FABM-NflexPD 1.0: Assessing an Instantaneous Acclimation Approach for Modelling Phytoplankton Growth

Onur Kerimoglu[1,2], Prima Anugerahanti[3], and Sherwood Lan Smith[3]

[1]Institute for Chemistry and Biology of the Marine Environment, University of Oldenburg, Germany
[2]Helmholtz Center for Coastal Research, Germany
[3]Earth SURFACE Research Center, Research Institute for Global Change, JAMSTEC, Japan

Correspondence: Onur Kerimoglu (kerimoglu.o@gmail.com)

Abstract. Coupled physical-biogeochemical models can generally reproduce large-scale patterns of primary production and biogeochemistry, but they often underestimate observed variability and gradients. This is partially caused by insufficient representation of systematic variations in the elemental composition and pigment density of phytoplankton. Although progress has been made through approaches accounting for the dynamics of phytoplankton composition with additional state variables,

5 formidable computational challenges arise when these are applied in spatially explicit setups. The Instantaneous Acclimation (IA) approach addresses these challenges by assuming that Chl:C:nutrient ratios are instantly optimized locally (within each modelled grid cell, at each timestep), such that they can be resolved as diagnostic variables. Here we present the first tests of IA in an idealized, 1D setup: we implemented the IA in the Framework for Aquatic Biogeochemical Models (FABM), and coupled it with the General Ocean Turbulence Model (GOTM) to simulate the spatio-temporal dynamics in a 1-D water

10 column. We show that the IA model and a fully dynamic, otherwise equivalently acclimative (DA) variant with an additional state variable behave similarly, and both resolve nutrient and growth dynamics not captured by a third, non-acclimative and fixed-stoichiometry (FS) variant.

1 Introduction

1.1 Modelling phytoplankton and their cellular composition:

15 In early ecosystem models, the elemental composition, i.e., proportion of carbon (C), nitrogen (N), and phosphorus (P) content of phytoplankton was generally assumed constant, and at least since the work of Dugdale (1967) their growth was typically described by the so-called 'Monod' model (Monod, 1949), which assumes a saturating response of the rate of carbon assimilation (and hence, of nutrient uptake) to the ambient nutrient concentration, described by a rectangular hyperbolic function. Similarly, specific chlorophyll (Chl) content, i.e., Chl:C ratio was assumed to be constant, when comparing the simulated phytoplankton

20 biomass against the *in-situ* or satellite based chlorophyll measurements. In many primary production modules coupled to general circulation models that are actively being used for various purposes to this date, phytoplankton C:Chl and/or C:N:P ratios are assumed to be constant (see e.g., the models in Laufkötter et al., 2015).

The inadequacy of these simplifying assumptions was made clear decades ago by the discovery that phytoplankton elemental composition (e.g., Gerloff and Skoog, 1954) and chlorophyll content (e.g., Platt and Jassby, 1976) are variable. Chl:C:N:P ratios

25 of phytoplankton have since been found to vary widely in many laboratory experiments (e.g., Kruskopf and Flynn, 2006) and field observations (e.g., Martiny et al., 2013; Burson et al., 2016). Since the work of Caperon (1968) and Droop (1968), the so called 'quota' (or variable internal stores, or 'Droop') model has been widely employed to describe the dynamics of carbon and nutrients bound to phytoplankton, using a separate state variable for each element or nutrient resolved. For describing variable Chl:C ratios, acclimation models such as that of Geider et al. (1998) are now commonly employed. Such models typically

30 couple a Droop-like description of variable N:C (or other nutrient:C) with photoacclimation, i.e., variation of Chl:C, using one more state variable for Chl bound to phytoplankton. Some models assume a constant N:C ratio, while describing the variations in Chl:C, e.g., using only the 'photoacclimation portion' (e.g., Moore et al., 2002) of the model by Geider et al. (1998) or using an empirical function (e.g., Oschlies and Schartau, 2005), that was earlier proposed by (Cloern et al., 1995).

Models that account for variations in cellular composition are indeed more likely to provide more realistic estimates of

35 phytoplankton biomass and biogeochemical fluxes. However, the mechanistic basis of some of these models are questionable, given their parameterisations of certain processes using heuristic or empirically inspired functions (Flynn et al., 2015). Moreover, schemes that require additional state variables, due to the need to calculate their transport as tracers, impose substantial computational costs. Especially for models that contain many clones/types (e.g., 350 in Dutkiewicz et al., 2020), such additional computational costs may severely limit the kinds of simulations and *in silica* experiments that can be conducted.

40 1.2 An optimality based resource allocation model

For the prediction of growth, nutrient uptake and acclimative variations of pigment and nutrient content of phytoplankton in response to changes in resource environment, 'Resource Allocation Models' (RAM) have been used (Shuter, 1979; Laws and Chalup, 1990; Armstrong, 1999; Klausmeier et al., 2004; Pahlow, 2005; Wirtz and Kerimoglu, 2016). This approach is based on the expectation that evolution has produced organisms that strive to maximize their net growth rate by optimally allocating their

45 resources to cellular functions. The dependence of all such functions on common resources therefore implies eco-physiological trade-offs (Smith et al., 2011). In this study, we specifically consider four physiological variables for describing the acclimative flexibilities involved in phytoplankton growth, as described by Pahlow et al. (2013):

Q: N quota (i.e., N:C ratio) of phytoplankton.

$f_{\mathbf{V}}$: fractional allocation to the nutrient uptake compartment (protoplast) to optimize the trade-off between photosynthesis (μ)
50 and nutrient uptake (V), as described by Pahlow and Oschlies (2013).

$f_{\mathbf{A}}$: fractional allocation to affinity to optimize the trade-off between nutrient affinity (A) and maximum uptake rate within the nutrient uptake compartment (V_{max}), as described by Pahlow (2005) and Smith et al. (2009).

$\hat{\theta}$: Chl:C ratio in chloroplasts ($\hat{\theta}$) to optimize the trade-off between energy gained by light harvesting and energetic costs of chlorophyll synthesis and maintenance, as described by Pahlow et al. (2013).

1.3 Instantaneous Acclimation (IA) approach

As in most previous models of flexible phytoplankton composition, the above mentioned model by Pahlow et al. (2013) explicitly resolved the dynamics of the carbon, nitrogen, and chlorophyll within phytoplankton biomass. This approach is well suited for simulating the short-term dynamics of growth, and hence for testing model assumptions against the results of batch culture experiments (e.g., Pahlow, 2005; Pahlow et al., 2013). Resolving the transient dynamics is important for such short-term experiments, where the response of phytoplankton may differ substantially in terms of nutrient uptake versus carbon-based growth and chlorophyll synthesis.

By contrast, oceanic (or even freshwater) observations are rarely available at such fine temporal resolution. The lack of observations at sufficient temporal resolution to test short-term model dynamics motivated the development of the Instantaneous Acclimation (IA) approach (Smith et al., 2016) as a way to potentially capture growth response at longer timescales while requiring substantially fewer calculations. IA is based on the balanced growth assumption, which Burmaster (1979) showed was able to reconcile the ability of the Droop, Monod and Michaelis-Menten models to capture phytoplankton growth response at steady state, as measured by continuous culture experiments. The key assumption is that growth and nutrient uptake are at all times strictly balanced (see Sect. 2.2 below for details). Based on this assumption, IA calculates only one specific rate for both growth and nutrient uptake. Smith et al. (2016) applied this assumption in a 0-D (box) model, adequate for reproducing sparse oceanic observations, but did not evaluate its performance compared to fully dynamic models of flexible composition.

Ward (2017) compared the results of a phytoplankton model with instantaneosly adjusting quota against a fully dynamic model with explicit state variables for each element resolved, and a fixed stoichiometry model, in a 0-D setup. He found that for a wide range of realistic forcing dynamics, the instantaneous approach yielded results practically indistinguishable from the fully dynamic model whereas these results differed considerably from those of the fixed stoichiometry model. To our knowledge, the IA approach has yet to be tested in a spatially explicit model, where the inclusion of transport terms may lead to additional complications.

1.4 Objectives of this study

This study presents a novel implementation of the IA-approach in the Framework for Aquatic Biogeochemical Models (FABM Bruggeman and Bolding, 2014) , and an assessment of its behavior compared to two other established variants (Fig. 1): the first is the widely used, non-acclimative, **Fixed-Stoichiometry (FS)** variant, which resolves only the N bound to phytoplankton explicitly. The second variant is the **Dynamic Acclimation (DA)** variant, which resolves the C and N bound to phytoplankton fully dynamically, with two state variables. The comparisons of the three model variants were conducted to answer the following two specific questions: *(i)* how do the simulations performed with the IA variant differ from those of the fully dynamic DA variant? and *(ii)* compared to the FS variant, do the results differ sufficiently to justify the additional complexities introduced by the IA variant?

In the following sections, we describe the general structure of the model, the details of the physiological flexibilities mentioned above for each model variant, and the setup to simulate the model. Then we show the results of the simulated patterns of
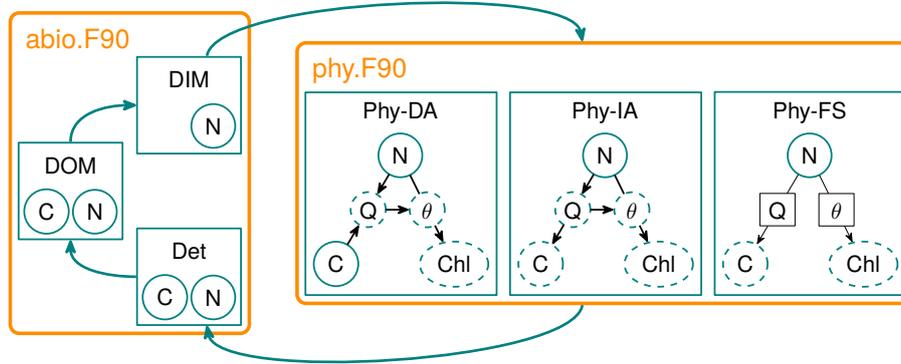
Figure 1. Diagram of the FABM-NflexPD model. Abiotic components, DOM, DIM and Det are calculated by the module `abio.F90`, which are then coupled to the phytoplankton simulated by the module `phy.F90` that simulates the dynamics of Phy_N, Phy_C and Phy_{Chl} by the DA, IA and FS variants (see section 2.2.2). Solid circles in the phytoplankton module represent state variables, dashed circles/ellipsoids represent diagnostically calculated variables and solid squares (for FS) represent prescribed values. The DA variant estimates the N, C and Chl content of phytoplankton based on a resource allocation scheme, whereas the FS variant estimates only N prognostically, while C and Chl are based on prescribed values of nitrogen quota (Q) and cellular Chl:C ratio (θ) (see the text).

phytoplankton in terms of carbon, nitrogen, and chlorophyll, cell quota (Q), Chl:C ratio, as well as the fractional allocations. We finally discuss the advantages, as well as the challenges and limitations of implementing the IA approach.

2 Model Description

2.1 General Structure

For describing the cycling of N, we consider a simple model structure (Fig. 1) with four compartments: C and N bound to phytoplankton (Phy_C, Phy_N), detritus (Det_C, Det_N), dissolved organic matter (DOC, DON) and Dissolved Inorganic Nitrogen (DIN). Note that our model does not resolve the dynamics of Dissolved Inorganic Carbon, DIC per se.

The coupled set of differential equations ($s(x)$ short for $\frac{dx}{dt}$) that describe the dynamics of state variables are provided in Eqs. (1–4). The definition and exact formulation of the trivial flux terms ($F_{\text{FROM-TO}}$) in Eqs. (1–4) are provided in Table 1. For equations applying only to a subset of our model variants, the variants are indicated near the equation number in curly braces ({}). In addition, Table 2 provides an overview of how the model variants differ.

$$s(Phy_N) = F_{DIN-Phy_N} - F_{Phy_N-Det_N} \tag{1a}$$

$$s(Phy_C) = F_{DIC-Phy_C} - F_{Phy_C-Det_C} \tag{\{DA\} \ (1b)}$$

$$s(Det_N) = F_{Phy_N-Det_N} - F_{Det_N-DON} \tag{2a}$$

$$s(Det_\text{C}) = F_{Phy_\text{C}-Det_\text{C}} - F_{Det_\text{C}-DOC} \tag{2b}$$

$$s(DON) = F_{Det_\text{N}-DON} - F_{DON-DIN} \tag{3a}$$

$$s(DOC) = F_{Det_\text{C}-DOC} - F_{DOC-DIC} \tag{3b}$$

$$s(DIN) = F_{DON-DIN} - F_{DIN-Phy_\text{N}} \tag{4}$$

The terms $F_{DIN-Phy_\text{N}}$ and $F_{DIC-Phy_\text{C}}$ have central importance to this study and deserve explanation. $F_{DIN-Phy_\text{N}}$ represents the net N flux from the DIN to phytoplankton, and is given by the product of the phytoplankton carbon biomass, Phy_C and the specific nutrient uptake rate, V:

$$F_{DIN-Phy_\text{N}} = V \cdot Phy_\text{C} \tag{5}$$

For the FS and IA variants, balanced growth (Burmaster, 1979) is assumed, such that V is directly linked to net growth rate, μ, via the nutrient quota, Q:

$$V = \mu \cdot Q \qquad\qquad \{\text{FS, IA}\} \tag{6}$$

whereas for the DA variant, V is calculated explicitly (Eq. 12). Net growth rate, μ, is obtained by subtracting the respiration costs associated with chlorophyll maintenance and synthesis, R_Chl, and nutrient uptake, R_N, from the cellular gross growth rate, μ_g (Eq. (13)):

$$\mu = \mu_\text{g} - R_\text{Chl} - R_\text{N} = \mu_\text{net} - \zeta_\text{N} \cdot V \tag{7}$$

where ζ_N is the cost of N assimilation (Table 3) and R_Chl is the cost of chlorophyll synthesis and maintenance (Section 2.2.4).

