a much more complex biogeochemistry model including a prognostic iron				
798	cycle. We have presented results that assess how these two models simulate the overall				
799	distribution of major tracers like DIC, alkalinity, nitrate and oxygen, as well as analyses of the				
800	interaction of the iron and nitrogen cycles, plankton community structure, export of organic				
801	matter from the euphotic zone, and historical trends over 1850-2014.				
802					
803	The overall distribution of major tracers indicates that both models do a reasonable job of				
804	simulating both biogeochemical (e.g., export and remineralization of organic matter) and				
805	physical (e.g., deep and intermediate ocean ventilation) processes. The volume of ocean with				
806	oxygen concentration below 6 or 60 µM compares favourably with other CMIP6 models (Figure				
807	5), and is among the most stable over historical time (Figure 22). CanESM5-CanOE has a				
808	substantially lower volume of water with [O ₂]<6 µM than CanESM5 and much closer to				
809	observation-based estimates (Figure 22a). Both models are biased slightly low in terms of				
810	historical uptake of anthropogenic CO ₂ , which may indicate weak Southern Ocean upwelling or				

811	too shallow remineralization of DIC or both (Figure 20). The spatial distribution of		
812	anthropogenic DIC is very similar between the two models (Supplementary Figure S7), which is		
813	expected as it is mainly a function of the physical ocean model circulation. However, CanESM5		
814	has higher concentrations in the main areas of accumulation, particularly the North Atlantic and		
815	the Southern Ocean. This probably indicates more efficient removal and export of 'natural' DIC		
816	by the plankton, particularly in the Southern Ocean upwelling zone (Figure 19), and deeper		
817	average remineralization, with the caveat that the preindustrial control simulations had different		
818	degrees of equilibration when the historical experiment was launched (cf. Séférian et al., 2016,		
819	Supplementary Table S6).		
820			
821	Analysis of phytoplankton and zooplankton biomass concentrations show that CanESM5 and		
822	CanESM5-CanOE compare somewhat favourably with available observational data but do have		
823	distinct biases. In particular, both zooplankton biomass and detrital organic matter concentration		
824	tend to be very low in CanESM5-CanOE; the total biomass of the plankton community and the		
825	standing crop of particulate organic matter are dominated by phytoplankton (e.g., Figure 17).		
826	Regional biases differ between the two models, with CanESM5-CanOE showing excessively		
827	large phytoplankton biomass in the tropics. We note, however, that the seasonal cycle of		
828	equatorial upwelling and the formation of the equatorial Pacific HNLC are reproduced rather		
829	well by our models (e.g., Figures 11, 15 and S6), and that CanESM5-CanOE is the first CanESM		
830	model to have genuinely simulated this as an emergent property (see section 3.3). In CanESM5-		
831	CanOE, decoupling of large and small phytoplankton populations associated with seasonal		
832	upwelling or convection (see below) is observed in some regions but not others.		
833			

834	Global export production is biased low, particularly in CanESM5-CanOE. This is due in part to
835	the biogeochemical model and in part to ocean circulation. CanESM5 has the same ocean
836	biology as CanESM2 but a different physical ocean model, and global ocean export production is
837	substantially lower in CanESM5. It is lower still in CanESM5-CanOE (Figure 19). We note that
838	CanESM5 performs better than CanESM2 on most metrics of physical ocean model evaluation
839	(Swart et al., 2019a), and shows a more realistic distribution of major tracers like DIC (Figure 8).
840	While the range of observation-based estimates of global ocean export production is large, and
841	encompasses the full range of CMIP5 and CMIP6 models, the change between CanESM2 and
842	CanESM5 is large. Changes in the physical ocean are not entirely independent of the
843	biogeochemistry model even when the latter is ostensibly identical. In CanESM2 and CanESM5,
844	iron limitation is specified as a spatially static 'mask' based on the observed distribution of
845	surface nitrate, and it is possible that in these two models ocean upwelling occurs in different
846	places relative to the specified boundary of the region of Southern Ocean iron imitation (Figure 3
847	of Zahariev et al., 2008). It is also possible that the lower export production in CanESM5-CanOE
848	is due to low iron supply to the surface waters of the Southern Ocean, but comparison with
849	available observations do not suggest that this is the case. Several biases are common to
850	CanESM5 and CanESM5-CanOE that relate to Southern Ocean upwelling (high Southern Ocean
851	surface nitrate concentration, low export production, weak anthropogenic CO ₂ uptake) and so are
852	probably more attributable to the physical ocean model than to the Fe submodel. The difference
853	between CanESM2 and CanESM5 bears this out.
854	
855	The development of CanOE was undertaken in response to some of the most severe limitations

s56 of CanESM^{1/2}, and in light of our collective experience. In addition to CMOC (Zahariev et al.,

857	2008), previous models developed by members of our group include Denman and Peña (1999;
858	2002), Christian et al. (2002a; 2002b), Christian (2005), and Denman et al. (2006). Christian et
859	al. (2002a) had a prognostic Fe cycle and multiple phytoplankton and zooplankton species, but
860	had fixed elemental ratios. Christian (2005) incorporated a cellular-regulation model, but only
861	for a single species and without Fe limitation. Christian (2005) had prognostic chlorophyll
862	whereas Denman and Peña (1999; 2002) and Christian et al. (2002a) used an irradiance-
863	dependent diagnostic formulation. Christian et al. (2002a) used multiplicative (Franks et al.,
864	1986) grazing, which creates stability in predator-prey interactions but severely limits
865	phytoplankton biomass accumulation under nutrient-replete conditions.
866	
867	One of the most important lessons from Christian et al. (2002a; 2002b) was that when a fixed
868	Fe/N ratio is employed, sensitivity to this parameter is extreme. Because Fe cell quotas are far
869	more variable than N, P, or Si quotas, treating this parameter as constant results in the specified
870	value influencing the overall solution far more than any other parameter. CanESM5-CanOE
871	largely succeeded in creating a prognostic Fe-N limitation model that produces HNLC conditions
872	in the expected regions (Figures 10, 11, 14, 15, <u>S6</u>), although surface nitrate concentration is low
873	relative to observation-based estimates in some cases. External Fe sources and scavenging
874	parameterizations will be revisited and refined in future versions. In CanESM5-CanOE the
875	scavenging model is very simple, with distinct regimes for concentrations greater or less than 0.6
876	nM; scavenging rates are very high above this threshold which causes deep-water concentrations
877	to converge on this value. The generally nutrient-like profile suggest that in CanOE the
878	scavenging rate is quite low for concentrations below 0.6 nM (Figure 13; see also Supplementary
879	Figure S9h). We note that the aeolian mineral dust deposition field employed here is derived

from the CanESM atmosphere model; these processes are not presently interactive but could bemade so in the future.

882

A particular issue with CanESM2 was that extremely high concentrations of nitrate occurred 883 884 under the Eastern Boundary Current (EBC) upwelling regions. This error resulted from spreading denitrification out over the ocean basin so that introduction of new fixed N from N₂ 885 fixation would balance denitrification losses within each vertical column, whereas in the real 886 world denitrification is highly localized in the low oxygen environments under the EBCs. 887 888 CanESM2 did not include oxygen, but CanESM5 CMOC incorporates oxygen as a 'downstream' 889 tracer that does not feed back on other biogeochemical processes. The incorporation of a more process-based denitrification parameterization in CanESM5-CanOE is independent of the many 890 other processes that are present in CanESM5-CanOE but not in CanESM5CMOC: a CMOC-like 891 892 model with prognostic denitrification is clearly an option. We chose not to include explicit, oxygen-dependent denitrification in CanESM5 because we wanted to maintain a CMOC-based 893 model as close to the CanESM2 version as possible, and because oxygen would not then be a 894 downstream tracer that does not affect other processes. 895

896

897 CanOE for the most part successfully reproduces the overall distribution of major tracers such as 898 nitrate, oxygen, DIC and alkalinity (and dFe, to the extent that its distribution is known). One 899 could argue that the gains made relative to CMOC are incremental. However, it is also important 900 to note that CanOE explicitly simulates important processes that are highly parameterized or 901 specified in CMOC. For example, the maintenance of HNLC regions is hardwired into CMOC 902 by specifying iron limitation as a function of the present day observed distribution of surface

nitrate. Both models show substantial gains in skill relative to CanESM2. These gains are similar
 in the two models and in coupled or ocean only (with CanESM2 forcing) mode (not shown) and
 are, therefore, attributable primarily to improvements in the ocean circulation model, although
 differences in initialization and spinup may also play a small role (e.g., Séférian et al., 2016).

