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Marseille, November 17, 2020

To the Editor, Geoscientific Model Development

Please find enclosed the second revised version of our research article entitled "Implementation and assessment of a carbonate system model (Eco3M-CarbOxv1.1) in a highly-dynamic Mediterranean coastal site (Bay of Marseille, France)". We considered all the valuable comments of the Dr. Munhoven and especially, we addressed the point on the poor scientific justification of the River intrusion experiment. We hope that our manuscript may now be accepted for final publication.

All authors approved the final version of the paper for submission.

Sincerely,

Frédéric DIAZ/Katixa LAJAUNIE-SALLA

Review of the first revised version of

"Implementation and assessment of a carbonate system model (Eco3M-CarbOxv1.1) in a highly-dynamic Mediterranean coastal site (Bay of Marseille, France)"

submitted for publication to Geoscientific Model Development by K. Lajaunie-Salla and co-authors

<u>General reply</u>: We thank again the Reviewer Dr Munhoven for his second evaluation of our work. We thank too him for the detailed and useful comments that contributed to greatly improve the manuscript. We consider all of his comments to improve the manuscript.

1. General comments

The authors' reply the revised manuscript are not very "reviewer-friendly." It is nowadays standard practice to provide a "track change" (or a LATEXDIFF) version of the manuscript clearly identifying the changes made to the text, right in the text. The equivalent information is seemingly given in the reply, except that the line numbers provided there do not match, so that one has to search manually for the exact location of the changes.

The preparation of the manuscript would also have benefited from some extra care. Page numbers restart at 1 after page 23 without any apparent reason.

Reply: The preparation of the revised ms has been reviewed with care and page numbers have been reprocessed. A new check of the different sections has been carefully performed.

1.1 Appreciation of the replies to reviewers

The authors have all in all well responded to the referees' comments, with one exception though. In the response to my comment 2.3, regarding the missing effect of river intrusions on DIC — TA perturbations are taken into account, but as these are carried mainly by HCO3⁻, they also generate DIC perturbations of the same magnitude, which are neglected — I read at the top of the fourth page (page numbers in the response would have been helpful) that

"Concerning the riverine inputs scenarii, we decide to focus on nitrate and alkalinity supply. In fact the model simulates the DIC increases, as is observed, which highlight that the carbonate system module is well resolved."

The reply to the comments is somewhat ambiguous as suggests that DIC changes are taken into account, while they are actually not, as stated in the manuscript at lines 413–414:

"[...] the experimental design on the Rhône River supply only considers the TA perturbation on the carbonate system but not that due to the DIC supply."

So, even if the model reproduces the observed DIC increases (as stated in the reply), this must obviously be for the wrong reasons, as only one half of the effects of the perturbation due to river intrusions is taken into account. By the way, no one argued that the carbonate system module was not well resolved.

<u>Reply</u>: We decided to remove the scenario of AT supply by river and all the text refered to it in the methods and results/discussion sections. See our detailed comment on this point hereafter.

1.2 Appreciation of the revised manuscript

The model description has been improved and the rationale behind the carbonate speciation calculations is now presented in a new appendix. Unfortunately, the layout of that appendix is rather chaotic which makes it difficult to read.

2. Specific comments

2.1 River intrusion experiments: poor justification

The justification added at lines 414–417 for taking the effect of river intrusions into account only in terms of the resulting TA but not DIC perturbations is rather cavalier and scientifically untenable. This is a completely unrealistic assumption that makes the outcome of the experiment meaningless and thus essentially invalidates any conclusion drawn from it.

I only see two options to address this shortcoming:

- 1. the river intrusion experiments are repeated with the effect on DIC included (which should be rather straightforward to correct) and the discussion of the results adapted;
- 2. these experiments are simply taken out of the paper as the current results are essentially unfounded.

Even in preliminary experiments, one must not chose to disregard one of two effects of a perturbation if these are of the same order of magnitude. Such arbitrary choices lead to arbitrary results.

Reply: We have chosen the second option proposed by the Reviewer. Mainly because it was not straightforward to consider DIC supply by River as considered for AT. In fact DIC is not as conservative as AT is regard to salinity. It is then impossible to use a relationship between DIC and salinity as that used between AT and salinity in the previous version of our manuscript. In the context of a 0D modelling developed here, this kind of relationship would have been the only mean to take into account the DIC supply by the Rhône River. In the revised version, we stressed (l 398-401) that rivers also supply TA and DIC and a consideration for these supplies in future works may sensibly modify the modeled carbonate balance in the BoM compared to that presented in this study. Concomitantly we slightly rewrote the end of section 4.1 on the model performance in Discussion.

As noted in the conclusion (1434), a coupling of the Eco3M-CarbOx model to a 3D hydrodynamics model is planed to better represent the complexity of functioning of the BOM. This implementation will enable, for example, to take into account actual DIC and AT supplies from the Rhône River by considering forcing values measured in the River.

3. Technical comments

Reply: All these technical comments have been taken into account in the new revised version of ms.

Page 1, line 22: "the year 2017 that is a period for which" should read "the year 2017 for which"

Page 1, line 25: "of most of variables of carbonate system except Total Alkalinity." should read "of most of the variables of the carbonate system except for Total Alkalinity."

Page 1, line 26: "experiments were also conducted" should read "experiments were conducted"

Page 1, line 26: "to (i) seawater" should read "to (i) a seawater"

Page 1, line 27: "Rhône River plume intrusion" should read "Rhône River plume intrusions"; by the way: the name of that river is sometimes spelled "Rhône", more often "Rhone" — please use the same spelling consistently throughout

Page 1, line 35: "external forcing have" should read "external forcings have"

Page 5, line 188: "a salinity threshold of 37 has been chosen" – is this correct? A threshold of 37 looks rather high to me.

Page 6, line 226: "during MWC period" should read "during the MWC period"

Page 7, line 271: "15 March and 6 May" should either read "15th March and 6th May" or "March 15th and May 6th" (as on line 277, p. 8)

Page 11, line 390: "Moreover, previous study" should read "Moreover, a previous study", or even better reformulate the sentence to read "Moreover, Fraysse et al. (2013) highlight that . . . " and discard the citation in brackets.

Page 11, line 402: "a longer period ca. 15 days" should read "a longer period of ca. 15 days"

Page 11, lines 402–403: "high atmospheric pCO2 value and wind speed" better had to read "high atmospheric pCO2 and high wind speeds"

Page 11, line 411: "due to some two" should read "due to two"

29th and 30th pages (pages nr. 6 and 7), throughout: "in the pH scale" should read "on the pH scale" 29th page (page nr. 6): "Concentration in Total Fluoride (TF) ions" should read "Total Fluoride concentration" (without "ions," as TF includes the non ionic HF)

29th page (page nr. 6): "Concentration in Total Sulphate (TS) ions" should read "Total Sulphate concentration" ("ions" is superfluous)

29th page (page nr. 6): "Concentration in Total Boron (TB)" should read "total boron concentration" (without "ions," as TB includes the non ionic B(OH)3)

29th page (page nr. 6): KF is the dissociation constant of hydrogen fluoride (or of hydrofluoric acid), not of fluoride ions

29th page (page nr. 6): in the expression for KF, the exponent for lons should be 0.5 (or 1/2) and not 1.5.

30th page (page nr. 7): "Every constant are corrected by the hydrostatic pressure" should read "All the constants are corrected for the effect of hydrostatic pressure"

Guy Munhoven Liège, 9th October 2020

Implementation and assessment of a carbonate system model (Eco3M-CarbOx v1.1) in a highly-dynamic Mediterranean coastal site (Bay of Marseille, France).

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- (frederic.diaz@univ-amu.fr)
- 16 Abstract. A carbonate chemistry balance module was implemented into a biogeochemical model of the planktonic
- 17 food web. The model, named Eco3M-CarbOx includes 22 state variables that are dispatched into 5 compartments:
- 18 phytoplankton, heterotrophic bacteria, detrital particulate organic matter, labile dissolved organic and inorganic matter.
- 19 This model is applied to and evaluated in the Bay of Marseille (BoM, France) that is a coastal zone impacted by the
- urbanized and industrialized Aix-Marseille Metropolis, and subject to significant increases in anthropogenic emissions 20
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- 22 The model was evaluated over the year 2017 that is a period for which in situ data of carbonate system are available
- 23 in the study site. The biogeochemical state variables of the model only change with time, to represent the time evolution
- 24 of a sea surface water cell in response to the implemented realistic forcing conditions. The model correctly simulates
- 25 the values ranges and seasonal dynamics of most of variables of carbonate system except Total Alkalinity-for the total
- 26 alkalinity. Several numerical experiments were also conducted to test the response of carbonate system to (i) a seawater
 - temperature increase, (ii) wind events, (iii) Rhône River plume intrusionintrusions and (iv) levels of atmospheric CO2
 - contents. This set of numerical experiments shows that the Eco3M-CarbOx model provides expected responses in the
- 29 alteration of the marine carbonate balance regarding each of the considered perturbation. When the seawater
- 30 temperature changes quickly, the behaviour of the BoM waters alters within a few days from a source of CO2 to the
- 31 atmosphere to a sink into the ocean. Moreover, the higher the wind speed is, the higher the air-sea CO2 gas exchange
- 32 fluxes are. The river intrusions with nitrate and alkalinity supplies lead to a decrease in the pCO2 value, favouring the
- 33 conditions of a sink for atmospheric CO2 into the BoM. A scenario of high atmospheric concentrations of CO2 also
- 34 favours the conditions of a sink for atmospheric CO2 into the waters of the BoM. Thus the model results suggest that
- 35 external forcingforcings have an important impact on the carbonate equilibrium in this coastal area.

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Mis en forme

1. Introduction

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38 Current climate change mostly originates from the carbon dioxide (CO2) increase in the atmosphere at a high annual 39 rate (+2.63 ppm from May 2018 to May 2019, https://www.esrl.noaa.gov/gmd/ccgg/trends/global.html). This 40 atmospheric CO₂ increase impacts the carbonate chemistry equilibrium of the oceanic water column (Allen et al., 2009; 41 Matthews et al., 2009). Oceans are known to act as a sink for anthropogenic CO2, i.e. 30% of emissions, which leads 42 to a marine acidification (Gruber et al., 2019; Orr et al., 2005; Le Quéré et al., 2018). 43 CO2 is a key molecule in the biogeochemical functioning of the marine ecosystem. Photo-autotrophic organisms, mainly phytoplankton and macro-algae, fix this gas through photosynthesis in the euphotic zone and, in turn, produce 44 45 organic matter and dissolved oxygen. Heterotrophic organisms, mainly heterotrophic protists and metazoans consume 46 organic matter and dissolved oxygen by aerobic respiration and, in turn, produce CO2. In the Ocean, the main processes 47 regulating CO₂ exchanges between the atmosphere and sea are the solubility pump and the biological pump at different 48 time-scales. Overall, the thermohaline gradients drive the solubility pump, while the metabolic processes of gross 49 primary production and respiration set the intensity of the biological pump (Raven and Falkowski, 1999). 50 The coastal zones, despite their small surface area and volume compared to those of the open ocean, have a large 51 influence upon carbon dynamics and represent 14 to 30% of the oceanic primary production (Gattuso et al., 1998). At 52 the interface between open-ocean and continents, these zones receive large inputs of nutrients and organic matter from 53 rivers, groundwater discharge, and from atmospheric depositions (Cloern et al., 2014; Gattuso et al., 1998). On the 54 eoastline, coastal areasOn coasts, shorelines are subject to an increasing density of population and associated 55 urbanization (Small and Nicholls, 2003). This rapid alteration of the coastlineshorelines all over the world accelerates 56 the emissions of greenhouses gases near the coastal ocean, and it also involves large discharges of material into the 57 seawater by wastewater runoff and/or rivers (Cloern, 2001). These anthropogenic forcing alter the biogeochemical 58 functioning of these zones and could lead to a growing eutrophication (Cloern, 2001). Moreover, these forcing could 59 affect the carbonate chemistry dynamics and amplify or attenuate the acidification in coastal zones. This alteration of 60 the marine environment may provoke further changes in the structure of the plankton community, including in fine 61 consequences on the populations with high trophic levels, such as teleosts (Esbaugh et al., 2012). At the global scale, coastal zones are considered to be a significant sink for atmospheric CO2, with an estimated flux converging to 0.2 62 63 PgC y-1 (Roobaert et al., 2019). However, some studies highlight that the status of these areas as a net sink or source 64 still remains uncertain due to the complexity of the interactions between biological and physical processes, and also 65 due to the lack of in situ measurements (Borges and Abril, 2011; Chen et al., 2013; Chen and Borges, 2009), Moreover, 66 the capacity for coastal zones to absorb atmospheric CO2 resulting from the increasing human pressure also remains 67 poorly known. There are few works which highlight, under future atmospheric CO2 levels, that shallow seaswhether coastal zones will become a net sink or a reduced source of CO₂ (Andersson and Mackenzie, 2012; Cai, 2011). 68 69 The current increase in the atmospheric CO_2 partial pressure (pCO_2) in the surface ocean is slowly shifting the marine 70 carbonate chemistry equilibrium towards increases in the seawater pCO_2 and bicarbonate ions (HCO₃⁻) and decreases 71 in pH and carbonate ions (CO₃²⁻) (Hoegh-Guldberg et al., 2018). These trends were already described in several coastal 72 and open-ocean locations worldwide (Cai et al., 2011). In a coastal Northwestern Mediterranean site, a 10-year time-73 series of in situ measurements highlights a trend of pH decrease and pCO2 increase (Kapsenberg et al., 2017). Low pH 74 values can inhibit the ability of many marine organisms to form the calcium carbonate (CaCO₃) used in the making of 75 skeletons and shells (Gattuso et al., 2015). In an extreme case, this shift may promote dissolution of CaCO₃ because 76 the water will become under-saturated with respect to CaCO₃ minerals (Doney et al., 2009).

Mis en forme : Centré

The present study is dedicated to the implementation of a carbonate system module into a preexisting biogeochemical model of planktonic food web. This new model, named Eco3M-CarbOx (v1.1) is then evaluated in a highly-dynamic

coastal site, *i.e.* Bay of Marseille (BoM) in the Northwestern Mediterranean Sea. This evaluation is performed on the seasonal dynamics of biogeochemical and carbonate modeled variables against that of the corresponding *in situ* data available over the year 2017. This study is extended by a fine analysis of the variability of the marine carbonate system (stocks, fluxes) in relation to physical (*e.g.* wind events, river intrusions, temperature increases, changes in the atmospheric *p*CO₂ levels) and biogeochemical processes (gross primary production (GPP) and respiration (R)) in the study site. The BoM is suitable to this kind of study because this coastal area is subject to high emissions of atmospheric CO₂ from the nearby urban area, and it also receives effluents from the Aix-Marseille metropolis. In addition, strong wind events (Mistral) regularly occur, which could lead to (i) strong latent heat losses at the surface (Herrmann et al., 2011) and upwelling along the coast with a common consequence of cooling effect and (ii) RhoneRhône River plume intrusion under specific wind conditions (Fraysse et al., 2013, 2014). In this regional context, many anthropogenic forcing can interact with the dynamics of the carbonate systems. Natural determinants of the composition of the marine planktonic community can also play a crucial role in these dynamics.

The Eco3M-CarbOx biogeochemical model was developed to represent the dynamics of the seawater carbonate system and plankton food web in the BoM. The model was implemented using the Eco3M (Ecological Mechanistic and

Modular Modelling) platform (Baklouti et al., 2006). The model structure used is based on an existing model of the

plankton ecosystem (Fraysse et al., 2013), including a description of Carbon (C), Nitrogen (N) and Phosphorus (P)

marine biogeochemical cycles. The Eco3M-CarbOx model includes 22 prognostic state variables that are split into 5

compartments: phytoplankton, heterotrophic bacteria, detrital particulate organic matter, labile and semi-labile

2. Materials & Methods

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2.1 Numerical model description

dissolved organic matter, nutrients (ammonia, nitrate and phosphate), dissolved oxygen, and carbonate system variables (Fig. 1). In this study, the state variables of the Eco3M-CarbOx model only change along time (i.e. usually termed "model 0D"), they are representative of the time evolution of a sea surface water cell but this biogeochemical model is not coupled with a hydrodynamic model. The model presented in this study includes a set of new developments and improvements in the realism of the plankton web structure and process formulations compared to the model of Fraysse et al. (2013). In order to improve the representation of chlorophyll concentration in the Bay of Marseille the phytoplankton is divided in two groups: one with some ecological and physiological traits of the Synechococcus cyanobacteria, which is one of the major constitutive members of pico-autotrophs in Mediterranean Sea (Mella-Flores et al., 2011), and another with traits of large diatoms, which are generally observed during spring blooms at mid-latitudes (Margalef, 1978). For both of the phytoplankton, there is a diagnostic chlorophyll-a variable related to the phytoplankton C-biomass, the phytoplankton N-to-C ratio, and the limiting internal ratio f_0^N (Faure et al., 2010; Smith and Tett, 2000; Tab. B2, Appendix B). The functional response of primary production was modified using another formulation of temperature limitation function which takes into account the optimal temperature of growth for each phytoplankton group. The exudation of phytoplankton was modified taking into account the intracellular phytoplankton ratio. For the uptake of matter by bacteria and the remineralization processes the dependence on intracellular bacteria ratio was modified. A temperature dependence of all biogeochemical processes was added to take into account the effects of rapid and strong variations of seawater temperature on plankton during episodes of upwelling for instance that are usually observed in the BoM. Also certain parameters in some formulations were modified owing to the alterations of some formulations. (Tabs. B4 & B5, Appendix B).