$F_{DIC-Phy_\text{C}}$ is required only by the DA variant that explicitly resolves the dynamics of Phy_C (Eq. 1b). It is given by the product of net growth rate, μ with Phy_C, as is typical in quota models (Caperon, 1968; Droop, 1968):

$$F_{DIC-Phy_\text{C}} = \mu \cdot Phy_\text{C} \qquad\qquad \{\text{DA}\} \tag{8}$$

2.2 Flexibilities Represented by the Model Variants

We compare the behavior of three model variants that differ in their representation of the physiological flexibilities. These variants are:

Dynamic Acclimation' (DA): explicitly describes the dynamics of nitrogen and carbon bound to phytoplankton, and the acclimation mechanisms mentioned in the Introduction.

Table 1. Definitions, expansions/values and units of terms/symbols regarding the fluxes between model compartments.

Term/Symbol	Definition	Expansion/Value	Units
$F_{Phy_N-Det_N}$	N flux from Phy_N to Det_N	$m \cdot Phy_N^2$	$\mathrm{mmolN\,m^{-3}d^{-1}}$
$F_{Phy_C-Det_C}$	C flux from Phy_C to Det_C	$F_{Phy_N-Det_N}/Q$	$\mathrm{mmolC\,m^{-3}d^{-1}}$
F_{Det_N-DON}	N flux from Det_N to DON	$r_{\mathrm{hyd}} \cdot Det_N$	$\mathrm{mmolN\,m^{-3}d^{-1}}$
F_{Det_C-DOC}	C flux from Det_C to DOC	$r_{\mathrm{hyd}} \cdot Det_C$	$\mathrm{mmolC\,m^{-3}d^{-1}}$
$F_{DON-DIN}$	N flux from DON to DIN	$r_{\mathrm{rem}} \cdot DON$	$\mathrm{mmolN\,m^{-3}d^{-1}}$
$F_{DOC-DIC}$	C flux from DOC to DIC	$r_{\mathrm{rem}} \cdot DOC$	$\mathrm{mmolN\,m^{-3}d^{-1}}$
m	Mortality rate coefficient	0.1	$\mathrm{m^3 mmolN^{-1}d^{-1}}$
r_{hyd}	Hydrolysis rate constant	0.1	$\mathrm{d^{-1}}$
r_{rem}	Remineralization rate constant	0.1	$\mathrm{d^{-1}}$

Instantaneous Acclimation (IA): assumes that the nitrogen quota (i.e., molar N:C ratio) adjusts instantaneously to its optimal
130   value locally (i.e., at any point in time and space), but is otherwise identical to the DA variant with respect to the acclimation mechanisms.

'Fixed Stoichiometry' (FS) which assumes no physiological acclimation or quota variability whatsoever.

In the following, representations of the acclimative flexibilities by each model variant are explained in detail.

2.2.1 Flexibility I: Nutrient Quota

135 Flexibility in the elemental composition of phytoplankton (Q) is a result of acclimation processes, such as synthesis of enzymes or pigments, which differ in elemental composition (e.g. Geider and La Roche, 2002), in response to changes in resource (light and nutrients) availability.

DA: For the dynamic-acclimation variant, Q, is the ratio of the phytoplankton N and C state variables:

$$Q = \frac{Phy_N}{Phy_C} \qquad \text{\{DA\} (9)}$$

140 **IA:** For the instantaneous-acclimation variant, Q is assumed to adjust instantaneously to its balanced-growth optimum (Q^o) according to Pahlow and Oschlies (2013):

$$Q^o = \frac{Q_0}{2}\left[1 + \sqrt{1 + \frac{2}{Q_0(\hat{\mu}_{\mathrm{net}}/\hat{V} + \zeta_N)}}\right] \qquad \text{\{IA\} (10)}$$

Note that this solution differs slightly from the solution proposed by Smith et al. (2016), where the cost of chlorophyll maintenance and synthesis was ignored. (see Appendix 1 for details).

145 **FS:** In the fixed-stoichiometry variant, Q is a prescribed parameter (Table 2).

Table 2. Summary of differences between model variants. NA denotes not applicable. (*): prescribed, see Table 3. (**): we present our results in units of gChl gC^{-1} for the sake of consistency with literature values.

Term	Definition	Equation/Definition			Units
		IA	DA	FS	
Phy_C	Carbon bound to phytoplankton	Phy_N/Q	Eq. (1b)	Phy_N/Q	mmolC m^{-3}
$F_{DIN-Phy_N}$	N flux from DIN to Phy_N		Eq. (5)		mmolN m^{-3} d^{-1}
$F_{DIC-Phy_C}$	C flux from DIC to Phy_N	NA	Eq. (8)	NA	mmolc m^{-3} d^{-1}
f_V	Fractional allocation to uptake		Eq. (14)	(*)	–
V	Specific N uptake rate	Eq. (6)	Eq. (12)	Eq. (6)	molN molC^{-1} d^{-1}
μ	Cellular net growth rate		Eq. (7)		d^{-1}
μ_g	Cellular gross growth rate		Eq. (13)		d^{-1}
μ_{net}	$\mu_g - R_{Chl}$		Eq. (7)		d^{-1}
$\hat{\mu}_g$	Gross growth rate within chloroplast		Eq. (21)		d^{-1}
$\hat{\mu}_{net}$	Net growth rate within chloroplast		$\hat{\mu}_g - \hat{R}_{Chl}$		d^{-1}
Q	N quota	Eq. (10)	Phy_N/Phy_C	(*)	molN molC^{-1}
$\hat{V}$	Protoplast-specific N uptake rate		Eq. (16)		molN molC^{-1} d^{-1}
f_A	Fractional allocation to affinity		Eq. (18)	(*)	–
$\hat{\theta}$	Chl:C in chloroplasts		Eq. (26)	(*)	gChl molC^{-1} (**)
θ	Chl:C in the entire cell		Eq. (24)		gChl molC^{-1} (**)
R_N	Cost of N uptake		Eq. (7)		d^{-1}
$\hat{R}_{Chl}$	Cost of light harvesting within chloroplast		Eq. (23)		d^{-1}
R_{Chl}	Cellular cost of light harvesting		Eq. (25)		d^{-1}
f_C	Fractional N allocation to C-fixation		Eq. (11)	NA	–
L_N	Nutrient limitation term		NA	Eq. (15)	–
L_I	Light limitation		Eq. (22)		–

2.2.2 Flexibility II: growth vs nutrient uptake

Given the high nitrogen content in the enzymes responsible for both CO_2 fixation and nutrient uptake and assimilation (Geider and La Roche, 2002), we consider a trade-off in the allocation of nitrogen between carbon fixation and nutrient uptake for the acclimative variants, whereas this trade-off is ignored for the FS variant.

150   **DA & IA:** For the acclimative variants, following Pahlow and Oschlies (2013), the trade-off is specified in terms of the fraction of cellular nitrogen reserves allocated to nitrogen uptake (f_V), which linearly increases V, and decreases μ_g, through decreasing the resources available for carbon fixation (equivalently, relative size of the chloroplast, following Pahlow

and Oschlies (2013)), f_{C}:

$$f_{\mathrm{C}} = \left(1 - \frac{Q_0}{2Q} - f_{\mathrm{V}} \right) \qquad \{\text{IA, DA}\} \quad (11)$$

155 where f_{V} is the fractional allocation towards nutrient uptake:

$$V = f_{\mathrm{V}} \cdot \hat{V} \qquad \{\text{IA, DA}\} \quad (12)$$

where $\hat{V}$ is the protoplast-specific N uptake rate (see below). The cellular gross growth rate is then determined by scaling the gross growth rate within the chloroplast $\hat{\mu}_{\mathrm{g}}$ (see Section 2.2.4) by the relative size of the chloroplast, f_{C}:

$$\mu_{\mathrm{g}} = f_{\mathrm{C}} \cdot \hat{\mu}_{\mathrm{g}} \qquad (13)$$

160 Note that, for calculating the effective flux from DIN to Phy_{N} (Eq. (5)), only the DA variant uses V as calculated by Eq. (12), while the IA variant calculates the uptake rate from the growth rate, based on the balanced growth assumption (Eq. (6)). However, the IA variant still needs the V as calculated by Eq. (12), for calculating the costs of nutrient uptake (Eq. (7)).

Both acclimative variants assume that f_{V} maximises the net specific growth rate under balanced growth conditions.
165 Following Pahlow and Oschlies (2013), this optimal value is found as (see Appendix 1):

$$\frac{\mathrm{d}\mu}{\mathrm{d}f_{\mathrm{V}}} = 0 \quad \Rightarrow \quad f_{\mathrm{V}} = \left(\frac{Q_0}{2Q} \right) - \zeta_{\mathrm{N}}(Q - Q_0) \qquad \{\text{IA, DA}\} \quad (14)$$

FS: For the fixed-stoichiometry variant, the gross growth rate, μ_{g} is obtained by the multiplication of $\hat{\mu}_{\mathrm{g}}$, for FS, interpreted as the light-limited potential growth rate, with a nutrient limitation term L_{N}, formulated as a hyperbolic function of ambient DIN concentration, following the Michaelis-Menten-Monod model (Johnson and Goody, 2011; Monod, 1949):

170 $$\mu_{\mathrm{g}} = \hat{\mu}_{\mathrm{g}} \cdot L_N = \hat{\mu}_{\mathrm{g}} \cdot \frac{DIN}{K_{\mathrm{N}} + DIN} \qquad \{\text{FS}\} \quad (15)$$

Thus, for the FS variant, μ (Eq. (7)), and hence, through the balanced growth assumption, V (Eq. (6)) are directly linked to the external nutrient concentration (Eq. (15)) as in typical fixed-stoichiometry models. Given the fact that both L_{N} (Eq. 15) for the FS variant and f_{C} (Eq. 11) for the acclimative variants have an equivalent role (in scaling $\hat{\mu}_{\mathrm{g}}$ to μ_{g}), and they both represent nutrient limitation, we consider them to be comparable, i.e., $L_{\mathrm{N}} \sim f_{\mathrm{C}}$.

175 ### 2.2.3 Flexibility III: nutrient affinity vs. maximum uptake rate

DA & IA: The protoplast-specific N uptake rate, $\hat{V}$ can be described by a function of maximal uptake rate, $\hat{V}_{\mathrm{max}}$, and nutrient affinity, $\hat{A}$:

$$\hat{V} = \frac{\hat{V}_{\mathrm{max}} \cdot \hat{A} \cdot DIN}{\hat{V}_{\mathrm{max}} + \hat{A} \cdot DIN} \qquad \{\text{IA, DA}\} \quad (16)$$

The acclimation variants introduce a trade-off between affinity vs. maximum uptake rate. This trade-off is captured by
the fractional allocation of resources to affinity (f_A), which increases affinity, $\hat{A} = f_A \hat{A}_0$, while decreasing maximum
uptake rate, $\hat{V}_{\max} = (1 - f_A)\hat{V}_0$, so that Eq. (16) becomes:

$$\hat{V} = \frac{(1 - f_A)\hat{V}_0 \cdot f_A \hat{A}_0 \cdot DIN}{(1 - f_A)\hat{V}_0 + f_A \hat{A}_0 \cdot DIN} \qquad\qquad \{\text{IA, DA}\} \quad (17)$$

f_A is set to its optimum value, which maximizes $\hat{V}$, and hence also V (Pahlow, 2005):

$$\frac{\mathrm{d}\hat{V}}{\mathrm{d}f_A} = 0 \quad \Rightarrow \quad f_A = \left[1 + \sqrt{\frac{\hat{A}_0 \cdot DIN}{\hat{V}_0}}\right]^{-1} \qquad\qquad \{\text{IA, DA}\} \quad (18)$$

FS: The fixed-stoichiometry variant ignores this trade-off entirely, by describing the nutrient limitation with the Michaelis-
Menten-Monod function (Eq. (15)). Following Button (1978) and Smith et al. (2009), the K_N parameter in Eq. (15), can
be expressed as a function of $V_{\max}$ and $\hat{A}$, according to:

$$K_N = \frac{\hat{V}_{\max}}{\hat{A}} = \frac{(1 - f_A) \cdot \hat{V}_0}{f_A \cdot \hat{A}_0} \qquad\qquad \{\text{FS}\} \quad (19)$$

Based on Eq. (19), corresponding K_N values were diagnosed from the solution of the IA variant (i.e., using the locally
optimized f_A values as calculated with Eq. (18), and $\hat{A}_0$ and $\hat{V}_0$ parameters specified for the IA and DA variants). The
biomass-weighted spatio-temporal average K_N value so obtained was prescribed for the FS variant (Table 3).