908 Plankton community structure in CanESM5-CanOE is somewhat biased toward high concentrations of phytoplankton, low concentrations of zooplankton and detritus, and low export 909 (Sect. 3.4). In the development phase, a fair number of experiments were conducted with various 910 values of the grazing rates and detritus sinking speeds. A wide range of values of these 911 912 parameters was tested, with no resulting improvement in the overall results. Possibly the detrital remineralization rates are too high, although primary production is also on the low end of the 913 CMIP6 range (not shown), and would probably decline further if these rates were decreased. The 914 915 model was designed around the Armstrong (1994) hypothesis of 'supplementation' vs 'replacement', i.e., small phytoplankton and their grazers do not become much more abundant in 916 more nutrient-rich environments but rather stay at about the same level and are joined by larger 917 species that are absent in more oligotrophic conditions (see also Chisholm, 1992; Landry et al., 918 1997; Friedrichs et al., 2007). The results presented here suggest that this was partially achieved 919 but further improvement is possible (Figure 17). 920

921

As to whether the gains in skill with <u>CanESM5-</u>CanOE justify the extra computational cost,
Taylor diagrams (Figures 4, 8, 9, and Supplementary Figure <u>S2S4</u>) show a modest but consistent
gain <u>in skill at simulating the major biogeochemical species (O2, DIC, alkalinity)</u> across
variables and depths, especially for alkalinity at mid-depths (Supplementary Figure <u>S2S4</u>), for

which CanESM5 displays the least skill relative to other fields or depths. Other processes that are 926 927 highly parameterized in CanESM5CMOC, such as calcification and CaCO₃ dissolution, were not 928 addressed in detail in this paper, but are an important factor in determining the subsurface 929 distribution of alkalinity. As noted above for maintenance of HNLC conditions, Again, we 930 emphasize that we are simulating as an emergent property something that is parameterized in CanESM5CMOC (as previously noted for surface nitrate concentration in HNLC regions), and 931 doing at least as well in terms of model skill. As a general rule, the potential for improving skill 932 and achieving better results in novel environments (e.g., topographically complex regional 933 934 domains like the Arctic Ocean and the boreal marginal seas), is expected to be greater in less parameterized models (e.g., Friedrichs et al., 2007; Tesdal et al., 2016). 935

936

937 An updated version of CanESM5 CMOC with prognostic denitrification is clearly possible. 938 However, for the reasons discussed above, a prognostic Fe cycle with a fixed phytoplankton Fe/N remains problematic, and the model would still have a single detritus sinking speed and 939 remineralization length scale. We are also developing CanOE for regional downscaling 940 applications (Hayashida, 2018; Holdsworth et al., 2021), and it is likely that the simplification of 941 942 having a single particle sinking speed is not well suited to a domain with complex topography 943 and prominent continental shelf and slope. The number of tracers in CanOE is not particularly large compared with other CMIP6 models. We expect to further refine CanOE and its 944 parameterizations, evaluate it against new and emerging ocean data sets (e.g., GEOTRACES, 945 946 biogeochemical Argo), and incrementally improve CMOC (which we will maintain for a wide 947 suite of physical-climate experiments for which ocean biogeochemistry is not central to the 948 purpose). For CMIP6, we chose to keep CMOC as close to the CanESM2 version as possible.

949	This strategy allows	us to quantify how n	nuch of the improv	ement in model s	kill is due to the
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950 physical circulation, which is in fact substantial as is illustrated by greater skill with respect to

951 DIC (e.g., Figure 8) and alkalinity (Supplementary Figure S4), particularly at intermediate depths

- 952 (400-900 m). The CanESM terrestrial carbon model is also undergoing important new
- 953 developments (e.g., Asaadi and Arora, 2021) and we expect CanESM to continue to offer a
- 954 credible contribution to global carbon cycle studies, as well as advancing regional downscaling
- 955 and impacts science.

956

- 957 *Code availability*. The full CanESM5 source code is publicly available at
- gitlab.com/cccma/canesm; within this tree the <u>CMOC/CanOE ocean biogeochemistry</u> code can
- 959 be found at gitlab.com/cccma/cannemo/-
- 960 /tree/v5.0.3/nemo/CONFIG/CCC_CANCPL_ORCA1_LIM_CMOC or
- 961 CCC CANCPL ORCA1 LIM CANOE (last access: 21 September 2021). The version of the
- 962 code which can be used to produce all the simulations submitted to CMIP6, and described in this
- paper, is tagged as v5.0.3 and has the associated DOI: https://doi.org/10.5281/zenodo.3251113
- 964 (Swart et al., 2019b).

965

Data availability. All CanESM5-simulations conducted for CMIP6, including those described in
 this paper, are publicly available via the Earth System Grid Federation-(ESGF) (source_id =
 <u>CanESM5 or CanESM5-CanOE</u>). All observational data and other CMIP6 model data used are
 publicly available.

971	Author contributions. Formulation of the overall research goals and aims: JRC, KLD, NS, NCS;				
972	Implementation and testing of the model code: JRC, HH, AMH, WGL, OGJR, AES, NCS;				
973	Carrying out the experiments: JRC, WGL, OGJR, AES, NCS; Creation of the published work:				
974	JRC, HH, AMH, AES, NS, NCS.				
975					
976	Competing interests. The authors declare that they have no conflict of interest.				
977					
978	Disclaimer. CanESM has been customized to run on the ECCC high-performance computer, and				
979	a significant fraction of the software infrastructure used to run the model is specific to the				
980	individual machines and architecture. While we publicly provide the code, we cannot provide				
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991 992 993 994 995	Literature Cited				

996 Arguez, A. and Vose, R.: The Definition of the Standard WMO Climate Normal The Key to 997 Deriving Alternative Climate Normals, Bulletin of the American Meteorological Society, 92, 699-704, 10.1175/2010BAMS2955.1, 2011. 998 999 1000 Armstrong, R.: Grazing limitation and nutrient limitation in marine ecosystems - steady-state 1001 solutions of an ecosystem model with multiple food-chains, Limnology and Oceanography, 39, 1002 597-608, 1994. 1003 Arora, V., Scinocca, J., Boer, G., Christian, J., Denman, K., Flato, G., Kharin, V., Lee, W., and 1004 1005 Merryfield, W.: Carbon emission limits required to satisfy future representative concentration pathways of greenhouse gases, Geophysical Research Letters, 38, 10.1029/2010GL046270, 1006 2011. 1007 1008 Arora, V., Boer, G., Christian, J., Curry, C., Denman, K., Zahariev, K., Flato, G., Scinocca, J., 1009 1010 Merryfield, W., and Lee, W.: The Effect of Terrestrial Photosynthesis Down Regulation on the 1011 Twentieth-Century Carbon Budget Simulated with the CCCma Earth System Model, Journal of Climate, 22, 6066-6088, 10.1175/2009JCLI3037.1, 2009. 1012 1013 1014 Arora, V., Katavouta, A., Williams, R., Jones, C., Brovkin, V., Friedlingstein, P., Schwinger, J., 1015 Bopp, L., Boucher, O., Cadule, P., Chamberlain, M., Christian, J., Delire, C., Fisher, R., Hajima, T., Ilyina, T., Joetzjer, E., Kawamiya, M., Koven, C., Krasting, J., Law, R., Lawrence, D., 1016 1017 Lenton, A., Lindsay, K., Pongratz, J., Raddatz, T., Seferian, R., Tachiiri, K., Tjiputra, J., Wiltshire, A., Wu, T., and Ziehn, T.: Carbon-concentration and carbon-climate feedbacks in 1018 CMIP6 models and their comparison to CMIP5 models, Biogeosciences, 17, 4173-4222, 1019 10.5194/bg-17-4173-2020, 2020. 1020 1021 Asaadi, A. and Arora, V.: Implementation of nitrogen cycle in the CLASSIC land model, 1022 1023 Biogeosciences, 18, 669-706, 10.5194/bg-18-669-2021, 2021. 1024 Aumont, O., Ethe, C., Tagliabue, A., Bopp, L., and Gehlen, M.: PISCES-v2: an ocean 1025 biogeochemical model for carbon and ecosystem studies, Geoscientific Model Development, 8, 1026 1027 2465-2513, 10.5194/gmd-8-2465-2015, 2015. 1028 Babbin, A., Keil, R., Devol, A., and Ward, B.: Organic Matter Stoichiometry, Flux, and Oxygen 1029 1030 Control Nitrogen Loss in the Ocean, Science, 344, 406-408, 10.1126/science.1248364, 2014. 1031 Chisholm, S.W.: Phytoplankton size, in: Primary productivity and biogeochemical cycles in the 1032 1033 sea, edited by: Falkowski, P.G., and Woodhead A.D., Plenum, New York, 213-237, 1992. 1034 1035 Christian, J.: Biogeochemical cycling in the oligotrophic ocean: Redfield and non-Redfield models, Limnology and Oceanography, 50, 646-657, 2005. 1036 1037 Christian, J. and Karl, D.: Microbial community structure at the United States Joint Global 1038 Ocean Flux Study Station ALOHA - inverse methods for estimating biochemical indicator ratios. 1039 1040 Journal of Geophysical Research-Oceans, 99, 14269-14276, 10.1029/94JC00681, 1994. 1041

