

# Explicit silicate cycling in the Kiel Marine Biogeochemistry Model, version 3 (KMBM3) embedded in the UVic ESCM version 2.9

Karin Kvale<sup>1</sup>, David P. Keller<sup>1</sup>, Wolfgang Koeve<sup>1</sup>, Katrin J. Meissner<sup>2,3</sup>, Chris Somes<sup>1</sup>, Wanxuan Yao<sup>1</sup>, and Andreas Oschlies<sup>1</sup>

<sup>1</sup>GEOMAR Helmholtz Centre for Ocean Research Kiel, Düsternbrooker Weg 20, D-24105 Kiel, Germany

<sup>2</sup>Climate Change Research Centre, Level 4 Mathews Bldg., UNSW, Sydney, NSW, AU

<sup>3</sup>ARC Centre of Excellence for Climate Extremes, Australia

**Correspondence:** K. Kvale (kkvale@geomar.de)

**Abstract.** We describe and test a new model of biological marine silicate cycling, implemented in the Kiel Marine Biogeochemical Model version 3 (KMBM3), embedded in the University of Victoria Earth System Climate Model (UVic ESCM) version 2.9. This new model adds diatoms, which are a key aspect of the biological carbon pump, to an existing ecosystem model. The new model performs well against important ocean biogeochemical indicators and captures the large-scale features of the marine silica cycle. Furthermore it is computationally efficient, allowing both fully-coupled, long-timescale transient simulations, as well as “offline” transport matrix spinups. We assess the fully-coupled model against modern ocean observations, the historical record since 1960, and a business-as-usual atmospheric CO<sub>2</sub> forcing to the year 2300. The model simulates a global decline in net primary production (NPP) of 1.8% having occurred since the 1960s, with the strongest declines in the tropics, northern mid-latitudes, and Southern Ocean. The simulated global decline in NPP reverses after the year 2100 (forced by the extended RCP 8.5 CO<sub>2</sub> concentration scenario), and NPP returns to 96% of the pre-industrial rate by 2300. This recovery is dominated by increasing primary production in the Southern Ocean, mostly by calcifying phytoplankton. Large increases in calcifying phytoplankton in the Southern Ocean offset a decline in the low latitudes, producing a global net calcite export in 2300 that varies only slightly from pre-industrial rates. Diatom [distribution moves](#) southward in our simulations, following the receding Antarctic ice front, but diatoms are out-competed by calcifiers across most of their pre-industrial Southern Ocean habitat. Global opal export production thus drops to 50% of its pre-industrial value by 2300. Model nutrients phosphate, silicate, and nitrate build up along the Southern Ocean particle export pathway, but dissolved iron (for which ocean sources are held constant) increases in the upper ocean. This different behaviour of iron is attributed to a reduction of low-latitude NPP (and consequently, a reduction in both uptake and export and particle, including calcite, scavenging), an increase in seawater temperatures (raising the solubility of particle forms), and stratification that “traps” the iron near the surface. These results are meant to serve as a baseline for sensitivity assessments to be undertaken with this model in the future.

*Copyright statement.* TEXT

# 1 Introduction

It has become apparent in recent decades that the representation of elemental cycles of nutritive elements (N, P, Si, Fe) is important in order to simulate critical biogeochemical feedbacks (Heinze et al., 2019). Here, we describe an updated representation of ocean biogeochemistry (the Kiel Marine Biogeochemical Model, version 3; KMBM3) embedded in an earth system model, the University of Victoria Earth System Climate Model version 2.9, second level updates (UVic ESCM; Weaver et al., 2001; Eby et al., 2009), to which we have added a silicate cycle.

Silicate (considered here as any anion that combines silicon, Si, and between 2 and 4 oxygen molecules) is important for simulating ocean biogeochemistry for several reasons. It regulates the growth of phytoplankton diatoms, which are responsible for 40% of the particulate carbon export in the modern ocean (Jin et al., 2006). Fluctuations in the relative abundance of diatoms might therefore affect air-sea CO<sub>2</sub> exchange, with a globally significant influence on atmospheric CO<sub>2</sub> concentrations over millennial and longer timescales (e.g. Matsumoto et al., 2002; Renaudie, 2016). High latitude diatoms with low N:P ratios have been demonstrated to exert global control on the nitrogen budget (Weber and Deutsch, 2012), and are speculated to exert seasonal control on carbonate chemistry through competition with calcifiers (Merico et al., 2006). Silicic acid (Si(OH)<sub>4</sub>, and SiO(OH)<sub>3</sub><sup>-</sup>) distributions are tightly controlled by diatom primary production (biogenic silica, or opal; SiO<sub>2</sub>) and subsequent dissolution (e.g., only about 34% global average ocean silicic acid is preformed, as opposed to biologically regenerated; Holzer et al. 2014). First attempts to constrain the marine silicate cycle demonstrate representations of the silica cycle are well suited to parameter optimisation (Holzer et al., 2014; Pasquier and Holzer, 2017). Due to diatoms' important role in biological carbon export, evaluating simulated silicate and diatoms presents a unique opportunity to constrain ocean carbon cycling.

In the following sections we describe our new model of explicit diatoms and silicate cycling in the ocean. As far as we are aware, this model is unique among ecosystem models embedded within earth system models in that it combines established parameterisations of a diatom functional type (i.e. fast growth rates and inefficient nutrient utilisation, Tréguer et al. 2018) with a novel temperature-dependent opal dissolution parameterisation and bottom-up and top-down competition for resources with explicit, fully independent calcifying phytoplankton and nitrogen fixing diazotrophs. These assumptions bias KMBM3 diatoms towards large species in the well-mixed high latitudes, and early seasonal succession. Modelled silicate boundary conditions include hydrothermal silicate inputs and a benthic transfer function to approximate sediment sequestration. A ballast model applied to the calcifiers (Kvale et al., 2015b) further differentiates KMBM3, as does the ability to operate in an “offline” parameter optimisation framework (Kvale et al., 2017; Yao et al., 2019).

## 2 Model Description

### 2.1 Physics

The University of Victoria Earth System Climate Model (UVic ESCM) version 2.9 is a coarse-resolution ( $1.8^{\circ} \times 3.6^{\circ} \times 19$  ocean depth layers) ocean-atmosphere-biosphere-cryosphere-geosphere model. Model structure and physics are described in

Weaver et al. (2001), Meissner et al. (2003) and Eby et al. (2009). We use the second level updates of version 2.9. Additionally, an ideal age tracer (Koeve et al., 2015) is merged into this version of the model to allow for explicit tracing of water mass age.

## 55 2.2 Biogeochemistry

Development of the Kiel Marine Biogeochemistry Model has progressed in recent years. Beginning with a base composition of nitrate, phosphate, DIC, and alkalinity as well as general phytoplankton, diazotrophs and one zooplankton functional type (Schmittner et al., 2005, 2008), Keller et al. (2012) updated zooplankton grazing and included a seasonally cycling iron mask (KMBM1). To this version prognostic iron tracers were implemented by Nickelsen et al. (2015) (KMBM2), and prognostic  
60  $\text{CaCO}_3$  and calcifying phytoplankton were added by Kvale et al. (2015b). Somes et al. (2013) included benthic denitrification and Muglia et al. (2017) included hydrothermal iron in MOBI, a separate branch of the UVic ESCM biogeochemistry. Kvale and Meissner (2017) examined the sensitivity of primary production in the model to light attenuation parameter value following a correction in the calculation of light availability (Partanen et al., 2016). To the Keller et al. (2012) version Kvale et al. (2017) added offline transport matrix method (TMM) capability, allowing for rapid matrix-based computation of model mean ocean  
65 tracer states and coupling to alternative physical models (Khatiwala et al., 2005; Khatiwala, 2007). This model description unites divergent code, assimilating the Nickelsen et al. (2015); Kvale et al. (2015b, 2017) models as well as the benthic denitrification code of Somes et al. (2013) and hydrothermal iron of Muglia et al. (2017), and expands the code to include explicit diatoms and a silicic acid tracer.

With up to 17 model tracers, KMBM3 biogeochemistry is now fairly complex. An abbreviated description is provided in  
70 the following sections, with only the relevant model developments described here. All simulations use the fully-coupled UVic ESCM framework.

We introduce a new phytoplankton functional type to which has been assigned key physiological attributes of fast growing phytoplankton diatoms. This new model version therefore contains phytoplankton types “slow growing low latitude phytoplankton (LP)”, “nitrogen fixing diazotrophs (DZ)”, “moderate growth phytoplankton, including calcifiers (CP)”, and “fast  
75 growing diatoms (DT)”. Zooplankton contribute to  $\text{CaCO}_3$  production. Prognostic tracers include nitrate, phosphate, oxygen, DIC, alkalinity, iron,  $\text{CaCO}_3$ , silicic acid, and particle forms of iron and organic detritus. The new model schematic is shown in Figure 1 and state variables are given in Table 1. Diatoms compete with the other phytoplankton types for light and nutrients. The diatoms’ growth is also limited by dissolved silicic acid availability; they implicitly produce opal that instantly remineralises back into dissolved silicic acid throughout the water column. To ensure absolute model conservation of silica, any opal  
80 that reaches the seafloor is replaced by external sources (prescribed atmospheric dust deposition, sediment release via a benthic transfer function, prescribed hydrothermal vent release, and river fluxes that can be set to compensate for any remainder; see below for details). The phytoplankton and detritus production and remineralisation are linked to nutrients through fixed Redfield stoichiometry using a base unit of  $\text{mmol nitrogen m}^{-3}$ .

In the following model description, notation will generally follow the symbols used in Kvale et al. (2015b), with the abbreviations LP, CP, DT, DZ representing the phytoplankton types, and “Z” representing zooplankton when a distinction is necessary.  
85 The most important biogeochemical model parameters are listed in Tables 2 to 5. The model description here covers only the

most relevant equations, and equations that have changed in this newest version; please see Kvale et al. (2015b); Nickelsen et al. (2015); Keller et al. (2012); Schmittner et al. (2005), and Schmittner et al. (2008) for original references and a complete description of the other equations.

### 90 2.2.1 Sources and Sinks

Tracer concentrations ( $C$ ) vary according to:

$$\frac{\partial[C]}{\partial t} = T + S + B \quad (1)$$

with  $T$  including all transport terms (advection, diffusion, and convection),  $S$  representing all source and sink terms, and  $B$  representing air-sea interface boundary (including virtual [evaporation-precipitation correction](#)) fluxes.

### 95 2.2.2 Phytoplankton

Phytoplankton ( $X$  representing all types except diazotrophs) biomass source and sink terms are:

$$S(X) = J_X X - G_X - \mu_X^* X - m_X X \quad (2)$$

where growth ( $J$ ), mortality ( $m$ ), and fast recycling ( $\mu^*$ ) [rates](#) are described below, and losses to zooplankton grazing ( $G$ ) are described in the zooplankton equations. The diazotroph equation is similar except that it does not include a linear mortality loss, only a loss to fast recycling.

As in Keller et al. (2012), the maximum possible growth rate of phytoplankton ( $J_{\max}$ ) is a modified Eppley curve (Eppley, 1972), and is a function of seawater temperature ( $T$ ) [in Celsius](#), an e-folding temperature parameter  $T_b$ , and iron availability ( $u_{\text{Fe}}$ ) [modifying a growth parameter \( \$a\$ \)](#). [This parameterisation assumes sufficient iron is required for the utilisation of other nutrients \(Galbraith et al., 2010; Keller et al., 2012; Nickelsen et al., 2015\).](#)

$$105 \quad J_{\max} = a \times \exp^{\frac{T}{T_b}} \times u_{\text{Fe}}. \quad (3)$$

As in earlier versions of the model, diazotroph growth rate is calculated following general phytoplankton (now LP) and then assigned an additional handicap.