2.2.4 Flexibility IV: photoacclimation

Photo-acclimation is based on the net carbon fixation rate within the chloroplast, $\hat{\mu}_{\text{net}}$ (equivalent to $\mathscr{A}$ in Pahlow and Oschlies
(2013)), which is obtained by subtracting the chloroplast specific synthesis and maintenance costs of chlorophyll, from the
gross growth rate within the chloroplast, i.e.,

$$\hat{\mu}_{\text{net}} = \hat{\mu}_g - \hat{R}_{\text{Chl}} \qquad\qquad (20)$$

where, $\hat{\mu}_g$ is given by the product of daylength as a fraction of 24 h,, L_D, potential turnover rate, $\hat{\mu}_0$, and the light-saturation of
the photosynthetic apparatus, L_I:

$$\hat{\mu}_g = L_D \hat{\mu}_0 L_I \qquad\qquad (21)$$

L_I is a saturating function of daytime average light, $\bar{I}$, and chlorophyll density in chloroplasts, $\hat{\theta}$:

$$L_I = 1 - \exp\left(\frac{-\alpha \hat{\theta} \bar{I}}{\hat{\mu}_0}\right) \qquad\qquad (22)$$

where α is light affinity. Returning to Eq. (20), $\hat{R}_{\text{Chl}}$ is given by:

$$\hat{R}_{\text{Chl}} = \left(\hat{\mu}_g + R_M^{\text{Chl}}\right) \zeta_{\text{Chl}} \hat{\theta} \qquad\qquad (23)$$

where $R_{\mathrm{M}}^{\mathrm{Chl}}$ and ζ_{Chl} are the costs of chlorophyll maintenance and synthesis, respectively.

Photo-acclimation is mainly represented in terms of the chlorophyll density in chloroplasts, $\hat{\theta}$. Increasing $\hat{\theta}$ reduces light limitation (Eq. (22)), but at the expense of greater respiration costs (Eq. (23)). In turn, for obtaining the cellular Chl:C ratio, θ, is calculated by multiplying $\hat{\theta}$ times f_{C}, i.e., size of the chloroplast:

$$\theta = f_{\mathrm{C}} \cdot \hat{\theta} \qquad \text{\{IA,DA\} (24)}$$

Similarly, the overall respiratory cost of maintaining cellular chlorophyll is obtained by multiplying the chloroplast-specific cost by the size of the chloroplast:

$$R_{\mathrm{Chl}} = f_{\mathrm{C}} \cdot \hat{R}_{\mathrm{Chl}} \qquad \text{\{IA,DA\} (25)}$$

Although θ (Eq. 24) is only a diagnostic quantity, R_{Chl} (Eq. 25) directly determines the net growth rate through Eq. (7). Therefore, scaling of the chloroplast-specific respiration rate, $\hat{R}_{\mathrm{Chl}}$ by f_{C} can considered to be an acclimative quality implied by variable f_{V} and Q, which, in combination (Eq. 11), determine the chlorophyll maintenance cost through Eq. (25)).

IA & DA: In the acclimation variants, $\hat{\theta}$ is assumed to adjust instantaneously to its optimal value, which maximizes $\hat{\mu}_{\mathrm{net}}$. Following Pahlow et al. (2013) this optimal value is:

$$\hat{\theta} = \begin{cases} \dfrac{1}{\zeta_{\mathrm{Chl}}} + \dfrac{\hat{\mu}_0}{\alpha \bar{\bar{I}}}\left(1 - W_0\left[\left(1 + \dfrac{R_{\mathrm{M}}^{\mathrm{Chl}}}{L_{\mathrm{D}}\hat{\mu}_0}\right)\exp\left(1 + \dfrac{\alpha \bar{I}}{\hat{\mu}_0 \zeta_{\mathrm{Chl}}}\right)\right]\right), & \bar{I} > \bar{I}_{\mathrm{C}} \\ 0, & \bar{I} \leq \bar{I}_{\mathrm{C}} \end{cases} \qquad \text{\{IA, DA\} (26)}$$

where W_0 is the 0-branch of the Lambert-W function, $\bar{I}$ is the daytime average irradiance (i.e., $\hat{I} = \bar{I}_{\mathrm{24h}}/L_{\mathrm{D}}$) and $\bar{I}_{\mathrm{C}}$ is the critical daytime average irradiance level, above which chlorophyll synthesis is worthwhile (Pahlow et al., 2013):

$$\bar{I}_{\mathrm{C}} = \frac{\zeta_{\mathrm{Chl}} R_{\mathrm{M}}^{\mathrm{Chl}}}{\alpha L_{\mathrm{D}}} \qquad (27)$$

FS: For the fixed-stoichiometry variant, $\hat{\theta}$ is prescribed as the biomass-weighted average value calculated by the IA variant. Considering that θ is typically a constant 'conversion factor' in classical, fixed stoichiometry and fixed Chl:C models, in Eqs. (24) and (25), we assume that the size of the chloroplast, f_{C}, is constant too. For the sake of consistency with the IA variant, f_{C} for FS is diagnosed from its expanded form, i.e., $1 - \frac{Q_0}{2Q} - f_{\mathrm{V}}$ (Eq. 11). Hence, in addition to the prescribed value of Q (see Section 2.2.1), the biomass-weighted mean of f_{V}, as calculated by the IA variant is prescribed (Table 3). Given the comparability of the terms (Section 2.2.2), diagnosing f_{C} from L_{N} comes into question, which is elaborated in Appendix B.

2.2.5   Temperature Scaling

Kinetic rate constants: m, r_{hyd}, r_{rem} in Table 1, and $\hat{V}_0$, $\hat{A}$, $\hat{A}_0$ and $R_{\mathrm{M}}^{\mathrm{Chl}}$ in Table 3 are prescribed for a reference temperature of $T_{\mathrm{r}} = 20\,°\mathrm{C} = 293.15\,\mathrm{K}$, and scaled to the ambient temperature in water, T (in K), according to the Arrhenius function:

$$f(T) = \exp\left(\frac{-E_{\mathrm{a}}}{R}\left[\frac{1}{T} - \frac{1}{T_{\mathrm{r}}}\right]\right) \qquad (28)$$

Table 3. Descriptions, values and units of model parameters regarding phytoplankton growth. Prescribed values for Q, K_N, f_V, and are based on the biomass-weighted averages estimated by the IA variant. All other parameter values are taken from Pahlow et al. (2013) and Smith et al. (2016).

Term/Symbol	Definition	Value	Unit	Used by
$\hat{\mu}_0$	Potential maximum growth rate	5.0	d^{-1}	all
Q_0	Subsistence quota	0.039	$\mathrm{mmolN\,molC}^{-1}$	IA, DA
$\hat{A}_0$	Potential maximum nutrient affinity	0.1	$\mathrm{m}^3\,\mathrm{mmolC}^{-1}\,\mathrm{d}^{-1}$	IA, DA
$\hat{V}_0$	Potential maximum N uptake rate	5.0	$\mathrm{molN\,molC}^{-1}\,\mathrm{d}^{-1}$	IA, DA
α	Chl-specific slope of P-I curve	1.0	$\mathrm{m}^2\,\mathrm{E\,molCgChl}^{-1}\,\mathrm{d}^{-1}$	all
$R_\mathrm{M}^\mathrm{Chl}$	Cost of chlorophyll maintenance	0.1	d^{-1}	all
ζ_Chl	Cost of chlorophyll synthesis	0.5	$\mathrm{mmolC\,gChl}^{-1}$	all
ζ_N	Cost of N uptake	0.6	$\mathrm{molC\,molN}^{-1}$	all
Q	N quota	0.084	$\mathrm{molN\,molC}^{-1}$	FS
K_N	Half saturation constant for N uptake	4.84	$\mathrm{mmolN\,m}^{-3}$	FS
f_V	Fractional allocation to uptake	0.32	–	FS
$\hat{\theta}$	Chl:C in chloroplasts	0.518	$\mathrm{gChl\,molC}^{-1}$	FS

where the gas constant $R = 8.3145\,\mathrm{J\,mol}^{-1}\mathrm{K}^{-1}$, and the activation energy, $E_\mathrm{a} = 4.82\cdot10^4\,\mathrm{J\,mol}^{-1}$, such that every 10 degrees increase/decrease in T approximately doubles/halves the reference rates.

2.3   Coupling with the Hydrodynamical Host

The model is implemented in the Framework for Aquatic Biogeochemical Models (i.e., FABM, Bruggeman and Bolding,
235 2014), so that it can be used, without modification, in combination with various hydrodynamical hosts. In this study, we performed simulations of an idealized water column, using the General Ocean Turbulence Model (i.e., GOTM Burchard et al., 2006). GOTM calculates and provides the relevant physical quantities, such as T (Eq. 23) and I (Eq. 17). I is attenuated with depth (z) by various substances in water, according to:

$$I(z) = I_0 \left[A \exp\left(\frac{-z}{\eta_1} \right) + (1 - A) \exp\left(\frac{-z}{\eta_2} - \int_z^0 \sum_i k_i c_i(z')\mathrm{d}z' \right) \right] \tag{29}$$

240 where A, η_1 and η_2 represent the differential attenuation length scales of red and blue light (Burchard et al., 2006), and k_i is the specific attenuation coefficient of the biological quantities, which we set as $0.03\,\mathrm{m}^2\,\mathrm{mmolN}^{-1}$ for Phy_N and Det_N. In order to account for background attenuation, we set the 'light extinction method' to 'Jerlov Type IB', corresponding to $A = 0.67$ $\eta_1 = 1.0\,\mathrm{m}$, $_2 = 17\,\mathrm{m}$, characterizing water of medium clarity (Paulson and Simpson, 1977). Our results are qualitatively

insensitive to these parameter settings. Besides providing necessary environmental variables, GOTM calculates the transport
245   rates of the biological quantities, according to the general equation (Burchard et al., 2006):

$$\frac{\partial c_i}{\partial t} + \frac{\partial}{\partial z}\left(w_i c_i - K_z \frac{\partial c_i}{\partial z}\right) = s(c_i) \tag{30}$$

where, K_Z is the eddy diffusivity calculated by GOTM, the source terms, $s(c_i)$ correspond to Eqs. (1-4) and advection rates,
w_i are all set to 0.0, except that of detritus for which a sinking rate of -2.0 m d^{-1} was specified.

2.4   Idealized Setup and Simulations

250   We consider an idealized water column of 100 m depth. In order to mimic an environment that is characterized by strong
seasonality, with deep mixed layers in spring and summer stratification, we force the model with astronomically calculated
short wave radiation at 60°N latitude, and a repeating annual cycle of air temperature that ranges between 4–20 °C as described
by a scaled sinusoidal function (Fig. 2).
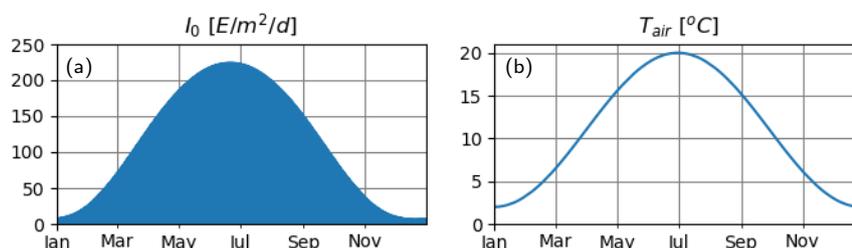
Figure 2. Atmospheric variables. (a) astronomically estimated instantaneus irradiance at the water surface, and (b) prescribed air temperature.