1042 Christian, J., Arora, V., Boer, G., Curry, C., Zahariev, K., Denman, K., Flato, G., Lee, W., 1043 Merryfield, W., Roulet, N., and Scinocca, J.: The global carbon cycle in the Canadian Earth system model (CanESM1): Preindustrial control simulation, Journal of Geophysical Research-1044 1045 Biogeosciences, 115, 10.1029/2008JG000920, 2010. 1046 1047 Christian, J. R., Verschell, M. A., Murtugudde, R., Busalacchi, A. J., and McClain, C. R.: 1048 Biogeochemical modelling of the tropical Pacific Ocean. I: Seasonal and interannual variability, 1049 Deep-Sea Research Part II-Topical Studies in Oceanography, 49, 509-543, 2002a. 1050 1051 Christian, J. R., Verschell, M. A., Murtugudde, R., Busalacchi, A. J., and McClain, C. R.: Biogeochemical modelling of the tropical Pacific Ocean. II: Iron biogeochemistry, Deep-Sea 1052 Research Part II-Topical Studies in Oceanography, 49, 545-565, 0.1016/s0967-0645(01)00111-4, 1053 1054 2002b. 1055 1056 Denman, K. and Pena, M.: A coupled 1-D biological/physical model of the northeast subarctic 1057 Pacific Ocean with iron limitation, Deep-Sea Research Part II-Topical Studies in Oceanography, 46, 2877-2908, 10.1016/S0967-0645(99)00087-9, 1999. 1058 1059 1060 Denman, K. and Pena, M.: The response of two coupled one-dimensional mixed layer/planktonic ecosystem models to climate change in the NE subarctic Pacific Ocean, Deep-Sea Research Part 1061 II-Topical Studies in Oceanography, 49, 5739-5757, 10.1016/S0967-0645(02)00212-6, 2002. 1062 1063 Denman, K., Voelker, C., Pena, M., and Rivkin, R.: Modelling the ecosystem response to iron 1064 fertilization in the subarctic NE Pacific: The influence of grazing, and Si and N cycling on CO₂ 1065 1066 drawdown, Deep-Sea Research Part II-Topical Studies in Oceanography, 53, 2327-2352, 10.1016/j.dsr2.2006.05.026, 2006. 1067 1068 1069 Devol, A.H.: Denitrification including anammox, in: Nitrogen in the Marine Environment, 2nd 1070 edition, edited by: Capone, D.G., Bronk D.A., Mulholland M.R., and Carpenter E.J., Elsevier, Amsterdam, 263-301, 2008. 1071 1072 1073 Dickson, A.G., Sabine, C.L., and Christian, J.R., eds.: Guide to best practices for ocean CO₂ measurements, PICES Special Publication #3, 2007 1074 1075 1076 Elser, J. and Urabe, J.: The stoichiometry of consumer-driven nutrient recycling: Theory, observations, and consequences, Ecology, 80, 735-751, 10.1890/0012-1077 9658(1999)080[0735:TSOCDN]2.0.CO;2, 1999. 1078 1079 1080 Franks, P., Wroblewski, J., and Flierl, G.: Behavior of a simple plankton model with food-level acclimation by herbivores, Marine Biology, 91, 121-129, 10.1007/BF00397577, 1986. 1081 1082 1083 Friedlingstein, P., O'Sullivan, M., Jones, M., Andrew, R., Hauck, J., Olsen, A., Peters, G., Peters, W., Pongratz, J., Sitch, S., Le Quere, C., Canadell, J., Ciais, P., Jackson, R., Alin, S., Aragao, L., 1084 Arneth, A., Arora, V., Bates, N., Becker, M., Benoit-Cattin, A., Bittig, H., Bopp, L., Bultan, S., 1085 Chandra, N., Chevallier, F., Chini, L., Evans, W., Florentie, L., Forster, P., Gasser, T., Gehlen, 1086 M., Gilfillan, D., Gkritzalis, T., Gregor, L., Gruber, N., Harris, I., Hartung, K., Haverd, V., 1087

1088	Houghton, R., Ilyina, T., Jain, A., Joetzjer, E., Kadono, K., Kato, E., Kitidis, V., Korsbakken, J.,					
1089	Landschutzer, P., Lefevre, N., Lenton, A., Lienert, S., Liu, Z., Lombardozzi, D., Marland, G.,					
1090	Metzl, N., Munro, D., Nabel, J., Nakaoka, S., Niwa, Y., O'Brien, K., Ono, T., Palmer, P., Pierrot,					
1091	D., Poulter, B., Resplandy, L., Robertson, E., Rodenbeck, C., Schwinger, J., Seferian, R.,					
1092	Skjelvan, I., Smith, A., Sutton, A., Tanhua, T., Tans, P., Tian, H., Tilbrook, B., Van der Werf,					
1093	G., Vuichard, N., Walker, A., Wanninkhof, R., Watson, A., Willis, D., Wiltshire, A., Yuan, W.,					
1094	Vue X and Zaehle S · Global Carbon Budget 2020 Farth System Science Data 12 2260_					
1095	3340 10 5194/essd-12-3269-2020 2020					
1096	55 10, 10.517 # 0564 12 5207 2020, 2020.					
1097	Friedrichs M Dusenberry I Anderson I Armstrong R Chai F Christian I Doney S					
1098	Dunne I Fuji M Hood R McGillicuddy D Moore I Schartau M Snitz V and					
1099	Wiggert, J.: Assessment of skill and portability in regional marine biogeochemical models: Role					
1100	of multiple planktonic groups, Journal of Geophysical Research-Oceans, 112,					
1101	10.1029/2006JC003852, 2007.					
1102						
1103	Garcia, H.E., Locarnini, R.A., Boyer, T.P., Antonov, J.I., Baranova, O.K., Zweng, M.M.,					
1104	Reagan, J.R., and Johnson, D.R.: World Ocean Atlas 2013, Volume 3: Dissolved Oxygen,					
1105	Apparent Oxygen Utilization, and Oxygen Saturation. S. Levitus, Ed., A. Mishonov Technical					
1106	Ed., NOAA Atlas NESDIS 75, 27 pp., 2014.					
1107						
1108	Garcia, H.E., Weathers, K., Paver, C.R., Smolyar, I., Boyer, T.P., Locarnini, R.A., Zweng,					
1109	M.M., Mishonov, A.V., Baranova, O.K., Seidov, D. and Reagan, J.R.: World Ocean Atlas 2018,					
1110	Volume 3: Dissolved Oxygen, Apparent Oxygen Utilization, and Oxygen Saturation. A.					
1111	Mishonov Technical Ed., NOAA Atlas NESDIS 83, 38pp., 2018a.					
1112						
1113	Garcia, H.E., Locarnini, R.A., Boyer, T.P., Antonov, J.I., Baranova, O.K., Zweng, M.M.,					
1114	Reagan, J.R., and Johnson, D.R.: World Ocean Atlas 2013, Volume 4: Dissolved Inorganic					
1115	Nutrients (phosphate, nitrate, silicate). S. Levitus, Ed., A. Mishonov Technical Ed.; NOAA Atlas					
1116	NESDIS 76, 25 pp., 2014.					
1117						
1118	Garcia, H.E., Weathers, K., Paver, C.R., Smolyar, I., Boyer, T.P., Locarnini, R.A., Zweng,					
1119	M.M., Mishonov, A.V., Baranova, O.K., Seidov, D. and Reagan, J.R.: World Ocean Atlas 2018,					
1120	Volume 4: Dissolved Inorganic Nutrients (phosphate, nitrate and nitrate+nitrite, silicate). A.					
1121	Mishonov Technical Ed., NOAA Atlas NESDIS 84, 35pp., 2018b.					
1122						
1123	Geider, R., MacIntyre, H., and Kana, T.: A dynamic regulatory model of phytoplanktonic					
1124	acclimation to light, nutrients, and temperature. Limnology and Oceanography, 43, 679-694.					
1125	1998.					
1126						
1127	Guilvardi E and Madec G · Performance of the OPA/ARPEGE-T21 global ocean-atmosphere					
1128	counled model Climate Dynamics 13 149-165 10 1007/s003820050157 1997					
1120	coupled model, emilate Dynamics, 15, 119 105, 10.1007/5005020050157, 1997.					
1120	Havashida, H: Modelling sea-ice and oceanic dimethylsulde production and emissions in the					
1121	Arctic PhD thesis University of Victoria 2018					
1127						
1122	Holdsworth A.M. Zhai I. I.u. V. and Christian I.P. Euture changes in accomparantly and					
1133	Tronosworun, A.m., Zhan, E., Eu, T., and Christian, J.K. Puture changes in oceanography and					

biogeochemistry along the Canadian Pacific continental margin, Frontiers in Marine Science, 1134 1135 10.3389/fmars.2021.602991, 2021. 1136 1137 Jetten, M., Wagner, M., Fuerst, J., van Loosdrecht, M., Kuenen, G., and Strous, M.: Microbiology and application of the anaerobic ammonium oxidation ('anammox') process, 1138 Current Opinion in Biotechnology, 12, 283-288, 10.1016/S0958-1669(00)00211-1, 2001. 1139 1140 Johnson, K. S., Gordon, R. M., and Coale, K. H.: What controls dissolved iron concentrations in 1141 the world ocean?, Marine Chemistry, 57, 137-161, 10.1016/s0304-4203(97)00043-1, 1997. 1142 1143 Johnson, K., Elrod, V., Fitzwater, S., Plant, J., Chavez, F., Tanner, S., Gordon, R., Westphal, D., 1144 Perry, K., Wu, J., and Karl, D.: Surface ocean-lower atmosphere interactions in the Northeast 1145 Pacific Ocean Gyre: Aerosols, iron, and the ecosystem response, Global Biogeochemical Cycles, 1146 17, 10.1029/2002GB002004, 2003. 1147 1148 1149 Key, R.M., Olsen, A., van Heuven, S., Lauvset, S. K., Velo, A., Lin, X., Schirnick, C., Kozyr, 1150 A., Tanhua, T., Hoppema, M., Jutterström, S., Steinfeldt, R., Jeansson, E., Ishii, M., Perez, F. F., 1151 and Suzuki, T.: 2015. Global Ocean Data Analysis Project, Version 2 (GLODAPv2), 1152 ORNL/CDIAC-162, NDP-093. Carbon Dioxide Information Analysis Center, Oak Ridge 1153 National Laboratory, US Department of Energy, Oak Ridge, Tennessee. doi:10.3334/CDIAC/OTG.NDP093 GLODAPv2 1154 1155 1156 Lambert, S. and Boer, G.: CMIP1 evaluation and intercomparison of coupled climate models, Climate Dynamics, 17, 83-106, 10.1007/PL00013736, 2001. 1157 1158 1159 Landry, M., Barber, R., Bidigare, R., Chai, F., Coale, K., Dam, H., Lewis, M., Lindley, S., McCarthy, J., Roman, M., Stoecker, D., Verity, P., and White, J.: Iron and grazing constraints on 1160 primary production in the central equatorial Pacific: An EqPac synthesis, Limnology and 1161 Oceanography, 42, 405-418, 10.4319/lo.1997.42.3.0405, 1997. 1162 1163 Lauvset, S., Key, R., Olsen, A., van Heuven, S., Velo, A., Lin, X., Schirnick, C., Kozyr, A., 1164 Tanhua, T., Hoppema, M., Jutterstrom, S., Steinfeldt, R., Jeansson, E., Ishii, M., Perez, F., 1165 Suzuki, T., and Watelet, S.: A new global interior ocean mapped climatology: the 1 degrees x 1 1166 degrees GLODAP version 2, Earth System Science Data, 8, 325-340, 10.5194/essd-8-325-2016, 1167 2016. 1168 1169 Li, Y.: Distribution patterns of the elements in the ocean - a synthesis, Geochimica et 1170 1171 Cosmochimica Acta, 55, 3223-3240, 1991. 1172 1173 Locarnini, R.A., Mishonov, A.V., Antonov, J.I., Boyer, T.P., Garcia, H.E., Baranova, O.K., 1174 Zweng, M.M., Paver, C.R., Reagan, J.R., Johnson, D.R., Hamilton, M., and Seidov, D.: World Ocean Atlas 2013, Volume 1: Temperature. S. Levitus, Ed., A. Mishonov Technical Ed., NOAA 1175 1176 Atlas NESDIS 73, 40 pp., 2013. 1177 1178 Locarnini, R.A., Mishonov, A.V., Antonov, J.I., Boyer, T.P., Garcia, H.E., Baranova, O.K., Zweng, M.M., and Johnson, D.R.: World Ocean Atlas 2009, Volume 1: Temperature. Levitus, 1179