119 Additionally, a carbonate system module was developed and three state variables were added: dissolved inorganic 120 carbon (DIC), total alkalinity (TA) and the calcium carbonate (CaCO₃) implicitly representing calcifying organisms. The knowledge of DIC and TA allows the calculation of the pCO2 and pH (total pH scale) diagnostic variables, 121 122 necessary for resolving all the equations of the carbonate system. These equations use apparent equilibrium constants, 123 which depend on temperature, pressure, and salinity (Dickson, 1990a, 1990b; Dickson and Riley, 1979; Lueker et al., 2000; Millero, 1995; Morris and Riley, 1966; Mucci, 1983; Riley, 1965; Riley and Tongudai, 1967; Uppström, 1974; 124 125 Weiss, 1974). The details of the resolution of carbonate system module are given in the Appendix A. For this module 126 three processes were also added: the precipitation and dissolution of calcium carbonate and the gas exchange of pCO_2 127 with the atmosphere. Based on the review of Middelburg (2019), it is considered that: (i) TA decreases by 2 moles for 128 each mole of CaCO3 precipitated, by 1 mole for each mole of ammonium nitrified, by 1 mole for each mole of 129 ammonium assimilated by phytoplankton, and TA increases by 2 moles for each mole of CaCO3 dissolved, and by 1 130 mole for each mole of organic matter mineralized by bacteria in ammonium (SeeTab. B2, Appendix B-Tab. B2). (ii) 131 DIC is consumed during the photosynthesis and calcification processes and is produced by respiration (of 132 phytoplankton, zooplankton, and bacteria) and the CaCO3 dissolution processes. Moreover, the dynamics of DIC are 133 altered by the CO2 exchanges with the atmosphere (Tab. B2, Appendix B). The air-sea CO2 fluxes are calculated from 134 the pCO₂ gradient across the air-sea interface and the gas transfer velocity (Tab. B3, Appendix B) estimated from the 135 wind speed and using the parametrization of Wanninkhof (1992).

136 In the Eco3M-CarbOx model, zooplankton is considered as an implicit variable. However, a closure term based on the 137 assumption that all of the matter grazed by the zooplankton and higher trophic levels returns as either organic or 138 inorganic matter by excretion, egestion and mortality processes is taken into account (Fraysse et al., 2013). The model 139 considers a "non-redfieldian" stoichiometry for phytoplankton and bacteria. All the biogeochemical model 140 formulations, equations and associated parameters values are detailed in the Appendix B.

Study area

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The RhoneRhône River, which flows into the Gulf of Lions, is the greatest source of freshwater and nutrients for the Mediterranean Sea, with a river mean flow of 1800 m³ s⁻¹ (Pont et al., 2002). Several studies highlight the eastward intrusion events from the RhoneRhône River plume in the BoM under East and South-easterly wind conditions, which favor biological productivity (Fraysse et al., 2014; Gatti et al., 2006; Para et al., 2010). The biogeochemistry of the BoM is complex and highly driven by hydrodynamics. For example, North-Northwesterly winds induce upwelling events which bring upward cold and nutrient-rich waters (Fraysse et al., 2013). Moreover, the oligotrophic Northern Current occasionally intrudes into the BoM (Petrenko, 2003; Ross et al., 2016). Despite the presence of several marine protected areas around the BoM (the Regional Park of Camargue, the Marine protected area Côte Bleue and the National Park of Calanques), it is strongly impacted by diverse anthropogenic forcing, because industrialized and urbanized areas are located all along the coast. From the land, the BoM receives

The BoM is located in the eastern part of the Gulf of Lions, in the Northwestern Mediterranean Sea (Fig. 2). The city

of Marseille, located on the coast of the BoM, is the second largest city of France, with a population of ca. 1 million.

153 154 nutrients and organic matter from the urban area of the Aix-Marseille metropolis (Millet et al., 2018), the industrialized 155 area of Fos-sur-Mer (one of the biggest oil-based industry areas in Europe), and the Berre Lagoon, which is eutrophized 156 (Gouze et al., 2008; Fig. 2C). From the atmosphere, the BoM is subject to fine particles deposition and greenhouse gas

157 emissions (including CO2) from the nearby urban area, and it also receives effluents from the Aix-Marseille metropolis.

2.3 Dataset

The modelled variables of the carbonate system (DIC, TA, *p*H and *p*CO₂) and chlorophyll-a are hereafter compared to observations collected at the SOLEMIO station (Figs. 2C & 3), which is a component of the French national monitoring network (Service d'Observation en Milieu Littoral - SOMLIT, http://somlit.epoc.u-bordeaux1.fr/fr/). Major biogeochemical parameters have been recorded since 1994. Carbonate chemistry variables (*p*H, *p*CO₂, DIC and TA) have been available since 2016, every two weeks.

In the present work, the Eco3M-CarbOx model was run for the whole year of 2017. This year was chosen because in

2.4 Design of numerical experiments

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situ data of carbonate systems (DIC, TA, pH and pCO₂) are available for the whole year at the SOLEMIO station (Fig. 166 2C). The biogeochemical variables were initialized using in situ data from winter conditions (Tab. B1, Appendix B). 167 The model was forced by time-series of sea surface temperature and salinity, wind (at 10 m), light, and atmospheric 168 169 CO2 concentrations. The sea temperature time-series is from in situ hourly data recorded at the Planier station (Fig. 2C). For salinity, hourly in situ data from the SOLEMIO station and from the CARRY buoy were used (Fig. 2C). Wind 170 171 and light hourly time-series were extracted from the WRF meteorological model at the SOLEMIO station (Yohia, 172 2017). Finally, we used hourly atmospheric CO2 values from in situ measurements recorded at the Cinq Avenues station 173 (CAV station, Fig. 2B) by the AtmoSud Regional Atmospheric Survey Network, France (https://www.atmosud.org). 174 This simulation is the reference simulation (noted S0). As highlighted previously, RhoneRhône River plume intrusions 175 (due to wind-specific conditions) have an impact on the dynamics of primary production (Fraysse et al., 2014; Ross et 176 al., 2016) and then on the seawater CO2-concentrations, carbonate system. Moreover, the seawater temperature and 177 atmospheric CO2 variations control the seawater CO2 dynamics via the solubility equilibrium and gas exchange with 178 the atmosphere (Middelburg, 2019). In order to quantify the impact of different forcing, several simulations (hereafter 179 noted S), which are summarized in Table 1, were conducted:

- 180 Impact of seawater temperature increase, S1: the forcing time-series of *in situ* temperatures was shifted by +1.5°C
 181 (Cocco et al., 2013).
 - Impact of wind events: a first simulation S2 was run with a constant wind intensity of 7 m s⁻¹ (2017 annual average wind speed) throughout the year and a second one (S3) with two three-day periods of strong wind speed (20 m s⁻¹) representative of short bursts of Mistral (data not shown) starting on May 15th and August 15th, and a constant value of 7 m s⁻¹ the rest of the year.
 - Impact of Rhonenutrient supply (nitrate) during a Rhône River plume intrusion (a salinityS4). A threshold of 37 has been chosen to identify the presence of low-salinity waters from the RhoneRhône River plume):
 - Nitrate inputs were simulated during in the river plume intrusions (\$4):forcing file of salinity. Here, the levelcontents of nitrate supplied by the river depends on the salinity-level. A relationship between these two variables was then established using NO₃ and salinity data atfor the SOLEMIO point from the MARS3D-RHOMA coupled physical and biogeochemical model (Fraysse et al., 2013; Pairaud et al., 2011), which. This relationship has already been used successfully to reproduce realistic observed conditions in the studies of Fraysse et al. (2014) and Ross et al. (2016): NO_{3intrusion} (mmol m⁻³) = -1.770 ×S + 65.
 - TA inputs were simulated during river plume intrusions (S5): the level of TA supply by the river depends on the salinity. A relationship was established using *in situ* data from the SOLEMIO station during river intrusion:

 TA_{intrusion} (µmol kg⁻¹) = -21.0×S + 3400.
 - Non-urban atmospheric CO₂ concentrations (S6S5): this simulation takes into account the forcing of atmospheric CO₂ values measured at the Observatoire de Haute Provence station (OHP, Fig. 2B) located outside of the Aix-

Mis en forme : Couleur de police : Couleur personnalisée(RVB(35:31:32))

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Marseille metropolis from the ICOS National Network, France (http://www.obs-hp.fr/ICOS/Plaquette-ICOS-201407_lite.pdf).

In this work, we calculated the daily mean values of state variables, the statistical parameters and mean fluxes of modeled processes throughout the year and over two main hydrological periods: the stratified and mixed water column periods. The stratified water column (SWC) is defined with a temperature difference between the surface and bottom of more than 0.5°C (Monterey and Levitus, 1997). For the simulated year (2017), the SWC period lasts from May 10th to October 20th. The mixed water column (MWC) period corresponds to the rest of the year.

3. Results

3.1 Model skills

Following the recommendations of Rykiel (1996), three criteria were considered to evaluate the performance of our model:

- Does the model reproduce the timing of the observed variations of carbonate system at the seasonal time scale?
- Does the model reproduce the observed pCO₂ and pH ranges at the seasonal time scale?
- Analysis of the Willmott Skill Score (WSS): this index is an objective measurement of the degree of
 agreement between the modeled results and the observed data. A correct representation of observations by
 the model is achieved when this index is higher than 0.70 (Willmott, 1982).

Over most of the studied period, the model simulates lower chlorophyll-a concentrations than the *in situ* observations, especially during the MWC period (Fig. 3A). Two maxima of chlorophyll-a concentrations are observed *in situ*: the first one at *ca*. 1.71 mg m⁻³ in March and the second one at *ca*. 0.68 mg m⁻³ in May. They are both linked to RhoneRhône River plume intrusions. Several *in situ* maxima between 0.50 and 0.70 mg m⁻³ are observed between March and April (at the end of the MWC period), and they signaled the spring bloom event (Tab. 2 & Fig. 3A). The biogeochemical model quantitatively reproduces the spring bloom observed at the end of the MWC period (Fig. 3A) with a maximum value of *ca*. 0.69 mg m⁻³. The model does not catch the two aforementioned maxima of chlorophyll, and it contains a low WSS and a strong bias (0.37 and +0.22 mg m⁻³, respectively - Tab. 2).

On the whole, the seasonal variations of the seawater $p\text{CO}_2$ are correctly simulated by the biogeochemical model (Fig. 3B), even if the values are rather overestimated during the MWC period. From January to February, the model reproduces the slight decrease in the observed $p\text{CO}_2$ and from February to March the increase in $p\text{CO}_2$ even if the latter modelled remains smaller. In mid-April, during the simulated spring bloom period, the observed drop in $p\text{CO}_2$ and increase in pH are also spotted in the model (Fig. 3B & 3C). The model especially succeeds in reproducing the observed increase in relation to high temperatures during the SWC period. The reduction of the CO₂ solubility due to thermal effects mostly explains the increase in $p\text{CO}_2$ during the SWC period. The strong standard deviation of modeled values during the SWC period can be explained by the rapid changes in temperature probably due to upwelling usually occurring at this time of the year (Millot, 1990). The range of modeled $p\text{CO}_2$ values (345 - 503 μ atm) encompasses

the range of observed values (358 - 471 µatm; Tab. 2). The statistical analysis provides a mean bias of +23 µatm, and

234 a WSS of 0.69 (Tab. 2).

The seasonal dynamic of pH is mostly reproduced by the model, and in particular, the decrease during the SWC period (Fig. 3C). However, the modelled pH is generally underestimated throughout the year, except during the SWC period,

with a mean bias of -0.015 (Tab. 2). The seasonal range is captured by the model with a minimum value during the

- SWC period (7.994 vs. 8.014 for observations; Tab.2) and a maximum one during the MWC period (8.137 vs. 8.114
- 239 for observations; Tab.2). The statistical analysis highlights an index of agreement between the in situ data and the
- 240 model outputs higher than 0.70 (Tab. 2).
- 241 The seasonal variations of DIC show the highest values during the MWC period and a decrease (resp. increase) during
- 242 the beginning (resp. the end) of the SWC period (Fig. 3D). The lowest values are observed during September. The
- 243 Eco3M-CarbOx model closely matches the seasonal dynamic by reproducing the range of extreme observed values
- 244 (Tab. 2). The mean bias is also small (-8.48 μmol kg⁻¹, Tab. 2). More than 70% (0.73, Tab. 2) of modeled DIC
- concentrations are in statistical agreement with the corresponding observations.
- 246 The seasonal cycle of measured TA does not show a clear pattern (Fig. 3E). Large variations of values ranging between
- 247 2561 and 2624 µmol kg-1 (Tab. 2) are observed, whatever the hydrological season is that is considered. The
- 248 biogeochemical model provides almost constant values around 2570 µmol kg⁻¹ all along the year, which is lower than
- 249 in situ data. With a low WSS index of agreement and a large mean bias (Tab. 2), the model is not able to confidently
- 250 reproduce the observed variations of TA (Fig. 3E & Tab. 2).

3.2 Carbon fluxes and budgets

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- 252 For the year 2017, the values of temperature vary between 13.3°C and 25.9°C (Fig. 4A). The DIC variations closely
- 253 match those of temperature (correlation coef. -0.75). For example, the spring increase in temperature leads to a decrease
 - in DIC concentrations (Figs. 4A & 4C), and the minimum values are reached at the end of SWC period. Over the
 - simulated period, the air-sea CO₂ fluxes (F_{aera}) vary between -14 and 17 mmol m⁻³ d⁻¹, with a weakly positive annual
 - budget of +6 mmol m⁻³ y⁻¹ (or +0.017 mmol m⁻³ d⁻¹, Tab. 3). Then, the BoM waters would act as a net source of CO₂
- 257 to the atmosphere on an annual basis. However, on a seasonal basis, the BoM waters would change from a net sink
- during the MWC period (F_{aera} <0; Tab. 3) to a net source during the SWC one (F_{aera} >0; Tab. 3).
- 259 On an annual basis, the gross primary production (GPP) and total respiration (R) are balanced, leading to a null average
- 260 net ecosystem production (NEP, NEP=GPP-R) (Fig. 4F & Tab. 3). The intensity of autotroph respiration (Ra) is lower
- than that of primary production (annual mean of 0.065 vs. -0.413 mmol m⁻³ d⁻¹, respectively Tab. 3). While the
- zooplankton and bacterial respiration account for an average of 0.348 mmol $m^{-3}d^{-1}$ (Tab. 3). On a seasonal basis, the
- 263 model highlights an ecosystem dominated by autotrophy during the MWC period (NEP>0; Tab. 3) and heterotrophy
- during the SWC period with higher fluxes values (NEP<0; Tab. 3). The biogeochemical fluxes show the strongest
- variations along the SWC period, following those of temperature (Fig. 4F). The maximum GPP occurs in April and is
- 266 correlated with the maximum chlorophyll concentration. At this time, the ecosystem is autotroph (NEP>0; Figs. 4B &
- 267 4F), and is a net sink for atmospheric CO₂, which explains the DIC and seawater pCO₂ decreases during the bloom
- 268 period (Figs. 4C, 4D & 4E)
- When looking in details at the temperature and salinity 2017 time-series (Fig. 4A), several crucial events can be seen
- 270 occurring, including freshwater intrusions (e.g. 15-March 15th and 6-May 6th) into the BoM and large variations of
- temperature in relation with upwelling events or latent heat losses due to wind bursts. The largest freshwater intrusion
- 272 from the RhoneRhône River plume occurs in mid-March, with a minimum observed salinity of ca. 32.5 at the
- 273 SOLEMIO station (Fig. 4A). During this event, the seawater pCO₂ decreases and pH increases concomitantly (Figs.
- 274 4C & 4D). Then, seawater appears to be temporarily under-saturated in CO2 and the BoM waters thus acts as a sink
- for atmospheric CO₂ at the time of intrusion (Fig. 4E).
- During the SWC period, upwelling events quickly cool the surface seawater. In two days, from July 25th to 27th, the
- water temperature drops from 24.7° C to 16.9° C (Fig. 4G). The decrease in temperature corresponds to the increase in
 - DIC concentrations (Fig. 4I). Concomitantly, the values of seawater pCO₂ decrease from 497 to 352 µatm and pH

- 279 increase from 7.99 to 8.12 (Figs. 4I & 4J). This event quickly changes the BoM waters from a source to a sink for
- 280 atmospheric CO₂ (from +17 to -14 mmol m⁻³ d⁻¹, Fig. 4K), and also from a net heterotroph to a net autotroph ecosystem
- 281 (Fig. 4L).