    All other meteorological conditions are assumed constant, and the model ignores precipitation and evaporation losses, as
255   well as tidal variations. Starting from initial conditions, and annually repeating meteorological forcing as described above,
each model variant was run for 3 years. The 3rd year results were nearly identical to those for the 2nd year, indicating that an
equilibrium annual cycle was reached. In the following, we elaborate the seasonal dynamics during the 3rd year.

3   Results

Daytime-averaged irradiance, $\bar{I}$ and water temperature T simulated by different model variants are very similar with subtle
260   differences (Fig. 3a,d vs. b,e, vs. c,f), because each variant calculates slightly different phytoplankton biomass (see below),
resulting in differences in attenuation of light and associated heating. Seasonal and vertical distributions of DIN as estimated
by the model variants are similar (Fig. 3g-i). DIN depletion (<1 mmolN m^{-3}) during summer is confined to the upper 25 m as
estimated by the FS variant, whereas it extends 5-10 m deeper as estimated by the IA and DA variants.

    With all three model variants phytoplankton growth patterns are characterized by an intense surface bloom in spring, fol-
265   lowed by a gradual deepening of the biomass maxima (Fig. 4a-c). Biomass concentration as estimated by the IA and DA
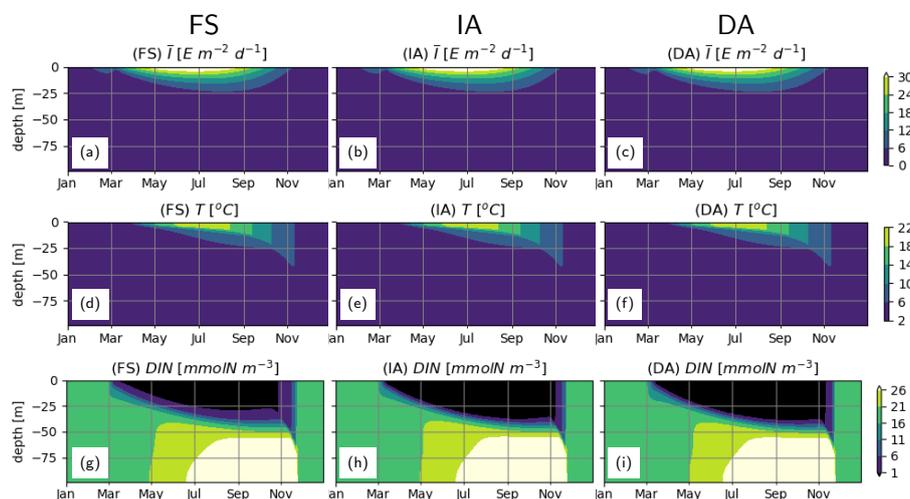
Figure 3. Abiotic environment. (a-c) daytime-averaged photosynthetically active radiation, $\bar{I}$ [E m^{-2} d^{-1}], (d-f) water temperature T [°C], and (g-i) DIN [mmolN m^{-3}], as simulated by the FS (left); IA (center) and DA (right) variants.

variants during summer is considerably greater, with steeper gradients, than with the FS variant (Fig. 4a,b,c). Compared to the FS variant, the acclimation response in the other two variants tends to produce steeper gradients over both depth and time, because of combined dependencies on the three dynamically optimized allocation factors (f_A, f_V, and $\hat{\theta}$). This effect is most pronounced for Phy_{Chl}, which differs the most between the FS and the other two variants. With the FS variant, given the

270 constant N:C (Q) and cellular Chl:C (θ) (Fig. 4g,m), C, N and Chl bound to phytoplankton clearly display identical patterns (Fig. 4a,d,j; note that apparent differences in contour plots are due to contour limits not matching these ratios). IA and DA on the other hand simulate slightly different patterns for C, N and Chl bound to phytoplankton (Fig. 4b,e,k and c,f,l), because of the seasonally and vertically variable Chl:C:N. Decoupling of Phy_N from Phy_C is mainly monotonic, and is driven by increasing Q with depth (Fig. 3h-i). On the other hand, decoupling of Phy_{Chl} from Phy_C follows a more complex pattern,

275 because of the uni-modal distribution of θ across the water column (Fig. 4n-o). As a result of this uni-modality, Chl simulated by the IA and DA variants forms a distinct, thin layer below the thermocline (Fig. 4k-l).

During summer, $\hat{\theta}$ follows a complex, but roughly uni-modal distribution across depth (Fig. 5b-c): intermediate values at the surface first increase with depth to reach a maximum and then sharply decrease with increasing depth. The low values of $\hat{\theta}$ towards the surface reflect the optimization, which reduces pigment density when light is abundantly available because

280 of the chloroplast-specific respiratory costs $\hat{\theta}$ (Eq. 23). This can be seen in the flattening of the light-saturation function L_I (Eq. 22). In the deep layers, as $\bar{I}$ approaches $\bar{I}_C$, irradiance becomes insufficient to support the synthesis and maintenance of chlorophyll, and $\hat{\theta}$ rapidly converges to 0. f_A and f_V simulated by the IA and DA variants (Fig. 5e-f,h-j) increase with nutrient limitation (Fig. 5n-o) as expected (Smith et al., 2016). The fraction of resources available for carbon fixation, f_C, displays a similar pattern in all model variants and is roughly the inverse of f_V: high during winter throughout the water column and in the

285 deeper layers during summer, low in the upper layers during summer (Fig. 5j-l). For the FS variant, the pattern of the nutrient
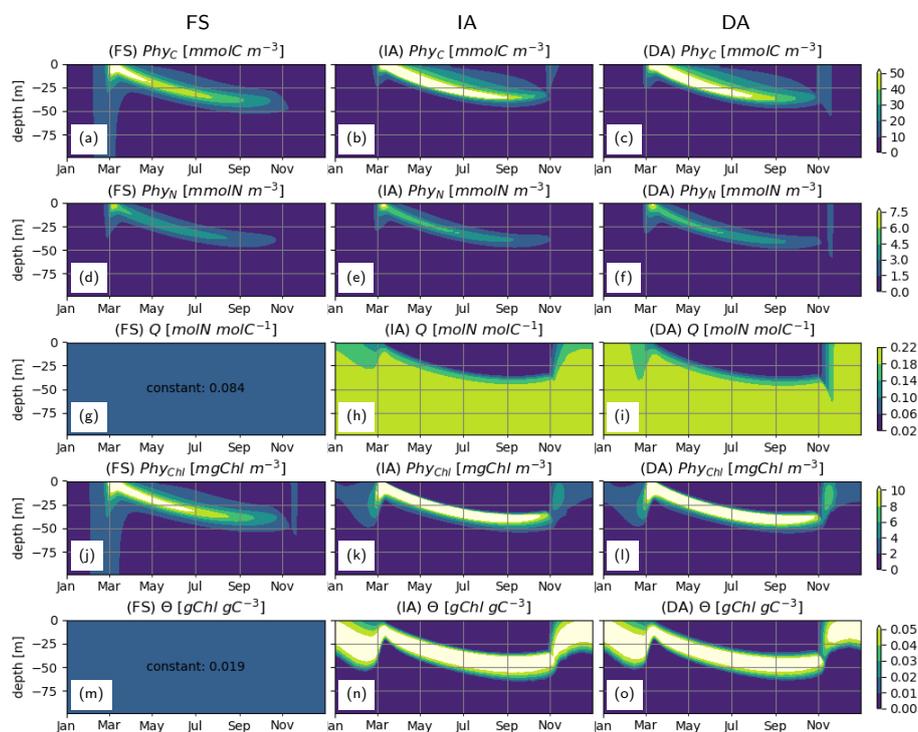
Figure 4. Phytoplankton C, N and Chl concentrations: (a-c) Phy_C [mmolC m^{-3}], (d-f) Phy_N [mmolN m^{-3}], (j-l) Phy_{Chl} [mgC m^{-3}]; and phytoplankton N:C (Q) and Chl:C (Θ) ratios: (g-i) Q [molN molC^{-1}] and (m-o) Θ [gChl gC^{-1}], as simulated by the FS (left); IA (center) and DA (right) variants.

limitation term, L_N, is similar to the patterns of f_C for IA and DA variants, Fig.5, although its magnitude in the summer is higher than other variants, as can be explained by the incomplete DIN depletion (Fig.3g). Light saturation of photosynthesis, L_I, displays a similar pattern in all variants (Fig. 5m-o), and mainly reflects irradiance levels (Fig. 3a-c). However, compared to the FS variant, the intermediate L_I values in the IA and DA variants penetrate deeper (Fig. 5n,o vs m), because the optimization
290   of $\hat{\theta}$ enhances light harvesting ability at these intermediate depths (Fig. 5b,c).

The IA and DA variants estimate higher net growth rates, μ, and nutrient uptake rates, v_N, in surface layers during the spring bloom, and in deeper layers during summer (Fig. 6a-c,d-f). Negative v_N in the bottom layers as estimated by the FS and IA variants is a direct result of the balanced growth assumption (Eq. 6) and can be interpreted as exudation. Respiratory costs of nutrient uptake, R_N, (Fig. 6h-i) are much lower than the chlorophyll maintenance and synthesis costs, R_{Chl} (Fig. 6j-l). High
295   R_{Chl} as estimated by the FS variant in the surface layers during the growing season is the cause of the relatively low μ estimated by this variant (Fig. 6j vs k-l): The IA and DA variants achieve much lower R_{Chl} by reducing $\hat{\theta}$ (Fig. 5b-c) at the surface, and low Q (Fig. 3h-i) and high f_V at the surface (Fig. 5h-i) further reduce θ (Eq. 24), and hence R_{Chl} (Eq. 25). For the FS variant, R_N drops below 0, implying negative respiration, which is a model artefact, as a result of $\hat{\mu}_{net}$ becoming negative (see Eq. A4
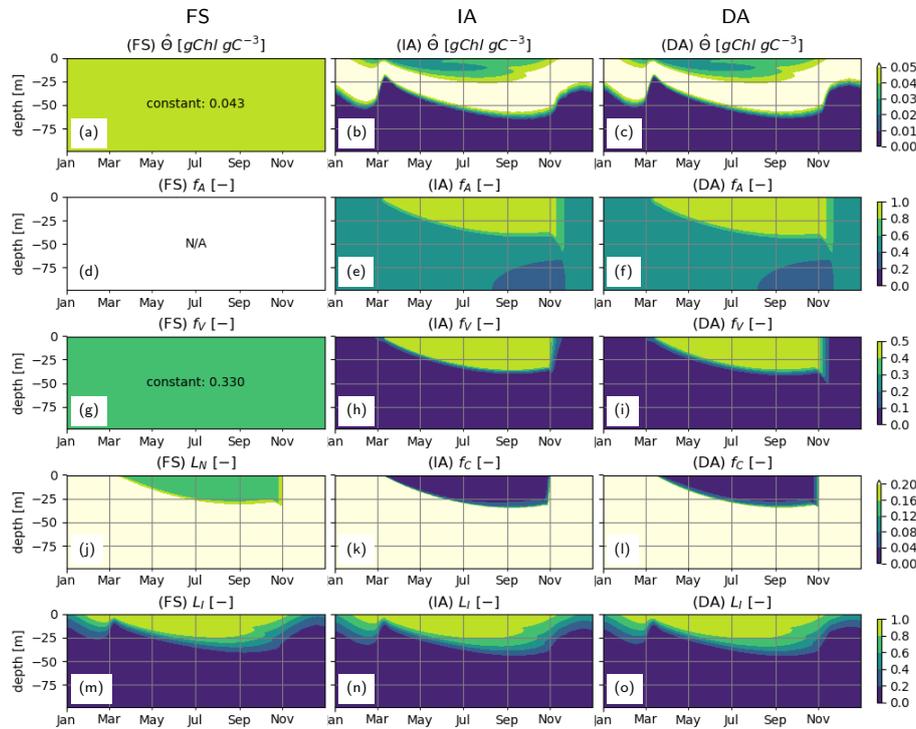
Figure 5. Phytoplankton physiological variables. (a-c) Chlorophyll density in chloroplasts, $\hat{\Theta}$ [gChl gC^{-1}]; (d-f) fractional allocation to affinity, f_A [-]; (g-i) fractional allocation to nutrient uptake, f_V [-]; nutrient limitation term of the FS variant, L_N [-] (j) and fractional allocation to carbon fixation of the IA and DA variants, f_C [-] (k-l); and (m-o) light saturation of photosyntehsis, L_I [-] as simulated by the FS (left); IA (center) and DA (right) variants.

in Sect.A1) due to the fixed $\hat{\theta}$. However these negative values are small, and therefore do not have a significant effect on the model results, as evidenced by a sensitivity experiment, where $\hat{\mu}_{\mathrm{net}}$ was constrained to positive values for the FS variant.