- 1180 <u>S., Ed., NOAA Atlas NESDIS 68, 184 pp., 2010.</u>
- 1181
- Locarnini, R.A., Mishonov, A.V., Baranova, O.K., Boyer, T.P., Zweng, M.M., Garcia, H.E.,
- 1183 Reagan, J.R., Seidov, D., Weathers, K., Paver, C.R., and Smolyar, I.: World Ocean Atlas 2018,
- 1184 <u>Volume 1: Temperature. A. Mishonov Technical Ed., NOAA Atlas NESDIS 81, 52pp., 2018.</u>
- 11851186 Loladze, I., Kuang, Y., and Elser, J.: Stoichiometry in producer-grazer systems: Linking energy
- 1187 flow with element cycling, Bulletin of Mathematical Biology, 62, 1137-1162,
- 1188 10.1006/bulm.2000.0201, 2000. 1189
- Lukas, R.: Pacific ocean equatorial currents, in: Encyclopedia of Ocean Sciences, edited by:
 Steele, J.H., Thorpe, S.A., and Turekian, K.K., Academic, San Diego, 2069-2076,
 10.1006/rwos.2001.0354, 2001.
- 1193
- Madec, G. and Imbard, M.: A global ocean mesh to overcome the North Pole singularity,
 Climate Dynamics, 12, 381-388, 10.1007/s003820050115, 1996.
- 1196

- Moriarty, R. and O'Brien, T.: Distribution of mesozooplankton biomass in the global ocean,
 Earth System Science Data, 5, 45-55, 10.5194/essd-5-45-2013, 2013.
- Nozaki, Y.: Elemental Distribution, in: Encyclopedia of Ocean Sciences, edited by: Steele, J.H.,
 Thorpe, S.A., and Turekian, K.K., Academic, San Diego, 840-845, 10.1006/rwos.2001.0402,
 2001.
- 1203
- Orr, J. C., Najjar, R. G., Aumont, O., Bopp, L., Bullister, J. L., Danabasoglu, G., Doney, S. C.,
 Dunne, J. P., Dutay, J. C., Graven, H., Griffies, S. M., John, J. G., Joos, F., Levin, I., Lindsay,
 K., Matear, R. J., McKinley, G. A., Mouchet, A., Oschlies, A., Romanou, A., Schlitzer, R.,
- 1207 Tagliabue, A., Tanhua, T., and Yool, A.: Biogeochemical protocols and diagnostics for the
- CMIP6 Ocean Model Intercomparison Project (OMIP), Geoscientific Model Development, 10,
 2169-2199, 10.5194/gmd-10-2169-2017, 2017.
- 1210
- Philander, S.G. and Chao, Y.: On the contrast between the seasonal cycles of the equatorialAtlantic and Pacific oceans, Journal of Physical Oceanography, 21, 1399-1406, 1991.
- 1213
 1214 Raven, J. and Geider, R.: Temperature and algal growth, New Phytologist, 110, 441-461,
 10.1111/j.1469-8137.1988.tb00282.x, 1988.
- 1216
- Riche, O. and Christian, J.: Ocean dinitrogen fixation and its potential effects on ocean primary
 production in Earth system model simulations of anthropogenic warming, Elementa-Science of
 the Anthropocene, 6, 10.1525/elementa.277, 2018.
- 1220
- 1221 Schlitzer, R., Anderson, R., Dodas, E., Lohan, M., Geibere, W., Tagliabue, A., Bowie, A.,
- 1222 Jeandel, C., Maldonado, M., Landing, W., Cockwell, D., Abadie, C., Abouchami, W.,
- 1223 Achterberg, E., Agather, A., Aguliar-Islas, A., van Aken, H., Andersen, M., Archer, C., Auro,
- 1224 M., de Baar, H., Baars, O., Baker, A., Bakker, K., Basak, C., Baskaran, M., Bates, N., Bauch, D.,
- 1225 van Beek, P., Behrens, M., Black, E., Bluhm, K., Bopp, L., Bouman, H., Bowman, K., Bown, J.,

Boyd, P., Boye, M., Boyle, E., Branellec, P., Bridgestock, L., Brissebrat, G., Browning, T., 1226 1227 Bruland, K., Brumsack, H., Brzezinski, M., Buck, C., Buck, K., Buesseler, K., Bull, A., Butler, E., Cai, P., Mor, P., Cardinal, D., Carlson, C., Carrasco, G., Casacuberta, N., Casciotti, K., 1228 1229 Castrillejo, M., Chamizo, E., Chance, R., Charette, M., Chaves, J., Cheng, H., Chever, F., Christl, M., Church, T., Closset, I., Colman, A., Conway, T., Cossa, D., Croot, P., Cullen, J., 1230 Cutter, G., Daniels, C., Dehairs, F., Deng, F., Dieu, H., Duggan, B., Dulaquais, G., 1231 Dumousseaud, C., Echegoyen-Sanz, Y., Edwards, R., Ellwood, M., Fahrbach, E., Fitzsimmons, 1232 J., Flegal, A., Fleisher, M., van de Flierdt, T., Frank, M., Friedrich, J., Fripiat, F., Frollje, H., 1233 Galer, S., Gamo, T., Ganeshram, R., Garcia-Orellana, J., Garcia-Solsona, E., Gault-Ringold, M., 1234 George, E., Gerringa, L., Gilbert, M., Godoy, J., Goldstein, S., Gonzalez, S., Grissom, K., 1235 Hammerschmidt, C., Hartman, A., Hassler, C., Hathorne, E., Hatta, M., Hawco, N., Hayes, C., 1236 Heimburger, L., Helgoe, J., Heller, M., Henderson, G., Henderson, P., van Heuven, S., Ho, P., 1237 Horner, T., Hsieh, Y., Huang, K., Humphreys, M., Isshiki, K., Jacquot, J., Janssen, D., Jenkins, 1238 W., John, S., Jones, E., Jones, J., Kadko, D., Kayser, R., Kenna, T., Khondoker, R., Kim, T., 1239 Kipp, L., Klar, J., Klunder, M., Kretschmer, S., Kumamoto, Y., Laan, P., Labatut, M., Lacan, F., 1240 Lam, P., Lambelet, M., Lamborg, C., Le Moigne, F., Le Roy, E., Lechtenfeld, O., Lee, J., 1241 1242 Lherminier, P., Little, S., Lopez-Lora, M., Lu, Y., Masque, P., Mawji, E., Mcclain, C., Measures, C., Mehic, S., Menzel Barraqueta, J., van der Merwe, P., Middag, R., Mieruch, S., Milne, A., 1243 Minami, T., Moffett, J., Moncoiffe, G., Moore, W., Morris, P., Morton, P., Nakaguchi, Y., 1244 1245 Nakayama, N., Niedermiller, J., Nishioka, J., Nishiuchi, A., Noble, A., Obata, H., Ober, S., Ohnemus, D., van Ooijen, J., O'Sullivan, J., Owens, S., Pahnke, K., Paul, M., Pavia, F., Pena, L., 1246 Petersh, B., Planchon, F., Planquette, H., Pradoux, C., Puigcorbe, V., Quay, P., Queroue, F., 1247 Radic, A., Rauschenberg, S., Rehkamper, M., Rember, R., Remenvi, T., Resing, J., Rickli, J., 1248 Rigaud, S., Rijkenberg, M., Rintoul, S., Robinson, L., Roca-Marti, M., Rodellas, V., Roeske, T., 1249 Rolison, J., Rosenberg, M., Roshan, S., van der Loaff, M., Ryabenko, E., Saito, M., Salt, L., 1250 1251 Sanial, V., Sarthou, G., Schallenberg, C., Schauer, U., Scher, H., Schlosser, C., Schnetger, B., Scott, P., Sedwick, P., Semiletov, I., Shelley, R., Sherrell, R., Shiller, A., Sigman, D., Singh, S., 1252 Slagter, H., Slater, E., Smethie, W., Snaith, H., Sohrin, Y., Sohst, B., Sonke, J., Speich, S., 1253 Steinfeldt, R., Stewart, G., Stichel, T., Stirling, C., Stutsman, J., Swarr, G., Swift, J., Thomas, A., 1254 Thorne, K., Till, C., Till, R., Townsend, A., Townsend, E., Tuerena, R., Twining, B., Vance, D., 1255 Velazquez, S., Venchiarutti, C., Villa-Alfageme, M., Vivancos, S., Voelker, A., Wake, B., 1256 Warner, M., Watson, R., van Weerlee, E., Weigand, M., Weinstein, Y., Weiss, D., Wisotzki, A., 1257 Woodward, E., Wu, J., Wu, Y., Wuttig, K., Wyatt, N., Xiang, Y., Xie, R., Xue, Z., Yoshikawa, 1258 H., Zhang, J., Zhang, P., Zhao, Y., Zheng, L., Zheng, X., Zieringer, M., Zimmer, L., Ziveri, P., 1259 Zunino, P., and Zurbrick, C.: The GEOTRACES Intermediate Data Product 2017, Chemical 1260 Geology, 493, 210-223, 10.1016/j.chemgeo.2018.05.040, 2018. 1261 1262 Séférian, R., Gehlen, M., Bopp, L., Resplandy, L., Orr, J., Marti, O., Dunne, J., Christian, J., 1263 1264 Doney, S., Ilyina, T., Lindsay, K., Halloran, P., Heinze, C., Segschneider, J., Tjiputra, J., Aumont, O., and Romanou, A.: Inconsistent strategies to spin up models in CMIP5: implications 1265