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3.3 Impact of external forcing on the dynamics of carbonate system

3.3.1 Temperature increase

- Here we compare the reference simulation S0 with the S1 simulation (seawater temperature elevation of 1.5°C Fig. 284
- 285 5). During the year, there are few changes on the carbonate system variables such as the pCO_2 and pH (data not shown).
- The main alterations occur during the blooms of phytoplankton. The simulated bloom of phytoplankton occurs later, 286
- 287 at beginning of May, for both diatoms and picophytoplankton, with a maximum value of chlorophyll at 1.4 and 0.4 mg
- 288 m⁻³, respectively (Figs. 5A & 5F).
- 289 As both the limitations due to light and nutrients remain about the same during the simulations S0 and S1, this
- 290 counterintuitive occurrence of bloom relative to changes in temperature is mainly explained by the temperature limiting
- 291 function, which depends on the optimal temperature of growth (Topt). For the picophytoplankton, from January to April,
- 292 the increase of 1.5°C drastically reduces the limitation by temperature (Fig. 5C), because the temperature is closer to
 - the optimal temperature (T_{opt} =16°C, Tab. A4) during S1 than S0. In the S0 simulation, the temperature reaches T_{opt} ca.
- 293 April 15th and it induces the bloom, while at the same time in S1 the temperature moves slightly away from Toot and it 294
- does not enable the triggering of a bloom. At the time of the bloom in S1, the opposite configuration occurs. In S0, the 295
- 296 ambient temperature is again far from Topt, explaining the absence of bloom, while in the S1 the ambient temperature
- 297 is closer to Topt enabling the occurrence of bloom. The picophytoplankton bloom then occurs later in the warm
- simulation S1 than in the reference simulation S0 (Fig. 5A). The duration and termination of bloom is controlled both 298
- 299 by the nutrients availability and the temperature (Figs. 5C & 5D). Inversely, from January to April, the diatoms' growth
- 300 limitation by temperature is strengthened in the warm simulation S1 (Fig. 5H), because the resulting ambient
- temperature is further from the optimum temperature (Topt=13°C, Tab. A4) than that in the reference simulation S0. 301
- This induces a slower growth of diatoms and a delay of the maximum concentration (Fig. 5F). Afterwards the 302
- 303 photosynthesis is mainly limited by temperature (Fig. 5H).
- 304 The ecosystem is net autotroph at the time of blooms whatever the simulation considered (NEP>0; Fig. 5E) and the
- 305 quantity of DIC (not shown) fixed through autotroph processes is larger than that released by heterotroph processes.
- 306 During the short period of bloom, the seawater pCO₂ decreases, leading to some negative air-sea fluxes (i.e. an oceanic
- 307 sink for atmospheric CO2). In the warm simulation, the later occurrence of bloom enables the period of the spring sink
- 308 to extend by ca. three weeks over May relative to the reference simulation (Fig. 5J).

309 3.3.2 Wind speed

- 310 The Bay of Marseille is periodically under the influence of strong wind events (Millot, 1990). Here we compare two
- 311 simulations: one with a constant wind value (S2) and the other one with two wind events that occur in May and August
- 312 (S3) (Figs. 6A & D). The result of this numerical experiment shows that the stronger the wind speed is, the higher the
- 313 air-sea fluxes are, mainly owing to the increase in gas transfer velocity. Depending on the gradient of CO2 between
- 314 seawater and the atmosphere, strong wind speeds will favor either the emission or uptake of CO2 (Figs. 6B & E). In
- 315 May, with the air-sea CO2 flux being positive, the outgassing of CO2 to the atmosphere is enhanced leading to a
- 316 decrease in seawater pCO₂ (Fig. 6C). On the contrary, in August the oceanic sink of atmospheric CO₂ is amplified
- 317 which leads to an increase in the seawater pCO2 value (Fig. 6F).

3.3.3 Supply in nitrate and alkalinity by river inputs

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According to the model results (Fig. 7), the occasional inputs of nitrate (S4) that are linked to RhoneRhône River plume intrusions favor primary production and they led to increased chlorophyll concentrations (Figs. 7B & 7C) five times during the SWC period. These blooms, as seen previously, lead to a decrease (resp. increase) in the seawater pCO_2 (resp. pH) (Figs. 7E & 7F). It can be noted that with the strongest river supply at mid-March (Figs. 7A & 7B) the occurrence of the spring bloom is earlier (Fig. 7C) than that occurring in the reference simulation (S0). The time lag between river nutrient supply and bloom is due to the temperature limitation (Fig. 4C). During blooms occurring within the SWC period following intrusions, the DIC concentrations are generally lower than those of the reference simulation, as in the case of the bloom of mid-May (decrease by ca. 15 μ mol kg⁻¹, Fig. 7J), due to the autotroph processes dominating the heterotroph ones. In turn, the seawater pCO_2 drops by ca. 30 μ atm (Fig. 7K) and pH increases by ca. 0.030 (Fig. 7L). Nitrate inputs, favoring primary production, reduce the source of CO_2 to the atmosphere or and

intensify the sink of atmospheric CO₂ into the waters of BoM (FigFigs. 7E & 7K),

The supply of alkalinity during the Rhone River plume intrusions (Fig. 8A) significantly increases the DIC

concentrations (ca. +50 µmol kg⁴, Figs. 8B & 8F), in every hydrological period considered. During the strongest

freshwater input at mid-March, the sharp TA increase by ca. +150 μmol kg⁴-(Fig. 8E) leads to a large pCO₂ drop by

ca. 92 μatm and a pH increase by 0.13 (Figs. 8G & 8H). The air-sea gradient of pCO₂ increases at mid-March, favoring

sink conditions for atmospheric CO2 into the waters of the BoM (Fig. 8G).

3.3.4 Urban air CO2 concentrations

The Aix-Marseille metropolis is strongly subject to urban emissions to the atmosphere (Xueref-Remy et al., 2018a).

The seasonal variability of atmospheric CO_2 concentrations at the urban site (CAV station, Fig. 2) is much higher than

that observed in a non-urban area (OHP station, Fig. 2), especially during the MWC period (Fig. 9A8A): CO2

concentrations vary between 379 and 547 µatm at the CAV station and between 381 and 429 µatm at the OHP station.

Moreover, in winter the atmospheric pCO₂ is higher in the urban area than non-urban area, whereas in summer those

of both areas are quite close. These differences in the seasonal pattern and between areas are usually explained by (i)

the thinner atmospheric boundary layer, (ii) the decreased fixation of CO_2 by terrestrial vegetation, and (iii) the greater

influence of anthropogenic activities by emissions from heating (Xueref-Remy et al., 2018b). Forcing the model by atmospheric pCO₂ values from urban or non-urban site can lead to significant differences in the values of the seawater

 pCO_2 during the MWC period especially. The air-sea gradient of pCO_2 is higher when using a forcing derived from

the CO2 concentrations originating from an urban area than from non-urban area, which strengthens the sink of

atmospheric CO_2 into the waters of BoM. The seawater pCO_2 is then lower with non-urban area pressure ($\frac{86S5}{1}$) than

with urban area pressure (S0), because of lower CO₂ solubility in the BoM (Fig. 9B8B).

4. Discussion

4.1 Model performance

The evaluation of model skill vs. in situ data highlights that the modeled pH, pCO₂, DIC are in acceptable agreement

with observations (Fig. 3). The seasonal variations observed for the different variables are captured by the model,

including for example the seasonal decrease in DIC and pH during the SWC period, in relation to the increase in pCO₂,

and the inverse scenario during the MWC period. The chlorophyll content variability is not well reproduced, especially

during spring (Fig. 3A), even taking into account the nitrate supply from the RhoneRhône River plume intrusion (Fig.

7C). This is due to the multiple origins of chlorophyll, organic matter, and nutrients in the BoM that are not accounted

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for in the Eco3M-CarbOx model: autochthonous marine production, and allochthonous origins from the RhoneRhône and Huveaune River plumes (Fraysse et al., 2013). The observed variations and levels of TA are not correctly simulated by the model (Fig. 3F), even taking into account the supply of TA coming from the Rhone River plume (Fig. 8). The formulation used in this study for TA inputs from rivers needs to be refined and compared with other works (Gemayel et al., 2015; Schneider et al., 2007).3F). The study of Soetaert et al. (2007) highlights that the main variations of TA in the marine coastal zones are linked to freshwater supplies and marine sediments. The present study does not take into account the inputs of TA from the Rhône River and the water-sediment interface, and it may explain why the TA variable is not correctly predicted by our model.

4.2 Contribution of physical and biogeochemical processes to the variability of carbonate system

The contribution of each biogeochemical process to the DIC variability can be assessed using the presented model: the aeration process contributes to 78% of the DIC variations and biogeochemical processes together to 22% (Tab. 3). As mentioned by Wimart-Rousseau et al. (2020), the model suggests that the seawater pCO₂ variations and associated fluxes would be mostly driven by the seawater temperature dynamics. Moreover, the seasonal variations of the air-sea CO₂ flux are in agreement with some previous field studies (De Carlo et al., 2013; Wimart-Rousseau et al., 2020),

which measured a weak oceanic sink for atmospheric CO₂ during winter and a weak source to the atmosphere during

372 summer.

The model results reveal that temperature would play a crucial role in controlling two counterbalanced processes: (1) the carbonate system equilibrium and (2) the phytoplankton growth. The increase in temperature during SWC leads to a higher pCO_2 in seawater due to the decrease in the CO_2 solubility (Middelburg, 2019) and, at the same time, the fixation of DIC by phytoplankton is favored, leading to a decrease in the pCO_2 level. The imbalance between the latter two processes leads to a change in the ecosystem status (autotrophic or heterotrophic) and the corresponding behavior as a sink or source to the atmosphere. In case of a 1.5°C rise over the whole year, the temperature variation has a very small impact on the carbonate system dynamics. However, it favors the autotrophic processes and strengthens the oceanic sink of atmospheric CO_2 during the bloom of phytoplankton (Figs. 5E & 5J).

4.3 Contribution of the external forcing to the variability of carbonate system

In line with several previous works on the Northwestern Mediterranean Sea (De Carlo et al., 2013; Copin-Montégut et al., 2004; Wimart-Rousseau et al., 2020), the model also suggests that the status of the Bay of Marseille regarding sink or source for CO₂ could change at high temporal frequency (*i.e.* hours to days). Bursts of North, Northwestern winds, lead to sudden and sharp decreases in seawater temperature (<2 days, Fig. 4G) either directly by latent heat loss through evaporation at the surface (Herrmann et al., 2011) or indirectly by creating upwelling (Millot, 1990), with the consequences of decrease in the seawater *p*CO₂ values (Fig. 4J) and *in fine*, an alteration of the CO₂ air-sea fluxes. Model results suggest that the fast variations of temperature could lead to rapid changes of the sink *vs.* source status in this coastal zone (Fig. 4K). Moreover, previous study on the BoM highlights that upwelling favors ephemeral blooms of phytoplankton by nutrient supplies up to euphotic layer (Fraysse et al., 2013) and would, in turn, contribute to the seawater *p*CO₂ decrease. Moreover, Fraysse et al. (2013) highlight that upwelling in the BoM favors ephemeral blooms of phytoplankton by nutrient supplies up to euphotic layer and would, in turn, contribute to the seawater *p*CO₂ decrease. North, and Northwestern winds through latent heat losses and/or upwelling events could then enhance the sink for atmospheric CO₂ due to the temperature drop and nutrients inputs. However, these results remain to be preliminary because in our experimental design only the cooling effect of upwelling on the carbonate balance is taken into account. But concomitantly, upwelling usually bring nutrients and DIC at the surface and these supplies could also perturb the

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balance of the carbonate system. A next coupling of the Eco3M-CarbOx model with a tridimensional hydrodynamic model would enable to certainly embrace the multiple effects of upwelling on the dynamics of the carbonate system in this area and refine the results presented in this study. High wind speeds (>7 m s⁻¹) amplified considerably the gaseous exchange of CO₂ (De Carlo et al., 2013; Copin-Montégut et al., 2004; Wimart-Rousseau et al., 2020). The model highlights that a strong wind event of 3 days has a significant impact on the seawater pCO_2 values during a longer period of ca. 15 days (Fig. 6). A combination of high atmospheric pCO2 value and high wind speed would then favor the sink for CO2 into the waters of the BoM. The aeration process depends also on the choice of the formulation of the gas transfer velocity (k600). In this study, the formulation of Wanninkhof (1992) is used and depends of the wind speed at 10 m above the water surface. However, the current velocity could favor the gas exchange and suspended matter concentration could limit the gas exchange (Abril et al., 2009; Upstill-Goddard, 2006; Zappa et al., 2003). Due to the important heterogeneity of physical and biogeochemical forcings in coastal zones, other factors that control the air-sea gas exchange should certainly be taken into account. The simulation with intrusions of the RhoneRhône River plume shows that inputs of nitrate and TA-cause a drop of seawater pCO₂ dueowing to some two concomitant effects: nutrients supply favors favoring the phytoplankton development and TA inputs(Fig. 7). In this scenario, the oceanic sink of atmospheric CO₂ is enhanced. But rivers also supply TA (e.g. Gemayel et al., 2015; Schneider et al., 2007) and DIC (e.g. Sempéré et al., 2000) that shift the carbonate system equilibrium leading totoward a pCO₂ decrease and a DIC increase (Middelburg, 2019) (Figs. 7 & 8). The consequence is that the oceanic sink of CO2 is enhanced. In fact, the experimental design on the Rhone River supply only considers the TA perturbation on the carbonate system but not that due to the DIC supply. To take into account the latter supply may further impact the DIC concentration when intrusions occur and it could sensibly modify the. Taking into account these further supplies may sensibly modify the modeled carbonate balance in the BoM. A next step to the present work will be to design more realistic numerical experiments to refine the results obtained in this preliminary study. The intrusions of RhoneRhône River plume also induce a salinity decrease of the BoM waters, which leads to drop the pCO2 levels in the model. This drop of pCO2 is due to the decrease in the CO2 solubility when salinity decreases (Middelburg, 2019). In the scenario of forcing the model by using urban atmospheric pCO2 time-series, the air-sea gradient increases and then, it enhances the status of the BoM as a sink for atmospheric CO2. As suggested by the in situ study of Wimart-Rousseau et al. (2020), the Eco3M-Carbox model highlights the crucial role of the coastal ocean in urbanized area, with an increase in atmospheric CO2, the CO2 uptake by the costal ocean may increase. This results is in line with

5. Conclusion

owing to high atmospheric CO2 levels.

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A marine carbonate chemistry module was implemented in the Eco3M-CarbOx biogeochemical model and evaluated against *in situ* data available in the Bay of Marseille (Northwestern Med. Sea) over the year 2017. The model correctly simulates the values ranges and seasonal dynamics of most of variables of the carbonate system except Total Alkalinityfor the total alkalinity. Several numerical experiments were also conducted to test the sensitivity of carbon balance to physical processes (temperature and salinity), biogeochemical processes (GPP and respiration processes) and external forcing (wind, river intrusion and atmospheric CO₂). This set of numerical experiments shows that the

studies of Andersson and Mackenzie (2004) and Cai (2011) that predict an increase in the intensity of CO2 sink in

coastal areas due to high atmospheric CO2 levels and a potential threat to coastal marine biodiversity- in coastal areas

Eco3M-CarbOx model provides expected responses in the alteration of the marine carbonate balance regarding each of the considered perturbation. On the whole, the model results suggest that the carbonate system is mainly driven by the seawater temperature dynamics. At a seasonal scale, the BoM marine waters appear to be a net sink of atmospheric CO2 and a dominant autotroph ecosystem during the MWC period, and a net source of CO2 to the atmosphere during the SWC period, which is mainly characterized by a dominance of heterotroph processes. However, the model results highlight that sharp seawater cooling observed within the SWC period and probably owing to upwelling events, eausescause the CO2 status of the BoM marine waters to change from a source to the atmosphere to a sink into the ocean within a few days. External forcing as the temperature increases leads to a delay in the bloom of phytoplankton. Strong wind events enhance the gas exchange of CO_2 with the atmosphere. A RhoneRhône River plume intrusion with input of nitrate and alkalinity favors pCO2 decreases, and the sink of atmospheric CO2 into the BoM waters is enhanced. The higher airatmospheric pCO2 values from the urban area intensify the oceanic sink of atmospheric CO2. The BoM biogeochemical functioning is mainly forced by wind-driven hydrodynamics (upwelling, downwelling), urban rivers, wastewater treatment plants, and atmospheric deposition (Fraysse et al., 2013). In addition, Northern Current and RhoneRhône River plume intrusions frequently occurred (Fraysse et al., 2014; Ross et al., 2016). Moreover, the BoM harbors the second bigger metropolis of France (Marseille) that is impacted by many harbor activities. The next step of this study will be to couple the Eco3M-CarbOx biogeochemical model to a 3D hydrodynamic model that will mirror the complexity of the BoM functioning. In this way, the contributions of hydrodynamic, atmospheric, anthropic, and biogeochemical processes to the DIC variability will be able to be determined with higher refinement and realism, and an overview of the air-sea CO2 exchange could be made at the scale of the Bay of Marseille. The main results of our study could be transposed to other coastal sites that are also impacted by urban and anthropic pressures. Moreover, in this paper we highlighted that fast and strong variations of

pCO₂ values occur, so thus it is essential to acquire more in situ values at high frequency (at least with an hourly

resolution) to understand the rapid variations of the marine carbon system at these short spatial and temporal scales.