For the most part, primary production and relevant dynamics take place within roughly the upper 50m in the simulated system (Figs. 4-6). A comparison of average quantities in this zone, as estimated by the three model variants (Fig. 7) reveals subtle differences between model variants that are not resolved by the contour plots (Fig. 4–6). In both the IA and DA variants, DIN concentrations are almost entirely depleted before the onset of winter mixing, whereas in the FS variant DIN remains substantially higher (>5 mmolN m^{-3}) (Fig. 7a). Q and f_C, as estimated by the IA and DA variants are nearly identical throughout the season (Fig. 7b,c), but slight differences arise during winter. For DA, Phy_C and Phy_N, hence Q, become vertically homogeneous due to rapid turbulent mixing (Fig. 4c,f,i). However under the instantaneous acclimation assumption in the IA variant, no matter how well mixed the water column may be, vertical gradients persist for the optimal Q values (Eq.9) between the surface and bottom layers (Fig. 4h). L_N as estimated by the FS variant as a direct function of DIN (Eq. 15) is lower than the functionally equivalent f_C estimated by the IA and DA variants, based on Q and f_V (Eq. 11) (Fig. 7c), except in the late summer where the DIN as estimated by FS becomes considerably higher relative to the other two variants. This reflects the multiplicative light
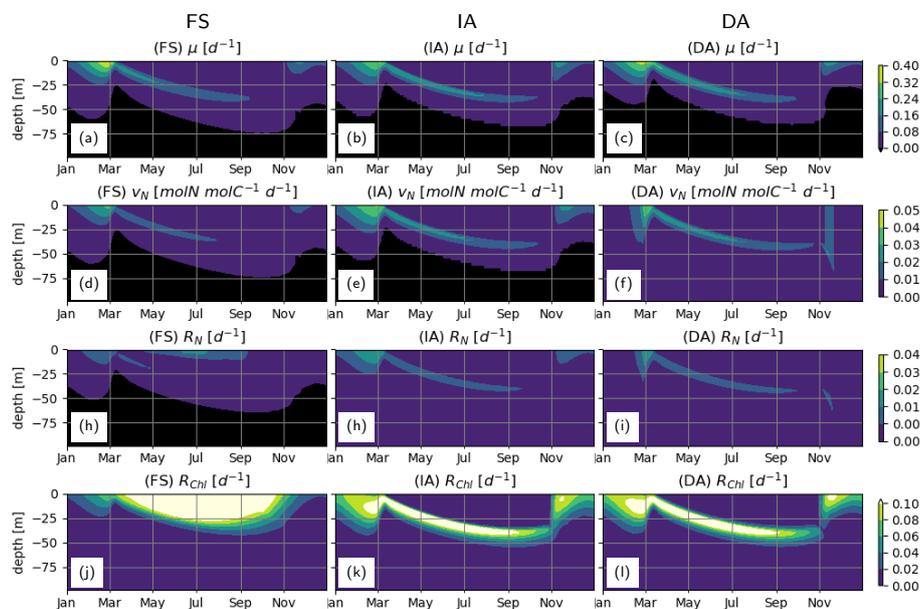
Figure 6. Phytoplankton growth, uptake and respiration rates. (a-c) Net growth rate, μ [d^{-1}], (d-f): specific uptake rate, v_N [mmolN mmolC^{-1} d^{-1}] and respiration costs of (g-i) N uptake, R_N [d^{-1}] and (j-l) chlorophyll maintenance and synthesis, R_{Chl} [d^{-1}] as simulated by the FS (left); IA (center) and DA (right) variants.

and temperature dependencies in the FS variant, compared to their interactive effects in the optimally-acclimating DA and IA variants. Compared to simple multiplicative dependencies, dynamic resource allocation allows faster growth and hence nutrient uptake under a broad range of conditions. This is because the continual re-balancing of intra-cellular resource allocation can to some degree ameliorate light (nutrient) limitation to the degree that nutrient (light) is replete (see Fig. 3 of Smith et al., 2016).

All three model variants produce similar Phy_{C} during winter, but important differences arise during the spring bloom (Fig. 7d), and especially during summer, when the FS variant produces considerably lower values compared to the IA and DA variants (Fig. 7d). Short-lived differences between the IA and DA variants during the spring and autumn blooms are attributable to a number of subtle differences in the two scenarios, foremost the unequal Q under increased vertical mixing as mentioned above, slight differences in irradiance levels as a result of attenuation by differing Phy_{N} concentrations (Eq. 29) during the spring bloom, and a higher Phy_{C} below 50m as estimated by DA just before the autumn bloom (not shown), which results in a larger entrainment of phytoplankton in the surface layers with the weakening of stratification (Fig. 3).

C:N of detritus as estimated by the FS variant approaches a constant equilibrium value throughout the water column by the end of the first year, and remains there during the third year (Fig. 8a,d). This is as expected, and this value is simply equal to the reciprocal of the prescribed constant (N:C) quota of phytoplankton, calculated as the biomass-weighted average of the Q estimated by the IA variant (Table 3). The C:N ratio of detritus, as estimated by the IA and DA variants, increases during summer (Fig. 8b,c ande,f), driven by the lower phytoplankton quotas during summer (Fig. 4).
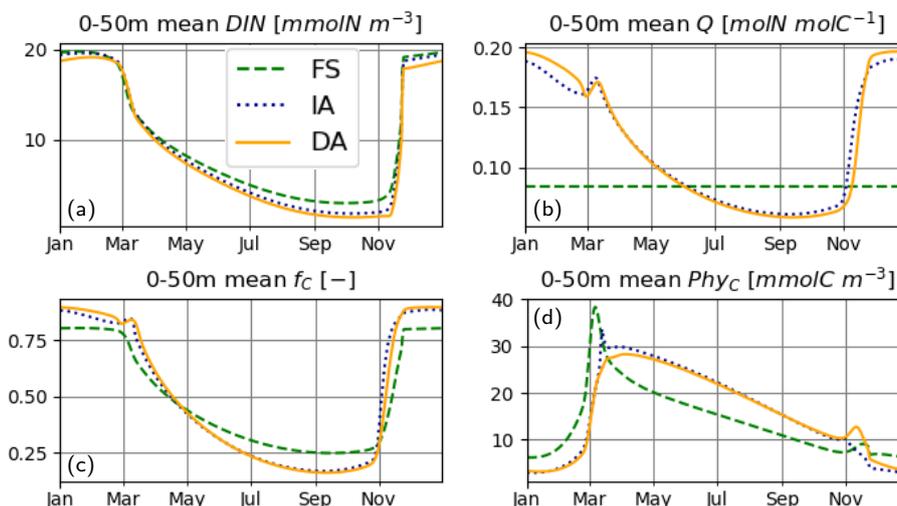
Figure 7. Upper 50m averages of critical variables. (a) DIN [mmolN m^{-3}], (b) phytoplankton Q [mmolN mmolC^{-1}], (c) resources available for carbon fixation, f_C [-] (L_N [-] for FS), and (d) Phy_C [mmolC m^{-3}], as simulated by the FS (dashed magenta line), IA (fine-dashed dark blue line) and DA (continuous orange line) variants.
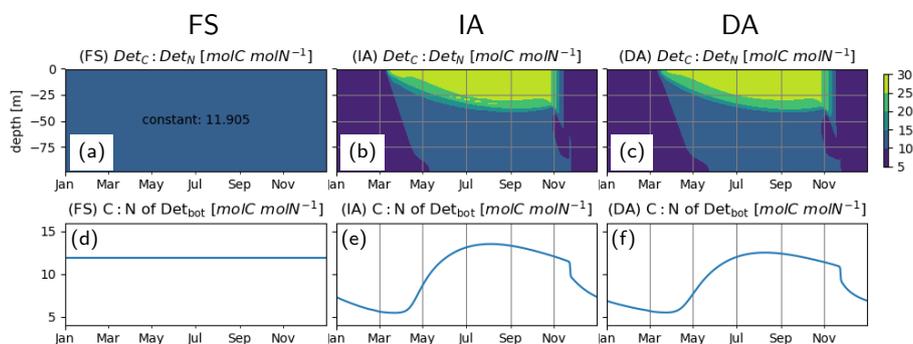
Figure 8. Detrital C:N [molC molN^{-1}] (a-c) in the entire water column, and (d-f) in the bottom layer (d-f), as simulated by the FS (left); IA (center) and DA (right) variants.

4 Discussion

4.1 Modelling variable phytoplankton composition

330   Elemental composition and pigment density of phytoplankton are known to vary, at both the organismal and community levels (Halsey and Jones, 2015), as demonstrated in the laboratory and under *in-situ* conditions (Moreno and Martiny, 2018). Such variations in phytoplankton and hence detrital C:nutrient ratios have implications for C and nutrient export fluxes, including the functioning of the biological carbon pump in the ocean. Notwithstanding, in many biogeochemical models coupled to GCM's, primary producers are still unrealistically represented with a constant 'Redfield' C:N:P ratio, and/or constant Chl:C

ratio (see, e.g. the models used in Laufkötter et al., 2015). More detailed 'Quota' models exist; however, these approaches are often challenged by two major limitations: *i*) dependence on formulations that lack a clear mechanistic basis, and *ii*) their requirement for additional state variables, which increase computational costs.

A concrete example of the first problem, i.e., dependence on heuristic formulations, is the down-regulation of nutrient uptake, which is needed to avoid unrealistically high nutrient quotas in a Droop scheme. Often, down-regulation is formulated as some function (linear, e.g., Grover (e.g. 1991) or non-linear, e.g., Geider et al. (1998)) of 'relative quota', with reference to a prescribed maximum value. The acclimation scheme used in this study (IA and DA variants), requires no such explicit down-regulation term, nor any prescribed maximum quota value. This is because the optimization of growth, subject to the growth vs nutrient uptake trade-off (sect. 2.2.2), accomplishes this regulation by balancing the marginal benefits of investing into nutrient uptake versus photosynthesis. This RAM approach, which links various cellular functions via trade-offs, has proven successful at reproducing various Chl:C:N:P measurements obtained in laboratory experiments (e.g. Klausmeier et al., 2004; Pahlow et al., 2013; Wirtz and Kerimoglu, 2016). Furthermore, given its mechanistic basis, this approach can be expected to reproduce biological feedbacks more realistically (Flynn et al., 2015), and thereby improve the generality and portability of models (Anugerahanti et al., in prep.). Recent applications of these models in 3D setups with realistic forcings (Kerimoglu et al., 2017; Pahlow et al., 2020) have indicated that accounting for acclimation enhances the ability of models to reproduce field observations. Moreover, a consistent representation of phytoplankton composition allows identification of potential alterations in trophic transfer efficiencies as mediated by changes in food quality of prey in response to environmental change (Kerimoglu et al., 2018; Kwiatkowski et al., 2018).

Regarding the second problem, i.e., the computational costs of resolving additional state variables, Smith et al. (2016) proposed the 'Instantaneous Acclimation' approach, which is nearly identical to the IA variant we presented here (see sect. 2.2.1). As in Smith et al. (2016), we considered the same specific acclimation mechanisms of (Pahlow et al., 2013), but under the assumption that the N quota adjusts instantaneously to an optimal value locally, under strictly 'balanced growth' (Burmaster, 1979). While at steady-state, this is a natural consequence of any 'Droop-like' model (Burmaster, 1979), assuming this behavior to hold under transient conditions is merely an approximation. Ward (2017), using a classical Droop-approach, showed that this approximation holds well under a wide range of conditions in a 0D (box) setup. Here, in an idealized 1D setup, we have shown that the original (i.e., fully acclimative) IA model behaves very similarly to the DA variant. Our preliminary experiments demonstrated that, even in an envrionment characterized by periodic perturbations of stratification during summer, behavior of the two variants remain similar (results not shown). This is significant, considering that IA requires only 1 state variable, whereas DA requires 2 state variables. Thus, it can be concluded that IA provides improved realism over the computationally equivalent FS approach. For simulating a few years of the dynamics of the single phytoplankton group in a 1D setup as we did here, differences in computational costs relative to the fully dynamic variant are nearly negligible, but for simulating decades/centuries or millennia in a 3D setup (e.g., as in Pahlow et al., 2020), and/or when multiple clones/types are considered (e.g., 350 in Dutkiewicz et al., 2020), differences in computational costs can indeed be substantial.