- for ocean biogeochemical model performance assessment, Geoscientific Model Development, 9,
 1267 1827-1851, 10.5194/gmd-9-1827-2016, 2016.
- 1268
- 1269 Siegel, D., Buesseler, K., Behrenfeld, M., Benitez-Nelson, C., Boss, E., Brzezinski, M., Burd, A.,
- 1270 Carlson, C., D'Asaro, E., Doney, S., Perry, M., Stanley, R., and Steinberg, D.: Prediction of the
- 1271 Export and Fate of Global Ocean Net Primary Production: The EXPORTS Science Plan,

- 1272 Frontiers in Marine Science, 3, 10.3389/fmars.2016.00022, 2016.
- 1273

Steinacher, M., Joos, F., Frolicher, T., Bopp, L., Cadule, P., Cocco, V., Doney, S., Gehlen, M.,
Lindsay, K., Moore, J., Schneider, B., and Segschneider, J.: Projected 21st century decrease in
marine productivity: a multi-model analysis, Biogeosciences, 7, 979-1005, 10.5194/bg-7-9792010, 2010.

1277 1278

Stramski, D., Reynolds, R., Babin, M., Kaczmarek, S., Lewis, M., Rottgers, R., Sciandra, A.,
Stramska, M., Twardowski, M., Franz, B., and Claustre, H.: Relationships between the surface
concentration of particulate organic carbon and optical properties in the eastern South Pacific
and eastern Atlantic Oceans, Biogeosciences, 5, 171-201, 10.5194/bg-5-171-2008, 2008.

1282 1283

Strous, M., Heijnen, J., Kuenen, J., and Jetten, M.: The sequencing batch reactor as a powerful
tool for the study of slowly growing anaerobic ammonium-oxidizing microorganisms, Applied
Microbiology and Biotechnology, 50, 589-596, 10.1007/s002530051340, 1998.

1287

1288 Swart, N., Cole, J., Kharin, V., Lazare, M., Scinocca, J., Gillett, N., Anstey, J., Arora, V.,

1289 Christian, J., Hanna, S., Jiao, Y., Lee, W., Majaess, F., Saenko, O., Seiler, C., Seinen, C., Shao,
1290 A., Sigmond, M., Solheim, L., von Salzen, K., Yang, D., and Winter, B.: The Canadian Earth

System Model version 5 (CanESM5.0.3), Geoscientific Model Development, 12, 4823-4873,
10.5194/gmd-12-4823-2019, 2019a.

1293

Swart, N. C., Cole, J., Kharin, S., Lazare, M., Scinocca, J., Gillett, N., Anstey, J., Arora, V.,
Christian, J., Hanna, S., Jiao, Y., Lee, W., Majaess, F., Saenko, O., Seiler, C., Seinen, C.,

Shao, A., Solheim, L., von Salzen, K., Yang, D., and Winter, B.: The Canadian Earth System
Model (CanESM), v5.0.3., https://doi.org/10.5281/zenodo.3251114, 2019b.

1298

Takeda, S., Chai, F., and Nishioka, J., Eds.: Report of Working Group 22 on Iron Supply and its
Impact on Biogeochemistry and Ecosystems in the North Pacific Ocean. PICES Sci. Rep. No.
42, 60 pp., 2013.

1302

1305

Taylor, K.: Summarizing multiple aspects of model performance in a single diagram., Journal of
Geophysical Research-Atmospheres, 106, 7183-7192, 10.1029/2000JD900719, 2001.

Tesdal, J., Christian, J., Monahan, A., and von Salzen, K.: Evaluation of diverse approaches for
estimating sea-surface DMS concentration and air-sea exchange at global scale, Environmental
Chemistry, 13, 390-412, 10.1071/EN14255, 2016.

1309

1310 von Salzen, K., Scinocca, J. F., McFarlane, N. A., Li, J. N., Cole, J. N. S., Plummer, D.,

1311 Verseghy, D., Reader, M. C., Ma, X. Y., Lazare, M., and Solheim, L.: The Canadian Fourth

1312 Generation Atmospheric Global Climate Model (CanAM4). Part I: Representation of Physical

- 1313 Processes, Atmosphere-Ocean, 51, 104-125, 10.1080/07055900.2012.755610, 2013.
- 1314

1315 White, J., Zhang, X., Welling, L., Roman, M., And Dam, H.: Latitudinal gradients in

1316 zooplankton biomass in the tropical Pacific At 140°W During The JGOFS Eqpac study – effects

1317 of El Nino, Deep-Sea Research Part II-Topical Studies in Oceanography, 42, 715-733,

10.1016/0967-0645(95)00033-M, 1995.

1319

1320 Wolf-Gladrow, D., Zeebe, R., Klaas, C., Kortzinger, A., and Dickson, A.: Total alkalinity: The

- explicit conservative expression and its application to biogeochemical processes, Marine
 Chemistry, 106, 287-300, 10.1016/j.marchem.2007.01.006, 2007.
- 1323

Zahariev, K., Christian, J., and Denman, K.: Preindustrial, historical, and fertilization simulations
using a global ocean carbon model with new parameterizations of iron limitation, calcification,
and N₂ fixation, Progress in Oceanography, 77, 56-82, 10.1016/j.pocean.2008.01.007, 2008.

1320

1328 Zweng, M.M., Reagan, J.R., Antonov, J.I., Locarnini, R.A., Mishonov, A.V., Boyer, T.P.,

- 1329 Garcia, H.E., Baranova, O.K., Johnson, D.R., Seidov, D., and Biddle, M.M.: World Ocean Atlas
- 1330 2013, Volume 2: Salinity. S. Levitus, Ed.; A. Mishonov, Technical Ed., NOAA Atlas NESDIS
- 1331 74, 39 pp., 2013. 1332
- 1333 Zweng, M.M., Reagan, J.R., Seidov, D., Boyer, T.P., Locarnini, R.A., Garcia, H. E., Mishonov,
- 1334 A.V., Baranova, O.K., Weathers, K., Paver, C.R., and Smolyar, I.: World Ocean Atlas 2018,
- 1335 Volume 2: Salinity. A. Mishonov Technical Ed., NOAA Atlas NESDIS 82, 50pp., 2018.
- 1336

Table 1 – Ecosystem model parameters.

Symbol	Description	Unit	
T _{ref}	Reference temperature	K	298.15
Eap	Activation energy for photosynthesis	kJ mol ⁻¹	37.4
Q ^N mins	Small phytoplankton minimum N quota	g N g C ⁻¹	0.04
Q ^N _{maxs}	Small phytoplankton maximum N quota	g N g C ⁻¹	0.172
Q ^N _{minl}	Large phytoplankton minimum N quota	g N g C ⁻¹	0.04
Q ^N _{maxl}	Large phytoplankton maximum N quota	g N g C ⁻¹	0.172
Q ^{Fe} mins	Small phytoplankton minimum Fe quota	μg Fe g C ⁻¹	4.65
Q ^{Fe} maxs	Small phytoplankton maximum Fe quota	μg Fe g C ⁻¹	93.
Q^{Fe}_{minl}	Large phytoplankton minimum Fe quota	μg Fe g C ⁻¹	6.5
Q ^{Fe} maxl	Large phytoplankton maximum Fe quota	μg Fe g C ⁻¹	70.
V ^N _{ref}	Reference rate of N uptake	g N g C ⁻¹ d ⁻¹	0.6
V ^{Fe} _{ref}	Reference rate of Fe uptake	μg Fe g C ⁻¹ d ⁻¹	79.
P ^C _{ref}	Reference rate of photosynthesis	$g C g C^{-1} d^{-1}$	3
k _{XU}	Rate coefficient for exhudation	d ⁻¹	1.7
k _{dgr}	Rate coefficient for chlorophyll degradation	d ⁻¹	0.02
ζ	Respiratory cost of biosynthesis	g C g N ⁻¹	2
α _{chl}	Initial slope of P-E curve	g C g CHL ⁻¹ h ⁻¹ (μ mol m ⁻² s ⁻¹) ⁻¹	1.08
$\Theta_{max}{}^{N}$	Maximum chlorophyll-nitrogen ratio	g g ⁻¹	0.18
K _{NiS}	Half-saturation for small phytoplankton nitrate uptake	mmol ⁻¹ m ³	0.1
K _{NaS}	Half-saturation for small phytoplankton ammonium uptake	mmol ⁻¹ m ³	0.05
K _{FeS}	Half-saturation for small phytoplankton iron uptake	nmol ⁻¹ m ³	100
K _{NiL}	Half-saturation for large phytoplankton nitrate uptake	mmol ⁻¹ m ³	1.0
K _{NaL}	Half-saturation for large phytoplankton ammonium uptake	mmol ⁻¹ m ³	0.05
K _{FeL}	Half-saturation for large phytoplankton iron uptake	nmol ⁻¹ m ³	200
m _{1S}	Small phytoplankton/zooplankton mortality rate (linear)	d ⁻¹	0.05
m _{2S}	Small phytoplankton/zooplankton mortality coefficient	$(mmol C m^{-3})^{-1} d^{-1}$	0.06
m _{lL}	Large phytoplankton/zooplankton mortality rate (linear)	d ⁻¹	0.1