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479 Code availability

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- 480 Eco3M is freely available under CeCILL license agreement (a French equivalent to the L-GPL license;
- 481 http://cecill.info/licences/Licence_CeCILL_V1.1-US.html; last access: 10 February 2020). The Eco3M-CarbOx
- 482 model is written in Fortran-90/95 and the plotting code is written in Matlab®. The exact version of the model used to
- 483 produce the results presented in this paper is archived on Zenodo (DOI: 10.5281/zenodo.3757677) (last access: 24
- 484 August 2020). A short User Manual is given in the Appendix C of this study.

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- 735

Table 1: Forcing of the different scenarios (S) simulated with the model. See section 2.4 for details of scenarios.

Atmospheric CO2

CAV station 2017

OHP station 2017

740

	Chl	seawater pCO ₂	pН	DIC	TA
Obs min-max	[0.10–1.71]	[358 – 471]	[8.014 - 8.114]	[2260 – 2348]	[2561 – 2624]
Mod min-max	[0.03 - 0.73]	[331 - 522]	[7.979 - 8.171]	[2220 - 2323]	[2560-2572]
Bias	-0.22	22.47	-0.016	-8.48	-24.91
WSS	0.36	0.69*	0.75*	0.71*	0.43
N	22	20	21	20	20

Table 2: Statistical evaluation of observations ps. model for 2017 year: observed and simulated minimum and maximum values, WSS = Wilmott Skill Score, N = number of measurements. Units of bias are those of modeled variables: chlorophyll \underline{a} (Chl, mg m⁻³), seawater pressure of CO₂ (seawater pCO₂, μ atm), pH, dissolved inorganic carbon (DIC, μ mol kg⁻¹) and total alkalinity (TA, μ mol kg⁻¹)- $\frac{1}{2}$ *significant value of WSS (> 0.70).

745

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742 743 744

		Aeration	GPP	$\mathbf{R}_{\mathbf{A}}$	$\mathbf{R}_{\mathbf{H}}$	R	NEP
	Year	0.017	-0.413	0.065	0.348	0.413	0
Mean flux	MWC	-0.245	-0.314	0.052	0.176	0.228	0.086
	SWC	0.405	-0.521	0.079	0.555	0.634	-0.113
Contribution	Year	78%	11%	2%	9%	11%	<u> </u>

Table 3: Mean flux values (mmol m^3 d^4) and the contribution of each process to the DIC variations for the reference simulation over the year and SWC/MWC periods. GPP: Gross primary production, R_A : Autotroph respiration, NEP: Net Ecosystem Production

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749 Figures

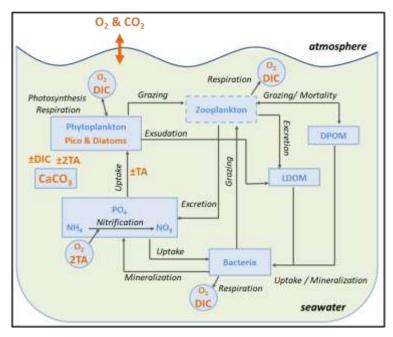


Figure 1: Schematic diagram of the biogeochemical model Eco3M-CarbOx. Explicit state variables of the model are indicated in continuous-line box or circles except the implicit variable for zooplankton (dotted line box). Orange-written state variables are added variables compared to the preexisting biogeochemical model of Fraysse et al. (2013). Arrows represent processes between two state variables. TA: Total Alkalinity. DIC: Dissolved Inorganic Carbon, CO2: carbon dioxide, O2: Oxygen, CaCO3: calcium carbonate.

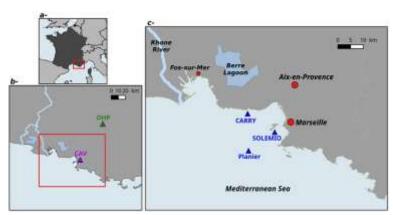


Figure 2: Map of study area: The Region Sud (A), Aix-Marseille Metropolis (B), the Bay of Marseille (C). CAV= Cinq AVenues Station (urban site), OHP: Observatoire de Haute Provence station (non-urban site), Carry, Solemio, Planier: Study Marine study sites at sea in the Bay of Marseille.

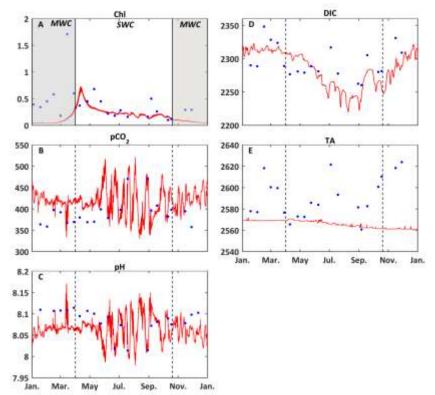


Figure 3: Comparison of model results (red) and in situ data (blue) at the surface of the SOLEMIO station. (A) Chlorophyll- ρ concentrations (mg m³), (B) ρ CO₂ (µatm), (C) ρ H, (D) DIC (µmol kg¹), (E) TA (µmol kg¹). The value of each state variable represents the mean around ±5 days of the sampling date, and the error bars are the standard deviation of values over this time period. The shaded area and dotted black line delimit the SWC and MWC periods.

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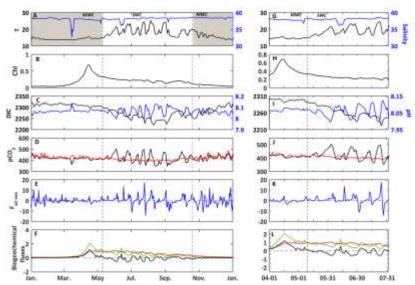


Figure 4: In the left panels: year 2017, right panels: temporal focus between April 1st and July 31th, 2017. In situ daily average of (A, G) temperature (°C, black line) and salinity (blue line) at the SOLEMIO station. (at the surface). Modeled daily average (B, H) chlorophyll \underline{a} concentrations (mg m³, black line) (C, I) DIC (µmol kg¹, black line) and $\underline{\mu}$ (blue line), (D, J) seawater $\underline{\mu}$ CO2 (µatm, black line) and atmosphere $\underline{\mu}$ CO2 from OHP (µatm, red line), (E, K) air-sea CO2 fluxes mmol m³ d¹), (F, L) Gross Primary Production (mmol m³ d¹, green line), total respiration (mmol m³ d¹, red line) and Net Ecosystem Production (mmol m³ d¹, black line). The shaded areas and dotted black lines delimit the SWC and MWC periods.

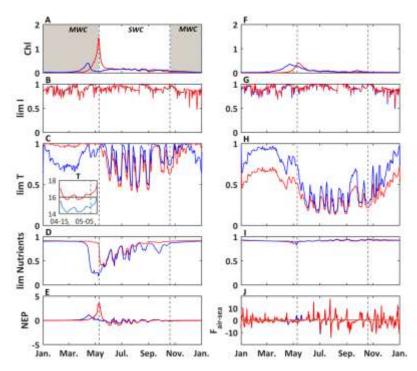


Figure 5: Modeled daily average chlorophyll \underline{a} concentrations (mg m⁻³) (A), light limitation (B), temperature limitation, and a zoom from April 15th to May 5th of temperature (C) and nutrient limitation (D) for picophytoplankton and the same set for diatoms (F, G, H and I). Modeled daily average NEP (mmol m⁻³ d⁻¹, E) and air-sea CO₂ flux (mmol m⁻³ d⁻¹, J). Reference simulation (S0, blue line) and temperature-shifted simulation by 1.5° C (S2, red line). The shaded area and dotted black lines delimit the SWC and MWC periods.

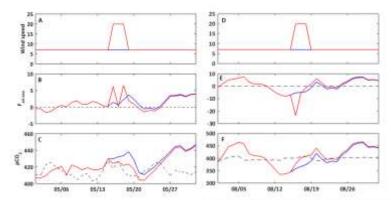


Figure 6: Temporal evolution for May (left panels) and August (right panels) 2017 of the wind speed (m s $^{-1}$, A, D); air-sea CO₂ fluxes (mmol m $^{-3}$ d $^{-1}$, B, E); seawater partial pressure of CO₂ (μ atm, C, F). Constant wind scenario (S2, blue line) and wind event scenario (S3, red line). On panels C and F, the dashed line represents the atmosphere partial pressure of CO₂ (μ atm) at the CAV station.

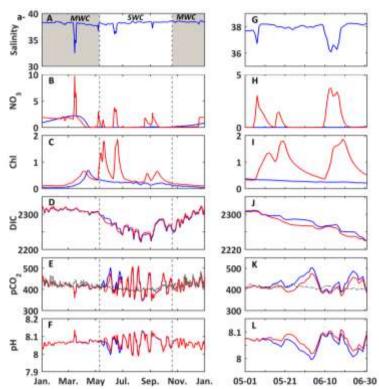
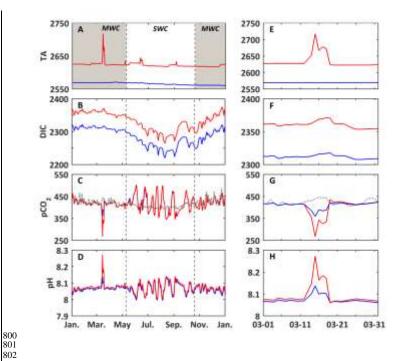


Figure 7: In the left panels: year 2017 and right panels: temporal focus between May 1st and July 1st, 2017. (A, G) *In situ* daily average of salinity. Modeled daily average (B, H) nitrate concentrations (mmol m⁻³); (C, I) chlorophyll \underline{a} concentrations (mg m⁻³); (D, J) DIC (μ mol kg⁻¹); (E, K) seawater ρ CO₂ (μ atm), and (F, L) ρ H. Reference simulation (S0, blue line) and nitrate supply simulation (S4, red line). On panels E and K, the dashed line represents the atmosphere partial pressure of CO₂ (μ atm) at the CAV station. The shaded area and dotted black lines delimit the SWC and MWC periods.



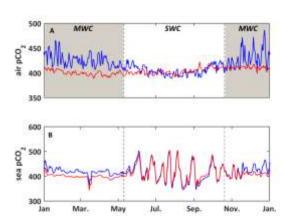


Figure 8: In the left for the whole year 2017 and in the right between March 1^{st} and April 1^{st} , 2017. Modelled daily average (A, E) TA (µmol kg $^{-t}$); (B, F) DIC (µmol kg $^{-t}$); (C, G) seawater ρCO_2 (µatm), and (D, H) ρH . Reference simulation (S0, blue line) and alkalinity supply simulation (S5, red line). On the panels C and G, the dashed line represents the atmosphere partial pressure of CO_2 (µatm) at the CAV station. The shaded area and dotted black lines delimit the SWC and MWC periods.

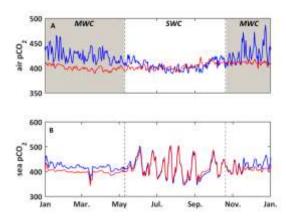


Figure 9:—(A) Temporal evolution for the year 2017 of the observed partial pressure of CO₂ (<u>matm</u>) in the atmosphere—in the the CAV station, called the "urban scenario" (S0, blue line), and at the OHP station, called the "non-urban scenario", and <u>in</u> seawater (S6, red line). (B)-Temporal evolution for the year 2017 of the modeled seawater partial pressure of CO₂ (<u>matm</u>) with forcings from the urban (S0, blue line) and non-urban (S6, red line) scenarios. The shaded area and dotted black lines delimit the SWC and MWC periods.

Appendix A: Details of resolution of carbonate system module

A.1. Calculation of carbonate systems constants:

• Concentration in Total Fluoride (TF) ionsconcentrations from Riley (1965) in mol kg⁻¹:

$$TF = \frac{0.000067}{18.998} \cdot \frac{S}{1.80655}$$

Concentration in Total Sulphate (TS) ionsconcentration from Morris and Riley (1966) in mol kg⁻¹:

$$TS = \frac{0.14}{96.062} \cdot \frac{S}{1.80655}$$

• Calcium ion concentration from Riley and Tongudai (1967) in mol kg-1:

$$Ca^{2+} = \frac{0.02128}{40.087} \cdot \frac{S}{1.80655}$$

• Concentration in Total Boron (TB) ionsconcentration from Uppström (1974) in mol kg⁻¹:

825
$$TB = \frac{0.000416 \cdot S}{35}$$

820

830

835

• Ionic Strength (IonS) from Millero (1982):

$$IonS = \frac{19.924 \cdot S}{1000 - 1.005 \cdot S}$$

The constants are calculated $\underline{\mathsf{inon}}$ the total pH scale except for K_S $\underline{\mathsf{inon}}$ free pH scale

If necessary, pH scale conversion factors are following:

From Seawater *pH* Scale (SWS) to total *pH* scale: SWStoTOT = $\frac{1 + \frac{TS}{K_S}}{1 + \frac{TS}{K_S} + \frac{TS}{K_F}}$

From Free pH Scale to Total pH scale: $FREE toTOT = 1 + \frac{TS}{K_S}$

• K_S equilibrium constant of dissociation of HSO_4 - from Dickson (1990a) in mol kg⁻¹:

$$K_S = \frac{-4276.1}{T_{(K)}} + 141.328 - 23.093 \cdot \log(T_{(K)}) + \left(324.57 - 47.986 \cdot \log(T_{(K)}) - \frac{13856}{T_{(K)}}\right) \cdot Ions^2$$

$$K_{S} = K_{S} + \left(-771.54 + 114.723 \cdot \log(T_{(K)}) + \frac{35474}{T_{(K)}}\right) \cdot lons + \frac{-2698}{T_{(K)}} \cdot \frac{lons^{\frac{1.5}{2}} + \frac{1776}{T_{(K)}}}{lons^{\frac{3}{2}}} + \frac{1776}{T_{(K)}} \cdot lons^{2}$$

$$K_S = e^{K_S} \cdot (1 - 0.001005 \cdot S)$$

K_F equilibrium constant of dissociation of <u>hydrogen</u> fluoride-ions (HF) formation from Dickson and Riley (1979) in mol kg
 1.