4.2 Qualitative versus quantitative differences between model variants

The Capacity to store nutrients is known to be an advantageous trait for phytoplankton in temporally fluctuating environments,
370   where greater nutrient storage capacity, e.g., by larger cells, during the nutrient-replete phase provides a competitive advantage during subsequent periods of nutrient scarcity (Grover, 1991; Litchman et al., 2009). Similarly, diffusion or active movement of nutrient-rich cells from the nutrient-replete to nutrient-rich environments, e.g., from bottom towards surface layers, has been shown to favor species with greater storage capacities (Kerimoglu et al., 2012; Grover, 2017). The IA model presented in this study cannot capture this effect like the DA variant, since in the former, immediate acclimation precludes the nutrients taken
375   up at a nutrient-rich environment to be used later, in a nutrient-poor environment. This may indeed be why the DA variant's Phy_C gradually exceeds that of the IA variant during summer (Fig. 7d) as the DIN gets depleted (Fig. 7a). However, under the quasi-realistic conditions we considered here, this effect is not large; i.e., the two acclimative variants simulate very similar phytoplankton biomass (Fig. 4) In fact, it is possible that the DA variant reproduces a weaker 'storage' advantage compared to a classical 'Droop' model (Grover, 1991; Kerimoglu et al., 2012). This is because in the latter nutrient uptake is only a function
380   of relative quota as explained above, whereas in our acclimative approach, just below the thermocline, where nutrients are plentiful, luxury uptake may be suppressed by low f_V, if allocation to f_C instead is beneficial (i.e., causes higher net growth rate). These remain open questions, and are subject to further investigation in future research.

    Some of the differences in phytoplankton growth dynamics, as simulated by the acclimative IA and DA variants and the non-acclimative FS variant, could be reconciled by tuning the parameters. For instance, the amount of phytoplankton biomass,
385   or the extent of nutrient depletion as simulated by the FS variant can be increased by specifying higher resource affinities (e.g., lower K_N or higher α, to make up for the deficiency in the formulation of light limited growth (Oschlies and Schartau, 2005). However, improvements in these specific aspects typically result in greater discrepancies in other aspects, such as the timing of the spring bloom, or winter concentrations of nutrients and phytoplankton. In other words, in terms of model performance trade-offs exist between multiple objectives. Such trade-offs become more obvious when attempting to simulate
390   multiple environments characterized by different resource conditions (e.g., multiple sites, or the same site in two different time periods) with a single parameter set (Anugerahanti et al.).

    The RAM approach used here, as in 'adaptive dynamics' approaches (Follows and Dutkiewicz, 2011), ambiguously reflects processes at multiple organismal scales. For instance, higher f_A and f_V and lower $\hat{\theta}$ at the surface layers during summer (Fig. 5), which agrees with lower light harvesting and higher nutrient harvesting investment as found by Bruggeman and
395   Kooijman (2007), can be attributed to: *i*) evolutionary adaptation of new species (which would be more relevant in a longer-term simulation), *ii*) selection among existing species that had been pre-adapted to these conditions, and *iii*) individual-level acclimation. Optimality-based acclimative models can thus capture some key community-level effects of evolutionary and ecological dynamics, without explicitly resolving competing species or groups (Smith et al., 2011). The same idea underlies the recent work of Chakraborty et al. (2020), where they described the changes in community composition by assuming that
400   the trophic strategy of the entire plankton community is optimized instantaneously.

Some features, such as the dense and thin chlorophyll layers at the thermocline as captured by the acclimative variants (Fig. 4) seem qualitatively irreproducible by the FS variant even for a single site and time period. This is because multiple dependencies are necessary for capturing this feature, namely the unimodal distribution of chlorophyll density over depth (Fig. 4) and the steep increase in chloroplast size with depth near the thermocline (Fig. 5), as well as the thermocline being the

405 best compromise between light- and nutrient- limitation (Fig.5). The FS variant includes only the last dependency, because it lacks acclimation, and is therefore unable to produce such thin chlorophyll layers. When the chloroplast size is assumed to be variable, and diagnosed by the nutrient limitation term, such that the vertical Chl:C increases monotonically with depth, the vertical distribution of Chl can be partially captured (Appendix B).

4.3 Physiological Flexibility and Environmental Feedbacks

410 The well known links between the composition of phytoplankton and the biogeochemistry of their ambient environments imply feedbacks, which are important in ecology, environmental science, and water quality studies. These feedbacks can be mediated by both physiological acclimation and evolutionary adaptation (Moreno and Martiny, 2018), with the latter typically understood to operate on much longer timescales. However, acclimation and adaptation do interact in eco-evolutionary dynamics, and for plankton they may even occur on similar timescales (Smith et al., 2011; Edelaar and Bolnick, 2019). Disentangling their

415 effects is challenging, and debate continues as to the relative roles of acclimation and evolutionary adaptation in determining the observed patterns of variation. For example, although Sharoni and Halevy (2020) attribute observed seasonal variations of detrital N:P ratios to seasonal sorting among various well adapted species, that conclusion was based on the assumption that acclimation implies a lack of nutrient limitation, which is not the assumption underlying most acclimative models, including ours. For example, the near-zero values of f_C in the upper 25 m during summer months (Fig. 5k,l) indicate extreme nutrient

420 limitation, which prevents growth in the surface layers (Fig. 6b,c). In any case, only models that account for the relevant flexibilities and variations in the composition of phytoplankton can be expected capture such feedbacks in a general, yet realistic sense, which is necessary to correctly assess the relative roles of plankton-related processes in biogeochemical cycles.

An important link between flexibility and environmental feedbacks is the role of phytoplankton in determining the elemental composition of particulate matter (Redfield, 1934). Key mechanisms involve the activities of nitrogen fixers and denitrifiers

425 (Redfield, 1958). However, given the differences in stoichiometry of macromolecules involved in various cellular functions (Geider and La Roche, 2002), a consistent description of the acclimation of phytoplankton is necessary to represent realistically the variabilities in elemental composition of particulate matter, hence, export fluxes. Fixed stoichiometry models erroneously predict constant elemental composition of detrital matter production, as demonstrated by our FS variant in this study. The so called 'Droop' models have been shown to capture the observed seasonal increase in detrital C:N ratios during summer,

430 reflecting nutrient limitation of phytoplankton (e.g., Mongin et al., 2003). Representing the growth and uptake terms consistently using the RAM framework, the DA variant resolves the seasonal and vertical variations in the elemental composition of particulate matter (Fig. 8). Estimates of the IA variant are nearly identical to those of the DA variant, thereby implying that a more realistic representation of these can be achieved at no additional computational cost compared to a fixed-stoichiometry models.

4.4 Present implementation, challenges and perspectives

Moving a coupled hydrodynamic-biogeochemical models from a 0-D setup to a spatially explicit setup can be error-prone and time consuming. The Framework for Aquatic Biogeochemical Models, (FABM Bruggeman and Bolding, 2014), provides an easy to use coupling layer that connects a hydrodynamic model with multiple biogeochemical sub-models. FABM specifies how the these models communicate by separating the hydrodynamics and biogeochemical models, with FABM acting as a glue layer in between. The biogeochemical model in this framework operates locally in space where the local source and sink terms are computed based on the local state and environment, making it feasible to scale up from 0-D to n-D, and swap different hydrodynamic models. FABM also provides mechanisms to pass other environmental data, such as temperature, salinity, and pH, from different submodules, as long as the biogeochemical models register any dependencies during initialisation. Therefore, complex description of the biogeochemical models can be partitioned into several sub-modules. The modular implementation of our model in FABM, specifically, the isolation of the phytoplankton module (Fig. 1) is expected to facilitate studies with multiple phytoplankton types. For example, without changing the model code nor recompiling, just through changing a configuration file, it is possible to include further types (see Bruggeman and Bolding, 2014), which can be parameterized, e.g., according to cell size (as in, e.g. Smith et al., 2016; Dutkiewicz et al., 2020). Moreover, the isolated phytoplankton module can be relatively easily coupled with or incorporated into existing models, especially those implemented in FABM.

Currently, the model simplistically accounts for the grazing losses to higher trophic levels with a quadratic mortality term (Table 1), without describing explicitly the dynamics of preditors. This limitation may prohibit realistic applications to highly productive ecosystems, where the strength of top-down control exhibits strong seasonality (e.g., Maar et al., 2014; Sailley et al., 2015). However, this problem can be easily resolved by adapting an existing zooplankton module available for FABM, such as the N-only resolving module in the 'NPZD' example provided in the standard FABM library (Bruggeman and Bolding, 2014).

When multiple nutrient elements in the dissolved inorganic material pool (e.g., C, N, and P) are resolved, maintaining mass balance becomes more complicated under the IA assumption. A carbon-based version of this model is available, where mass balance for both carbon and nitrogen is satisfied through additional nutrient flux terms (as presented by Smith et al., 2016). We plan to present the FABM-implementation and document the behavior of that model in a separate study. For simplicity, we have traced only N here, so that the IA variant need not maintain mass balance for C. For many ecological applications, especially those resolving multiple phytoplankton types, tracing only one nutrient element may be sufficient and more convenient.

In the current study, we focused on the differences between the fully acclimative IA and DA variants, and an entirely non-acclimative variant. Our acclimation scheme consists of four acclimative flexibilities: variability of internal nutrient quota, optimization of uptake vs growth trade-off, optimization of maximum uptake vs. affinity trade-off, and optimization of chlorophyll density in chloroplast density (and as an additional half-step, size of the chloroplast, see Appendix B). In a future study, we are planning to investigate the relative importance of each of these flexibilities for the organismal fitness under various environmental conditions: such an assessment would not only help the model developers to prioritize the research needs, but may also provide insights into the evolution of these acclimative flexibilities.

5 Conclusions

In this study, we present a FABM-implementation of the 'NflexPD' model, which contains a phytoplankton submodule that can
470 behave as the following three variants, which differ with respect to the representation of acclimative flexibilities and number
of state variables:

DA (Dynamic Acclimation): acclimative flexibilities represented, two state variables (C, N).

IA (Instantaneous Acclimation): acclimative flexibilities represented, one state variable (N).

FS (Fixed Stoichimetry): no acclimative flexibility represented, one state variable (N).

475 The acclimative flexibilities comprise:

Q : cellular N quota.

$f_{\mathbf{V}}$: for optimizing growth vs. nutrient uptake trade-off.

$f_{\mathbf{A}}$: for optimizing affinity vs max. uptake rate trade-off.

$\hat{\theta}$: optimal chlorophyll density in chloroplasts.

480 Coupled to an idealized, 1D water column model, we show that behavior of IA is stable and although not identical, very
similar to that of DA. In comparison to a model with fixed Chl:C:N, represented here by the FS variant, the acclimative variants
are found to estimate sustained growth during summer and stronger nitrogen depletion in the surface layers; steeper chlorophyll
layers at the thermocline; and unlike the FS variant, they can reproduce the variabilities in C:N of particulate matter. Therefore,
our study provides proof-of-concept that the IA approach is applicable in spatially-explicit setups. The model implementation
485 presented here tracks only N as dissolved nutrient, which restricts its utility in biogeochemical contexts, but it can be readily
used in various ecological contexts.

Code availability. For running the model and reproducing the results presented in this study, FABM and GOTM need to be downloaded
and installed. See https://github.com/fabm-model/fabm/wiki/GOTM for the instructions. The FABM-NflexPD is available from the 'Nbased'
branch of the git repository:https://github.com/OnurKerimoglu/fabm-nflexpd.git. Instructions for compiling FABM-NflexPD for GOTM-
490 FABM and a 0D setup are provided in README.md. The 'src' folder contains the Fortran codes. The model was implemented as two
separate modules: the 'phy.F90' module that describes phytoplankton growth and the 'abio.F90' module that describes everything other than
phytoplankton (Fig. 1). The phytoplankton module can reproduce the behavior of all three different variants considered in the manuscript
through optional parameters. The 'testcases' folder contains the configuration (yaml) file that was used to produce the results presented in
this manuscript, thereby providing examples of how each variant can be initiated.