Nickelsen et al. (2015) assigned a constant iron half saturation ( $k_{\text{Fe}}$ ) to diazotrophs but in general phytoplankton this parameter varied as a function of biomass ( $X$ ) to implicitly represent different cell sizes in the model. Because KMBM3 contains multiple phytoplankton types we revert to a prescribed but unique  $k_{\text{Fe}}$  for all phytoplankton types and calculate iron availability as:

$$110 \quad u_{\text{Fe}} = \frac{[\text{Fe}]}{k_{\text{Fe}} + [\text{Fe}]}. \quad (4)$$

The maximum potential growth rate is then multiplied by a nutrient availability ( $u$ ) for nitrate, phosphate, and dissolved silicic acid (the latter for diatoms only) to calculate potential growth under nutrient limitation but replete light, where  $k_{\text{N}}$ , and  $k_{\text{P}}$

115 are fixed half saturation constants unique to each phytoplankton type (and  $k_P$  is calculated from 16 P to 1 N Redfield stoichiometry). These equations are applied to obtain maximum possible growth rates as a function of temperature and nutrients:

$$u_{\text{NO}_3^-} = \frac{[\text{NO}_3^-]}{k_N + [\text{NO}_3^-]} \quad (5)$$

120 
$$u_{\text{PO}_4^{3-}} = \frac{[\text{PO}_4^{3-}]}{k_P + [\text{PO}_4^{3-}]} \quad (6)$$

$$u_{\text{Si}} = \frac{[\text{Si}]}{k_{\text{Si}} + [\text{Si}]} \quad (7)$$

Silica uptake uses the [empirical](#) Aumont et al. (2003) scaling of  $k_{\text{Si}}$  in  $\text{mol Si m}^{-3}$ :

$$k_{\text{Si}} = 8e^{-4} + 7.2e^{-3} \times \frac{[\text{Si}]}{k_{\text{Si}}^* + [\text{Si}]} \quad (8)$$

125 with a  $k_{\text{Si}}^*$  value adopted from Aumont et al. (2003) of  $3e^{-2} \text{ mol Si m}^{-3}$ .

The potential growth rate under limited light availability ( $J_I$ ) but replete nutrients is calculated as:

$$J_I = \frac{J_{\text{max}} \alpha^{\text{chl}} \theta I}{(J_{\text{max}}^2 + (\alpha^{\text{chl}} \theta I)^2)^{\frac{1}{2}}} \quad (9)$$

where  $\alpha^{\text{chl}}$  is the initial slope of the photosynthesis versus irradiance ( $I$ ) curve in chlorophyll units:

$$\alpha^{\text{chl}} = \alpha_{\text{min}}^{\text{chl}} + (\alpha_{\text{max}}^{\text{chl}} - \alpha_{\text{min}}^{\text{chl}}) u_{\text{Fe}}, \quad (10)$$

130 and  $\theta$  is a Chl:C ratio ([noting, chlorophyll is not a prognostic tracer](#), Nickelsen et al., 2015):

$$\theta = \theta_{\text{min}} + (\theta_{\text{max}} - \theta_{\text{min}}) u_{\text{Fe}}. \quad (11)$$

For simplicity, the same maximum and minimum values of  $\alpha^{\text{chl}}$  and  $\theta$  are used for all phytoplankton types.

Light attenuation by coccoliths is included in the calculation of available irradiance at each depth level:

$$I = I_{z=0} \text{ PAR } e^{-k_w \tilde{z} - k_c \int_0^{\tilde{z}} (LP + CP + DZ + DT) dz - k_{\text{CaCO}_3} \int_0^{\tilde{z}} [\text{CaCO}_3] dz} (1 + a_i (e^{-k_i (h_i + h_s)} - 1)) \quad (12)$$

135 where PAR stands for the photosynthetically available radiation,  $k_w$ ,  $k_c$ ,  $k_{\text{CaCO}_3}$ , and  $k_i$  are the light attenuation coefficients for water, all phytoplankton,  $\text{CaCO}_3$ , and ice,  $\tilde{z}$  is the effective vertical coordinate,  $a_i$  is the fractional sea ice cover, and  $h_i$  and  $h_s$  are calculated sea ice and snow cover thickness. Opal generated by diatoms is not explicitly traced and is therefore not included in the underwater light field.

The actual growth rate ( $J_X$ ) of phytoplankton is taken to be the minimum of the growth functions described above:

140 
$$J_{LP} = \min(J_I, J_{\text{max}} u_{\text{NO}_3^-}, J_{\text{max}} u_{\text{PO}_4^{3-}}), \quad (13)$$

$$J_{CP} = \min(J_I, J_{\max} u_{\text{NO}_3^-}, J_{\max} u_{\text{PO}_4^{3-}}), \quad (14)$$

$$J_{DT} = \min(J_I, J_{\max} u_{\text{NO}_3^-}, J_{\max} u_{\text{PO}_4^{3-}}, J_{\max} u_{\text{Si}}), \quad (15)$$

145

$$J_{DZ} = \min(J_I, J_{\max} u_{\text{PO}_4^{3-}}). \quad (16)$$

In Equation 2, two loss terms other than predation are considered. **Non-grazing mortality** is parameterised using a linear mortality rate ( $m$ ). Temperature-dependent fast remineralisation is a loss term used to implicitly account for the microbial loop and dissolved organic matter cycling, and is parameterised using a temperature dependency multiplied by a constant ( $\mu_0^*$ ) (Schmittner et al., 2008):

$$\mu^* = \mu_0^* \times \exp \frac{T}{T_b}. \quad (17)$$

With this formulation, increasing seawater temperature increases respiration and the return of nutrients to the upper ocean.

### 2.2.3 Zooplankton

Changes in zooplankton population ( $Z$ ) are calculated as the total available food (phytoplankton, zooplankton, and organic detritus) scaled with a growth efficiency coefficient ( $\varpi$ ) minus mortality. In addition to **non-grazing mortality** calculated with a quadratic mortality function ( $m_Z Z^2$ ), zooplankton mortality also encompasses losses from higher trophic level predation ( $G_Z$ ) (Keller et al., 2012).

$$S(Z) = \varpi (G_{LP} + G_{CP} + G_{DZ} + G_{DT} + G_{Detr_{\text{tot}}} + G_Z) - m_Z Z^2 - G_Z. \quad (18)$$

Zooplankton grazing ( $G$ ) follows Keller et al. (2012). Relevant parameters are listed in Table 4. Grazing of each food source is calculated using a Holling II function, where a calculated maximum zooplankton grazing rate ( $\mu_Z^{\max}$ ) is reduced by a scaling that is weighted by a **relative** food preference ( $\psi_X$ , where “ $X$ ” stands for any of the food sources **and the sum of all preferences must equal 1**), the total prey population and a half saturation constant for zooplankton ingestion ( $k_z$ ):

$$G_X = \mu_Z^{\max} \times Z \times \frac{\psi_X X}{\psi_{LP} LP + \psi_{CP} CP + \psi_{DZ} DZ + \psi_{DT} DT + \psi_{Det} Detr_{\text{tot}} + \psi_Z Z + k_z}. \quad (19)$$

The calculated maximum potential grazing rate is a function of a maximum potential grazing rate at  $0^\circ\text{C}$  ( $\mu_Z^\theta$ ), temperature, and oxygen, where grazing activity is capped when temperatures exceed  $20^\circ\text{C}$  (Keller et al., 2012):

$$\mu_Z^{\max} = \mu_Z^\theta \times \max(0, r_{\text{sox}}^{\text{O}_2} \times b^{c \cdot \min(20, T)}). \quad (20)$$

Grazing is also reduced under **suboxic** conditions ( $r_{\text{sox}}^{\text{O}_2}$ ):

$$r_{\text{sox}}^{\text{O}_2} = 0.5 (\tanh([\text{O}_2] - 8) + 1) \quad (21)$$

where  $\text{O}_2$  is dissolved oxygen in  $\text{mmol m}^{-3}$ .

## 170 2.2.4 Organic Detritus

As was introduced in Kvale et al. (2015b), organic carbon detritus sources and sinks are split into “free” ( $Detr_{\text{free}}$ ) and “ballast” ( $Detr_{\text{bal}}$ ) pools using a fixed ratio ( $R_{\text{bal:tot}}$ ). Ballast detritus is formed of the  $\text{CaCO}_3$ -protected portion of calcifying phytoplankton ( $CP$ ), and zooplankton. This protected portion does not interact with nutrient pools directly, and instead transfers from the “ballast” to the “free” detrital pool at the rate of  $\text{CaCO}_3$  dissolution ( $\lambda_{\text{CaCO}_3}$ ):

$$175 \quad Detr_{\text{tot}} = Detr_{\text{bal}} + Detr_{\text{free}} \quad (22)$$

$$\begin{aligned} S(Detr_{\text{bal}}) = & (1 - \gamma) (G_{Detr_{\text{bal}}} + (G_Z + G_{CP})R_{\text{bal:tot}}) + (m_Z Z + m_{CP} CP)R_{\text{bal:tot}} - G_{Detr_{\text{bal}}} \\ & - \frac{R_{\text{bal:tot}} \lambda_{\text{CaCO}_3} [\text{CaCO}_3]}{R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}}} - w_C \frac{\partial Detr_{\text{bal}}}{\partial z} \end{aligned} \quad (23)$$

where  $\gamma$  is the food assimilation efficiency,  $R_{\text{CaCO}_3:\text{POC}}$  is a fixed production ratio of  $\text{CaCO}_3$  to organic detritus,  $R_{\text{C:N}}$  is a  
180 Redfield molar ratio, and  $w_C$  is the sinking speed of calcite. Free detritus is described as:

$$\begin{aligned} S(Detr_{\text{free}}) = & (1 - \gamma) (G_Z + G_{DZ} + G_{DT} + G_{Detr_{\text{free}}} + G_Z(1 - R_{\text{bal:tot}}) + G_{CP}(1 - R_{\text{bal:tot}})) \\ & + m_Z Z^2(1 - R_{\text{bal:tot}}) + m_{DZ} DZ + m_{DT} DT + m_{LP} LP \\ & + m_{CP} CP(1 - R_{\text{bal:tot}}) - \mu_D Detr_{\text{free}} - G_{Detr_{\text{free}}} \\ & + \frac{R_{\text{bal:tot}} \lambda_{\text{CaCO}_3} [\text{CaCO}_3]}{R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}}} - w_D \frac{\partial Detr_{\text{free}}}{\partial z} \end{aligned} \quad (24)$$

185 where  $\mu_D$  is the detrital remineralisation rate and  $w_D$  is the sinking speed of organic detritus.

## 2.2.5 Detrital Iron

Iron in detritus follows Nickelsen et al. (2015), but with the additional contribution of calcifiers and diatoms:

$$\begin{aligned} S(Detr_{\text{Fe}}) = & R_{\text{Fe:N}} ((1 - \gamma)(G_{LP} + G_{DZ} + G_{DT} + G_{Detr_{\text{tot}}} + G_Z + G_{CP}) \\ & + m_{LP} LP + m_{DZ} DZ + m_{DT} DT + m_Z Z^2 + m_{CP} CP - G_{Detr_{\text{tot}}}) \\ 190 \quad & + [\text{Fe}]_{\text{orgads}} + [\text{Fe}]_{\text{orgads}_{\text{ca}}} + [\text{Fe}]_{\text{col}} - \mu_D Detr_{\text{Fe}} - w_D \frac{\partial Detr_{\text{Fe}}}{\partial z}. \end{aligned} \quad (25)$$

$R_{\text{Fe:N}}$  is a fixed iron to nitrogen ratio that converts total organic carbon detritus sources into iron units. The remineralisation and sinking of detrital iron occurs at the same rates as free organic detritus. Two scavenging processes are considered: adsorption of dissolved iron onto organic detritus ( $[\text{Fe}]_{\text{orgads}}$ ):

$$[\text{Fe}]_{\text{orgads}} = k_{\text{Fe}_{\text{org}}} \times [\text{Fe}]_{\text{prime}} \quad (26)$$

195 and particulate  $\text{CaCO}_3$  ( $[\text{Fe}]_{\text{orgads}_{\text{ca}}}$ ):

$$[\text{Fe}]_{\text{orgads}_{\text{ca}}} = k_{\text{Fe}_{\text{ca}}} \times [\text{Fe}]_{\text{prime}} \quad (27)$$

calculated as a function of free organic detritus and carbonate particles, respectively, a scavenging rate constant ( $k_{\text{Fe}_{\text{org}}}$  and  $k_{\text{Fe}_{\text{ca}}}$ ), iron available for scavenging ( $[\text{Fe}]_{\text{prime}}$ ), and inorganic precipitation of dissolved iron into colloidal material ( $[\text{Fe}]_{\text{col}}$ ), which is calculated as a linear function independent of organic particle concentration and unchanged from Nickelsen et al. (2015). While calcite is known to be a powerful scavenger of iron and other trace metals (Olsson et al., 2014), the strength of plankton-derived calcite in scavenging dissolved iron is unquantified. A test of this sensitivity is planned at a later stage, so for now the calcite scavenging parameter value is set equal to that of organic detritus.

### 2.2.6 Calcite

As in Kvale et al. (2015b), the source and sink terms for  $\text{CaCO}_3$  include both phytoplankton calcifier and zooplankton sources from grazing and mortality, and losses from dissolution and sinking:

$$S([\text{CaCO}_3]) = ((1 - \gamma)(G_{CP} + G_Z) + m_{CP}CP + m_ZZ) R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}} - \lambda_{\text{CaCO}_3}[\text{CaCO}_3] - w_C \frac{\partial[\text{CaCO}_3]}{\partial z} \quad (28)$$

where  $w_C$  is the sinking speed of calcite, and the dissolution specific rate ( $\lambda_{\text{CaCO}_3}$ ) is calculated using a parameterisation from Aumont et al. (2003); Kvale et al. (2015b).