840
$$K_F = e^{\frac{1590.2}{T(\kappa)} - 12.641 + 1.525 \cdot lons^{\frac{1}{5}}} \cdot (1 - 0.001005 \cdot S) K_F = e^{\frac{1590.2}{T(\kappa)} - 12.641 + 1.525 \cdot lons^{\frac{1}{2}}} \cdot (1 - 0.001005 \cdot S)$$

 \bullet K_B equilibrium constant of dissociation of boric acid from Dickson (1990b) in mol kg⁻¹

$$K_B = (-8966.9 - 2890.53 \cdot S^{\frac{1}{2}} - 77.942 \cdot S + 1.728 \cdot S^{\frac{3}{2}} - 0.0996 \cdot S^2) / T_{(K)}$$

$$K_B = K_B + 148.0248 + 137.1942 \cdot S^{\frac{1}{2}} + 1.62142 \cdot S + (-24.4344 - 25.085 \cdot S^{\frac{1}{2}} - 0.2474 \cdot S) \cdot \log(T) + 0.053105 \cdot S^{\frac{1}{2}} \cdot T$$

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29

$$K_0 = \exp\left(-60.2409 + 93.4517 \cdot \frac{100}{T_{(K)}} + 23.3585 \cdot \log\left(\frac{T_{(K)}}{100}\right) + S \cdot \left(0.023517 - 0.023656 \cdot \frac{T_{(K)}}{100} + 0.0047036 \cdot \left(\frac{T_{(K)}}{100}\right)^2\right)\right)$$

• K_e : Dissociation constant of water from Millero (1995) in (mol kg⁻¹)²:

$$K_e = \exp(\frac{-13847.26}{T_{(K)}} + 148.9802 - 23.6521 \cdot \log(T_{(K)}) + \left(-5.977 + \frac{118.67}{T_{(K)}} + 1.0495 \cdot \log(T_{(K)})\right) \cdot \frac{5^{\frac{1}{2}}}{S^{\frac{1}{2}}} \cdot S^{\frac{1}{2}} - 0.01615 \cdot S)$$

850 $K_e = K_e \cdot SWStoTOT$, inon total pH scale in mol kg⁻¹

• K_1 and K_2 from Lueker et al. (2000) in mol kg⁻¹:

$$\begin{split} K_1 &= 10^{\frac{-3633.86}{T(K)} + 61.2172 - 9.6777 \cdot \log\left(T_{(K)}\right) + 0.011555 \cdot S - 0.0001152 \cdot S^2} \\ K_2 &= 10^{\frac{-471.78}{T(K)} + 251.929 - 3.16967 \cdot \log\left(T_{(K)}\right) + 0.01781 \cdot S - 0.0001122 \cdot S^2} \end{split}$$

• K_{ca} for calcite from Mucci (1983) in (mol kg⁻¹)²:

855 K_{ca}

$$=\frac{171.9065-0.077993 \cdot T_{(K)}}{10} + \frac{2839.319}{T_{(K)}} + 71.595 \cdot log 10 \left(T_{(K)}\right) + \left(-0.77712+0.0028426 \cdot T_{(K)}\right) + \frac{178.34}{T_{(K)}} s^{\frac{1}{2}} - 0.07711 \cdot S + 0.0041249 \cdot S^{\frac{1.5}{2}}}{10} - 171.9065 - 0.077993 \cdot T_{(K)} + \frac{2839.319}{T_{(K)}} + 71.595 \cdot log 10 \left(T_{(K)}\right) + \left(-0.77712+0.0028426 \cdot T_{(K)}\right) + \frac{178.34}{T_{(K)}} s^{\frac{1}{2}} + 20.07711 \cdot S + 0.0041249 \cdot S^{\frac{1.5}{2}} + 20.07711 \cdot S + 0.0041249 \cdot S + 0$$

• Every constant All the constants are corrected by the effect of hydrostatic pressure:

R = 83.1451 in ml bar-1 K-1 mol-1

$$lnK_{1}fac = \frac{\left(25.5 - 0.1271 \cdot T_{(^{\circ}C)} + 0.5 \cdot \left(\frac{-3.08 + 0.0877 \cdot T_{(^{\circ}C)}}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R^{*}T_{(K)}}; K_{1} = K_{1} \cdot e^{lnK_{1}fac}$$

$$lnK_{2}fac = \frac{\left(15.82 - 0.0219 \cdot T_{(^{\circ}C)} + 0.5 \cdot \left(\frac{1.13 + 0.1475 \cdot T_{(^{\circ}C)}}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}} : K_{2} = K_{2} \cdot e^{lnK_{2}fac}$$

$$860 \quad lnK_2 fac = \frac{\left(\frac{15.82 - 0.0219 \cdot T_{(^{\circ}C)} + 0.5 \cdot \left(\frac{1.13 + 0.1475 \cdot T_{(^{\circ}C)}}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}} \cdot K_2 = K_2 \cdot e^{lnK_2 fac}$$

$$lnK_{B}fac = \frac{\left(\frac{29.48 - 0.1622 \cdot T_{(^{\circ}C)} + 0.002608 \cdot T_{(^{\circ}C)}^{2} + 0.5 \cdot \left(\frac{-2.84}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}}; K_{B} = K_{B} \cdot e^{lnK_{B}fac}$$

$$lnK_{e}fac = \frac{\left(\frac{20.02 - 0.1119 \cdot T_{(^{\circ}C)} + 0.001409 \cdot T_{(^{\circ}C)}^{2} + 0.5 \cdot \left(\frac{-5.13 + 0.0794 \cdot T_{(^{\circ}C)}}{1000}\right) P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}}; K_{e} = K_{e} \cdot e^{lnK_{e}fac}$$

$$lnK_{F}fac = \frac{\left(\frac{9.78 - 0.009 \cdot T_{(^{\circ}C)} + 0.0009429 \cdot T_{(^{\circ}C)}^{\circ}^{2} + 0.5 \cdot \left(\frac{-3.91 + 0.054 \cdot T_{(^{\circ}C)}}{10000}\right) \cdot p_{bar}\right) \cdot p_{bar}}{R \cdot T_{(K)}}; K_{F} = K_{F} \cdot e^{lnK_{F}fac}$$

$$lnK_{S}fac = \frac{\left(18.03 - 0.0466 \cdot T_{(^{\circ}C)} + 0.000316 \cdot T_{(^{\circ}C)}^{2} + 0.5 \cdot \left(\frac{-4.53 + 0.009 \cdot T_{(^{\circ}C)}}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}}; K_{S} = K_{S} \cdot e^{lnK_{S}fac}$$

$$lnK_{ca}fac = \frac{\left(48.76 - 0.5304 \cdot T_{(^{\circ}C)} + 0.5\left(\frac{-11.76 + 0.3692 \cdot T_{(^{\circ}C)}}{1000}\right) \cdot P_{bar}\right) \cdot P_{bar}}{R \cdot T_{(K)}}; K_{ca} = K_{ca} \cdot e^{lnK_{ca}fac}$$

• Calculation of the Fugacity factor:

We suppose that the pressure is at one atmosphere or close to it (Weiss, 1974):

$$P_{atm} = 1.01325 \ bar$$

$$delta = 57.7 - 0.118 \cdot T \text{ in cm}^3 \text{ mol}^{-1}$$

870
$$b = -1636.75 + 12.0408 \cdot T - 0.0327957 \cdot T^2 + 3.16528 \cdot 0.00001 \cdot T^3 \text{ in cm}^3 \text{ mol}^{-1}$$

$$FugFac = exp^{\frac{(b+2\cdot delta)\cdot P_{atm}}{R\cdot T}}$$

A.2. Resolution of carbonate system

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To resolve the carbonate system, we calculate the deltapH, which is the difference of pH between two iterations of the model. We initialize the run by imposing a pH value of 8.

875 if (nbiter < 1) pH = 8pHtol = 0.001! tolerance for iterations end deltapH = pHtol + 1hile (abs(deltapH) > 0.0001) $H = 10^{-pH}$ $Denom = H^2 + K_1 \cdot H + K_1 \cdot K_2$ $CAlk = DIC \cdot K_1 \cdot \left(\frac{H + 2 \cdot K_2}{Denom}\right)$ $BAlk = \frac{TB \cdot K_B}{K_B + H}$ $OH = \frac{K_e}{H}$ $FreetoTot = 1 + \frac{TS}{K_S}$ $Hfree = \frac{H}{FreetoTot}$ $HSO_4 = \frac{TS}{1 + \frac{K_S}{Hfree}}$ $HF = \frac{TF}{1 + \frac{K_F}{Hfree}}$ $Residual = TA - CAlk - BAlk - OH + Hfree + HSO_4 + HF$ $Slope = DIC \cdot H \cdot K_1 \cdot (H^2 + K_1 \cdot K_2 + 4 \cdot H \cdot K_2)$ $Slope - \frac{Slope}{1 + OH + H} + \frac{BAlk \cdot H}{1 + BAlk \cdot H}$ $do\ while\ (abs(deltap H)>0.0001)$ 880 885 890

$$Slope = \frac{Slope}{Denom^2} + OH + H + \frac{BAlk \cdot H}{K_B + H}$$

 $Slope = log10 \cdot Slope$

deltapH = Residual/Slope ! this is Newton's method

do while (abs(deltapH) > 1) deltapH = $\frac{deltapH}{2}$! to keep the jump from being too big

895 enddo

pH = pH + deltapH! Is on the same scale as K_1 and K_2 were calculated, i.e. total pH scale

$$pCO_2 = \left(\frac{DIC \cdot H^2}{H^2 + K_1 \cdot H + K_1 \cdot K_2}\right) \cdot \frac{10^6}{K_0 \cdot FugFac} ! \text{ in } \mu\text{atm}$$

$$CO_2 = \frac{1 + \frac{K_1}{H} + \frac{K_1 \cdot K_2}{H^2}}{1 + \frac{K_1}{H} + \frac{K_2}{H^2}}$$

$$HCO_3 = \frac{K_1 \cdot CO_2}{H}$$

$$HCO_{3} = \frac{K_{1} \cdot CO_{2}}{H}$$

$$900 \quad CO_{3} = \frac{K_{2} \cdot HCO_{3}}{H}$$

$$Omega = \frac{Ca \cdot CO_{3} \cdot 10^{-6}}{K_{ca}}$$

$$Omega = \frac{Ca \cdot CO_3 \cdot 10^{-6}}{K_{ca}}$$

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Appendix B: Biogeochemical model variables and parameters

 $Table\ B1:\ Initial\ conditions\ of\ the\ state\ variables\ of\ Eco3M-CarbOx\ model\ (*diagnostic\ variables)$

Variables	Name	Unit	values
	PicoC	mmolC m ⁻³	0.0480
Picophytoplankton	PicoN	mmolN m ⁻³	0.0092
	PicoP	mmolP m ⁻³	0.0003
	DiaC	mmolC m ⁻³	0.0571
Diatom	DiaN	mmolN m ⁻³	0.0089
	DiaP	mmolP m ⁻³	0.0007
	BacC	mmolC m ⁻³	0.1083
Bacteria	BacN	mmolN m ⁻³	0.0379
	ВасР	mmolP m ⁻³	0.0039
DPOM	DPOC	mmolC m ⁻³	0.1252
Detrital Particulate organic matter	DPON	mmolN m ⁻³	0.0307
Detritar Farticulate organic matter	DPOP	mmolP m ⁻³	0.0021
LDOM	LDOC	mmolC m ⁻³	1.0990
Labile Dissolved organic matter	LDON	mmolN m ⁻³	8.7980
Labile Dissolved organic matter	LDOP	mmolP m ⁻³	0.0018
	$\mathrm{NH_4}$	mmolN m ⁻³	0.3375
	NO_3	mmolN m ⁻³	0.6723
DIM Dissolved inorganic matter	PO_4	mmolP m ⁻³	0.7150
Ü	DO	mmolO m ⁻³	257.00
	DIC	μmolC kg ⁻¹	2358.4
Total alkalinity	TA	μmolC kg ⁻¹	2660.5
Sea water partial pressure of CO ₂	pCO_2	μatm	371.28
$p\mathbf{H}$	pH	<u></u>	8.1099
calcium carbonate	CaCO ₃	mmol m ⁻³	1.0000
Picophytoplankton chlorophyll*	PicoChl	mgChl m ⁻³	0.0193
Diatom chlorophyll*	DiaChl	mgChl m ⁻³	0.0229
Number of bacteria*	NBA	10 ¹² cell m ⁻³	0.2000

Table B2: Balance equations of Eco3M-CarbOx model

Variables	Balance equation	
Pico- phytoplankton	$ \begin{split} \frac{\partial PicoC}{\partial t} &= R_{PP}^{PicoC} - R_{resp}^{PicoC} - R_{Gr}^{PicoC} - R_{Gr} \\ \frac{\partial PicoN}{\partial t} &= R_{uptPicoN}^{NH_4} + R_{uptPicoN}^{NO_3} - R_{exu}^{PicoN} - R_{Gr} \\ \frac{\partial FicoP}{\partial t} &= R_{uptPicoP}^{PicOP} - R_{exu}^{PicoP} - R_{Gr} \end{split} $	
Diatom	$\begin{aligned} \operatorname{PicoChl} &= Q_{C}^{N} \cdot \left(Q_{N,\min}^{\operatorname{Chla}} + f_{Q} \cdot \left(Q_{N,\min}^{\operatorname{Chla}} - Q_{N,\min}^{\operatorname{Chla}}\right)\right) \cdot \operatorname{PicoC} \\ & \frac{\partial \operatorname{DiaC}}{\partial t} = R_{PP}^{\operatorname{DiaC}} - R_{resp}^{\operatorname{DiaC}} - R_{Gr}^{\operatorname{DiaC}} - R_{Gr} \\ & \frac{\partial \operatorname{DiaN}}{\partial t} = R_{uptDiaN}^{\operatorname{H}_{4}} + R_{uptDiaN}^{\operatorname{Hos}_{2}} - R_{exu}^{\operatorname{DiaN}} - R_{Gr} \\ & \frac{\partial \operatorname{DiaP}}{\partial t} = R_{uptDiaP}^{\operatorname{PO_{4}}} - R_{exu}^{\operatorname{DiaP}} - R_{Gr} \end{aligned}$	\ \ \ \
Bacteria	$\begin{split} DiaChl &= Q_{N}^{C} \cdot \left(Q_{N,min}^{Chla} + f_{Q} \cdot \left(Q_{N,max}^{Chla} - Q_{N,min}^{Chla}\right)\right) \cdot DiaC \\ \\ \frac{\partial BacC}{\partial t} &= R_{uptBac}^{DPOC} + R_{uptBac}^{LDOC} - R_{BR} - R_{Gr}^{BacC} \\ \\ \frac{\partial BacN}{\partial t} &= R_{uptBac}^{DPON} + R_{uptBac}^{LDON} + R_{uptBac}^{NH4} - R_{Gr}^{BacN} \\ \\ \frac{\partial BacP}{\partial t} &= R_{uptBac}^{DPOP} + R_{uptBac}^{LDOP} + R_{uptBac}^{PO4} - R_{Gr}^{BacP} \\ \\ \\ \frac{\partial BacP}{\partial t} &= R_{uptBac}^{DPOP} + R_{uptBac}^{LDOP} + R_{uptBac}^{PO4} - R_{Gr}^{BacP} \\ \end{split}$	ر ال ر
рром	$ \frac{\partial DPOC}{\partial t} = R_{pf} + R_m - R_{Gr} - R_{uptBac}^{DPOC} $ $ \frac{\partial DPON}{\partial t} = R_{pf} + R_m - R_{Gr} - R_{uptBac}^{DPON} $ $ \frac{\partial DPOP}{\partial t} = R_{pf} + R_m - R_{Gr} - R_{uptBac}^{DPON} $	_ _} _}
LDOM	$\begin{split} \frac{\partial LDOC}{\partial t} &= R_{exu}^{picoC} + R_{exu}^{DiaC} + R_{exu}^{LDOC} - R_{uptBac}^{LDOC} \\ \frac{\partial LDON}{\partial t} &= R_{exu}^{picoP} + R_{exu}^{DiaN} + R_{exu}^{LDON} - R_{uptBac}^{LDON} \\ \frac{\partial LDOP}{\partial t} &= R_{exu}^{picoP} + R_{exu}^{DiaP} + R_{exu}^{LDOP} - R_{uptBac}^{LDOP} \end{split}$	_ ⊃
NH4	$\frac{\partial NH_4}{\partial t} = R_{excr}^{NH_4} + R_{miner}^{NH_4} - R_{nit} - \sum R_{uptPhyN}^{NH_4} - R_{uptBac}^{NH_4}$	
NO ₃	$\frac{\partial NO_3}{\partial t} = R_{nit} - \sum_{i} R_{uptPhyN}^{NO_3}$	_
PO ₄	$\frac{\partial PO_4}{\partial t} = R_{excr}^{PO_4} + R_{miner}^{PO_4} - \sum_{uptPhyP} R_{uptPhyP}^{PO_4} - R_{uptBac}^{PO_4}$	
ро	$\frac{\partial DO}{\partial t} = R_{aera} + \left(\frac{O}{C}\right) \cdot R_{pp}^{PhyC} + \left(\frac{O}{N}\right) \cdot R_{uptPhyN}^{NO_3} - \left(\frac{O}{C}\right) \cdot R_{resp}^{PhyC} - \left(\frac{O}{C}\right) \cdot R_{excr}^{DIC} - \left(\frac{O}{C}\right) \cdot R_{BR} - \left(\frac{O}{N}\right) \cdot R_{nit}$	7
DIC	$\frac{\partial DIC}{\partial t} = R_{aera} + R_{resp}^{PhyC} + R_{BR} + R_{excr}^{DIC} - R_{pp}^{PhyC} - R_{precip} + R_{diss}$	` _
TA	$\frac{\partial TA}{\Delta \partial t} = 2 \cdot R_{diss} + \left(R_{uptPhyN}^{NO_3} + R_{uptPhyP}^{PO_4} - R_{uptPhyN}^{NH_4} \right) + R_{miner}^{NH_4} - 2 \cdot R_{precip} - 2 \cdot R_{nit}$	/ F

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 $\,$ Table B3: Biogeochemical processes simulated by the Eco3M-CarbOx model