Appendix A: Details of Derivations

A1 R_N for FS variant

According to Eq. (7), $R_\mathrm{N} = \zeta_\mathrm{N} \cdot V$. For the DA and IA variants, V can be calculated externally (Eq. (12)), hence so can be R_N. For the FS variant on the other hand, there is no explicit solution for V, but it can only be calculated as a function of μ, ($V = \mu \cdot Q$, Eq.(6)), and since μ in turn, depends on R_N ($\mu = \mu_\mathrm{net} - R_\mathrm{N}$, Eq. (7)), R_N cannot be directly calculated. Expanding the terms in Eq. (7) according to Eqs. (6), (13) and (20):

$$\mu = \hat{\mu}_\mathrm{net} \cdot L_\mathrm{N} - \zeta_{\mathrm{N} \cdot \mu \cdot Q} \tag{A1}$$

reorganizing:

$$\mu = \frac{\hat{\mu}_\mathrm{net} \cdot L_\mathrm{N}}{1 + \zeta_\mathrm{N} \cdot Q} \tag{A2}$$

substituting this with μ in:

$$R_\mathrm{N} = \zeta_\mathrm{N} \cdot V = \zeta_\mathrm{N} \cdot \mu \cdot Q \tag{A3}$$

we obtain a V-independent expression for R_N:

$$R_\mathrm{N} = \zeta_\mathrm{N} \cdot \frac{\hat{\mu}_\mathrm{net} \cdot L_\mathrm{N}}{1 + \zeta_\mathrm{N} \cdot Q} \cdot Q \tag{A4}$$

It can be verified that, when this term is substituted in $\mu = \mu_\mathrm{net} - R_N$, it yields $\mu = \mu_\mathrm{net} - \zeta_\mathrm{N} \cdot \mu \cdot Q = \mu_\mathrm{net} - \zeta_\mathrm{N} \cdot V$, i.e., Eq. (7), implying that using R_N in Eq. (A4) for the FS variant makes Eq. (7) valid for the FS variant as well.

A2 Optimal Q and f_V

In Eq. (7), substituting μ_g, R_N and and R_{Chl} with the expanded forms in Eqs. (13), (20), and (25), respectively, and subsequently expanding θ, using Eq. (24)):

$$\mu = f_C \hat{\mu}_g - \zeta_N f_V \hat{V} - (\hat{\mu}_g + R_M^{Chl}) \zeta_{Chl} \hat{\theta} f_C \tag{A5}$$

Reorganizing:

$$\mu = f_C \left[\hat{\mu}_g (1 - \zeta_{Chl} \hat{\theta}) - \zeta_{Chl} \hat{\theta} R_M^{Chl} \right] - \zeta_N f_V \hat{V} \tag{A6}$$

Substituting the term in square brackets with $\hat{\mu}_\mathrm{net}$ based on Eq. (7) and expanding f_C using Eq. (11)):

$$\mu = \left(1 - \frac{Q_0}{2Q} - f_V \right) \hat{\mu}_\mathrm{net} - \zeta_N f_V \hat{V} \tag{A7}$$

At this point, it can be readily recognized that Eq. (A7)) is equivalent to the Eq. (5) in Pahlow and Oschlies (2013), only difference being their denotion of $\hat{\mu}_{\text{net}}$ as $\hat{\mu}^I$. Note that their formulation of respiration losses within the chloroplast as a fraction

520 of gross growth with respect to chloroplast (i.e., $\hat{\mu}^I = \hat{\mu}_g^I(1-\zeta^C)$ in their notation), differs from the more precise formulation we used here, that considers a base loss rate independent of gross growth. However, considering that $\hat{\mu}_{\text{net}}$ (just like their $\hat{\mu}^I$) is independent of Q and f_V, the solutions provided by Pahlow and Oschlies (2013) for f_V^o (i.e., their Eq. (9), our Eq. (14)) and Q (their Eq. (10), our Eq. (10)) can be directly used, only after replacing $\hat{\mu}^I$ in the original solutions with $\hat{\mu}_{\text{net}}$ for the latter.

Appendix B: FS variant with a variable chloroplast size

525 Given the similar roles of f_C in the IA and DA variants and the nutrient limitation term, L_N, in the FS variant for calculating μ_g (see Section 2.2.2), L_N can be considered as a proxy for the relative size of the chlorplast. Therefore, f_C in Eq. (24) and (25) can be replaced by L_N for scaling the chloroplast-specific chlorophyll density and respiration costs in order to represent spatio-temporal variations of the cellular Chl:C ratio and proportional respiration costs.

When this is done, unlike the original results shown in the main text (Fig. 4m), a spatio-temporally variable Chl:C ratio

530 (Fig. B1c) is obtained. Monotonically increasing L_N with depth during summer (Fig. 5j) reduces Chl at the surface, and enhances it at the deeper layers relative to the Chl pattern obtained with constant Chl:C (compare Fig. 4m vs. Fig. B1a). However, due to the missing unimodal signal through $\hat{\theta}$ as accounted for by the IA and DA variants (see Fig. 5b,c), the resulting Chl pattern is still qualitatively different from those estimated by the truly acclimative variants (compare Fig. B1a vs. Fig. 4k,l). Furthermore, the relatively higher value of L_N during the spring bloom under nutrient-rich conditions (Fig. 5j)

535 relative to the prescribed, constant value of f_C=0.44 used for the case with constant chloroplast size (hence, constant Chl:C) shown in the main text as yielded by the prescribed values of f_V, Q and Q_0 (Table 3 and Eq. (11)), results in greater R_{Chl} (compare Fig. B1d vs Fig. 6j). Hence, net cellular growth rate, μ becomes slightly lower than in the constant chloroplast case during the spring bloom (compare Fig. B1b vs Fig. 6a). On the other hand, during summer, relatively lower values of L_N make R_{Chl} lower, and μ greater compared to the constant chloroplast case.

540 Dynamics of the Phy_C within the top 50m as simulated with this flavor of the FS variant with variable chloroplast size are almost identical to those simulated by the standard, 'vanilla' version with constant chloroplast size (compare Fig. B2d with Fig. 7d). Relatively higher R_{Chl} at nutrient-rich conditions during winter and early spring makes the winter Phy_C concentrations (Fig. B2d) lower in comparison to the standard case (Fig. 7d). On the other hand, relatively lower R_{Chl} at nutrient-scarce summer conditions make the Phy_C concentrations (Fig. B2d) slightly higher than the standard case (Fig. 7d). As a result, the

545 the average DIN concentrations in the surface 50m become slightly lower than the standard case (Fig. B2a vs. Fig. 7a), which is, better observed in lower L_N (Fig. B2c vs. Fig. 7c), due to the strong response of the function at low concentrations.

Despite the differences in details explained above, especially based on the preserved qualitative differences in simulated Phy_C concentrations between the FS and acclimative variants, it can be concluded that the overall conclusions are insensitive to the assumption regarding the size of the chloroplast of the FS variant.
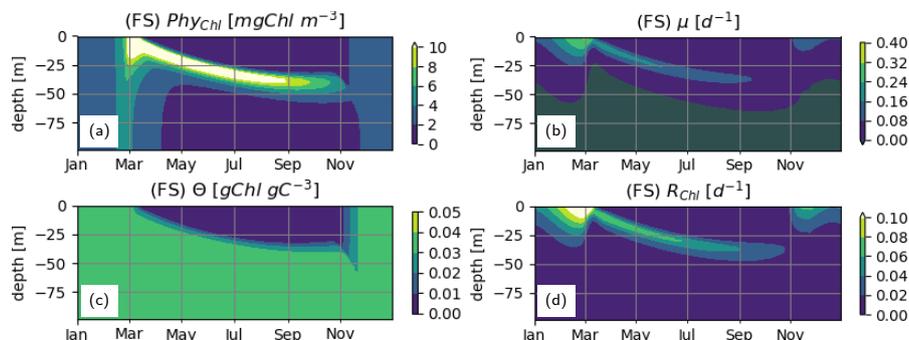
Figure B1. Phytoplankton (a) Chl concentration, Phy_{Chl} [$mgC\ m^{-3}$]; (b) net growth rate, μ [d^{-1}]; (c) Chl:C, Θ [$gChl\ gC^{-1}$]; (d) respiration cost of chlorophyll maintenance and synthesis, R_{Chl} [d^{-1}] as simulated by the FS variant, when the prescrebed Θ (Table 3) is scaled with f_C, according to Eq. (24).
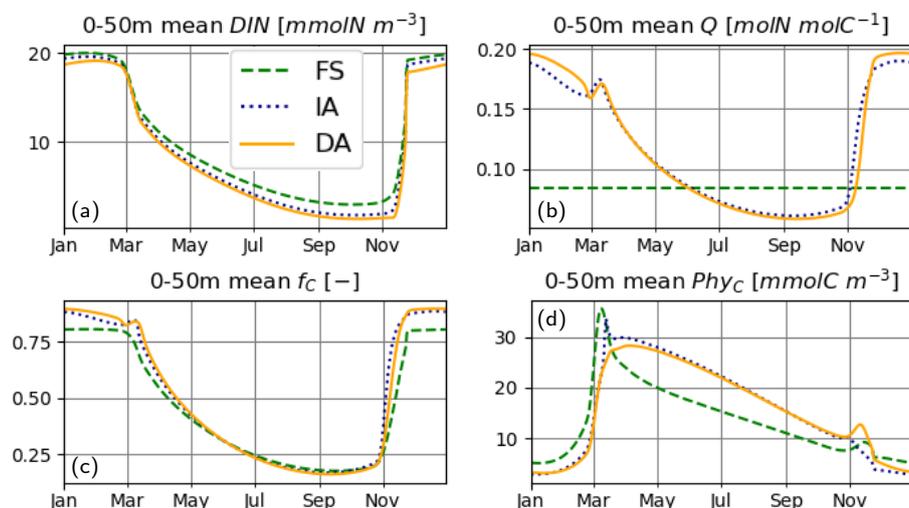
Figure B2. Like Fig. 7, but when for the FS variant, prescribed $\hat{\theta}$ (Table 3) is scaled with L_N, i.e., replacing f_C with L_N in Eq. (24).

550 *Author contributions.* OK and SLS designed the study, OK implemented the model code, performed the model runs and prepared the figures, PA tested the model, all authors contributed to the writing of the manuscript.

Competing interests. No competing interests are present.

Acknowledgements. We are grateful to Markus Pahlow for critically reviewing the model description, helping to correct the implementation of IA and DA variants, and providing the code for the model diagram. OK, SLS and PA were supported through a bilateral research project,

555   funded jointly by the German Research Foundation DFG (grant no. KE 1970/2-1, P.I.: OK) and the Japan Society for the Promotion of Science, JSPS (P.I.: SLS). At an early stage, OK was additionally supported by the Federal Agency of Environment (Bundesumweltamt, UBA; Grant no. 3718252110, P.I.: OK). We acknowledge the developers of the open-source software used in this study, foremost FABM and GOTM.

References

560   Anugerahanti, P., Kerimoglu, O., and Smith, S. L.: Enhancing ocean biogeochemical model performance and generality with phytoplankton photo-acclimation.

Arhonditsis, G. B. and Brett, M. T.: Evaluation of the current state of mechanistic aquatic biogeochemical modeling, Marine Ecology Progress Series, 271, 13–26, https://doi.org/10.3354/meps271013, 2004.

Armstrong, R. A.: An optimization-based model of iron—light-–ammonium colimitation of nitrate uptake and phytoplankton growth, Lim-

565   nol. Oceanogr., 44, 1436–1446, https://doi.org/10.4319/lo.1999.44.6.1436, 1999.

Bruggeman, J. and Bolding, K.: A general framework for aquatic biogeochemical models, Environmental Modelling & Software, 61, 249–265, https://doi.org/10.1016/j.envsoft.2014.04.002, 2014.

Bruggeman, J. and Kooijman, S. a. L. M.: A biodiversity-inspired approach to aquatic ecosystem modeling, Limonology And Oceanography, 52, 1533–1544, https://doi.org/10.4319/lo.2007.52.4.1533, 2007.

570   Burchard, H., Bolding, K., Kühn, W., Meister, A., Neumann, T., and Umlauf, L.: Description of a flexible and extendable physical-biogeochemical model system for the water column, Journal of Marine Systems, 61, 180–211, https://doi.org/10.1016/j.jmarsys.2005.04.011, 2006.

Burmaster, D. E.: The Continuous Culture of Phytoplankton: Mathematical Equivalence Among Three Steady-State Models, The American Naturalist, 113, 123–134, https://doi.org/10.1086/283368, 1979.

575   Burson, A., Stomp, M., Akil, L., Brussaard, C. P. D., and Huisman, J.: Unbalanced reduction of nutrient loads has created an offshore gradient from phosphorus to nitrogen limitation in the North Sea, Limnology and Oceanography, 61, 869–888, https://doi.org/10.1002/lno.10257, 2016.

Button, D. K.: On the theory of control of microbial growth kinetics by limiting nutrient concentrations, Deep-Sea Research, 25, 1163–1177, https://doi.org/10.1016/0146-6291(78)90011-5, 1978.

580   Caperon, J.: Population growth response of Isochrysis Galbana to nitrate variation at limiting concentrations, Ecology, 49, 866–872, 1968.

Chakraborty, S., Cadier, M., Visser, A. W., Bruggeman, J., and Andersen, K. H.: Latitudinal Variation in Plankton Traits and Ecosystem Function, Global Biogeochemical Cycles, 32, e2020GB006 564, https://doi.org/10.1029/2020GB006564, 2020.