Calcite associated with living plankton is calculated separately (but not traced) as:

$$S[\text{CaCO}_3]_{\text{liv}} = (S(CP) + S(Z)) R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}}. \quad (29)$$

### 2.2.7 Opal

The production of biogenic opal is calculated as a function of the diatom grazing and linear mortality loss terms:

$$\text{Pr}(\text{Opal}) = ((1 - \gamma) G_{DT} + m_{DT}DT) R_{\text{Opal:POC}} R_{\text{C:N}} \quad (30)$$

where  $R_{\text{Opal:POC}}$  is a production ratio that varies as:

$$R_{\text{Opal:POC}} = R_{\text{Opal:POC},0} \left( \min\left(\frac{[\text{Si}]}{k_{\text{Si}}}, 1\right) (4 - 3 \times \min\left(\frac{[\text{Fe}]}{k_{\text{FeDT}}}, 1\right)) \right). \quad (31)$$

This parameterisation was introduced by Aumont et al. (2003) and yields an average surface opal:free detritus export value of around 1 across the Southern Ocean, using a fixed average ratio ( $R_{\text{Opal:POC},0}$ ) of 0.5. Production of lithogenic opal occurs mostly on land (Tréguer and De La Rocha, 2013), so its contribution to marine silicate cycling is included simplistically via the dissolved silicate river flux calculation.

We present a novel opal dissolution parameterisation based on evidence that opal dissolution rates increase when organic coatings are stripped away by strong bacterial activity (Sarmiento and Gruber, 2006). We approximate an exponential flux function and apply our e-folding temperature parameterisation to represent microbially enhanced dissolution in  $\text{mol Si m}^{-3}$  units:

$$\text{Di}(\text{Opal}) = \int \text{Pr}(\text{Opal}) dz \frac{d}{dz} \left( \frac{\lambda_{\text{Opal}}}{w_{\text{Opal}}} \exp\left(\frac{T}{T_b}\right) \right). \quad (32)$$

225 This parameterisation results in greater dissolution at warm temperatures and is similar to the instant-sinking-and-dissolution function applied to model calcite (Schmittner et al., 2008) (although, the function for calcite was replaced when a prognostic tracer was added by Kvale et al., 2015b). We find this parameterisation offers simplicity and consistency with other temperature-dependent rates in KMBM3, as well as adding mechanistic realism, and improving model fit to World Ocean Atlas silica distributions (relative to other parameterisations that we tested, e.g. the temperature-dependent parameterisation of Gnanadesikan (1999) or the temperature and oxygen-dependent parameterisation of Enright et al. (2014)). The Gnanadesikan (1999) parameterisation yields lower dissolution rates at low temperatures than the Enright et al. (2014) parameterisation, which is similarly formulated but which includes an additional oxygen scaling. The Enright et al. (2014) oxygen scaling is not justified in their model description, but it has the effect of increasing Si dissolution rates in the deep ocean (exacerbating the overestimation of Si dissolution in this region by the Gnanadesikan (1999) scaling described in Ridgwell et al. (2002)) and decreasing Si dissolution rates (to a lesser extent) in the near-surface. Our temperature scaling has the effect of raising dissolution rates at the surface.

### 2.2.8 Particle Sinking

Detritus (Schmittner et al., 2005), calcite (Kvale et al., 2015b), and iron (Nickelsen et al., 2015) particles are exported from the surface with a sinking speed ( $w$ ) that increases linearly ( $wdd$ ,  $wdc$ ) with depth ( $z$ ; Berelson 2001) for calcite and ballasted organic detritus:

$$w_C = w_{C,0} + wdc \times z \quad (33)$$

and free detritus and associated iron:

$$w_D = w_{D,0} + wdd \times z. \quad (34)$$

Alternative parameterisations exist and their effects on fluxes and model performance make for interesting comparisons (e.g., Cael and Bisson, 2018), but we do not explore them here. The initial surface sinking speeds of particles are assigned different values to represent the denser structure of  $\text{CaCO}_3$  relative to that of organic detritus. Ballasted detritus sinks at the  $\text{CaCO}_3$  speed, but once it enters the free pool it uses the organic detrital sinking speed and remineralisation rate. Any organic detritus reaching the sediments is dissolved back in to the water column to ensure conservation of carbon and phosphate in the model domain. Calcite particles that reach the seafloor enter the sediment model (Kvale et al., 2015b) if it is active, though we do not use it here (in this case the particles dissolve). Iron detritus reaching the sediments is lost from the ocean, unless bottom water oxygen falls below  $5 \text{ mmol m}^{-3}$ , whereupon detrital iron reaching the bottom is dissolved back into the water column (Nickelsen et al., 2015). By definition, iron is not mass-conserving in the model, but is formulated with open boundary conditions (atmosphere and sediments) - hence it is not just the oxygen threshold that can cause a loss or gain of marine iron. In the model the oxygen threshold is only met along coastal boundaries in the north Pacific under modern conditions. A sedimentary release of dissolved iron ( $[\text{Fe}]_{\text{sed}}$ ) is prescribed as in Nickelsen et al. (2015):

$$[\text{Fe}]_{\text{sed}} = R_{\text{Fe:P}_{\text{sed}}} F_{\text{POP}} \exp \frac{T}{T_b} \quad (35)$$

where  $R_{Fe:P_{sed}}$  is a ratio of iron released from sediment in proportion to the flux of phosphorus in the organic detritus ( $F_{POP}$ , which includes free and ballast detritus) reaching the bottom.

Opal reaching the seafloor is either returned to the silicic acid tracer or considered lost to the sediments (bSi) according to criteria laid out in Sarmiento and Gruber (2006):

$$\text{For } F(\text{Opal}) > 2 \text{ mmol m}^{-2} \text{d}^{-1} \quad b\text{Si} = 0.3 \times F(\text{Opal})$$

$$\text{For } F(\text{Opal}) \leq 2 \text{ mmol m}^{-2} \text{d}^{-1} \quad b\text{Si} = 0.05 \times F(\text{Opal}) \quad (36)$$

Opal flux,  $F(\text{Opal})$ , is calculated as [depth-integrated](#) opal production ( $Pr(\text{Opal})$ ) less the [depth-integrated](#) instantaneous dissolution term ( $Di(\text{Opal})$ ). The model conserves silica when river fluxes are allowed to compensate for the net change due to the ocean sedimentary sink, hydrothermal input, and atmospheric deposition.

The sub-grid bathymetric scaling that was introduced for the iron model in Nickelsen et al. (2015) is extended here to apply to all particle fluxes reaching the bottom ocean grid. This scaling feature calculates a sedimentary exchange factor based on the proportion of each grid cell that falls outside of the model grid's depth according to a high-resolution bathymetric dataset. This scaling is used to account for high-relief bathymetric features, such as ridges and troughs, in the sediment transfer functions. We do not use the sediment model in this manuscript.

## 2.2.9 Dissolved Inorganic Tracers

Ocean nutrient sources and sinks ([in concentration/time units](#)) follow:

$$\begin{aligned} S([PO_4^{3-}]) = & (\mu_D Detr_{free} + \mu_{LP}^* LP + \mu_{CP}^* CP + \mu_{DT}^* DT + (\gamma - \varpi)(G_{LP} + G_{CP} + G_{DT} + G_{DZ} \\ & + G_{Detr_{free}} + G_Z) - J_{LP} LP - J_{CP} CP - J_{DT} DT - J_{DZ} DZ) R_{P:N} \end{aligned} \quad (37)$$

$$\begin{aligned} S([NO_3^-]) = & (\mu_D Detr_{free} + \mu_{LP}^* LP + \mu_{CP}^* CP + \mu_{DT}^* DT + (\gamma - \varpi)(G_{LP} + G_{CP} + G_{DT} + G_{DZ} \\ & + G_{Detr_{free}} + G_Z) - J_{LP} LP - J_{CP} CP - J_{DT} DT - u_{NO_3} J_{DZ} DZ)(1 - 0.8 R_{O:N} r_{sox}^{NO_3^-}) \end{aligned}$$

where  $R_{P:N}$  and  $R_{O:N}$  are Redfield molar ratios and  $u_{NO_3}$  is [nitrate availability \(diazotrophs use nitrate when available\)](#). In suboxic water (less than  $5 \text{ mmol m}^{-3}$ ), oxygen consumption is replaced by denitrification ( $r_{sox}^{NO_3^-}$ ):

$$r_{sox}^{NO_3^-} = \max(0, 0.5(1 - \tanh([O_2] - 5))). \quad (38)$$

There are no additions of phosphate, nitrate, or oxygen along the boundary (with the exception of air/sea gas exchange in the case of oxygen) (Keller et al., 2012).

Dissolved iron includes sources and sinks of particulate iron mentioned above, as well as prescribed dust deposition ( $[Fe]_{dust}$ ) as in Nickelsen et al. (2015) and hydrothermal iron ( $[Fe]_{hydr}$ ) (Muglia et al., 2017) boundary terms ([also in concentration/time](#)

units):

$$\begin{aligned}
285 \quad S([\text{Fe}]) &= R_{\text{Fe:N}} (\mu_D \text{Detr}_{\text{tot}} + \mu_{LP}^* LP + \mu_{CP}^* CP + \mu_{DT}^* DT + (\gamma - \varpi)(G_{LP} + G_{CP} + G_{DT} + G_{DZ} \\
&\quad + G_{\text{Detr}_{\text{tot}}} + G_Z) - J_{LP} LP - J_{CP} CP - J_{DT} DT - J_{DZ} DZ) \\
&\quad - [\text{Fe}]_{\text{orgads}} - [\text{Fe}]_{\text{col}} + \mu_D \text{Detr}_{\text{Fe}}
\end{aligned} \tag{39}$$

$$B([\text{Fe}]) = [\text{Fe}]_{\text{sed}} + [\text{Fe}]_{\text{dust}} + [\text{Fe}]_{\text{hydr}} \tag{40}$$

The Nickelsen et al. (2015) model applied the iron dust flux to the surface ocean after the biological routine and before the  
290 mixing routine. This resulted in low model sensitivity to iron dust inputs, because the dust was mixed away [faster than the biological processes](#) had a chance to access it. In this version, the iron dust flux is added prior to the biological routine (which operates on a shorter timestep than ocean mixing), and results in a greater biological sensitivity to iron dust flux.

DIC and alkalinity tracer sources and sinks are a function of sources and sinks of prognostic  $\text{CaCO}_3$  (Kvale et al., 2015b).

$$\begin{aligned}
295 \quad S([\text{DIC}]) &= S([\text{PO}_4^{3-}]) R_{\text{C:P}} + \lambda_{\text{CaCO}_3} [\text{CaCO}_3] - S[\text{CaCO}_3]_{\text{liv}} \\
&\quad - ((1 - \gamma)(G_Z + G_{CP}) + m_Z Z + m_{CP} CP) R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}}
\end{aligned} \tag{41}$$

$$\begin{aligned}
S([\text{Alk}]) &= -S([\text{PO}_4^{3-}]) R_{\text{C:P}} + 2 (\lambda_{\text{CaCO}_3} [\text{CaCO}_3] - S[\text{CaCO}_3]_{\text{liv}}) \\
&\quad - 2 ((1 - \gamma)(G_Z + G_{CP}) + m_Z Z + m_{CP} CP) R_{\text{CaCO}_3:\text{POC}} R_{\text{C:N}}
\end{aligned} \tag{42}$$

With the exception of air/sea gas exchange of  $\text{CO}_2$  there are no boundary additions of DIC or alkalinity when the sediment  
300 model is deactivated (as is presented here).

Dissolved silicic acid tracer sources, sinks, and boundary terms (all in concentration/time units) follow:

$$S([\text{Si}]) = \text{Di}(\text{Opal}) - \text{Pr}(\text{Opal}) \tag{43}$$

$$B([\text{Si}]) = [\text{Si}]_{\text{riv}} + [\text{Si}]_{\text{dust}} - [\text{Si}]_{\text{sed}} + [\text{Si}]_{\text{hydr}} \tag{44}$$

where [discharge](#) from rivers ( $[\text{Si}]_{\text{riv}}$ ) is used as a budget balancing term to compensate for any remainder in the other  
external sources and sinks: windborne dust ( $[\text{Si}]_{\text{dust}}$ ), hydrothermal silicate ( $[\text{Si}]_{\text{hydr}}$ ), and loss to the sediments ( $[\text{Si}]_{\text{sed}}$ ). Dust  
deposition from the atmosphere is prescribed using an interpolated monthly pre-industrial dust flux derived from the NCAR's  
Community Climate System Model (Mahowald et al., 2006). The silica content of the dust is derived from maps (Zhang  
et al., 2015), with a global average dust solubility of 3% assumed to produce annual bioavailable fluxes in agreement with  
estimates (Sarmiento and Gruber, 2006; Tréguer and De La Rocha, 2013). Observations demonstrate wide spatial variability  
310 in the solubility parameter (Tréguer and De La Rocha, 2013) so our single value represents a simplification. Silicate from  
hydrothermal sources are prescribed using a static mask scaled from the hydrothermal iron mask (Muglia et al., 2017) using a  
Fe:Si ratio to obtain the estimated annual total contribution from hydrothermal sources in Sarmiento and Gruber (2006).