Notation	Biogeochemical processes	Unit	Formulation	-
R_{pp}^{Phy}	Primary production	molC m ⁻³ s ⁻¹	$R_{PP}^{PhyC} = P_{max} \cdot f_T^{PP} \cdot f_I \cdot PhyC$	$\begin{split} f_Q &= min[f_Q^N, f_Q^P]; f_Q^X = \frac{Q_C^X - Q_{C,min}^X}{Q_C^X - Q_{C,min}^X + \beta_X} \\ f_T^{PP} &= \\ max \left(\frac{\frac{2 \cdot (1-b) - \frac{(T-T_{let})}{(T_{opt} - T_{let})}}{\left(\frac{(T-T_{let})}{(T_{opt} - T_{let})}\right)^2 - 2 \cdot b \cdot \frac{(T-T_{let})}{(T_{opt} - T_{let})} + 1}}; 0 \right) \\ f_I &= \left[1 - exp \left(\frac{-\alpha_{Chia} \cdot E_{PAR} \cdot Q_C^{Chia}}{p_{max} \cdot f_Q \cdot f_T^{PP}} \right) \right] \end{split}$
R_{resp}^{Phy}	Phytoplankton respiration	$molC \ m^{3} \ s^{1}$	$R_{resp}^{PhyC} = k_r^{PhyC} \cdot PhyC$	
$R_{uptPhy}^{NH_4}$	NH ₄ uptake by phytoplankton		$R_{uptPhyN}^{NH_4} = V_{N,max} \cdot \frac{NH_4}{NH_4 + K_{NH_4}}$	$V_{N,max} = Q_{C,max}^{N} \cdot R_{PP}^{Phy}$
$R_{uptPhy}^{NO_3}$	NO ₃ uptake by phytoplankton	molN m ⁻³ s ⁻¹	$R_{uptPhyN}^{NO_3} = V_{N,max} \cdot \frac{NO_3}{NO_3 + K_{NO_3}} \cdot \left(1 - \frac{I_{in}.NH_4}{NH_4 + K_{in}}\right)$	
$R_{uptPhy}^{PO_4}$	PO ₄ uptake by phytoplankton	$molP\ m^{-3}\ s^{-1}$	$R_{uptPhyP}^{PO_4} = V_{P,max} \cdot \frac{PO_4}{PO_4 + K_{PO_4}}.$	$V_{P,max} = Q_{C,max}^P \cdot R_{PP}^{Phy}$
R_{exu}^{PhyC}	Phytoplankton exudation as LDOC	molC m ⁻³ s ⁻¹	$R_{exu}^{PhyC} = \left(1 - f_Q\right) \cdot R_{PP}^{Phy}$	
R_{exr}^{PhyX}	Phytoplankton exudation as LDON or LDOP	$molX \; m^{-3} \; s^{-1}$	$R_{exu}^{PhyX} = \left(1 - h_Q^X\right) \cdot R_{uptX}^{Phy}$	$h_Q^X = \frac{Q_{C,max}^X - Q_C^X}{Q_{C,max}^X - Q_{C,min}^X}$
R_{BP}	Bacterial production	cell m ⁻³ s ⁻¹	$R_{BP} = \mu_{max}^{Ba} \cdot f_Q^{Ba} \cdot f_T^{Ba} \cdot NBA$	$\begin{split} f_T^{Ba} &= Q_{10}^{\frac{(T-T_{rem})}{10}}; \\ f_Q^{BA} &= min\left[1 - \frac{Q_{C,min}^{BA}}{Q_C^{BA}}, 1 - \frac{Q_{N,min}^{BA}}{Q_N^{BA}}, 1 - \frac{Q_{N,min}^{BA}}{Q_N^{BA}}\right] \end{split}$
R_{BR}	Bacterial respiration	molC m ⁻³ s ⁻¹	$\begin{split} R_{BR} &= \rho_g^{Ba} \cdot Q_{\overline{C}}^{Ba} \cdot R_{BF} \cdot Q_C^{Ba} \cdot R_{BP} + \\ \rho_r^{Ba} \cdot \left(Q_{\overline{C}}^{Ba} - Q_{\overline{C},min}^{Ba}\right) \cdot \left(Q_C^{Ba} - Q_{C,min}^{Ba}\right) \cdot NBA \end{split}$	
R_{uptBac}^{X}	X uptake by bacteria	molX m ⁻³ s ⁻¹	$R_{uptBac}^{X} = V_{max}^{BA} \cdot \frac{x}{x + \kappa_{X}^{Ba}} \cdot f_{T}^{Ba} \cdot NBA$	
R_{Gr}^{Phy}	Phytoplankton grazing by zooplankton	molX m ⁻³ s ⁻¹	$R_{Gr}^{Phy} = g_{Phy} \cdot f_{Gr} \cdot Phy$	$f_{Gr} = \frac{_{Phy}}{_{Phy+DPOM}}$
R_{Gr}^{DPOM}	DPOM grazing by zooplankton	molX m ⁻³ s ⁻¹	$R_{Gr}^{DPOM} = g_{DPOM} \cdot f_{Gr} \cdot DPOM$	$g_{DPOM} = \frac{g_{Pico} \cdot Pico + g_{Dia} \cdot Dia}{Pico + Dia};$ $f_{Gr} = \frac{DPOM}{Phy + DPOM}$

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Table B4: Value of parameters

Parameters		Pico	Dia	Unit	Reference
P_m^C	Maximal production	1.815	1.057	d-1	Sarthou et al. (2005)
m_1	Fraction of the solar energy flux photosynthetically available	0.43	0.43	4	Tett (1987)
m_2	Sea surface reflection	0.95	0.95	/=	Tett (1987)
m_3	More rapid attenuation of polychromatic light near the sea surface	1.0	1.0	4	Tett (1987)
$lpha_{Chla}$	Chlorophyll-specific light absorption coefficient	8 10-6	5 10-6	$m^2molC(gChlaJ)^{\text{-}1}$	Leblanc et al. (2018)
T_{opt}	Temperature optimal of growth	16.0	13.0	°C	<u> </u>
T_{let}	Lethal temperature	11.0	9.0	°C	<u> </u>
b	Shape factor for temperature curve	0.5	0.8	£	Lacroix and Grégoire (2002)
$\boldsymbol{\beta}_N$	Coefficient in the quota function	0.0072	0.002	molN molC-1	Leblanc et al. (2018)
β_P	Coefficient in the quota function	0.0002	0.0005	molP molC-1	Leblanc et al. (2018)
$Q_{C,min}^N$	Minimum phytoplankton N:C ratio	0.115	0.07	molN molC-1	Leblanc et al. (2018)
$Q_{C,max}^N$	Maximum phytoplankton N:C ratio	0.229	0.18	molN molC-1	Leblanc et al. (2018)
$Q_{C,min}^{P}$	Minimum phytoplankton P:C ratio	0.0015	0.006	molP molC-1	Auger et al. (2011); Campbell et al. (2013)
$Q_{C,max}^{P}$	Maximum phytoplankton P:C ratio	0.0068	0.016	molP molC-1	Auger et al. (2011); Campbell et al. (2013)
$Q_{N,min}^{Chla}$	Minimum phytoplankton Chl:N ratio	1.0	1.0	gChl molN-1	Leblanc et al. (2018)**
$Q_{N,max}^{Chla}$	Maximum phytoplankton Chl:N ratio	2.2	2.7	gChl molN ⁻¹	Leblanc et al. (2018)
k_r^{PhyC}	Phytoplankton respiration rate	0.099	0.099	d^{-1}	Faure et al. (2010)
K_{NO_3}	Half saturation constant for NO ₃	0.73	1.0	mmolN m ⁻³	Leblanc et al. (2018)
K_{NH_4}	Half saturation constant for NH ₄	0.07	0.015	mmolN m ⁻³	Leblanc et al. (2018)
K_{PO_4}	Half saturation constant for PO ₄	0.008	0.01	mmolP m ⁻³	Leblanc et al. (2018)**
I_{in}	Factor of inhibition	0.82	0.82	4	Harrison et al. (1996)
K_{in}	Amount of NH ₄ from which assimilation by NO ₃ is reduced.	0.578	0.578	mmolN m ⁻³	Harrison et al. (1996)
g	Grazing rate	1.452	0.846	d-1	Gutiérrez-Rodríguez et al. (2011)

^{**} calibrated from parameter used in the cited article

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 $915 \quad \textbf{Table B5: Value of parameters (continue)} \\$