m_{2L}	Large phytoplankton/zooplankton mortality coefficient	$(mmol C m^{-3})^{-1} d^{-1}$	0.06
X _{minp}	Minimum phytoplankton concentration for linear mortality	mmol C m ⁻³	0.01
a _L	Large zooplankton grazing parameter	$(\text{mmol C m}^{-3})^{-1}$	0.25
GL0	Large zooplankton maximum grazing rate	d ⁻¹	0.85
as	Small zooplankton grazing parameter	(mmol C m ⁻³) ⁻¹	0.25
Gso	Small zooplankton maximum grazing rate	d ⁻¹	1.7
λ	Assimilation efficiency	n.d.	0.8
r _{zs}	Microzooplankton specific respiration rate at T _{ref}	d ⁻¹	0.3
r _{zl}	Mesozooplankton specific respiration rate at T_{ref}	d ⁻¹	0.1
r 1	Small detritus remineralization rate at T _{ref}	d ⁻¹	0.25
r ₂	Large detritus remineralization rate at T _{ref}	d ⁻¹	0.25
E _{ar}	Activation energy for detritus remineralization	kJ mol ⁻¹	54.0
Ws	Small detritus sinking speed	m d ⁻¹	2.
Wl	Large detritus sinking speed	m d ⁻¹	30.
W _{Ca}	CaCO ₃ sinking speed	m d ⁻¹	20.
P _{Ca}	CaCO ₃ production as fraction of mortality	mol CaCO ₃ molC ⁻¹	0.05
k _{Ca}	CaCO ₃ dissolution rate	d ⁻¹	0.0074
S _{Fe1}	Dissolved iron scavenging loss rate (Fe≤L _{Fe})	d ⁻¹	0.001
S _{Fe2}	Dissolved iron scavenging loss rate (Fe>L _{Fe})	d ⁻¹	2.5
L_{Fe}	Ligand concentration	nmol Fe m ⁻³	600.
P _{Fe}	POC-dependence parameter for Fe scavenging	$(\text{mmolC m}^{-3})^{-1}$	0.66
K-NH40xkNH40x	Nitrification rate in darknessconstant	d ⁻¹	0.05
K _E	Half-saturation for irradiance inhibition of nitrification	W m ⁻²	1.
k_{dnf}	Light and nutrient saturated rate of N ₂ fixation at 30°C	mmol m ⁻³ d ⁻¹	0.0225
a	Initial slope for irradiance-dependence of N ₂ fixation	$(W m^{-2})^{-1}$	0.02
K _{Fe}	Half-saturation for Fe dependence of N ₂ fixation	nmol m ⁻³	100.
K _{NO3}	Half-saturation for DIN inhibition of N ₂ fixation	mmol m ⁻³	0.1
O _{mxd}	O ₂ concentration threshold for denitrification	mmol m ⁻³	6.
Af	Anammox fraction of N loss to denitrification	n.d.	0.25

Figure 1 - Schematic of the CanOE biology model. Model currencies including chlorophyll (Chl) are indicated by coloured boxes except oxygen (O₂) and carbonate (CaCO₃). Arrows indicate flows of carbon (C), nitrogen (N) and iron (Fe) between compartments containing small (S) and large (L) phytoplankton (P), zooplankton (Z), and detritus (D) components; counterflows of oxygen are not shown.

Figure 2 - Global distribution of oxygen (O_2) concentration in mmol m⁻³ at 400, 900, and 1400 m (rows) for CanESM5-CanOE, CanESM5, the mean for other (non-CanESM) CMIP6 models, and World Ocean Atlas 20132018 (WOA20132018) observations (columns). Difference from the observation-based fields are shown in Supplementary Figure S3.

Figure 3 - Latitude-depth distribution (surface to 1750 m) of zonal mean oxygen concentration (O₂), oxygen concentration at saturation (O₂(sat)), and apparent oxygen utilization (AOU) in mmol m⁻³ for CanESM5-CanOE, CanESM5, the mean for other CMIP6 models, and observations (WOA20132018). Note different colour scales for different rows. Difference from the observation-based fields are shown in Supplementary Figure S3.

Figure 4 - Taylor diagrams (Taylor, 2001) comparing modelled and observed distributions of oxygen at specific depths from 100 to 3500 m. Angle from the vertical indicates spatial pattern correlation. Distance from the origin indicates ratio of standard deviation in modelled vs. observed (WOA20132018) fields. Red dots represent CanESM5-CanOE, blue dots CanESM5, small grey dots other CMIP6 models, and large grey dots the model ensemble mean for all CMIP6 models except CanESM5 and CanESM5-CanOE.

Figure 5 - Total volume of ocean with oxygen (O₂) concentration less than (a) 6 mmol m⁻³ (mean for last 30 years of the historical experiment) and (b) 60 mmol m⁻³. Observation are from $WOA_{20132018}$.

Figure 6 - Global distribution of aragonite saturation (Ω_A) at 400, 900, and 3500 m for CanESM5-CanOE, CanESM5, the mean for other CMIP6 models, and observations (GLODAP<u>v2</u> + WOA<u>20132018</u>). Note different colour scales for different depths. Difference from the observation-based fields are shown in Supplementary Figure S3.

Figure 7 - Latitude-depth distribution of zonal mean (surface to 1150 m) aragonite saturation state (Ω_A), calcite saturation state (Ω_C), and carbonate ion concentration ([CO₃⁻⁻]) in mmol m⁻³ for CanESM5-CanOE, CanESM5, the mean for other CMIP6 models, and observations (GLODAP<u>v2</u> + WOA20132018). Difference from the observation-based fields are shown in Supplementary Figure S3.

Figure 8 - Taylor diagrams comparing modelled and observed distributions of DIC at specific depths from 100 to 3500 m. Observations are from GLODAPv2 (Lauvset et al., 2016). Red dots represent CanESM5-CanOE, blue dots CanESM5, magenta dots CanESM2, small grey dots other CMIP6 models, and large grey dots the model ensemble mean for all CMIP6 models except CanESM5 and CanESM5-CanOE.

Figure 9 - Taylor diagrams comparing modelled and observed (GLODAP $\underline{v2}$ + WOA20132018) distributions of Ω_A at specific depths from 100 to 3500 m. Symbol colours as in Figure 8.

Figure 10 - Climatological seasonal cycle of surface nitrate concentration averaged for selected ocean regions. Thick red line represents CanESM5-CanOE, thick blue line CanESM5, thick black line observations (WOA20132018), thin grey lines individual CMIP6 models, and thick grey line the model ensemble mean (excluding CanESM5 and CanESM5-CanOE). Regional boundaries are given in Supplementary Table S4S5 and Supplementary Figure S5.

Figure 11 - Climatological seasonal cycle of zonal mean surface nitrate concentration for a selection of CMIP6 models, a model ensemble mean (MEM) excluding CanESM5 and CanESM5-CanOE, and an observation-based data product (WOA20132018). An alternate version showing only for (a) all latitudes and (b) latitudes <20° is given in Supplementary Figure S6.

Figure 12 - Global distribution of dissolved iron (dFe) concentration (log10 of concentration in nmol m⁻³) at the ocean surface for CanESM5-CanOE and other CMIP6 models that published this field. Concentrations exceeding 1000 nmol m⁻³ are masked white. <u>CanESM5 is not included</u> because it does not have prognostic iron.

Figure 13 - Global mean depth profiles of dissolved iron concentration for CanESM5-CanOE and other CMIP6 models that published this field. GFDL-CM4 is excluded because it has very high concentrations (>2000 nmol m⁻³) near the surface. Thick red line represents CanESM5-CanOE, thin grey lines individual CMIP6 models, and the thick grey line the model ensemble mean (excluding CanESM5-CanOE and GFDL-CM4).

Figure 14 - Mean surface nitrate (NO₃) vs. dissolved iron (dFe) concentrations in different oceans, including the major high nutrient / low chlorophyll (HNLC) regions. CanESM5-CanOE is shown as a red dot and other CMIP5 models as grey dots (CanESM5 is not included because it does not have iron). Observed NO₃ is shown as a vertical black line as there are no observational estimates of dFe concentration. For GFDL-CM4, nitrate is estimated as phosphate x 16. Region definitions are given in Supplementary Table S4S5 and Supplementary Figure S5.

Figure 15 - Surface nitrate (NO₃) concentrations along the Pacific equator (mean from 2°S-2°N) during the upwelling season (June-October) for CanESM5-CanOE (red), CanESM5 (blue), and WOA20132018 observations (black).

Figure 16 - Annual mean surface ocean concentration of large and small phytoplankton and zooplankton in CanESM5-CanOE (red) and of phytoplankton and zooplankton in CanESM5 (blue) for the representative ocean regions shown in Figure 14. Observational estimates (black) are for phytoplankton biomass calculated from satellite ocean colour estimates of surface chlorophyll (SeaWiFS/MODIS; Tesdal et al. 2016), assuming a carbon-to-chlorophyll ratio of 50 g/g. Region definitions are given in Supplementary Table S<u>54 and Supplementary Figure S5</u>.

Figure 17 - Mean annual cycle of surface chlorophyll for the representative ocean regions shown in Figures 14 and 16. CanESM5-CanOE large and small phytoplankton concentrations are shown separately and combined (red) along with CanESM5 (blue) and observational estimates (black). Region definitions are shown in Supplementary Table S<u>54 and Supplementary Figure S5</u>.