### 3 Model Assessment in a Modern Climate

315 The model is spun up to equilibrium (greater than 15,000 years) at year 1765 boundary conditions, then forced with historical data for comparison to observational datasets. Forcing includes historical atmospheric CO<sub>2</sub> concentrations, agricultural land cover, volcanic radiative forcing, sulphate aerosol and CFC concentrations, changes in land ice and solar forcing (Machida et al., 1995; Battle et al., 1996; Etheridge et al., 1996, 1998; Flückiger et al., 1999, 2004; Ferretti et al., 2005; Meure et al., 2006). Solar insolation at the top of the atmosphere, wind stress, and wind fields vary seasonally (Kalnay et al., 1996), and  
320 wind fields are geostrophically adjusted to air temperature anomalies (Weaver et al., 2001).

Table 6 lists key biogeochemical properties diagnosed by the model for a modern climate, as well as corresponding properties diagnosed from the Keller et al. (2012) version. We compare model output at year 2004, although data sources reflect a range of collection and publication dates. Net primary production (NPP) is similar to previous model versions (52.3 Pg C y<sup>-1</sup>, e.g. compared to 54.3 Pg C y<sup>-1</sup> in Keller et al. 2012), and still within the literature range of 44–78 Pg C y<sup>-1</sup> (e.g., Carr et al., 2006;  
325 Jin et al., 2006). Calcite production is similar to and nitrogen fixation rates are somewhat improved with respect to earlier model versions, though remain too low, as do deep particle fluxes of particulate organic carbon (POC) and shallow and deep calcite (also referred to as PIC, for particulate inorganic carbon). Diagnosed surface opal production and deep ocean (2 km depth) opal flux are about 50% less than the Tréguer and De La Rocha (2013) estimates. Accordingly, there is too little dissolution of opal throughout the water column (less than half of the observational estimate in Tréguer and De La Rocha, 2013). However, the  
330 calculated flux of silicate through the seafloor is nearly 3 times higher than estimated by Tréguer and De La Rocha (2013). An overestimation of seafloor flux is potentially corrected with an increase in open ocean opal dissolution rates or an increase in benthic return. However, a reduction of seafloor silicate flux would result in an even smaller river silicate input (which is already less than half the observational estimate). Another potential model correction could be the addition of a siliceous sponge sink in the model, which could reduce the seafloor flux without reducing the river flux. The strength of the sponge sink has been  
335 estimated at  $3.7 \pm 3.6$  Tmol Si y<sup>-1</sup> (Tréguer and De La Rocha, 2013), but the authors prefer to delay implementation until these large error bars can be reduced. It should be noted that the global silica cycle may be unbalanced between sources and sinks at present and observations of global input and output rates are both affected by anthropogenic perturbations and have large uncertainties (Tréguer and De La Rocha, 2013). Atmospheric and hydrothermal silica inputs are fixed in these simulations, but each source and sink of silicate produces a unique spatial distribution in the water column. These terms are slated for automated  
340 calibration in the future (e.g., Yao et al., 2019; Kriest, 2017; Kriest et al., 2017), thus, our hand-tuned simulations are meant to serve as a baseline for future improvements to the model parameter values.

Total global phytoplankton biomass is on the low end, but within the range of, previous estimates (0.80 Pg C, increased from 0.53 in Keller et al. 2012, compared to 0.5–2.4 Pg C from Buitenhuis et al. 2013; Table 6). KMBM3 explicitly represents only a fraction of the ecological complexity found in the real ocean, which causes our biomass estimates for the phytoplankton  
345 types to look dissimilar to the Buitenhuis et al. (2013) biomass estimates (e.g., with low latitude mixed phytoplankton (a model-specific category, LP) having a biomass only 25% of the lowest picophytoplankton estimate and both diazotrophs (DZ) and calcifiers (CP) having a biomass 4 times greater than the upper observational estimate). These phytoplankton types proved

particularly difficult to tune by hand, with small variations in parameter values causing extinction. An over-estimate of calcifiers is compensated by the low PIC:POC production ratio (0.07), which is meant to represent 7% of the phytoplankton class having a PIC:POC production ratio of 1 (compare to a ratio for *Emiliana huxleyi* of 0.51–2.30 from Paasche 2001). Overestimated diazotroph biomass results in an increase ( $0.21 \text{ Pg N y}^{-1}$ ) in nitrogen fixation compared to earlier model versions, which is still improved with respect to, but still lower than the observational independent estimate. In KMBM3, diazotrophs use preformed nitrate when available. Thus in our modelling context, this phytoplankton type can be considered “slow-growing phytoplankton capable of fixing nitrogen when necessary”. Constraints on this phytoplankton type will be explored in the future. Diatom biomass estimates are comparatively reasonable and are within the Buitenhuis et al. (2013) range.

Looking next at spatial distributions of biological rates, Figure 2 compares KMBM3 NPP at year 2014 to the Westberry et al. (2008) model applied to MODIS (NASA, 2018) climatology from 2012-2018. As is repeatedly found in the KMBM (see plot of the Keller et al. 2012 model NPP), open-ocean primary production rates are generally too high, particularly in the eastern equatorial Pacific and northern Indian Ocean. These very high-production zones compensate in the globally integrated rate estimate for the diffuse and more widespread production calculated from satellites. The spatial pattern in NPP has changed very little from earlier model versions, with a continued under-estimate of NPP occurring within gyres and an over-estimate of NPP in the 40-60 degree N and S ranges.

Spatial biases in NPP are also held in surface calcite concentrations. Figure 3 compares KMBM3  $\text{CaCO}_3$  averaged between 2004-2014 with the 2002-2018 climatology data from MODIS (NASA, 2018). Model calcite is too high in the North Pacific and Southern Ocean between 40 and 60 S, and too low south of 60 S. Upwelling zones and the Indian Ocean have an over-estimated role in calcite production, and coastal calcite production is either not resolved, or under-estimated.

Phytoplankton biomass is compared to the MAREDAT datasets (Leblanc et al., 2012; Luo et al., 2012; O'Brien, 2012) in Figure 4. Globally integrated diatom biomass is within observational estimates and the diatom geographical distributions agree roughly with the very sparse Leblanc et al. (2012) dataset (highest concentrations in the Southern Ocean, North Pacific, and North Atlantic), though the model appears to over-estimate the North Atlantic biomass. Like the globally integrated biomass estimate, calcifiers (CP) are universally over-estimated compared to O'Brien (2012). As explained above, this phytoplankton type is meant to represent a variety of moderate growth, moderate nutrient affinity phytoplankton types, including those that calcify. Therefore it is not unexpected to have an over-estimate of the biomass. Diazotroph biomass is primarily concentrated in the tropics, which agrees spatially but not in magnitude with the limited data of Luo et al. (2012). Mixed low-latitude phytoplankton (LP) is a model-specific category with no clear analogue in the MAREDAT dataset.

Interior ocean particle flux performance is encouraging. Figure 5 compares model POC, PIC, and opal fluxes at 2 km depth to the Honjo et al. (2008) data compilation. The model shows a general under-estimate with respect to the observations for all particle fluxes, particularly for intermediate rates. Root mean square error is improved beyond Kvale et al. (2015a) for PIC (103.1 compared to 147.1), and POC (96.4 versus 98.0 in Kvale et al. 2015a). Root mean square error for opal flux at 2 km depth is 221.0. However, the model captures large-scale features of high flux rates in the north west Pacific and Southern Ocean, as well as moderate flux rates in the Indian and North Atlantic basins.

Particle flux rates, alongside the model's ocean circulation, impact ocean nutrient distributions. Figure 6 shows carbon and nutrient profiles, globally and for each basin, compared to GLODAP (Key et al., 2015; Lauvset et al., 2016) and World Ocean Atlas (WOA) (Garcia et al., 2014a, b) data. Nutrients are generally too high at depth, and oxygen is too low, as a result of over-estimated POC flux rates in the Southern Ocean and North Atlantic. Global root mean squared error is listed in the figure for each model dissolved inorganic tracer. All tracers except phosphate and oxygen show improvement with respect to the Keller et al. (2012) model version. Alkalinity is primarily affected by PIC flux rates, and therefore shows relatively good agreement (global RMSE value of 0.332) with observations. Silicic acid has a global root mean squared error of 0.389, making it one of the better-performing dissolved tracers. However, two problematic ocean basins are apparent in profile; concentrations are too high in the Atlantic by as much as 50 mmol Si m<sup>-3</sup> and too low in the Indian by as much as 50 mmol Si m<sup>-3</sup>.

Greater basin and surface detail in carbon and nutrient concentrations is displayed in Figures 7-12. Interior ocean alkalinity is too high by as much as 50  $\mu\text{mol kg}^{-1}$ , and surface alkalinity too low by as much as 50  $\mu\text{mol kg}^{-1}$ , in the Atlantic, as it was with previous model versions (Eby et al., 2009; Keller et al., 2012; Kvale et al., 2015a). Alkalinity in the interior Pacific is now slightly too low, and the surface too high by as much as 100  $\mu\text{mol kg}^{-1}$  along the Equator, whereas previously it was also over-estimated at depth and under-estimated at the surface (Kvale et al., 2015a). The physical circulation has not changed since previously published model versions, hence the new anomalies can be attributed to shifts in phytoplankton biogeography and calcification rates, and are consistent with a now lower calcite production rate in the model compared to Kvale et al. (2015a). The Indian Ocean interior alkalinity is similarly under-estimated, by as much as 50  $\mu\text{mol kg}^{-1}$ . Deep ocean DIC is improved with respect to past model versions (Figure 8), though (at least compared to Kvale et al. (2015a)) this improvement might be at least partly attributed to the use of a transient year 2014 model output that includes an anthropogenic signal, instead of the pre-industrial spinup. The deep north Pacific shows a low bias of up to 50  $\mu\text{mol kg}^{-1}$ , similar to previous versions. Surface DIC anomalies are also similar to previous model versions, with DIC being too low in the western Pacific (a consequence of too high export production in the model and physical biases), and too high in the surface Southern Ocean (possibly due to upstream production in the 40-60 S band being too high, which can increase regional surface concentrations). As with earlier model versions, deep ocean phosphate and nitrate concentrations (Figures 9 and 10) are also generally too high, particularly in the Southern Ocean-sourced deep and intermediate water masses in the Indian and Pacific sectors. Oxygen anomalies mirror nutrient biases (Figure 11), with oxygen being up to 50 mmol m<sup>-3</sup> too low along the Southern Ocean particle conveyor belt, and up to 50 mmol m<sup>-3</sup> too high in the sub-surface tropical ocean. The oxygen bias in the deep water masses can be addressed with better tuning of the biological production, export and flux parameters, but the tropical ocean deficiencies will require improvements to both the biogeochemistry as well as the physics (Oschlies et al., 2017).

Silicic acid distributions are reasonably well captured by the model (Figure 12). The model simulates the deep Southern Ocean maximum to within 40 mmol Si m<sup>-3</sup>, with a high bias in the deep Atlantic, Pacific, and especially Indian sectors. Silicic acid distributions are almost entirely controlled by diatom biogeography in KMBM3 (with hydrothermal, dust, and riverine fluxes having little impact on overall patterns), so this spatial weighting to the Indian sector might represent the high bias in diatom biomass in this basin (also clearly seen in the opal flux plots). North Atlantic silicic acid gradients are well represented, while a low bias is apparent in the deep North Pacific of around 20 mmol Si m<sup>-3</sup>. A low bias is also simulated in the surface

North Pacific, which possibly suggests deficiencies in the circulation within and between regional marginal seas (Nishioka et al., 2020). The North Pacific appears to perform well with respect to deep opal flux (Figure 5), but small biases in fluxes can compound with time in dissolved nutrient fields. It might also be that sedimentary processes that influence deep water silicic acid concentrations would be important to resolve in this region. Our low bias result in the deep North Pacific is interesting because the origins of the silicic acid-rich deep ocean plume in this region are still under debate (Tréguer and De La Rocha, 2013).