$V_{LDON,max}^{BA}$ Maximum LDON uptake by bacteria 0.32 mmolN (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010) $V_{NH_4,max}^{BA}$ Maximum NH ₄ uptake by bacteria 0.32 mmolN (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010)	Parameters		Value	Unit	Reference
QPA_{Cmin}^{R} Minimum bacteria Ccell ratio 0.49 mmolC (10¹² cell)¹ Fukuda et al. (1998) QPA_min Minimum bacteria N:cell ratio 0.09 mmolN (10¹² cell)¹ Fukuda et al. (1998) QPA_min Minimum bacteria N:cell ratio 0.005 mmolN (10¹² cell)¹ Fukuda et al. (1998) QPA_min Maximum bacteria P:cell ratio 0.005 mmolP (10¹² cell)¹ Fraysse et al. (2013) QPA_min Maximum bacteria P:cell ratio 0.02 mmolP (10¹² cell)¹ Fraysse et al. (2013) QPA_min Maximum bacteria P:cell ratio 0.02 mmolP (10¹² cell)¹ d¹ Fraysse et al. (2013) QPA_min Maximum DPO Maximum DPO mmolP (10¹² cell)¹ d¹ Thingstad (1987) VPA_DEOLAM Maximum DPO uptake by bacteria 0.02 mmolN (10¹² cell)¹ d¹ Faure et al. (2013) VPB_DEOLAM Maximum DPO uptake by bacteria 0.32 mmolN (10¹² cell)¹ d¹ Faure et al. (2010) VPB_DEOLAM Maximum DPO uptake by bacteria 0.48 mmolN (10¹² cell)¹ d¹ Faure et al. (2010) VPB_DEOLAM Maximum LDO Duptake by bacteria	NBA	Number of bacteria	0.20	$10^{12}\mathrm{cell}\;\mathrm{m}^{-3}$	Moran (2015)
QBA Nmin Maximum Minimum bacteria N:cell ratio 0.09 mmolN (10 ¹² cell) ⁻¹ Fukuda et al. (1998) QBA Nmax Pamax Maximum bacteria N:cell ratio 0.23 mmolN (10 ¹² cell) ⁻¹ Fukuda et al. (1998) QBA Pamax Maximum bacteria P:cell ratio 0.005 mmolP (10 ¹² cell) ⁻¹ Fraysse et al. (2013) PBA PBA PBO POC.max Respiration rate of bacteria 0.01 d-1 Thingstad (1987) VPADOC.max PBOOR.max Maximum DPOC uptake by bacteria 0.029 cell) ⁻¹ d ⁻¹ Campbell et al. (2013) VPBOOR.max PBOOR.max Maximum LDOC uptake by bacteria 0.05 mmolN (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010) VBADOP.max PBOOP.max Maximum DPOD uptake by bacteria 0.32 mmolN (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010) VBADOP.max PBOOP.max Maximum DPOD uptake by bacteria 0.01 mmolP (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010) WBADOP.max PBOOP.max Maximum DPOD uptake by bacteria 0.01 mmolP (10 ¹² cell) ⁻¹ d ⁻¹ Faure et al. (2010) Maximum DPOP uptake by bacteria 0.01 mmolP (10 ¹² cell) ⁻¹ d ⁻¹ Thingstad (1987) WBADOP.max PBOOP.max	μ_{max}^{Ba}	Bacterial production rate	8.36	d^{-1}	Fraysse et al. (2013)
OR™ max (PR min) Maximum bacteria P:cell ratio 0.23 mmolN (10¹² cell)¹¹ Fukuda et al. (1998) OR™ min (Pr min) Minimum bacteria P:cell ratio 0.005 mmolP (10¹² cell)¹¹ Fraysse et al. (2013) OR™ max (PR min) Maximum bacteria P:cell ratio 0.02 mmolP (10¹² cell)¹¹ Fraysse et al. (2013) OR max (PR min) Maximum Deoc to carbon respired by bacteria 0.00 d³¹ Thingstad (1987) VBADOLOMAN (PBPON max) Maximum DPOC uptake by bacteria 0.029 cell)¹ d³¹ Campbell et al. (2013) VBADOLOMAN (PBPON max) Maximum DPON uptake by bacteria 0.05 mmolN (10¹² cell)¹ d³¹ Faure et al. (2010) VBADOLOMAN (PBPON max) Maximum DPON uptake by bacteria 0.05 mmolN (10¹² cell)¹ d³¹ Faure et al. (2010) VBADOLOMAN (PBPON max) Maximum DPOP uptake by bacteria 0.03 mmolN (10¹² cell)¹ d³¹ Faure et al. (2010) VBADOLOMAN (PBPON max) Maximum DPOP uptake by bacteria 0.01 mmolN (10¹² cell)¹ d³¹ Faure et al. (2010) VBADOLOMAN (PBPON max) Maximum DPOP uptake by bacteria 0.48 mmolP (10¹² cell)¹ d³¹ Faure et al. (2010) VBADOLOMAN (PBPON max) Maximum DPOP uptake by bacteria 0.48 mmolP (10¹² cell)¹ d³¹ Thingstad (1987) <tr< td=""><td>$Q_{C,min}^{BA}$</td><td>Minimum bacteria C:cell ratio</td><td>0.49</td><td>mmolC (1012 cell)-1</td><td>Fukuda et al. (1998)</td></tr<>	$Q_{C,min}^{BA}$	Minimum bacteria C:cell ratio	0.49	mmolC (1012 cell)-1	Fukuda et al. (1998)
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$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$Q_{P,min}^{BA}$	Minimum bacteria P:cell ratio	0.005	mmolP (1012 cell)-1	Fraysse et al. (2013)
ρ ^{Ba} P ^{Ba} Respiration rate of bacteria 0.01 d³ mmolC (46²cell¹0¹² call)¹¹ d¹ campbell et al. (2013) Thingstad (1987) V ^{BA} P ^{BOD, max} Maximum LDOC uptake by bacteria 16.33 mmolC (10¹² cell)¹¹ d¹¹ campbell et al. (2013) Campbell et al. (2013) V ^{BA} P ^{BON, max} Maximum LDON uptake by bacteria 0.05 mmolN (10¹² cell)¹¹ d¹¹ Faure et al. (2010) Faure et al. (2010) V ^{BA} P ^{BON, max} Maximum LDON uptake by bacteria 0.32 mmolN (10¹² cell)¹¹ d¹¹ Faure et al. (2010) Faure et al. (2010) V ^{BA} P ^{BON, max} Maximum DPOP uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Faure et al. (2010) V ^{BA} P ^{BON, max} Maximum LDOP uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Thingstad (1987) V ^{BA} P ^{BON, max} Maximum PO4 uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Thingstad (1987) V ^{BA} P ^{BON, max} Maximum PO4 uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Thingstad (1987) V ^{BA} P ^{BON, max} Maximum PO4 uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Thingstad (1987) V ^{BA} P ^{BON, max} Maximum PO4 uptake by bacteria 0.48 mmolP (10¹² cell)¹ d¹¹ Thingstad (1987) Thingstad (1987) V ^{BA} P ^{BON} M ^{BON}	$Q_{P,max}^{BA}$	Maximum bacteria P:cell ratio	0.02	mmolP (1012 cell)-1	Fraysse et al. (2013)
$V_{DPOC,max}^{R}$ Maximum DPOC uptake by bacteria 0.029 mmolC $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Campbell et al. (2013) $V_{BPOC,max}^{R}$ Maximum LDOC uptake by bacteria 16.33 mmolC $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Campbell et al. (2013) $V_{BPON,max}^{R}$ Maximum DPON uptake by bacteria 0.05 mmolN $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Faure et al. (2010) $V_{BPON,max}^{R}$ Maximum LDON uptake by bacteria 0.32 mmolN $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Faure et al. (2010) $V_{BPOP,max}^{R}$ Maximum DPOP uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum DPOP uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum PO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum PO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum PO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum DPO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum DPO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) $V_{BO,max}^{R}$ Maximum DPO4 uptake by bacteria 0.48 mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$ Thingstad (1987) <t< td=""><td>$oldsymbol{ ho}_g^{Ba}$</td><td>Factor of carbon respired by bacteria</td><td>0.60</td><td><u></u></td><td>Thingstad (1987)</td></t<>	$oldsymbol{ ho}_g^{Ba}$	Factor of carbon respired by bacteria	0.60	<u></u>	Thingstad (1987)
$V_{BBOC,max}$ Maximum LDOC uptake by bacteria 16.33 mmolC (10^{12} cell) $^{-1}$ d $^{-1}$ Campbell et al. (2013) $V_{BBON,max}^{BON,max}$ Maximum DPON uptake by bacteria 0.05 mmolN (10^{12} cell) $^{-1}$ d $^{-1}$ Faure et al. (2010) $V_{BAO,max}^{BON,max}$ Maximum DPON uptake by bacteria 0.32 mmolN (10^{12} cell) $^{-1}$ d $^{-1}$ Faure et al. (2010) $V_{BAO,max}^{BON,max}$ Maximum DPOP uptake by bacteria 0.01 mmolN (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DPOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DPOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maximum DOP uptake by bacteria 0.48 mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{BAO,max}^{BON,max}$ Maxi	$oldsymbol{ ho}_r^{Ba}$	Respiration rate of bacteria	0.01	d^{-1}	Thingstad (1987)
$V_{PPON,max}^{N}$ Maximum DPON uptake by bacteria0.05mmolN $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Faure et al. (2010) $V_{BDON,max}^{N}$ Maximum LDON uptake by bacteria0.32mmolN $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Faure et al. (2010) $V_{BA}^{N}_{M_{A},max}$ Maximum DPOP uptake by bacteria0.01mmolN $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BOP,max}^{R}$ Maximum LDOP uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum PO4 uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum PO4 uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum PO4 uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum PO4 uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum DO4 uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum LDOP uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum LDOP uptake by bacteria0.48mmolP $(10^{12} \text{ cell})^1 \text{ d}^{-1}$ Thingstad (1987) $V_{BA}^{R}_{A,max}$ Maximum LDOP uptake by bacteria0.50mmolD molP m^3 Faure et al. (2010) $V_{BA}^{R}_{A,max}$ Half-saturation constant for LDON0.50 <td< td=""><td>,</td><td>Maximum DPOC uptake by bacteria</td><td>0.029</td><td></td><td>Campbell et al. (2013)</td></td<>	,	Maximum DPOC uptake by bacteria	0.029		Campbell et al. (2013)
$V_{BDON,max}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{Max}}$ $V_{BA_{DOP,max}}$ $V_{BDOP,max}$ $V_{BA_{DOP,max}}$ $V_{BA_{DOP,max}}$ $V_{BA_{DOP,max}}$ $V_{BA_{DOP,max}}$ $V_{BA_{DOP,max}}$ $V_{BA_{DOR}}$ V_{BA_{DO		Maximum LDOC uptake by bacteria	16.33	mmolC (10 ¹² cell) ⁻¹ d ⁻¹	Campbell et al. (2013)
$V_{BA_{4},max}$ Maximum NH4 uptake by bacteria0.32mmolN (10^{12} cell)- 1 d- 1 Faure et al. (2010) $V_{BPOP,max}$ Maximum DPOP uptake by bacteria0.01mmolP (10^{12} cell)- 1 d- 1 Thingstad (1987) $V_{BA_{DOP,max}}$ Maximum LDOP uptake by bacteria0.48mmolP (10^{12} cell)- 1 d- 1 Thingstad (1987) $V_{BA_{Max}}$ Maximum PO4 uptake by bacteria0.48mmolP (10^{12} cell)- 1 d- 1 Thingstad (1987) $V_{BA_{DOC}}$ Half-saturation constant for DPOC10.0mmolC m- 3 Faure et al. (2010) K_{BDOC} Half-saturation constant for LDOC25.0mmolC m- 3 Faure et al. (2010) K_{BDON}^{BDO} Half-saturation constant for DPON0.50mmolN m- 3 $\frac{1}{2}$ K_{BDON}^{BDO} Half-saturation constant for DPOP0.08mmolN m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{BDOP}^{BA} Half-saturation obstant for DPOP0.08mmolP m- 3 Leblanc et al. (2018) K_{B	$V_{DPON,max}^{BA}$	Maximum DPON uptake by bacteria	0.05	mmolN (10 ¹² cell) ⁻¹ d ⁻¹	Faure et al. (2010)
$V_{DPOP,max}$ Maximum DPOP uptake by bacteria0.01mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{DOP,max}$ Maximum LDOP uptake by bacteria0.48mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) $V_{PO_{4,max}}$ Maximum PO ₄ uptake by bacteria0.48mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) V_{DPOC} Half-saturation constant for DPOC10.0mmolP (10^{12} cell) $^{-1}$ d $^{-1}$ Thingstad (1987) K_{DPOC} Half-saturation constant for DPOC10.0mmolC m $^{-3}$ Faure et al. (2010) K_{DPOC} Half-saturation constant for LDOC25.0mmolN m $^{-3}$ $\frac{1}{2}$ K_{DDON}^{BA} Half-saturation constant for DPON0.50mmolN m $^{-3}$ $\frac{1}{2}$ K_{DDON}^{BA} Half-saturation constant for DPOP0.08mmolP m $^{-3}$ Leblanc et al. (2018) K_{DPOP}^{BA} Half-saturation constant for DPOP0.08mmolP m $^{-3}$ Leblanc et al. (2018) K_{DDO}^{BA} Half-saturation constant for PO40.02mmolP m $^{-3}$ Leblanc et al. (2018) E_{DLC} fraction excretion of DIC0.31 $\frac{1}{2}$ Faure et al. (2010) E_{NH4} fraction excretion of PO40.50 $\frac{1}{2}$ Faure et al. (2010) E_{PO4} Fraction of C assimilated0.92 $\frac{1}{2}$ Gerber and Gerber (1979) d_{N} Fraction of P assimilated0.95 $\frac{1}{2}$ Fraysse et al. (2013) $d_{P,200}$ Net N growth efficiency0.40 $\frac{1}{2}$ Le Borgne and Rodic (1997	$V_{LDON,max}^{BA}$	Maximum LDON uptake by bacteria	0.32	mmolN (10 ¹² cell) ⁻¹ d ⁻¹	Faure et al. (2010)
$V_{DDOP,max}^{BA}$ Maximum LDOP uptake by bacteria0.48mmolP (10^{12} cell)^{-1} d^{-1}Thingstad (1987) $V_{PO_{A,max}}^{BA}$ Maximum PO ₄ uptake by bacteria0.48mmolP (10^{12} cell)^{-1} d^{-1}Thingstad (1987) K_{DPOC}^{BA} Half-saturation constant for DPOC10.0mmolC m³Faure et al. (2010) K_{DDOC}^{BD} Half-saturation constant for LDOC25.0mmolN m³ \leftarrow K_{DDON}^{BA} Half-saturation constant for DPON0.50mmolN m³ \leftarrow K_{DDON}^{BA} Half-saturation constant for LDON0.50mmolN m³ \leftarrow K_{DDON}^{BA} Half-saturation constant for DPOP0.08mmolP m³ \leftarrow K_{DDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m³ \leftarrow K_{DDOP}^{BA} Half-saturation constant for PO40.02mmolP m³ \leftarrow Leblanc et al. (2018) K_{DDO}^{BA} Half-saturation constant for PO40.02mmolP m³ \leftarrow Campbell et al. (2013) ε_{DIC} fraction excretion of DIC0.31 \leftarrow Faure et al. (2010) ε_{NH4} fraction excretion of PO40.50 \leftarrow Fraysse et al. (2010) ε_{PO4} Fraction of C assimilated0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 \leftarrow Fraysse et al. (2010) d_P Fraction of P assimilated0.95 \leftarrow Frayse et al. (2013) d_P Fraction of P assimilated0.95 \leftarrow Frayse et al. (2013) d_P Frayse	$V_{NH_4,max}^{BA}$	Maximum NH4 uptake by bacteria	0.32	mmolN (10 ¹² cell) ⁻¹ d ⁻¹	Faure et al. (2010)
$V_{PO_{4,max}}^{BA}$ Maximum PO ₄ uptake by bacteria0.48mmolP (10^{12} cell) ⁻¹ d ⁻¹ Thingstad (1987) K_{DPOC}^{BA} Half-saturation constant for DPOC10.0mmolC m ⁻³ Faure et al. (2010) K_{DPOC}^{BA} Half-saturation constant for LDOC25.0mmolC m ⁻³ $\frac{1}{2}$ K_{DPON}^{BA} Half-saturation constant for DPON0.50mmolN m ⁻³ $\frac{1}{2}$ K_{DDON}^{BA} Half-saturation constant for DPON0.50mmolN m ⁻³ Leblanc et al. (2018) K_{DPOD}^{BA} Half-saturation constant for DPOP0.08mmolP m ⁻³ Leblanc et al. (2018) K_{DDOP}^{BA} Half-saturation constant for DPOP0.08mmolP m ⁻³ Leblanc et al. (2018) K_{DDOP}^{BA} Half-saturation constant for PO40.02mmolP m ⁻³ Leblanc et al. (2013) \mathcal{E}_{DIC} fraction excretion of DIC0.31 $\frac{1}{2}$ Faure et al. (2010) \mathcal{E}_{NH4} fraction excretion of PO40.50 $\frac{1}{2}$ Faure et al. (2010) \mathcal{E}_{PO4} Fraction excretion of PO40.50 $\frac{1}{2}$ Fraysse et al. (2013) d_C Fraction of C assimilated0.92 $\frac{1}{2}$ Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 $\frac{1}{2}$ Faure et al. (2010) d_P Fraction of P assimilated0.95 $\frac{1}{2}$ Faure et al. (2013) $k_{C,200}$ Net P growth efficiency0.40 $\frac{1}{2}$ Le Borgne and Rodic (1997) $k_{N,200}$ Net P growth efficiency0.37 $\frac{1}{2}$ <	$V_{DPOP,max}^{BA}$	Maximum DPOP uptake by bacteria	0.01	$mmolP\ (10^{12}\ cell)^{1}\ d^{1}$	Thingstad (1987)
K_{DPOC}^{BA} Half-saturation constant for DPOC 10.0 mmolC m ⁻³ Faure et al. (2010) K_{DPOC}^{BA} Half-saturation constant for LDOC 25.0 mmolC m ⁻³ $\frac{1}{2}$ K_{DPON}^{BA} Half-saturation constant for DPON 0.50 mmolN m ⁻³ $\frac{1}{2}$ K_{DDON}^{BA} Half-saturation constant for LDON 0.50 mmolN m ⁻³ $\frac{1}{2}$ K_{BA}^{BA} Half-saturation constant for DPOP 0.08 mmolP m ⁻³ Leblanc et al. (2018) K_{DDOP}^{BA} Half-saturation constant for DPOP 0.08 mmolP m ⁻³ Leblanc et al. (2018) K_{POA}^{BA} Half-saturation constant for PO4 0.02 mmolP m ⁻³ Campbell et al. (2013) E_{DIC} fraction excretion of DIC 0.31 $\frac{1}{2}$ Faure et al. (2010) E_{NH4} fraction excretion of PO4 0.50 $\frac{1}{2}$ Faure et al. (2010) E_{PO4} Fraction excretion of PO4 0.50 $\frac{1}{2}$ Fraysse et al. (2013) d_C Fraction of C assimilated 0.92 $\frac{1}{2}$ Gerber and Gerber (1979) d_N Fraction of P assimilated 0.95 $\frac{1}{2}$ Frayse et al. (2013) d_C Net N growth efficiency 0.40 $\frac{1}{2}$ Gerber and Gerber (1979) $k_{N,Z00}$ Net P growth efficiency 0.44 $\frac{1}{2}$ Le Borgne and Rodic (1997) $k_{N,Z00}$ Net P growth efficiency 0.44 $\frac{1}{2}$ Le Borgne (1982) Q_{10} Temperature coefficient 2.0 $\frac{1}{2}$ $\frac{1}{2}$ $T_$	$V_{LDOP,max}^{BA}$	Maximum LDOP uptake by bacteria	0.48	mmolP $(10^{12} \text{ cell})^{-1} \text{ d}^{-1}$	Thingstad (1987)
K_{DDO}^{BO} Half-saturation constant for LDOC25.0mmolC m-3 \leftarrow K_{DDON}^{BA} Half-saturation constant for DPON0.50mmolN m-3 \leftarrow K_{LDON}^{BA} Half-saturation constant for LDON0.50mmolN m-3 \leftarrow K_{NH4}^{BA} Half-saturation constant for NH40.15mmolN m-3Leblanc et al. (2018) K_{DPOP}^{BA} Half-saturation constant for DPOP0.08mmolP m-3 \leftarrow K_{DOD}^{BA} Half-saturation constant for LDOP0.08mmolP m-3Leblanc et al. (2018) K_{POA}^{BA} Half-saturation constant for PO40.02mmolP m-3Campbell et al. (2013) E_{DIC} fraction excretion of DIC0.31 \leftarrow Faure et al. (2010) E_{NH4} fraction excretion of PO40.50 \leftarrow Faure et al. (2010) E_{PO4} Fraction excretion of PO40.50 \leftarrow Fraysse et al. (2013) d_C Fraction of C assimilated0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of P assimilated0.95 \leftarrow Faure et al. (2010) d_P Fraction of P assimilated0.95 \leftarrow Fraysse et al. (2013) $k_{C,Z00}$ Net C growth efficiency0.40 \leftarrow Gerber and Gerber (1979) $k_{N,Z00}$ Net P growth efficiency0.44 \leftarrow Le Borgne and Rodic (1997) $k_{P,Z00}$ Net P growth efficiency0.37 \leftarrow Le Borgne (1982)Q10Temperature coefficient2.0 \leftarrow \leftarrow $t_{P,Z00}$ Net P growth	$V_{PO_4,max}^{BA}$	Maximum PO ₄ uptake by bacteria	0.48	$mmolP\ (10^{12}\ cell)^{1}\ d^{1}$	Thingstad (1987)
K_{DPON}^{BA} Half-saturation constant for DPON0.50mmolN m³ \leftarrow K_{LDON}^{BA} Half-saturation constant for LDON0.50mmolN m³ \leftarrow K_{NH4}^{BA} Half-saturation constant for NH40.15mmolN m³Leblanc et al. (2018) K_{DPOP}^{BA} Half-saturation constant for DPOP0.08mmolP m³ \leftarrow K_{LDOP}^{BA} Half-saturation constant for LDOP0.08mmolP m³Leblanc et al. (2018) K_{POA}^{BA} Half-saturation constant for PO40.02mmolP m³Campbell et al. (2013) ϵ_{DIC} fraction excretion of DIC0.31 \leftarrow Faure et al. (2010) ϵ_{NH4} fraction excretion of NH40.50 \leftarrow Faure et al. (2010) ϵ_{PO4} Fraction excretion of PO40.50 \leftarrow Fraysse et al. (2013) d_C Fraction of C assimilated0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 \leftarrow Faure et al. (2010) d_P Fraction of P assimilated0.95 \leftarrow Fraysse et al. (2013) d_C Net C growth efficiency0.40 \leftarrow Gerber and Gerber (1979) $k_{N,200}$ Net N growth efficiency0.44 \leftarrow Le Borgne and Rodic (1997) $k_{P,200}$ Net P growth efficiency0.37 \leftarrow Le Borgne (1982) Q_{10} Temperature coefficient2.0 \leftarrow \leftarrow $T_{r,m}$ Reference temperature for mineralization20.0 \sim \leftarrow $T_{r,m}$ Reference temperature for	K_{DPOC}^{BA}	Half-saturation constant for DPOC	10.0	mmolC m ⁻³	Faure et al. (2010)
K_{LDON}^{BA} Half-saturation constant for LDON 0.50 mmolN m ⁻³ Leblanc et al. (2018) K_{NH4}^{BA} Half-saturation constant for DPOP 0.08 mmolP m ⁻³ Leblanc et al. (2018) K_{DPOP}^{BA} Half-saturation constant for DPOP 0.08 mmolP m ⁻³ Leblanc et al. (2018) K_{DOD}^{BA} Half-saturation constant for LDOP 0.08 mmolP m ⁻³ Leblanc et al. (2018) K_{DOD}^{BA} Half-saturation constant for PO4 0.02 mmolP m ⁻³ Campbell et al. (2013) ε_{DIC} fraction excretion of DIC 0.31 \leftarrow Faure et al. (2010) ε_{NH4} fraction excretion of PO4 0.50 \leftarrow Faure et al. (2010) ε_{PO4} Fraction excretion of PO4 0.50 \leftarrow Faure et al. (2013) d_C Fraction of C assimilated 0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of N assimilated 0.95 \leftarrow Faure et al. (2010) d_P Fraction of P assimilated 0.95 \leftarrow Faure et al. (2013) $k_{C,ZOO}$ Net C growth efficiency 0.40 \leftarrow Gerber and Gerber (1979) $k_{N,ZOO}$ Net N growth efficiency 0.44 \leftarrow Gerber and Gerber (1979) $k_{P,ZOO}$ Net P growth efficiency 0.37 \leftarrow Le Borgne and Rodic (1997) $k_{P,ZOO}$ Net P growth efficiency 0.37 \leftarrow Le Borgne (1982) Qto Temperature coefficient 2.0 \leftarrow C c Trem Reference temperature for mineralization rate 0.05 c d-1 Lacroix and Grégoire (2002) c Reference temperature for Reference t	K_{LDOC}^{BA}	Half-saturation constant for LDOC	25.0	mmolC m ⁻³	⊭
R_{DDON} Main attention constant for NH40.15mmolN m³3Leblanc et al. (2018) R_{PA}^{BA} Half-saturation constant for DPOP0.08mmolP m³3Leblanc et al. (2018) R_{DOP}^{BA} Half-saturation constant for LDOP0.08mmolP m³3Leblanc et al. (2018) R_{POA}^{BA} Half-saturation constant for PO40.02mmolP m³3Campbell et al. (2018) R_{DIC}^{BA} Half-saturation constant for PO40.02mmolP m³3Campbell et al. (2010) E_{DIC} fraction excretion of DIC0.31 $\frac{1}{2}$ Faure et al. (2010) E_{NH4} fraction excretion of PO40.50 $\frac{1}{2}$ Faure et al. (2010) E_{PO4} Fraction excretion of PO40.50 $\frac{1}{2}$ Fraysse et al. (2013) d_C Fraction of C assimilated0.92 $\frac{1}{2}$ Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 $\frac{1}{2}$ Faure et al. (2010) d_P Fraction of P assimilated0.95 $\frac{1}{2}$ Fraysse et al. (2013) $R_{C,ZOO}$ Net C growth efficiency0.40 $\frac{1}{2}$ Fraysse et al. (2013) $R_{N,ZOO}$ Net N growth efficiency0.44 $\frac{1}{2}$ Le Borgne and Rodic (1997) $R_{P,ZOO}$ Net P growth efficiency0.37 $\frac{1}{2}$ Le Borgne (1982) $R_{P,ZOO}$ Net P growth efficient2.0 $\frac{1}{2}$ Le Borgne (1982) $R_{P,ZOO}$ Net P growth efficient2.0 $\frac{1}{2}$ Le Borgne (1982) $R_{P,ZOO}$ Net P growth efficient<	K_{DPON}^{BA}	Half-saturation constant for DPON	0.50	mmolN m ⁻³	⊭
K_{PODP}^{BA} Half-saturation constant for DPOP0.08mmolP m ⁻³ $\not=$ K_{DODP}^{BA} Half-saturation constant for LDOP0.08mmolP m ⁻³ Leblanc et al. (2018) K_{POA}^{BA} Half-saturation constant for PO40.02mmolP m ⁻³ Campbell et al. (2013) ε_{DIC} fraction excretion of DIC0.31 $\not=$ Faure et al. (2010) ε_{NH4} fraction excretion of NH40.50 $\not=$ Faure et al. (2010) ε_{PO4} Fraction excretion of PO40.50 $\not=$ Fraysse et al. (2013) d_C Fraction of C assimilated0.92 $\not=$ Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 $\not=$ Faure et al. (2010) d_P Fraction of P assimilated0.95 $\not=$ Fraysse et al. (2013) $k_{C,ZOO}$ Net C growth efficiency0.40 $\not=$ Gerber and Gerber (1979) $k_{N,ZOO}$ Net N growth efficiency0.44 $\not=$ Le Borgne and Rodic (1997) $k_{P,ZOO}$ Net P growth efficiency0.37 $\not=$ Le Borgne (1982)Q10Temperature coefficient2.0 $\not=$ $\not=$ T_{PEM} Reference temperature for mineralization20.0 \circ C $\not=$ K_{RIL} Nitrification rate0.05d ¹ Lacroix and Grégoire (2002) T_{PEM} Reference temperature for mineralization10.0 \circ C $\not=$	K_{LDON}^{BA}	Half-saturation constant for LDON	0.50	mmolN m ⁻³	₽
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ε_{DIC} fraction excretion of DIC0.31 \leftarrow Faure et al. (2010) ε_{NH_4} fraction excretion of NH40.50 \leftarrow Faure et al. (2010) ε_{PO_4} Fraction excretion of PO ₄ 0.50 \leftarrow Fraysse et al. (2013) d_C Fraction of C assimilated0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 \leftarrow Faure et al. (2010) d_P Fraction of P assimilated0.95 \leftarrow Fraysse et al. (2013) $k_{C,ZOO}$ Net C growth efficiency0.40 \leftarrow Gerber and Gerber (1979) $k_{N,ZOO}$ Net N growth efficiency0.44 \leftarrow Le Borgne and Rodic (1997) $k_{P,ZOO}$ Net P growth efficiency0.37 \leftarrow Le Borgne (1982) Q_{10} Temperature coefficient2.0 \leftarrow \leftarrow T_{rem} Reference temperature for mineralization20.0 \circ C \leftarrow k_{nit} Nitrification rate0.05 d^{-1} Lacroix and Grégoire (2002) T_{rem} Reference temperature for Reference temperature f	K_{LDOP}^{BA}	Half-saturation constant for LDOP	0.08	mmolP m ⁻³	Leblanc et al. (2018)
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ε_{PO4} Fraction excretion of PO40.50 \leftarrow Fraysse et al. (2013) d_C Fraction of C assimilated0.92 \leftarrow Gerber and Gerber (1979) d_N Fraction of N assimilated0.95 \leftarrow Faure et al. (2010) d_P Fraction of P assimilated0.95 \leftarrow Fraysse et al. (2013) $k_{C,zoo}$ Net C growth efficiency0.40 \leftarrow Gerber and Gerber (1979) $k_{N,zoo}$ Net N growth efficiency0.44 \leftarrow Le Borgne and Rodio (1997) $k_{P,zoo}$ Net P growth efficiency0.37 \leftarrow Le Borgne (1982) Q_{10} Temperature coefficient2.0 \leftarrow \leftarrow T_{rem} Reference temperature for mineralization20.0 \circ C \leftarrow k_{nit} Nitrification rate0.05 d^{-1} Lacroix and Grégoire (2002) T_{rem} Reference temperature for Reference temperature	ε_{DIC}	fraction excretion of DIC	0.31	<u></u>	Faure et al. (2010)
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d_P Fraction of P assimilated0.95 \leftarrow Fraysse et al. (2013) $k_{C,200}$ Net C growth efficiency0.40 \leftarrow Gerber and Gerber (1979) $k_{N,200}$ Net N growth efficiency0.44 \leftarrow Le Borgne and Rodie (1997) $k_{P,200}$ Net P growth efficiency0.37 \leftarrow Le Borgne (1982)Q10Temperature coefficient2.0 \leftarrow \leftarrow T_{rem} Reference temperature for mineralization20.0°C \leftarrow k_{nit} Nitrification rate0.05 d^{-1} Lacroix and Grégoire (2002) T_{rem} Reference temperature for10.0°C	$d_{\mathcal{C}}$	Fraction of C assimilated	0.92	<u></u>	Gerber and Gerber (1979)
$k_{C,zoo}$ Net C growth efficiency 0.40 \leftarrow Gerber and Gerber (1979) $k_{N,zoo}$ Net N growth efficiency 0.44 \leftarrow Le Borgne and Rodie (1997) $k_{P,zoo}$ Net P growth efficiency 0.37 \leftarrow Le Borgne (1982) Q10 Temperature coefficient 2.0 \leftarrow \leftarrow T_{rem} Reference temperature for mineralization 20.0 $^{\circ}$ C \leftarrow k_{nit} Nitrification rate 0.05 d^{-1} Lacroix and Grégoire (2002) T_{rem} Reference temperature for Re	d_N	Fraction of N assimilated	0.95	<u></u>	Faure et al. (2010)
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$k_{N,200}$ Net N growth efficiency 0.44 (1997) $k_{P,200}$ Net P growth efficiency 0.37 \leftarrow Le Borgne (1982) Q10 Temperature coefficient 2.0 \leftarrow \leftarrow T_{rem} Reference temperature for mineralization 20.0 $^{\circ}$ C \leftarrow k_{nit} Nitrification rate 0.05 d^{-1} Lacroix and Grégoire (2002) T_{rem} Reference temperature for 10.0 $^{\circ}$ C	$k_{C,zoo}$	Net C growth efficiency	0.40	<i></i> <u></u>	Gerber and Gerber (1979)
Q10 Temperature coefficient 2.0 \leftarrow \leftarrow \leftarrow Trem Reference temperature for mineralization 20.0 °C \leftarrow \leftarrow Nitrification rate 0.05 d-1 Lacroix and Grégoire (2002 True Reference temperature for 10.0 °C \leftarrow \leftarrow	$k_{N,zoo}$	Net N growth efficiency	0.44		C
Trem Reference temperature for mineralization 20.0 °C \leftarrow k_{ntt} Nitrification rate 0.05 d ⁻¹ Lacroix and Grégoire (2002) The Reference temperature for 10.0 °C \leftarrow	$k_{P,zoo}$	Net P growth efficiency	0.37	+_	Le Borgne (1982)
Trem mineralization 20.0 $^{\circ}$ C $^{\circ}$ Multiplication rate 0.05 $^{\circ}$ d ⁻¹ Lacroix and Grégoire (2002) $^{\circ}$ Reference temperature for 10.0 $^{\circ}$ C $^{\circ}$	Q10	•	2.0	+ <u>-</u>	
Reference temperature for 10.0 °C /	T_{rem}	1			
	k_{nit}		0.05	d^{-1}	Lacroix and Grégoire (2002
	T _{nit}		10.0	°C	4