Figure 18 - Climatological surface particulate organic carbon (POC) vs. chlorophyll for CanESM5-CanOE (red) and observations (black). Data are for all ocean grid points ($2x2^{\circ}$ uniform global grid) for all months of the year where observational data are available. Model POC is offset 17 mg m⁻³ for illustrative purposes. <u>Observed chlorophyll concentrations >1 mg m-3</u> are excluded as they largely represent coastal areas poorly resolved by coarse resolution global ocean models.

Figure 19 - (a) Global total export production (epc100) in PgC y^{-1} (b) and zonal mean export production in molC $m^{-2} y^{-1}$ according to selected CMIP6 models (mean for 1985-2014 of historical experiment). Thick red line represents CanESM5-CanOE, thick blue line CanESM5, thin grey lines individual CMIP6 models, and thick grey line the model ensemble mean (excluding CanESM5 and CanESM5-CanOE).

Figure 20 - Cumulative ocean uptake of carbon dioxide (CO_2) as anthropogenic dissolved inorganic carbon (AnthDIC) in PgC over the course of the historical experiment (1850-2014). Data are shown as successive five-year means. CMIP6 mean (thick grey line) indicates ensemble mean for CMIP6 models (thin grey lines) excluding CanESM5 (blue) and CanESM5-CanOE (red). An observation-based estimate of 145±20 PgC (Friedlingstein et al., 2020) is shown for nominal year 2014 (black).

Figure 21 - Change in export production (epc100) over the course of the historical experiment (1850-2014), normalized to the 1850-1900 mean. Data are shown as successive five-year means. Thick red line represents CanESM5-CanOE, thick blue line CanESM5, thin grey lines other CMIP6 models, and thick grey line the ensemble mean of non-CanESM models.

Figure 22 - (a) Change in total ocean volume with oxygen (O_2) concentration less than (a) 6 mmol m⁻³ and (b) 60 mmol m⁻³ over the course of the historical experiment (1850-2014), normalized to the 1850-1870 mean. Data are shown as successive five-year means. Thick red line represents CanESM5-CanOE, thick blue line CanESM5, and thin grey lines other CMIP6 models..

Dissolved iron model comparison to observations

As there is no gridded global data product for dissolved iron (dFe) we present here some comparisons with individual bottle samples from the GEOTRACES Intermediate Data Product 2017 (Schlitzer et al., 2018), the MBARI data compilation (www3.mbari.org/chemsensor/Data/), and the Pacific data set compiled by PICES WG22 (meetings.pices.int/members/working-groups/disbanded/wg22). The MBARI data include both profile (the "Global Iron Data") and surface transect ("MBARI SOLAS") data (the SOLAS data also include some profiles but are primarily underway surface measurements). The GEOTRACES data are primarily profile data (393 profiles total, only ~10% of 7519 data are <50 m). Most of the MBARI data are also in the PICES data set; these were preprocessed to remove redundant data. Data from the underway surface transects were excluded to avoid overweighting these regions due to autocorrelation, leaving a total of 3575 data points. Concentrations > 2.5 nM, which comprise 1.3% of GEOTRACES data and 3.3% of PICES/MBARI data, were excluded from the model/data comparisons below, except for the individual GA-02 profiles.

Evaluating the CanOE model beyond what has been discussed in the main text identified several key points, most of which have already been made to some degree. CanESM5-CanOE compares favourably with other CMIP6 models but is quite biased towards a 'nutrient-type' rather than 'scavenged-type' profile (Figures S4S9b, c, e-hg). In CanOE rates of Fe scavenging are very high above 0.6 nM dFe and very low below this concentration, resulting in an almost constant deep water concentration of 0.6 nM. The collected data (of which there are >10000 vs only a few hundred in 1997) show that the basic hypothesis articulated by Johnson et al. (1997) still holds: deep-water concentrations are generally close to 0.6 nM, although much higher concentrations are sometimes observed at depths of thousands of metres (Figure \$4\$9b, see e.g. Resing et al., 2014). High concentrations are also sometimes observed in near-surface waters (Figure \$4\$9b) and CanESM5-CanOE is among the models least able to reproduce these. CanESM5-CanOE consistently overestimates the lowest observed concentrations and underestimates the higher ones (Figure \$4\$9d). One might argue that other models do little better in a statistical sense. However, in several models the deviations from the 1:1 line are less systematic than in CanESM5-CanOE; the residuals may be larger but they are more homogeneous. This comparison also largely confirms the results of Séférian et al. (2020, their Figure 5), although only two of the four models shown here were included in their analysis. CNRM-ESM2-1 shows a capacity to simulate the full range of dFe concentrations, as was the case in Séférian et al., although in no case is the spatial pattern correlation very large (in Séférian et al., CNRM-ESM2-1 was the second highest at 0.21). Nonetheless, this analysis indicates that Therefore purely statistical comparisons as in Figures S4S9e-g can be misleading. CanESM5-CanOE shows fairly good skill by these metrics but has systematic biases. These biases can clearly be attributed to the rather simplistic scavenging model employed, and will be addressed in future versions.

We made direct comparisons of our modelled dissolved iron concentrations with GEOTRACES transect data for GA-02 in the Atlantic (e.g., Middag et al. 2015), which was the most spatially extensive transect available (Figure S9a). We show depth profiles from 47°S to 47°N. Mostly this confirms what we already knew from the other analyses presented: our model has a very low scavenging rate below 0.6 nM and a very high rate above, so that deep water concentrations are quite uniform and near-surface concentrations are biased low in high-deposition regions like the northern tropical Atlantic. For the most part, the model reproduces the observed concentrations quite well, given these known biases. What we learn from including this additional analysis is (a) the model is biased high in the Antarctic Bottom Water (Middag et al. 2015 give an excellent presentation of the location and biogeochemical properties of this water mass), and (b) the seasonal biological drawdown in the mid-latitude North Atlantic is weak. The former is probably due mainly to the low scavenging rate at concentrations <0.6 nM, although it may also indicate a high bias in surface waters of the source region (see section 3.3). The latter is probably related to the generally low rate of export production (Figure 19) and the weak North Atlantic spring/summer bloom (Figures 16 and 17). Model annual mean data are shown, but seasonal minima are not much less, compared to the strong summer drawdown in the observations.

Figure <u>S4S9</u>a – Global distribution of dissolved iron (dFe) measurements in the GEOTRACES (black/green) and PICES/MBARI (blue) data compilations. GEOTRACES Pacific data north of 25°S are indicated in green. Blue dots outside the Pacific are MBARI data. <u>Large symbols indicate stations along GA-02 that are compared in Figure S9h.</u>



Figure S4S9b - All observations of dissolved iron (dFe) concentration (<2 nM) plotted against sampling depth. GEOTRACES data are in black and PICES/MBARI data are blue. Model data are global mean profiles of annual mean data as in Figure 13 in the main text.



Figure S4S9c - Mean of observations of dissolved iron (dFe) concentration (<2 nM) for various depth strata, including both GEOTRACES and PICES/MBARI data. Means of all data available in that depth range; spatial coverage is very incomplete and inconsistent among the depth strata. Averaging layers are 50 m (0-1000 m), 100 m (1000-3000 m) or 300 m (>3000 m) thick. Model data are global mean profiles of annual mean data as in Figure 13 in the main text.



Figure <u>S4S9</u>d - Modelled and observed mean dissolved iron (dFe) concentrations (excluding those >2.5 nM) in the upper 50 m at locations where a depth profile was taken (N=1122). Observed data are means of all measurements made at depths <50 m within a given profile. Model data are climatological surface values for the month in which the observed data were collected (red: CanESM5-CanOE; black CNRM-ESM2-1; blue: GFDL-CM4; magenta: MPI-ESM1-2-LR). Selection of models is somewhat arbitrary but includes the models that show the highest overall skill according to the metrics shown in Figures <u>S4S9</u>e-g. Thin lines are linear regressions for individual models; thick black line is 1:1.



Figure \$4\$9e - Root mean square error and correlation coefficient for surface dissolved iron data (upper 50 m mean with maximum of 2.5 nM, as in Figure \$4\$9d) for CMIP6 models for which seasonal data were available.



Figure $\frac{84S9}{5}$ f - As $\frac{84S9}{5}$ e but for the Pacific only (north of 25°S).



Figure $\frac{54S9}{2}$ - As $\frac{54S9}{2}$ e but excluding the Pacific north of 25°S.



Figure S9h - Depth profiles for modelled and observed dissolved iron along the GA-02 transect of the Atlantic Ocean. Station locations are shown in Figure S9a (every third station, sorted by latitude). Model data are annual means.



References

Middag, R., M.M.P. van Hulten, H.M. Van Aken, M.J.A. Rijkenberg, L.J.A. Gerringa, P. Laan, and H.J.W. de Baar, 2015. Dissolved aluminium in the ocean conveyor of theWest Atlantic Ocean: Effects of the biological cycle, scavenging, sediment resuspension and hydrography. Marine Chemistry 177: <u>69–86.</u>

Resing, J.A., P.N. Sedwick, C.R. German, W.J. Jenkins, J.W. Moffett, B.M. Sohst and A. Tagliabue, 2015. Basin-scale transport of hydrothermal dissolved metals across the South Pacific Ocean. Nature 523: 200–203.

Séférian, R., et al., 2020. Tracking improvement in simulated marine biogeochemistry between CMIP5 and CMIP6, Current Climate Change Reports, 6: 95-119.

Schlitzer, R., et al., 2018. The GEOTRACES Intermediate Data Product 2017. Chemical Geology 493: 210-223.

The data file for the PICES WG22 data compilation requests that data users cite the original publications. As all of the data are used except for the SOLAS underway data, all of the original references are listed here:

Boyle, E.A., B.A. Bergquist, R.A. Kayser and N. Mahowald, 2005. Iron, manganese and lead at Hawaii Ocean Time-Series station ALOHA: Temporal variability and an intermediate water hydrothermal plume. Geochimca et Cosmochimica Acta, 69: 933-952.