Seasonal succession in phytoplankton types follows the general progression of zonal maxima in diatoms preceding calcifiers by a few weeks (albeit, in separate zonal ranges, Figure 13). This succession is due to the higher nutrient requirements, and faster growth rates, of the diatoms, which are able to take advantage of winter mixing early in the growing season. Once surface nutrient concentrations start to decline, the calcifying phytoplankton become relatively more successful. This pattern is most pronounced in the Southern Ocean. In the low latitudes, diazotroph biomass peaks earlier in the growing season than the low-latitude, non-calcifying phytoplankton (LP). Diazotrophs have a growth handicap with respect to LP, but their ability to fix nitrate gives them an advantage in more stratified summer conditions. This fixed nitrate is then used by the LP in the winter months.

## 4 Model Assessment Under Climate Change

In addition to the historical model forcings described earlier, from year 2005 to 2300 the simulations are forced using increasing CO<sub>2</sub> and non-CO<sub>2</sub> greenhouse gas concentrations, projected changes to the fraction of the land surface devoted to agricultural uses (calculated to year 2100 by Hurtt et al. (2011), and then held constant after), and changes in the direct effect of sulphate aerosols following “business-as-usual” RCP scenario 8.5 (RCP8.5, Riahi et al., 2007; Meinshausen et al., 2011). The wind fields continue to be geostrophically adjusted to air temperature anomalies.

### 4.1 Historical Changes (1964-2014)

We next explore model trends from the 1960s to the 2010s and compare to available data from this period. Significant changes are simulated to have already occurred in NPP and phytoplankton biomass, with most of the change over this period occurring since the 1980s (Figure 14). Global NPP is simulated to have declined 1.8% between 1964 and 2014, with the strongest declines in the tropics, northern mid-latitudes, and Southern Ocean due to increasing thermal stratification. That decline is dominated by the simulated loss of diatom biomass (a 8.2% decline) due to their high nutrient requirements, largely in the Southern Ocean (Figure 15). Diazotrophs are simulated to have experienced a 2.6% loss in biomass globally over this period. Calcifying phytoplankton and low latitude non-calcifying phytoplankton are simulated to have experienced a 2% and 1% gain, respectively, in net biomass. Increases in calcifying phytoplankton are simulated to have occurred in the Arctic and in the Southern Ocean and decreases are simulated to have occurred in the middle latitudes, where LP have increased their biomass. Warming, and a lengthening growing season (but increased stratification; see Arctic temperature trend in Figure 16) in the Arctic benefits calcifiers. Expansion of coccolithophores into the Arctic has been observed over recent decades (Neukermans

et al., 2018). In KMBM3 the Southern Ocean is also simulated to have experienced increasingly favourable conditions for calcifiers, with diatom biomass declining as calcifiers increase.

KMBM3 simulates a slower global decline in productivity and biomass than the scarce, and controversial, satellite and in-situ chlorophyll record reconstructed by Boyce et al. (2010), who calculated a 1% per year decline in chlorophyll. KMBM3 also does not simulate a large decline prior to the 1950s (Figure 17). Wernand et al. (2013) found no historical trend in chlorophyll in their Forel-Ule ocean colour scale proxy from 1899 to 2000. However, both chlorophyll reconstructions (and KMBM3) suggest strong regional variation in the historical trends. KMBM3 also simulates a smaller decline in global NPP than an earlier modelling effort, which obtained a 6.5% decline between 1960-2006 (Laufkötter et al., 2013). Their simulation similarly resulted in large declines in production in the low latitudes, which they attributed to warming and stratification over the historical period. KMBM3 simulates strong warming in the low latitudes (up to 0.5 degree warming in the zonal mean between 1964 and 2014; Figure 16). Zonal mean trends in natural radiocarbon (also Figure 16) are also positive in the upper ocean, suggesting enhanced stratification and reduced vertical mixing, particularly in the tropical and subtropical Pacific and Indian Ocean basins (this is without inclusion of the Seuss effect from fossil fuel emissions). Laufkötter et al. (2013) also found strong changes in phytoplankton biogeography in the Southern Ocean and north Atlantic, which they attributed to increasing zonal wind stress and vertical mixing, and surface freshening inducing stratification, respectively. However, our models do not agree in the response of diatoms and high-latitude calcifiers, for which their model simulates high latitude increases in diatoms. Our results are more similar to those of Rousseaux and Gregg (2015), who combined a model with satellite data to reconstruct ocean surface changes from 1998-2012. They simulated a global decline in diatoms over this period, which they attributed to an increase in nutrient limitation and photosynthetically available radiation, which favours other phytoplankton types. However, the Southern Ocean in their model showed no clear trend in phytoplankton community composition between 1998-2012 (Rousseaux and Gregg, 2015). The model we present here is fully competition-driven. Variable particle sinking and remineralisation rates, and explicit  $\text{CaCO}_3$  ballasting, further differentiate KMBM3. All of these factors may contribute to the different behaviour reported here.

Trends in biomass and NPP affect deep particle fluxes. Low latitude declines in primary production result in less deep ocean particle export in the western Pacific and Indian Ocean basins, while deep POC and PIC export is simulated to have slightly increased in the Indian and Pacific sectors of the Southern Ocean, and in the Arctic. Trends in deep ocean carbon particle export in KMBM3 generally follow trends in calcifier (CP) biomass (Figure 15), as  $\text{CaCO}_3$  (PIC) ballasting contributes significantly to deep export (Kvale et al., 2015b, a, 2019). Declining POC export in the North Atlantic is also simulated to have occurred, though in this region the trend is driven by an increase in diatoms, which are less efficient exporters of organic carbon. Deep ocean opal export is simulated to have declined almost globally, with the largest loss in the Southern Ocean. Opal dissolution is temperature-dependent, therefore regional warming is almost certainly contributing to the reduction of deep ocean opal flux, but the loss of diatom biomass is the major driver of this trend in the Southern Ocean. Global losses of particle export across 2 km depth between 1964 and 2014 are calculated at 1.8% (POC), 1.0% (PIC), and 7.8% (opal).

Unfortunately there is no comparable historical reconstruction of deep particle fluxes over this time period. Just as with NPP, Laufkötter et al. (2013) simulated a larger decline (8%) in export production (POC) between 1960 and 2006, which they

calculated at 100 m depth. They also simulated the largest declines in the Indian Ocean and west-central Pacific basins, driven  
485 by declines in NPP. The models disagree with respect to trends in the north Atlantic, with KMBM3 producing a decline in POC  
and PIC deep export (due to a decrease in calcifiers), and only a small increase in opal (due to an increase in diatoms). Both  
Laufkötter et al. (2013) and KMBM3 simulate a historical increase in diatoms in this region; the difference in export trends  
can be explained by which phytoplankton type (calcifiers or diatoms) is more efficient at POC export, with calcifiers being the  
more efficient carbon exporter in our formulation. Likewise, KMBM3 simulates different POC export trends in the Southern  
490 Ocean as Laufkötter et al. (2013), due to differences in model structure. Increasing calcifiers in the Indian sector in KMBM3  
increase POC and PIC export there, while in Laufkötter et al. (2013) diatoms regionally increase.

Changes in carbon and nutrient profiles between 1964 and 2014 (Figure 18) reflect a combination of physical-chemical  
uptake of anthropogenic CO<sub>2</sub> and the changes in ocean circulation on tracer accumulation (Figures 16 and 20). Anthropogenic  
carbon intrudes particularly quickly into the north Atlantic and Southern Ocean, where dissolved inorganic carbon (DIC) is  
495 simulated to have increased by more than 10 mmol C m<sup>-3</sup> in zonal mean over this time period. Both regions, but particularly the  
Southern Ocean, are thought to be the primary regions for anthropogenic carbon uptake into the ocean interior (e.g. Khatiwala  
et al., 2009). The low latitudes are primary regions of carbon storage (Frölicher et al., 2015), and strong (greater than 25 mmol  
C m<sup>-3</sup>) increases in DIC are also seen here, in all ocean basins, in the zonal mean profiles.

Declines in phosphate and nitrate are simulated in the upper ocean in all basins, with the largest zonal mean declines of  
500 up to 0.1 mmol P m<sup>-3</sup> and 0.5 mmol N m<sup>-3</sup>). Silicic acid **increases by up to 20 mmol Si m<sup>-3</sup> in the central North Pacific  
surface (Figure 19) due to a regional shift from DT to CP dominance**. Dissolved iron shows an increasing trend in the tropical  
surface. Surface and subsurface (300 m depth) concentration trends (Figure 19) reveal spatial heterogeneity; with declining  
surface and increasing sub-surface phosphate occurring in the subarctic North Pacific (a result of increasing particle export),  
and increasing concentrations along the Humboldt Current (driven by a regional reduction in nitrate, not shown). These results  
505 compare favourably with surface declines in phosphate recorded in the North Pacific between 1961 and 2012 (Yasunaka et al.,  
2016). They also compare favourably with the recent data compilation of Stramma et al. (2020), which shows a decline in  
surface nutrients, and an increase in subsurface nutrients, in the subarctic Pacific. In KMBM3, this pattern is produced by the  
replacement of diatoms with calcifiers, who are more efficient exporters of nutrients. The UVic ESCM lacks a fully dynamic  
atmosphere model, and therefore does not simulate multi-decadal oscillations in climate, which have been implicated in recent  
510 Pacific interior nutrient trends (Stramma et al., 2020). This may be why KMBM3 does a poor job reproducing observations  
of nutrient trends observed in the central Pacific (Stramma et al., 2020). Also, increases in nitrate are found in the observed  
record, which KMBM3 does not simulate. This may be because KMBM3 does not include anthropogenic sources of nitrate  
(summarized by Stramma et al., 2020).

Significant increases in North Atlantic deep ocean concentrations (below 3000 m depth) occur in phosphate, nitrate, and  
515 silicic acid (Figure 18). A decline in maximum meridional overturning (MOC) of about 1 Sv is apparent over this time, and  
the water has warmed more than 0.3 degrees (Figure 20). Warming of the Gulf Stream increases particle remineralisation rates,  
thereby raising nutrient concentrations along the North Atlantic Deep Water pathway.

Southern Ocean-sourced intermediate and deep water is simulated to have increased nutrient concentrations from 1964 to 2014, with a positive trend in phosphate and nitrate outcropping along the Antarctic ice margin, in qualitative agreement with the limited observations of an increasing trend in the Indian sector from 1965-2008 (Iida et al., 2013). The Southern Ocean is simulated to have experienced sub-surface increases in phosphate in the regions also experiencing increases in silicic acid and iron; this is due to the less efficient export of particles by diatoms (locally increasing over this time), relative to calcifiers.

Taken together, these results suggest declines in NPP and export production upstream of the Southern Ocean have introduced excess nutrients to the basin, raising the nutrient concentrations in Antarctic Bottom and Intermediate water masses despite declining Southern Ocean NPP. These production and export effects have been exacerbated by physical changes in the circulation; Figure 16 shows a decline in ideal age (greater than 5 years in the zonal mean) in Pacific Intermediate water and the Indian Ocean, which leads to less particle remineralisation (and hence, lower nutrient concentrations) there. Increasing water mass age in Southern Ocean-sourced intermediate and deep water masses (upwards of 30 years in the deep Pacific), likewise has the effect of producing more complete particle remineralisation, resulting in higher nutrient concentrations.

Oxygen is simulated to have declined in all ocean basins, with the exception of the sub-surface low latitudes. It has been previously estimated that the global ocean lost more than 2% of its oxygen since the 1960s (Schmidtko et al., 2017). KMBM3 simulates a 0.6% decline in total oxygen content from 1964-2014, which is an underestimate resulting from physical biases in our model (i.e., deficiencies in simulating low latitude ventilation), though biogeochemical deficiencies might also be contributing (Oschlies et al., 2017). In the deep ocean, the ageing of water masses (and associated more complete particle remineralisation) contributes to the simulated decline in oxygen. In the upper ocean, warming has reduced oxygen solubility, lowering near-surface concentrations.

## 4.2 Long-term Future Changes (2014-2294)

More significant changes in ocean biogeochemistry are still to come, if applied boundary forcing assumptions hold over the next centuries. Figures 21 to 24 extend the previous analysis to year 2294, with respect to year 2014 biogeochemistry. Critically, spatial patterns in NPP trends reverse, with strong increases, in places exceeding  $80 \text{ gC m}^{-2} \text{ y}^{-1}$  in the zonal mean, in the Southern Ocean (Figure 21). This trend is dominated by the increase in calcifiers (CP), and to a lesser geographical extent, diatoms (DT) (Figure 15). Calcifiers also continue their historical expansion into the Arctic, while diazotrophs are simulated to significantly increase in the middle latitudes after year 2100. Low latitude phytoplankton (LP) also broadly increase in biomass between 20 S and 60 N, though the net NPP trend in the low latitudes remains negative.