Tableau mis en forme

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Tableau mis en forme

K_{DO}	Half-saturation constant DO	30.0	$mmolO_2m^{-3}$	Tett (1990)
k_{diss}	Dissolution rate	10.9	d ⁻¹	Gehlen et al. (2007)
k_{precip}	Fraction of PIC to LPOC	0.02	⊭	Marty et al. (2002)
K_{C}	Half-saturation constant of CaCO ₃ precipitation	0.40	$(\mu mol \ kg^{-l})^2$	
$\left(\frac{o}{c}\right)$	Ratio O:C	1.0	/ _	4
$\left(\frac{o}{N}\right)$	Ratio O:N	2.0	£	₽

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Appendix C: Short User Manual

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After uploading the whole archive on the zenodo site (ref. doi: 10.5281/zenodo.3757677), the exact version of the Eco3M-CarbOx code used in this study can be be run as following:

make !two executable files will be created : eco3M_ini.exe and eco3M.exe

- the file config.ini allows to define: the time, time step, and save time of simulation variables biogeochemical process
- Results files are stocked in "SORTIES" directory
- Boundary conditions and forcings data are stocked in "DATA" directory
- All subroutines of biogeochemical processes are stocked in "F_PROCESS" directory

For further information, please contact Dr. Frédéric DIAZ (frederic.diaz@univ-amu.fr) or Dr. Christel PINAZO (christel.pinazo@univ-amu.fr)

Review of the first revised version of

930 "Implementation and assessment of a carbonate system model (Eco3M-CarbOxv1.1) in a highly-dynamic Mediterranean coastal site (Bay of Marseille, France)"

submitted for publication to Geoscientific Model Development

by K. Lajaunie-Salla and co-authors

935

<u>General reply</u>: We thank again the Reviewer Dr Munhoven for his second evaluation of our work. We thank too him for the detailed and useful comments that contributed to greatly improve the manuscript. We consider all of his comments to improve the manuscript.

940

1. General comments

The authors' reply the revised manuscript are not very "reviewer-friendly." It is nowadays standard practice to provide a "track change" (or a LATEXDIFF) version of the manuscript clearly identifying the changes made to the text, right in the text. The equivalent information is seemingly given in the reply, except that the line numbers provided there do not match, so that one has to search manually for the exact location of the changes.

The preparation of the manuscript would also have benefited from some extra care. Page numbers restart at 1 after page 23 without any apparent reason.

Reply: The preparation of the revised ms has been reviewed with care and page numbers have been reprocessed. A new check of the different sections has been carefully performed.

950

${\bf 1.1 \; Appreciation \; of \; the \; replies \; to \; reviewers}$

The authors have all in all well responded to the referees' comments, with one exception though. In the response to my comment 2.3, regarding the missing effect of river intrusions on DIC — TA perturbations are taken into account, but as these are carried mainly by HCO3°, they also generate DIC perturbations of the same magnitude, which are neglected — I read at the top of the fourth page (page numbers in the response would have been helpful) that

"Concerning the riverine inputs scenarii, we decide to focus on nitrate and alkalinity supply. In fact the model simulates the DIC increases, as is observed, which highlight that the carbonate system module is well resolved."

The reply to the comments is somewhat ambiguous as suggests that DIC changes are taken into account, while they are actually not, as stated in the manuscript at lines 413–414:

60 "[...] the experimental design on the Rhône River supply only considers the TA perturbation on the carbonate system but not that due to the DIC supply."

So, even if the model reproduces the observed DIC increases (as stated in the reply), this must obviously be for the wrong reasons, as only one half of the effects of the perturbation due to river intrusions is taken into account. By the way, no one argued that the carbonate system module was not well resolved.

965 <u>Reply:</u> We decided to remove the scenario of AT supply by river and all the text referred to it in the methods and results/discussion sections. See our detailed comment on this point hereafter.

1.2 Appreciation of the revised manuscript

The model description has been improved and the rationale behind the carbonate speciation calculations is now presented in a new appendix. Unfortunately, the layout of that appendix is rather chaotic which makes it difficult to read.

2. Specific comments

2.1 River intrusion experiments: poor justification

The justification added at lines 414–417 for taking the effect of river intrusions into account only in terms of the resulting TA but not DIC perturbations is rather cavalier and scientifically untenable. This is a completely unrealistic assumption that makes the outcome of the experiment meaningless and thus essentially invalidates any conclusion drawn from it.

I only see two options to address this shortcoming:

- 1. the river intrusion experiments are repeated with the effect on DIC included (which should be rather straightforward to correct) and 980 the discussion of the results adapted;
 - 2. these experiments are simply taken out of the paper as the current results are essentially unfounded.

Even in preliminary experiments, one must not choose to disregard one of two effects of a perturbation if these are of the same order of magnitude. Such arbitrary choices lead to arbitrary results.

Reply: We have chosen the second option proposed by the Reviewer. Mainly because it was not straightforward to consider DIC supply by River as considered for AT. In fact, DIC is not as conservative as AT is regard to salinity. It is then impossible to use a relationship between DIC and salinity as that used between AT and salinity in the previous version of our manuscript. In the context of a 0D modelling developed here, this kind of relationship would have been the only mean to take into account the DIC supply by the Rhône River. In the revised version, we stressed (1 398-401) that rivers also supply TA and DIC and a consideration for these supplies in future works may sensibly modify the modeled carbonate balance in the BoM compared to that presented in this study.

Concomitantly we slightly rewrote the end of section 4.1 on the model performance in Discussion.

As noted in the conclusion (1434), a coupling of the Eco3M-CarbOx model to a 3D hydrodynamics model is planned to better represent

As noted in the conclusion (1434), a coupling of the Eco3M-CarbOx model to a 3D hydrodynamics model is planned to better represent the complexity of functioning of the BOM. This implementation will enable, for example, to take into account actual DIC and AT supplies from the Rhône River by considering forcing values measured in the River.

995 3. Technical comments

Reply: All these technical comments have been taken into account in the new revised version of ms.

- Page 1, line 22: "the year 2017 that is a period for which" should read "the year 2017 for which"
- Page 1, line 25: "of most of variables of carbonate system except Total Alkalinity." should read "of most of the variables of the 1000 carbonate system except for Total Alkalinity."
 - Page 1, line 26: "experiments were also conducted" should read "experiments were conducted"
 - Page 1, line 26: "to (i) seawater" should read "to (i) a seawater"
 - Page 1, line 27: "Rhône River plume intrusion" should read "Rhône River plume intrusions"; by the way: the name of that river is sometimes spelled "Rhône", more often "Rhone" please use the same spelling consistently throughout
- 1005 Page 1, line 35: "external forcing have" should read "external forcings have"
 - Page 5, line 188: "a salinity threshold of 37 has been chosen" is this correct? A threshold of 37 looks rather high to me.
 - Page 6, line 226: "during MWC period" should read "during the MWC period" $\,$
 - Page 7, line 271: "15 March and 6 May" should either read "15th March and 6th May" or "March 15th and May 6th" (as on line 277, p. 8)
- 1010 Page 11, line 390: "Moreover, previous study" should read "Moreover, a previous study", or even better reformulate the sentence to read "Moreover, Fraysse et al. (2013) highlight that . . . " and discard the citation in brackets.
 - Page 11, line 402: "a longer period ca. 15 days" should read "a longer period of ca. 15 days"

Page 11, lines 402–403: "high atmospheric pCO2 value and wind speed" better had to read "high atmospheric pCO2 and high wind speeds"

1015 Page 11, line 411: "due to some two" should read "due to two"

29th and 30th pages (pages nr. 6 and 7), throughout: "in the pH scale" should read "on the pH scale"

29th page (page nr. 6): "Concentration in Total Fluoride (TF) ions" should read "Total Fluoride concentration" (without "ions," as TF includes the non ionic HF)

29th page (page nr. 6): "Concentration in Total Sulphate (TS) ions" should read "Total Sulphate concentration" ("ions" is superfluous)

1020 29th page (page nr. 6): "Concentration in Total Boron (TB)" should read "total boron concentration" (without "ions," as TB includes the non ionic B(OH)3)

29th page (page nr. 6): KF is the dissociation constant of hydrogen fluoride (or of hydrofluoric acid), not of fluoride ions

29th page (page nr. 6): in the expression for KF, the exponent for Ions should be 0.5 (or 1/2) and not 1.5.

30th page (page nr. 7): "Every constant are corrected by the hydrostatic pressure" should read "All the constants are corrected for the
1025 effect of hydrostatic pressure"

Guy Munhoven Liège, 9th October 2020