Cloern, J., Grenz, C., and Vidergar-Lucas, L.: An empirical model of the phytoplankton chlorophyll : carbon ratio - the conversion factor between productivity and growth rate, Limnol. Oceanogr., 40, 1313–1321, https://doi.org/10.4319/lo.1995.40.7.1313, 1995.

585   Dore, J. E., Letelier, R. M., Church, M. J., Lukas, R., and Karl, D. M.: Summer phytoplankton blooms in the oligotrophic North Pacific Subtropical Gyre: Historical perspective and recent observations, Progress in Oceanography, 76, 2–38, https://doi.org/10.1016/j.pocean.2007.10.002, 2008.

Droop, M.: Vitamin B12 and marine ecology. IV. The kinetics of uptake, growth and inhibition in \textit{Monochrysis lutheri}., Journal of the Marine Biological Association of the United Kingdom, 48, 689–733, 1968.

590   Dugdale, R.: Nutrient Limitation in the Sea: Dynamics, Identification and Significance, Limnology and Oceanography, 12, 685–695, https://doi.org/10.4319/lo.1967.12.4.0685, 1967.

Dutkiewicz, S., Cermeno, P., Jahn, O., Follows, M. J., Hickman, A. E., Taniguchi, D. A. A., and Ward, B. A.: Dimensions of marine phytoplankton diversity, Biogeosciences, 17, 609–634, https://doi.org/10.5194/bg-17-609-2020, 2020.

Edelaar, P. and Bolnick, D. I.: Appreciating the Multiple Processes Increasing Individual or Population Fitness, Trends in Ecology and

595   Evolution, 34, 435–446, https://doi.org/10.1016/j.tree.2019.02.001, https://doi.org/10.1016/j.tree.2019.02.001, 2019.

Flynn, K. J., St John, M., Raven, J. A., Skibinski, D. O., Allen, J. I., Mitra, A., and Hofmann, E. E.: Acclimation, adaptation, traits and trade-offs in plankton functional type models: Reconciling terminology for biology and modelling, Journal of Plankton Research, 37, 683–691, https://doi.org/10.1093/plankt/fbv036, 2015.

Follows, M. J. and Dutkiewicz, S.: Modeling Diverse Communities of Marine Microbes, Annual review of marine science, 3, 427–451, https://doi.org/10.1146/annurev-marine-120709-142848, 2011.

Geider, R. and La Roche, J.: Redfield revisited: variability of C:N:P in marine microalgae and its biochemical basis, European Journal of Phycology, 37, 1–17, https://doi.org/10.1017/S0967026201003456, 2002.

Geider, R., Maclntyre, H., and Kana, T.: A dynamic regulatory model of phytoplanktonic acclimation to light, nutrients, and temperature, Limnol. Oceanogr., 43, 679–694, https://doi.org/10.4319/lo.1998.43.4.0679, 1998.

Gerloff, G. C. . and Skoog, F.: Cell Contents of Nitrogen and Phosphorous as a Measure of Their Availability for Growth of Microcystis Aeruginosa., Ecology, 35, 348–353, http://www.jstor.org/stable/193009, 1954.

Grover, J.: Resource competition in a variable environment: phytoplankton growing according to the variable-internal-stores model, The American Naturalist, 138, 811–835, https://www.jstor.org/stable/2462435, 1991.

Grover, J. P.: Sink or swim? Vertical movement and nutrient storage in phytoplankton, Journal of Theoretical Biology, 432, 38–48, https://doi.org/10.1016/j.jtbi.2017.08.012, 2017.

Halsey, K. H. and Jones, B. M.: Phytoplankton strategies for photosynthetic energy allocation, Annual Review of Marine Science, 7, 265–297, https://doi.org/10.1146/annurev-marine-010814-015813, 2015.

Johnson, K. and Goody, R.: The Original Michaelis Constant: Translation of the 1913 Michaelis–Menten Paper, Biochemistry, 50, 8264–8269, https://doi.org/10.1021/bi201284u, 2011.

Kerimoglu, O., Straile, D., and Peeters, F.: Role of phytoplankton cell size on the competition for nutrients and light in incompletely mixed systems., Journal of Theoretical Biology, 300, 330–43, https://doi.org/10.1016/j.jtbi.2012.01.044, http://www.ncbi.nlm.nih.gov/pubmed/22342936, 2012.

Kerimoglu, O., Hofmeister, R., Maerz, J., Riethmüller, R., and Wirtz, K. W.: The acclimative biogeochemical model of the southern North Sea, Biogeosciences, 14, 4499–4531, https://doi.org/10.5194/bg-14-4499-2017, 2017.

Kerimoglu, O., Große, F., Kreus, M., Lenhart, H.-J., and van Beusekom, J. E.: A model-based projection of historical state of a coastal ecosystem: relevance of phytoplankton stoichiometry, Science of the Total Environment, 639, 1311–1323, https://doi.org/10.1016/j.scitotenv.2018.05.215, 2018.

Klausmeier, C., Litchman, E., Daufresne, T., and Levin, S.: Optimal nitrogen-to-phosphorus stoichiometry of phytoplankton, Nature, 429, 171–174, https://doi.org/1.1029/2001GL014649, 2004.

Kruskopf, M. and Flynn, K. J.: Chlorophyll content and fluorescence responses cannot be used to gauge reliably phytoplankton biomass, nutrient status or growth rate, New Phytologist, 169, 525–536, https://doi.org/10.1111/j.1469-8137.2005.01601.x, 2006.

Kwiatkowski, L., Aumont, O., Bopp, L., and Ciais, P.: The Impact of Variable Phytoplankton Stoichiometry on Projections of Primary Production, Food Quality, and Carbon Uptake in the Global Ocean, Global Biogeochemical Cycles, 32, https://doi.org/10.1002/2017GB005799, 2018.

Laufkötter, C., Vogt, M., Gruber, N., Aita-Noguchi, M., Aumont, O., Bopp, L., Buitenhuis, E., Doney, S. C., Dunne, J., Hashioka, T., Hauck, J., Hirata, T., John, J., Quéré, C. L., Lima, I. D., Nakano, H., Seferian, R., Totterdell, I., Vichi, M., and Völker, C.: Drivers and uncertainties of future global marine primary production in marine ecosystem models, Biogeosciences, 12, 6955–6984, https://doi.org/10.5194/bg-12-6955-2015, 2015.

Laws, E. A. and Chalup, M. S.: A microalgal growth model, Limnology and Oceanography, 35, 597–608, https://doi.org/10.4319/lo.1990.35.3.0597, 1990.

Litchman, E., Klausmeier, C. A., and Yoshiyama, K.: Contrasting size evolution in marine and freshwater diatoms, Proceedings of the National Academy of Sciences, 106, 2665–2670, 2009.

Maar, M., Rindorf, A., Møller, E. F., Christensen, A., Madsen, K. S., and van Deurs, M.: Zooplankton mortality in 3D ecosystem modelling considering variable spatial–temporal fish consumptions in the North Sea, Progress in Oceanography, 124, 78–91, https://doi.org/10.1016/j.pocean.2014.03.002, 2014.

Martiny, A. A. C., Pham, C. C. T. a., Primeau, F. F. W., Vrugt, J. a., Moore, J. K., Levin, S. a., and Lomas, M. W.: Strong latitudinal patterns in the elemental ratios of marine plankton and organic matter, Nature Geoscience, 6, 279–283, https://doi.org/10.1038/ngeo1757, 2013.

Mattern, J. P., Fennel, K., and Dowd, M.: Estimating time-dependent parameters for a biological ocean model using an emulator approach, Journal of Marine Systems, 96-97, 32–47, https://doi.org/10.1016/j.jmarsys.2012.01.015, 2012.

Mongin, M., Nelson, D. M., Pondaven, P., Brzezinski, M. A., and Tréguer, P.: Simulation of upper-ocean biogeochemistry with a flexible-composition phytoplankton model: C, N and Si cycling in the western Sargasso Sea, Deep-Sea Research Part I: Oceanographic Research Papers, 50, 1445–1480, https://doi.org/10.1016/j.dsr.2003.08.003, 2003.

Monod, J.: The growth of bacterial cultures, Annual Review of Microbiology, 3, 371–394, 1949.

Moore, J. K., Doney, S. C., Kleypas, J. A., Glover, D. M., and Fung, I. Y.: An intermediate complexity marine ecosystem model for the global domain, Deep-Sea Research Part II: Topical Studies in Oceanography, 49, 403–462, https://doi.org/10.1016/S0967-0645(01)00108-4, 2002.

Moreno, A. R. and Martiny, A. C.: Ecological Stoichiometry of Ocean Plankton, Annual Review of Marine Science, 10, 43–69, https://doi.org/10.1146/annurev-marine-121916-063126, 2018.

Oschlies, A. and Schartau, A.: Basin-scale performance of a locally optimized marine ecosystem model, Research, Journal of Marine, 63, 335–358, https://doi.org/10.1016/b978-0-408-70700-8.50018-5, 2005.

Pahlow, M.: Linking chlorophyll – nutrient dynamics to the Redfield N : C ratio with a model of optimal phytoplankton growth, Marine Ecology Progress Series, 287, 33–43, https://doi.org/10.3354/meps287033, 2005.

Pahlow, M. and Oschlies, A.: Optimal allocation backs Droop's cell-quota model, Marine Ecology Progress Series, 473, 1–5, https://doi.org/10.3354/meps10181, 2013.

Pahlow, M., Dietze, H., and Oschlies, A.: Optimality-based model of phytoplankton growth and diazotrophy, Marine Ecology Progress Series, 489, 1–16, https://doi.org/10.3354/meps10449, 2013.

Pahlow, M., Chien, C. T., Arteaga, L. A., and Oschlies, A.: Optimality-based non-Redfield plankton-ecosystem model (OPEM v1.1) in UVic-ESCM 2.9 - Part 1: Implementation and model behaviour, Geoscientific Model Development, 13, 4663–4690, https://doi.org/10.5194/gmd-13-4663-2020, 2020.

Paulson, C. and Simpson, J.: Irradiance measurements in the upper ocean, Journal of Physical Oceanography, 7, 952–956, 1977.

Platt, T. and Jassby, A. D.: the Relationship Between Photosynthesis and Light for Natural Assemblages of Coastal Marine Phytoplankton, https://doi.org/10.1111/j.1529-8817.1976.tb02866.x, 1976.

Redfield, A.: On the proportions of organic derivatives in sea water and their relation to the composition of plankton, in: James Johnstone Memorial Volume, edited by Daniel, R., pp. 177–192, University Press of Liverpool, 1934.

Redfield, A.: The biological control of chemical factors in the environment, American Scientist, 46, 205–221, 1958.

Sailley, S. F., Polimene, L., Mitra, A., Atkinson, A., and Allen, J. I.: Impact of zooplankton food selectivity on plankton dynamics and nutrient cycling, Journal of Plankton Research, 37, 519–529, https://doi.org/10.1093/plankt/fbv020, 2015.

Sharoni, S. and Halevy, I.: Nutrient ratios in marine particulate organic matter, Science Advances, 6, 1–10, https://doi.org/10.1126/sciadv.aaw9371, 2020.

675 Shuter, B.: A model of physiological adaptation in unicellular algae, Journal of Theoretical Biology, 78, 519–552, 1979.

Smith, S. L., Yamanaka, Y., Pahlow, M., and Oschlies, A.: Optimal uptake kinetics: physiological acclimation explains the pattern of nitrate uptake by phytoplankton in the ocean, Marine Ecology Progress Series, 384, 1–12, https://doi.org/10.3354/meps08022, 2009.

Smith, S. L., Pahlow, M., Merico, A., and Wirtz, K. W.: Optimality-based modeling of planktonic organisms, Limnology and Oceanography, 56, 2080–2094, https://doi.org/10.4319/lo.2011.56.6.2080, 2011.

680 Smith, S. L., Pahlow, M., Merico, A., Acevedo-Trejos, E., Sasai, Y., Yoshikawa, C., Sasaoka, K., Fujiki, T., Matsumoto, K., and Honda, M. C.: Flexible phytoplankton functional type (FlexPFT) model: size-scaling of traits and optimal growth, Journal of Plankton Research, 38, 977–992, https://doi.org/10.1093/plankt/fbv038, 2016.

Ward, B. A.: Assessing an efficient "Instant Acclimation" approximation of dynamic phytoplankton stoichiometry, J Plankton Res, 39, 803–814, https://doi.org/10.1093/plankt/fbx040, 2017.

685 Wirtz, K. W. and Kerimoglu, O.: Autotrophic Stoichiometry Emerging from Optimality and Variable Co-limitation, Frontiers in Ecology and Evolution, 4:131, https://doi.org/10.3389/fevo.2016.00131, 2016.