Brown, M. T., W. M. Landing, and C. I. Measures, 2005. Dissolved and particulate Fe in the western and central North Pacific: Results from the 2002 IOC cruise. Geochem, Geophys. Geosyst., 6, Q10001, doi: 10.1029/2004GC000893.

Bruland, K., Orians, K. and Cowen, P. ,1994. Reactive trace metals in the stratified central North Pacific. Geochem. Cosmochim. Acta. 58, 3171-3182.

Coale, K. H., Fitzwater, S. E., Gordon, R. M., Johnson, K. S. and Barber, R. T., 1996a. Control of community growth and export production by upwelled iron the equatorial Pacific Ocean. Nature, 379:621-624.

Ezoe M, Ishita T, Kinugasa M, Lai X, Norisuye K, Sohrin Y, 2004, Distributions of dissolved and acid-dissolvable bioactive trace metals in the North Pacific Ocean. Geochemical Journal, 38(6), 535-550.

Fitzwater, S.E., K.H. Coale, R.M. Gordon, K.S. Johnson, M.E. Ondrusek, 1996, Iron deficiency and phytoplankton growth in the equatorial Pacific, Deep-Sea Res. II, 43, 995-1015

Fujishima, Y., K. Ueda, M. Maruo, E. Nakayama, C.Tokutome, H.Hasegawa M. Matsui and Y. Sohrin. Distribution of Trace Bioelements in the Subarctic North Pacific Ocean and the Bering Sea (the R/V Hakuhou Maru Cruise KH-97-2), Journal of Oceanography, Vol. 57, pp.261 to 273, 2001

Gordon, R. M., Johnson, K. S. and Coale, K. H., 1998. The behavior of iron and other trace elements during the IronEx I and PlumEx experiments in the equatorial Pacific. Deep-Sea Res. II 45: 995-1041. Gordon, R.M., J.H. Martin, G.A. Knauer, 1982, Iron in north-east Pacific waters, Nature, 299, 611-612 Johnson - MLML World Iron Data Set: Appendix A to Johnson, K. S., Gordon, R. M. and Coale, K. H., 1997, What controls dissolved iron concentrations in the world ocean? Mar. Chem. 57: 137-161. Johnson, K.S., Elrod, V., Fitzwater, S., Plant, J.N., Chavez, F.P., Tanner, S.J., Gordon, M., Westphal, D.L., Perry, K.D., Wu, J., and Karl, D.M., 2003. Surface ocean-lower atmosphere interactions in the

Northeast Pacific Ocean Gyre: Aerosols, iron, and the ecosystem response, GBC(17), 10.1029/2002GB002004

Johnson, K.S., F.P. Chavez, V.A. Elrod, S.E. Fitzwater, J.T. Pennington, K.R. Buck, and P.M. Walz, 2001. The annual cycle of iron and the biological response in central California coastal waters. Geophys. Res. Letters 28: 1247-1250.

Johnson, WK, Miller, LA, Sutherland, ND, Wong, CS, 2005. Iron transport by mesoscale Haida eddies in the Gulf of Alaska, Deep-Sea Res. II, 52: 933-953.

Kitayama, S., K. Kuma, E. Manabe, K. Sugie, H. Takata, Y.

Isoda,K.Toda,S.Saitoh,S.Takagi,Y.Kamei,and K.Sakaoka,2009.Controls on iron distributions in the deep water column of the North Paciffic Ocean:iron(?) hydroxide solubility and humictype fluorescent dissolved organic matter.J.Geophys.Res.,114:C08019,doi:10.1029/2008JC004754

Kondo, Y., 2007. Dynamics of the organic Fe complexing ligands and phytoplankton in the Pacific Ocean. The University of Tokyo. Ph.D. Thesis, 256pp.

Kondo, Y., S. Takeda and K. Furuya, 2012. Distinct trends in dissolved Fe speciation between shallow and deep waters in the Pacific Ocean. Marine Chemistry, 134–135, 18–28

Kuma, K, J. Nishioka, K. Matsunaga, 1996, Controls on iron (III) hydroxide solubility in seawater: The influence of pH and natural organic chelators, Limnol. Oceanogr., 41, 396-407

Kuma, K., A. Katsumoto, H. Kawakami, F. Takatori, K. Matsunaga, 1998, Spatial variability of Fe(III) hydroxide solubility in the water column of the northern North Pacific Ocean, Deep-Sea Res. I, 45, 91-113

Kuma, K., Isoda, Y. and Nakabayashi, S., 2003, Control on dissolved iron concentrations in deep waters in the western North Pacific: Iron(III) hydroxide solubility. J. Geophys. Res., 108 (C9): 3289, doi:10.1029/2002JC001481 (2003).

Landing, W.M., K.W. Bruland, 1987, The contrasting biogeochemistry of iron and manganese in the Pacific Ocean, Geochim. et Cosmo. Acta, 51, 29-43

Mackey, D.J., J.E. O'Sullivan, R.J. Watson, 2002, Iron in the western Pacific: A riverine or hydrothermal source for iron in the Equatorial Undercurrnet?, Deep-Sea Res. I, 49, 877-893 Martin, J.H. and P.M. Gordon, 1988, Northeast Pacific iron distributions in relation to phytor

Martin, J.H. and R.M. Gordon, 1988, Northeast Pacific iron distributions in relation to phytoplankton productivity, Deep-Sea Res. I, 35, 177-196

Martin, J.H., R.M. Gordon, S. Fitzwater, W.W. Broenkow, 1989, VERTEX: phytoplankton/iron studies in the Gulf of Alaska, Deep-Sea Res., 36, 649-680

Nakabayashi, S., K. Kuma, K. Sasaoka, S. Saitoh, M. Mochizuki, N. Shiga, and M. Kusakabe, 2002. Variation in iron(III) solubility and iron concentration in the northwestern North Pacific Ocean. Limnol. Oceanogr. 47: 885-892

Nakabayashi, S., M. Kusakabe, K. Kuma, and I. Kudo, 2001. Vertical distributions of Iron(III) hydroxide solubility and dissolved iron in the northwestern North Pacific Ocean. Geophys. Res. Letters 28: 4611-4614.

Nishioka, J., S. Takeda, I. Kudo, D. Tsumune, T. Yoshimura, K. Kuma, A. Tsuda, 2003, Size-fractionated iron distributions and iron-limitation processes in the subarctic NW Pacific, GRL, 30, NO. 14, 1730, doi:10.1029/2002GL016853.

Nishioka, J., T. Ono, H. Saito, K. Sakaoka, and T. Yoshimura (2011),Oceanic iron supply mechanisms which support the spring diatom bloom in the Oyasio region,western subarctic Pacific,J.Geophys.Res.,116,C02021,doi:10.1029/2010JC006321

Nishioka, J., T. Ono, H. Saito, T. Nakatsuka, S. Takeda, T. Yoshimura, K. Suzuki, K. Kuma, S. Nakabayashi, D. Tsumune, H. Mitsudera, W. K. Johnson, A. Tsuda, (2007) Iron input into the western subarctic Pacific, importance of iron export from the Sea of Okhotsk, Journal of Geophysical Research, 112, C10012, doi:10.1029/2006JC004055.

Nishioka, J., Takeda, S., Wong, C., Johnson, W. 2001. Size-fractionated iron concentrations in the northeast Pacific Ocean: Distribution of soluble and small colloidal iron. Mar. Chem. 74, 157-179

Obata H., 1997. Development of an automated analytical method of iron in seawater and studies on the behavior of iron in the ocean.Kyoto University.Ph.D.Thesis,109pp.

Obata, H., H. Karatani and E. Nakayama, 1993. Automated determination of iron in seawater by chelating resin concentration and chemiluminescence detection. Analytical Chemistry, 65:1524-1528 Obata, H., H. Karatani, M. Matsui and E. Nakayama, 1997. Fundamental studies for chemical speciation of iron in seawater with an improved analytical method. Marine Chemistry, 56:97-106 Roy, E., 2009. The detection and biogeochemistry of trace metals in natural waters. Ph.D. Thesis, University of Maine.

Rue, E.L. and K.W. Bruland, 1995. Complexation of iron(III) by natural organic ligands in the central North Pacific as determined by a new competitive ligand equilibration/adsorptive cathodic stripping voltammetric method. Mar. Chem. 50: 117-138.

Takata, H, Kuma, K, Saitoh, Y, Chikira, M, Saitoh, S, Isoda, Y, Takagi, S, Sakaoka, K, 2006. Comparing the vertical distribution of iron in the eastern and western North Pacific Ocean. Geophys. Res. Lett., 33, L02613, doi:10.1029/2005GL024538, 2006.

Takata, H., K. Kuma, S. Iwade, Y. Yamajyoh, A. Yamaguchi, S. Takagi, K. Sakaoka, Y. Yamashita, E. Tanoue, T. Midorikawa, K. Kimura, J. Nishioka, 2004, Spatial variability of iron in the surface water of the northwestern North Pacific Ocean, Mar. Chem., 86, 139-157

Takeda, S., H. Obata, 1995, Response of equatorial Pacific phytoplankton to subnanomolar Fe enrichment, Mar. Chem, 50, 219-227

Tsuda, A., S. Takeda, H. Saito, J. Nishioka, Y. Nojiri, I. Kudo and others, 2003, A mesoscale iron enrichment in the western Subarctic Pacific induces a large centric diatom bloom, Science, 300, 958-961

Wu, J., E. Boyle, W. Sunda, and L.-S. Wen, 2001. Soluble and colloidal iron in the oligotrophic North Atlantic and North Pacific. Science 293: 847-849.