This new ecosystem model responds differently to forcing than previous versions. Kvale et al. (2015a) compared the responses of the KMBM1 biogeochemistry (no calcifiers) to two versions of calcifier model, also integrated to 2300 using RCP8.5 forcing, and found the application of calcifiers eliminated the global reduction of NPP found in the Keller et al. (2012) version until around the year 2100. However, the introduction of diatoms and iron, and reorganisation of the phytoplankton community structure, produces an even larger decline in global NPP to 2100 (close to  $5 \text{ Pg C y}^{-1}$ ) in KMBM3 than found in KMBM1 (less than  $1 \text{ Pg C y}^{-1}$ ). The difference appears to be in the low-latitude response, where calcifiers with low nutrient requirements in the previous version maintained NPP despite stratification, partly by supporting diazotrophic nitrogen fixation

through efficient ballast removal of surface nitrate (Kvale et al., 2019). In the current model, calcifiers have a lower biomass in the low latitudes, and do not establish this symbiotic relationship with diazotrophs to the same extent. As a consequence, diazotrophs decline more strongly than previously. After 2100, NPP increases abruptly (Figure 17). Rising NPP over the long-term is a long-standing feature of the KMBM biogeochemical model formulation, and occurs because of the acceleration of nutrient recycling by the temperature-sensitive microbial loop in the low and middle latitudes (Kvale et al., 2015a). In this latest model version, this increase in NPP is much smaller (less than 6 Pg C y<sup>-1</sup> by 2300, compared to 11-13 Pg C y<sup>-1</sup> in previous model versions). Again, this reduced sensitivity in the low latitudes is due to the reduction of low-latitude ballast-forming calcifiers in the new version. The response of calcifiers outside of the low latitudes to ocean changes is also changed from previous versions, in that our model now simulates increases in biomass in the Southern Ocean as well as the Arctic. Whether these differences are competition effects, or due to other model changes, such as the correction of light attenuation, is difficult to assess, but it confirms the finding of Fu et al. (2016) of a strong dependence on phytoplankton community structure in model response to climate change.

Trends in phytoplankton biomass and productivity can be explained by the physical changes in the model over this time (Figures 20 and 22). Maximum meridional overturning circulation (MOC) declines from 18 to 11 Sv over the period 1800 to 2200, before starting to increase after 2200. Slowing overturning helps to accelerate surface warming, and zonal mean temperatures in the upper north Atlantic rise more than 5 degrees by 2294. Globally increased radiocarbon in the upper ocean suggests widespread increased stratification, as well as a more complete separation between upper and lower water masses globally (as previously reported by Kvale et al., 2018, with only slightly different forcing conditions). Ideal age trends similarly show the lengthening Southern Ocean-sourced deep water pathway that extends to ventilate all ocean basins from the south, replacing the shoaled northern ventilation pathways (also described in detail in Kvale et al., 2018). Ideal ages increase more than 300 years in the deep southern Pacific and Atlantic basins, and in the deep north Atlantic, where North Atlantic Deep Water formation has declined. The net effect of these physical changes is an overall decline in low-latitude productivity (driven by increased nutrient limitation) but a strong increase in Southern Ocean productivity, with a faster biogeochemical connection between the surface ocean south of the Polar Front, and the abyssal basins (see the improved ventilation in the radiocarbon plots). At the poles, fast-growing, nutrient-demanding phytoplankton types (DT, CP) thrive, while in the lower latitudes it is the more efficient nutrient consumers (DZ and LP) who benefit.

Long-term particle export trends generally follow the historical trend, but with increasing magnitude (Figures 17 and 23). Globally integrated particle fluxes decline, and remain suppressed with respect to pre-industrial rates, for POC and opal. PIC surface export rates change very little and deep export rates increase with climate forcing as a response to increasing surface calcifier POC export fluxes (e.g., Kvale et al., 2015a). The production of both opal and PIC are scaled against their respective plankton types' POC production, so it is expected that PIC and opal fluxes follow the POC export production trend. Just as with NPP, the POC export production decline in KMBM3 is larger than in previous versions (about 2.0 Pg C y<sup>-1</sup> by 2100, rather than 1.5 PgC y<sup>-1</sup>).

POC and PIC fluxes increase where calcifying phytoplankton biomass also increases; south of 40 S, in the eastern equatorial Pacific upwelling zone, and along the Kuroshio Current into the North Pacific (Figure 23). Strong decreases in calcifiers, and

associated deep carbon export, occur in the Indian Ocean and North Atlantic. Opal export declines by more than 100 mmol Si m<sup>-2</sup> y<sup>-1</sup> both south of 40 S and north of 40 N (with only small changes in the Arctic). However, opal export (and diatoms) increase south of 60 S, where increased nutrients (Figure 24), particularly iron, and a short growing season favours diatoms over calcifiers.

The historical trend in carbon and nutrients is similarly extended, with continuing increases in DIC in the upper ocean (as atmospheric CO<sub>2</sub> continues to enter the ocean), declines in low latitude upper ocean nutrients phosphate, nitrate, and silicic acid (due to decreasing resupply from the deep ocean), increases in the deep ocean in the same nutrients, and widespread declines in oxygen (Figure 24). Oxygen declines along the Southern Ocean- abyssal global ocean pathway due to both warming and increasing particle remineralisation, which is also responsible for the increasing nutrient concentrations in the deep ocean. Decreasing phosphate and nitrate concentrations in the sub-surface tropical ocean basins are a product of declining particle remineralisation there, brought about by both warming, which shoals remineralisation and increases respiration rates, and a shift to less efficiently exporting phytoplankton (LP).

The striking trend in dissolved iron that emerges in these future projections of strongly increasing (more than 80 nmol m<sup>-3</sup> in the zonal mean) concentrations in the upper ocean was previously described by Nickelsen et al. (2015). They attributed the increase to stratification “trapping” aerosol iron near the surface. However, the regions showing the greatest increases in dissolved iron are also the regions experiencing both strong declines in NPP (and hence, lower iron uptake) and strong declines in particle export (and hence, less particulate iron scavenging and removal). The loss of calcifiers in the Indian Ocean and central Pacific particularly increases iron concentrations there, because of the dual effect of reduced POC and PIC scavenging of iron.

Our future simulation results broadly agree with other long-term simulations in the sustained, and significant, increase in Southern Ocean primary production that couples with a reorganisation of deep ocean circulation to produce a long term “nutrient trapping” effect in Southern Ocean-sourced interior water masses (e.g., Moore et al., 2018; Kvale et al., 2019). Near-surface increases in iron, and decreases in nitrate, phosphate, and silicic acid, have also been observed to 2100 in a comparison of 9 other earth system models by Fu et al. (2016). These same models also simulate weak to strong increases in diatoms in the Southern Ocean to 2100, though in most, if not all, of them, diatoms are the most efficient exporters of carbon and nutrients (unlike in KMBM3). Phytoplankton community composition and export formulation was discussed by Fu et al. (2016) to be of critical importance in determining trends in NPP, nutrients, and particle export over the coming century, thus a diversity of model formulations benefits our understanding of how the global ocean ecosystem might change in the future.

## 615 5 Conclusions

Our manuscript describes a new model of the marine silicate cycle (KMBM3), evaluates its performance against previous KMBM versions as well as key biogeochemical data derived from observations of the ocean, and compares long-term ecosystem projections to similar models available in the literature. We find our new model shows general improvement in the repre-

620 sentation of nutrients and particle fluxes and is mechanistically more realistic, with the added complexity of iron, calcite, and silicate merged into a single model code.

Simulations using our new model suggest diatoms have been, and will continue to be, the losers as the earth system warms. Their high nutrient requirements prove a disadvantage as the upper ocean stratifies, and small gains in productivity provided by sea ice retreat cannot compensate for the fact that their southern bound is ultimately limited geographically. Calcifying phytoplankton with more moderate nutrient requirements are the big winners across the high latitudes, while in the tropics 625 slow-growing, less nutrient-hungry phytoplankton are projected to thrive. From a deep ocean carbon sequestration perspective, the loss of diatom export production is of transient importance, as the calcifying phytoplankton increase their role in carbon export, efficiently sinking organic carbon as well as carbonate.

Our simulations also reveal the past may not accurately portray future trends, as evidenced by simulated historical declines in NPP in the Southern Ocean that reverse as conditions become more favourable for calcifiers. Significant and rapid increases in 630 dissolved iron in the low latitude tropical ocean is another potential biogeochemical “surprise”, still to come, if anthropogenic emissions of carbon follow the present trajectory.

Several novel aspects of KMBM3, including iron scavenging by calcite, silicate source and sink strengths, and different zooplankton grazing preferences are slated for further study. The impact of variable stoichiometry is another important potential aspect of biogeochemical modelling that is not explored here. More complete parameter assessment is planned in the context 635 of offline parameter optimisation and model calibration experiments (e.g., Kriest, 2017; Kriest et al., 2017; Yao et al., 2019) in the future, as is merging this new biogeochemical model into the latest UVic ESCM version 2.10 (Mengis et al., 2020). We look forward to further refinements, and the many applications of this model to come.

*Code and data availability.* Data and model code used in the writing of the manuscript is available on the OPeNDAP GEOMAR server at <https://dx.doi.org/20.500.12085/34412098-27f9-4cbb-992d-12d0d342aa45>. The KMBM3 code released with our manuscript is only part of 640 all model code required to use the UVic ESCM version 2.9, Updates 02. The UVic ESCM model code is found at <http://terra.seos.uvic.ca/model/>. The KMBM3 code is provided freely, but with the requirement that prospective users contact K Kvale with their research plans to avoid parallel projects emerging.

## Instructions for model use

The KMBM3 code described, and released, with our manuscript is only part of all code needed to use the UVic ESCM version 645 2.9. UVic ESCM 2.9 Updates 02 must be downloaded from <http://terra.seos.uvic.ca/model/>. Please follow the instructions on this webpage for installation and use.

KMBM3 code released with our manuscript is available at <https://dx.doi.org/20.500.12085/34412098-27f9-4cbb-992d-12d0d342aa45>. The code should be called first in the “mk.in” control file, with subsequent calls to the base model code.

*Author contributions.* KK designed the model, and wrote the model code and paper. DPK, WK, CS, and WY contributed model code and  
650 bug fixes from previous model versions. KJM and AO contributed to the design of the model. All co-authors contributed to the analysis of  
the transient simulations and edited the manuscript.

*Competing interests.* All authors declare no competing interests.

*Acknowledgements.* The authors would like to acknowledge computer resources made available by Kiel University, as well as topical dis-  
cussion with M. Eby at the University of Victoria, plotting scripts from A. Schmittner (Oregon State University) and I. Kriest (GEOMAR),  
655 plotting scripts and data from M. Pahlow (GEOMAR), H. Geisinger who created a hydrothermal silicate mask, H. Saini for code testing,  
and the rest of the Biogeochemical Modelling Department at GEOMAR for their kindness and expertise. KJM acknowledges support from  
the Australian Research Council (DP180100048 and DP180102357). Model tuning was performed in part at the NCI National Facility at  
the Australian National University, through awards under the National Computational Merit Allocation Scheme, the Intersect allocation  
scheme, and the UNSW HPC at NCI Scheme. Figure plotting used the Ferret plotting program. Ferret is a product of NOAA's Pacific Marine  
660 Environmental Laboratory.

## References

- Aumont, O., Maier-Reimer, E., Blain, S., and Monfray, P.: An ecosystem model of the global ocean including Fe, Si, P colimitations, *Global Biogeochemical Cycles*, 17, 2003.
- Balch, W. M.: The Ecology, Biogeochemistry, and Optical Properties of Coccolithophores, *Annual Review of Marine Science*, 10, 71–98, <https://doi.org/10.1146/annurev-marine-121916-063319>, <https://doi.org/10.1146/annurev-marine-121916-063319>, PMID: 29298138, 2018.
- Balch, W. M. and Utgoff, P. E.: Potential Interactions Among Ocean Acidification, Coccolithophores, and the Optical Properties of Seawater, *Oceanography*, 22, <https://doi.org/10.5670/oceanog.2009.104>, 2009.
- Battle, M., Bender, M., Sowers, T., Tans, P., Butler, J., Elkins, J., Ellis, J., Conway, T., Zhang, N., Lang, P., and Clarke, A.: Atmospheric gas concentrations over the past century measured in air from firn at the South Pole, *Nature*, 383, 231–235, <https://doi.org/10.1038/383231a0>, 1996.
- Berelson, W. M.: Particle settling rates increase with depth in the ocean, *Deep Sea Research Part II: Topical Studies in Oceanography*, 49, 237–251, [https://doi.org/10.1016/S0967-0645\(01\)00102-3](https://doi.org/10.1016/S0967-0645(01)00102-3), 2001.
- Boyce, D. G., Lewis, M. R., and Worm, B.: Global phytoplankton decline over the past century, *Nature*, 466, 591–596, <https://doi.org/10.1038/nature09268>, <https://doi.org/10.1038/nature09268>, 2010.
- Buitenhuis, E. T., Vogt, M., Moriarty, R., Bednaršek, N., Doney, S. C., Leblanc, K., Le Quéré, C., Luo, Y.-W., O'Brien, C., O'Brien, T., Peloquin, J., Schiebel, R., and Swan, C.: MAREDAT: towards a world atlas of MARine Ecosystem DATA, *Earth System Science Data*, 5, 227–239, 2013.
- Cael, B. B. and Bisson, K.: Particle Flux Parameterizations: Quantitative and Mechanistic Similarities and Differences, *Frontiers in Marine Science*, 5, 395, <https://doi.org/10.3389/fmars.2018.00395>, <https://www.frontiersin.org/article/10.3389/fmars.2018.00395>, 2018.
- Carr, M.-E., Friedrichs, M. A. M., Schmeltz, M., Aita, M. N., Antoine, D., Arrigo, K. R., Asanuma, I., Aumont, O., Barber, R., Behrenfeld, M., Bidigare, R., Buitenhuis, E. T., Campbell, J., Ciotti, A., Dierssen, H., Dowell, M., Dunne, J., Esaias, W., Gentili, B., Gregg, W., Groom, S., Hoepffner, N., Ishizaka, J., Kameda, T., Le Quéré, C., Lohrenz, S., Marra, J., Melin, F., Moore, K., Morel, A., Reddy, T. E., Ryan, J., Scardi, M., Smyth, T., Turpie, K., Tilstone, G., Waters, K., and Yamanaka, Y.: A comparison of global estimates of marine primary production from ocean color, *Deep-Sea Research Part II-Topical Studies in Oceanography*, 53, 741–770, 2006.
- Eby, M., Zickfeld, K., Montenegro, A., Archer, D., Meissner, K. J., and Weaver, A. J.: Lifetime of Anthropogenic Climate Change: Millennial Time Scales of Potential CO<sub>2</sub> and Surface Temperature Perturbations, *Journal of Climate*, 22, 2501–2511, <https://doi.org/10.1175/2008JCLI2554.1>, 2009.
- Enright, C., Buitenhuis, E., and Le Quere, C.: Description of the PlankTOM10 equations, [http://greenocean-data.uea.ac.uk/model/PlankTOM10\\_Manual.pdf](http://greenocean-data.uea.ac.uk/model/PlankTOM10_Manual.pdf), last accessed 05.06.2019, 2014.
- Eppley, R.: Temperature and phytoplankton growth in the sea, *Fishery Bulletin*, 70, 1063—1085, [http://lmgmacweb.env.uea.ac.uk/green\\_ocean/publications/Nano/Eppley72.pdf](http://lmgmacweb.env.uea.ac.uk/green_ocean/publications/Nano/Eppley72.pdf), 1972.
- Etheridge, D., Steele, L., Langenfelds, R., Francey, R., Barnola, J., and Morgan, V.: Natural and anthropogenic changes in atmospheric CO<sub>2</sub> over the last 1000 years from air in Antarctic ice and firn, *Journal of Geophysical Research-Atmospheres*, 101, 4115–4128, <https://doi.org/10.1029/95JD03410>, 1996.

- Etheridge, D., Steele, L., Francey, R., and Langenfelds, R.: Atmospheric methane between 1000 AD and present: Evidence of anthropogenic emissions and climatic variability, *Journal of Geophysical Research-Atmospheres*, 103, 15 979–15 993, <https://doi.org/10.1029/98JD00923>, 1998.
- 700 Ferretti, D., Miller, J., White, J., Etheridge, D., Lassey, K., Lowe, D., Meure, C., Dreier, M., Trudinger, C., van Ommen, T., and Langenfelds, R.: Unexpected changes to the global methane budget over the past 2000 years, *Science*, 309, 1714–1717, <https://doi.org/10.1126/science.1115193>, 2005.
- Flückiger, J., Dällenbach, A., Blunier, T., Stauffer, B., Stocker, T. F., Raynaud, D., and Barnola, J.-M.: Variations in atmospheric N<sub>2</sub>O concentration during abrupt climatic changes, *Science*, 285, 227–230, <https://doi.org/10.1126/science.285.5425.227>, 1999.
- 705 Flückiger, J., Blunier, T., Stauffer, B., Chappellaz, J., Spahni, R., Kawamura, K., Schwander, J., Stocker, T. F., and Dahl-Jensen, D.: N<sub>2</sub>O and CH<sub>4</sub> variations during the last glacial epoch: Insight into global processes, *Global Biogeochemical Cycles*, 18, <https://doi.org/10.1029/2003GB002122>, 2004.
- Frölicher, T. L., Sarmiento, J. L., Paynter, D. J., Dunne, J. P., Krasting, J. P., and Winton, M.: Dominance of the Southern Ocean in Anthropogenic Carbon and Heat Uptake in CMIP5 Models, *Journal of Climate*, 28, 862–886, <https://doi.org/10.1175/JCLI-D-14-00117.1>, <https://doi.org/10.1175/JCLI-D-14-00117.1>, 2015.
- 710 Fu, W., Randerson, J. T., and Moore, J. K.: Climate change impacts on net primary production (NPP) and export production (EP) regulated by increasing stratification and phytoplankton community structure in the CMIP5 models, *Biogeosciences*, 13, 5151–5170, <https://doi.org/10.5194/bg-13-5151-2016>, <https://www.biogeosciences.net/13/5151/2016/>, 2016.
- Galbraith, E. D., Gnanadesikan, A., Dunne, J. P., and Hiscock, M. R.: Regional impacts of iron-light colimitation in a global biogeochemical model, *Biogeosciences*, 7, 1043–1064, <https://doi.org/10.5194/bg-7-1043-2010>, <https://bg.copernicus.org/articles/7/1043/2010/>, 2010.
- 715 Garcia, H., Locarnini, R. A., Boyer, T. P., Antonov, J., Baranova, O., Zweng, M. M., Reagan, J., and Johnson, D.: World Ocean Atlas 2013, Volume 3: Dissolved Oxygen, Apparent Oxygen Utilization, and Oxygen Saturation, Tech. rep., NOAA Atlas NESDIS 75, U.S. Government Printing Office, Washington, D.C., 2014a.
- Garcia, H., Locarnini, R. A., Boyer, T. P., Antonov, J., Zweng, M. M., Reagan, J., Baranova, O., and Johnson, D.: World Ocean Atlas 2013, Volume 4: Dissolved Inorganic Nutrients (phosphate, nitrate, silicate), Tech. rep., NOAA Atlas NESDIS 76, U.S. Government Printing
- 720 Office, Washington, D.C., 2014b.
- Gnanadesikan, A.: A global model of silicon cycling: Sensitivity to eddy parameterization and dissolution, *Global Biogeochemical Cycles*, 13, 199–220, <https://doi.org/10.1029/1998GB900013>, <http://dx.doi.org/10.1029/1998GB900013>, 1999.
- Heinze, C., Eyring, V., Friedlingstein, P., Jones, C., Balkanski, Y., Collins, W., Fichefet, T., Gao, S., Hall, A., Ivanova, D., Knorr, W., Knutti, R., Löw, A., Ponater, M., Schultz, M. G., Schulz, M., Siebesma, P., Teixeira, J., Tselioudis, G., and Vancoppenolle, M.: ESD Reviews: Climate
- 725 feedbacks in the Earth system and prospects for their evaluation, *Earth System Dynamics*, 10, 379–452, <https://doi.org/10.5194/esd-10-379-2019>, <https://esd.copernicus.org/articles/10/379/2019/>, 2019.
- Holzer, M., Primeau, F. W., DeVries, T., and Matear, R.: The Southern Ocean silicon trap: Data-constrained estimates of regenerated silicic acid, trapping efficiencies, and global transport paths, *Journal of Geophysical Research: Oceans*, 119, 313–331, <https://doi.org/10.1002/2013JC009356>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1002/2013JC009356>, 2014.
- 730 Honjo, S., Manganini, S. J., Krishfield, R. A., and Francois, R.: Particulate organic carbon fluxes to the ocean interior and factors controlling the biological pump: A synthesis of global sediment trap programs since 1983, *Progress in Oceanography*, 76, 217–285, 2008.
- Hurtt, G. C., Chini, L. P., Frolicking, S., Betts, R. A., Feddema, J., Fischer, G., Fisk, J. P., Hibbard, K., Houghton, R. A., Janetos, A., Jones, C. D., Kindermann, G., Kinoshita, T., Klein Goldewijk, K., Riahi, K., Shevliakova, E., Smith, S., Stehfest, E., Thomson, A., Thornton, P.,

- van Vuuren, D. P., and Wang, Y. P.: Harmonization of land-use scenarios for the period 1500–2100: 600 years of global gridded annual  
735 land-use transitions, wood harvest, and resulting secondary lands, *Climatic Change*, 109, 117, <https://doi.org/10.1007/s10584-011-0153-2>,  
<http://dx.doi.org/10.1007/s10584-011-0153-2>, 2011.
- Iida, T., Odate, T., and Fukuchi, M.: Long-Term Trends of Nutrients and Apparent Oxygen Utilization South of the Polar Front in Southern  
Ocean Intermediate Water from 1965 to 2008, *PLOS ONE*, 8, 1–7, <https://doi.org/10.1371/journal.pone.0071766>, <https://doi.org/10.1371/journal.pone.0071766>, 2013.
- 740 Jin, X., Gruber, N., Dunne, J. P., Sarmiento, J. L., and Armstrong, R. A.: Diagnosing the contribution of phytoplankton functional groups to  
the production and export of particulate organic carbon,  $\text{CaCO}_3$ , and opal from global nutrient and alkalinity distributions, *Global Biogeo-  
chemical Cycles*, 20, <https://doi.org/10.1029/2005GB002532>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1029/2005GB002532>,  
2006.
- Kalnay, E., Kanamitsu, M., Kistler, R., Collins, W., Deaven, D., Gandin, L., Iredell, M., Saha, S., White, G., Woollen, J., Zhu, Y., Chelliah,  
745 M., Ebisuzaki, W., Higgins, W., Janowiak, J., Mo, K., Ropelewski, C., Wang, J., Leetmaa, A., Reynolds, R., Jenne, R., and Joseph, D.: The  
NCEP/NCAR 40-year reanalysis project, *Bulletin of the American Meteorological Society*, 77, 437–471, [https://doi.org/10.1175/1520-0477\(1996\)077<0437:TNYRP>2.0.CO;2](https://doi.org/10.1175/1520-0477(1996)077<0437:TNYRP>2.0.CO;2), 1996.
- Keller, D. P., Oschlies, A., and Eby, M.: A new marine ecosystem model for the University of Victoria Earth System Climate Model,  
*Geoscientific Model Development*, 5, 1195–1220, <https://doi.org/10.5194/gmd-5-1195-2012>, 2012.
- 750 Key, R., Olsen, A., van Heuven, S., Lauvset, A., Velo, Lin, X., Schirnick, C., Kozyr, A., Tanhua, T., Hoppema, M., Jutterström, S., Steinfeldt,  
R., Jeansson, E., Ishi, Perez, and Suzuki.: Global Ocean Data Analysis Project, Version 2 (GLODAPv2), Tech. rep., US Department of  
Energy, Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory, US Department of Energy, Oak Ridge, Tennessee.,  
[https://doi.org/10.3334/CDIAC/OTG.NDP093\\_GLODAPv2](https://doi.org/10.3334/CDIAC/OTG.NDP093_GLODAPv2), 2015.
- Khatiwala, S.: A computational framework for simulation of biogeochemical tracers in the ocean, *GLOBAL BIOGEOCHEMICAL CY-  
755 CLES*, 21, <https://doi.org/10.1029/2007GB002923>, 2007.
- Khatiwala, S., Primeau, F., and Hall, T.: Reconstruction of the history of anthropogenic  $\text{CO}_2$  concentrations in the ocean, *Nature*, 462,  
346–349, <https://doi.org/10.1038/nature08526>, <https://doi.org/10.1038/nature08526>, 2009.
- Khatiwala, S., Visbeck, M., and Cane, M.: Accelerated simulation of passive tracers in ocean circulation models, *OCEAN MODELLING*, 9,  
51–69, <https://doi.org/10.1016/j.ocemod.2004.04.002>, 2005.
- 760 Koeve, W., Wagner, H., Kähler, P., and Oschlies, A.:  $\text{C}$ -age tracers in global ocean circulation models, *Geoscientific Model  
Development*, 8, 2079–2094, <https://doi.org/10.5194/gmd-8-2079-2015>, <http://www.geosci-model-dev.net/8/2079/2015/>, 2015.
- Kriest, I.: Calibration of a simple and a complex model of global marine biogeochemistry, *Biogeosciences*, 14, 4965–4984,  
<https://doi.org/10.5194/bg-14-4965-2017>, <https://www.biogeosciences.net/14/4965/2017/>, 2017.
- Kriest, I., Sauerland, V., Khatiwala, S., Srivastav, A., and Oschlies, A.: Calibrating a global three-dimensional biogeochemical ocean model  
765 (MOPS-1.0), *Geoscientific Model Development*, 10, 127–154, <https://doi.org/10.5194/gmd-10-127-2017>, <https://www.geosci-model-dev.net/10/127/2017/>, 2017.
- Kvale, K. F. and Meissner, K. J.: Primary production sensitivity to phytoplankton light attenuation parameter increases with transient forcing,  
*Biogeosciences*, 14, 4767–4780, <https://doi.org/10.5194/bg-14-4767-2017>, <https://bg.copernicus.org/articles/14/4767/2017/>, 2017.
- 770 Kvale, K. F., Khatiwala, S., Dietze, H., Kriest, I., and Oschlies, A.: Evaluation of the transport matrix method for simulation of ocean biogeo-  
chemical tracers, *Geoscientific Model Development*, 10, 2425–2445, <https://doi.org/10.5194/gmd-10-2425-2017>, <https://gmd.copernicus.org/articles/10/2425/2017/>, 2017.

- Kvale, K. F., Turner, K. E., Keller, D. P., and Meissner, K. J.: Asymmetric dynamical ocean responses in warming icehouse and cooling greenhouse climates, *Environmental Research Letters*, 13, 125 011, <https://doi.org/10.1088/1748-9326/aaedc3>, <https://doi.org/10.1088/1748-9326/2Faaedc3>, 2018.
- 775 Kvale, K. F., Turner, K., Landolfi, A., and Meissner, K. J.: Phytoplankton calcifiers control nitrate cycling and the pace of transition in warming icehouse and cooling greenhouse climates, *Biogeosciences*, 16, 1019–1034, <https://doi.org/10.5194/bg-16-1019-2019>, <https://www.biogeosciences.net/16/1019/2019/>, 2019.
- Kvale, K. F., Meissner, K. J., and Keller, D. P.: Potential increasing dominance of heterotrophy in the global ocean, *Environmental Research Letters*, 10, <https://doi.org/10.1088/1748-9326/10/7/074009>, 2015a.
- 780 Kvale, K. F., Meissner, K. J., Keller, D. P., Eby, M., and Schmittner, A.: Explicit Planktic Calcifiers in the University of Victoria Earth System Climate Model, Version 2.9, *ATMOSPHERE-OCEAN*, 53, 332–350, <https://doi.org/10.1080/07055900.2015.1049112>, 2015b.
- Laufkötter, C., Vogt, M., and Gruber, N.: Long-term trends in ocean plankton production and particle export between 1960–2006, *Biogeosciences*, 10, 7373–7393, <https://doi.org/10.5194/bg-10-7373-2013>, <https://www.biogeosciences.net/10/7373/2013/>, 2013.
- Lauvset, S. K., Key, R. M., Olsen, A., van Heuven, S., Velo, A., Lin, X., Schirnack, C., Kozyr, A., Tanhua, T., Hoppema, M., Jutterström, S.,  
785 Steinfeldt, R., Jeansson, E., Ishii, M., Perez, F. F., Suzuki, T., and Watelet, S.: A new global interior ocean mapped climatology: the  $1^\circ \times 1^\circ$  GLODAP version 2, *Earth System Science Data*, 8, 325–340, <https://doi.org/10.5194/essd-8-325-2016>, <https://www.earth-syst-sci-data.net/8/325/2016/>, 2016.
- Leblanc, K., Arístegui Ruiz, J., Armand, L. K., Assmy, P., Beker, B., Bode, A., Breton, E., Cornet, V., Gibson, J., Gosselin, M.-P., Kopczynska, E. E., Marshall, H. G., Peloquin, J. M., Piontkovski, S., Poulton, A. J., Quéguiner, B., Schiebel, R., Shipe, R., Ste-  
790 felds, J., van Leeuwe, M. A., Varela, M., Widdicombe, C. E., and Yallop, M.: Global distributions of diatoms abundance, biovolume and biomass - Gridded data product (NetCDF) - Contribution to the MAREDAT World Ocean Atlas of Plankton Functional Types, <https://doi.org/10.1594/PANGAEA.777384>, <https://doi.org/10.1594/PANGAEA.777384>, 2012.
- Luo, Y., Doney, S. C., Anderson, L. A., Benavides, M., Berman-Frank, I., Bode, A., Bonnet, S., Boström, K. H., Böttjer, D., Capone, D. G., Carpenter, E. J., Chen, Y.-L., Church, M. J., Dore, J. E., Falcón, L. I., Fernández, A., Foster, R. A., Furuya, K., Gomez, F., Gundersen,  
795 K., Hynes, A. M., Karl, D. M., Kitajima, S., Langlois, R., LaRoche, J., Letelier, R. M., Marañón, E., McGillicuddy Jr, D. J., Moisander, P. H., Moore, C. M., Mouriño-Carballido, B., Mulholland, M. R., Needoba, J. A., Orcutt, K. M., Poulton, A. J., Rahav, E., Raimbault, P., Rees, A., Riemann, L., Shiozaki, T., Subramaniam, A., Tyrrell, T., Turk-Kubo, K. A., Varela, M., Villareal, T. A., Webb, E. A., White, A. E., Wu, J., and Zehr, J. P.: Global distributions of diazotrophs abundance, biomass and nitrogen fixation rates - Gridded data product (NetCDF) - Contribution to the MAREDAT World Ocean Atlas of Plankton Functional Types, <https://doi.org/10.1594/PANGAEA.774851>,  
800 <https://doi.org/10.1594/PANGAEA.774851>, supplement to: Luo, Y et al. (2012): Database of diazotrophs in global ocean: abundance, biomass and nitrogen fixation rates. *Earth System Science Data*, 4, 47–73, <https://doi.org/10.5194/essd-4-47-2012>, 2012.
- Luo, Y.-W., Lima, I. D., Karl, D. M., Deutsch, C. A., and Doney, S. C.: Data-based assessment of environmental controls on global marine nitrogen fixation, *Biogeosciences*, 11, 691–708, <https://doi.org/10.5194/bg-11-691-2014>, <https://www.biogeosciences.net/11/691/2014/>, 2014.
- 805 Machida, T., Nakazawa, T., Fujii, Y., Aoki, S., and Watanabe, O.: Increase in the atmospheric nitrous-oxide concentration during the last 250 years, *Geophysical Research Letters*, 22, 2921–2924, <https://doi.org/10.1029/95GL02822>, 1995.
- Mahowald, N. M., Muhs, D. R., Levis, S., Rasch, P. J., Yoshioka, M., Zender, C. S., and Luo, C.: Change in atmospheric mineral aerosols in response to climate: Last glacial period, preindustrial, modern, and doubled carbon dioxide climates, *JOURNAL OF GEOPHYSICAL RESEARCH-ATMOSPHERES*, 111, <https://doi.org/10.1029/2005JD006653>, 2006.

810 Matsumoto, K., Sarmiento, J. L., and Brzezinski, M. A.: Silicic acid leakage from the Southern Ocean: A possible explanation for glacial atmospheric pCO<sub>2</sub>, *Global Biogeochemical Cycles*, 16, 5–1–5–23, <https://doi.org/10.1029/2001GB001442>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1029/2001GB001442>, 2002.

Meinshausen, M., Smith, S. J., Calvin, K., Daniel, J. S., Kainuma, M. L. T., Lamarque, J.-F., Matsumoto, K., Montzka, S. A., Raper, S. C. B., Riahi, K., Thomson, A., Velders, G. J. M., and van Vuuren, D. P. P.: The RCP greenhouse gas concentrations and their extensions from 1765 to 2300, *Climatic Change*, 109, 213–241, <https://doi.org/10.1007/s10584-011-0156-z>, 2011.

815 Meissner, K., Weaver, A., Matthews, H., and Cox, P.: The role of land surface dynamics in glacial inception: A study with the UVic Earth System Model, *Climate Dynamics*, 21, 515–537, <https://doi.org/10.1007/s00382-003-0352-2>, 2003.

Mengis, N., Keller, D. P., MacDougall, A., Eby, M., Wright, N., Meissner, K. J., Oeschies, A., Schmittner, A., Matthews, H. D., and Zickfeld, K.: Evaluation of the University of Victoria Earth System Climate Model version 2.10 (UVic ESCM 2.10), *Geoscientific Model Development Discussions*, 2020, 1–28, <https://doi.org/10.5194/gmd-2019-373>, <https://gmd.copernicus.org/preprints/gmd-2019-373/>, 2020.

820 Merico, A., Tyrrell, T., and Cokacar, T.: Is there any relationship between phytoplankton seasonal dynamics and the carbonate system?, *Journal of Marine Systems*, 59, 120–142, 2006.

Meure, M. C., Etheridge, D., Trudinger, C., Steele, P., Langenfelds, R., van Ommen, T., Smith, A., and Elkins, J.: Law Dome CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O ice core records extended to 2000 years BP, *Geophysical Research Letters*, 33, <https://doi.org/10.1029/2006GL026152>, 2006.

825 Moore, J. K., Fu, W., Primeau, F., Britten, G. L., Lindsay, K., Long, M., Doney, S. C., Mahowald, N., Hoffman, F., and Randerson, J. T.: Sustained climate warming drives declining marine biological productivity, *Science*, 359, 1139–1143, <https://doi.org/10.1126/science.aao6379>, <https://science.sciencemag.org/content/359/6380/1139>, 2018.

Muglia, J., Somes, C. J., Nickelsen, L., and Schmittner, A.: Combined Effects of Atmospheric and Seafloor Iron Fluxes to the Glacial Ocean, *Paleoceanography*, 32, 1204–1218, <https://doi.org/10.1002/2016PA003077>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1002/2016PA003077>, 2017.

830 NASA: Sea-viewing Wide Field-of-view Sensor (SeaWiFS) Ocean Color Data products, <https://doi.org/10.5067/AQUA/MODIS/L3M/PIC/2018>, 2018.

Neukermans, G., Oziel, L., and Babin, M.: Increased intrusion of warming Atlantic water leads to rapid expansion of temperate phytoplankton in the Arctic, *Global Change Biology*, 24, 2545–2553, <https://doi.org/10.1111/gcb.14075>, <https://onlinelibrary.wiley.com/doi/abs/10.1111/gcb.14075>, 2018.

835 Nickelsen, L., Keller, D. P., and Oeschies, A.: A dynamic marine iron cycle module coupled to the University of Victoria Earth System Model: the Kiel Marine Biogeochemical Model 2 for UVic 2.9, *GEOSCIENTIFIC MODEL DEVELOPMENT*, 8, 1357–1381, 2015.

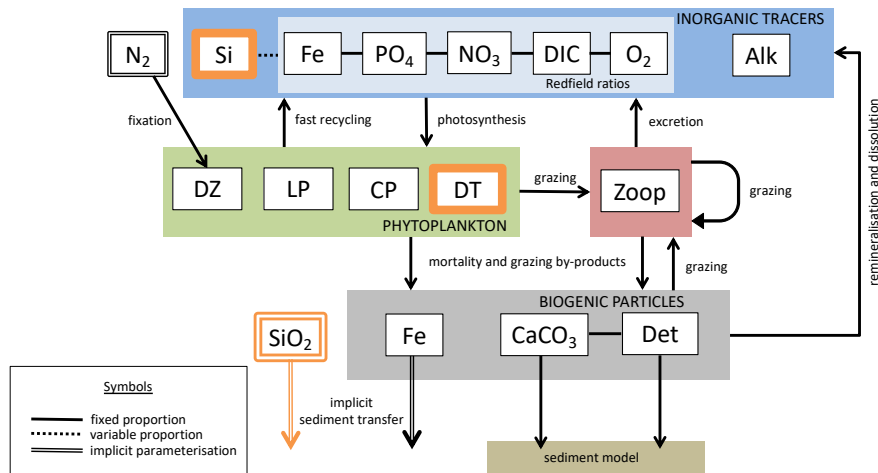
Nishioka, J., Obata, H., Ogawa, H., Ono, K., Yamashita, Y., Lee, K., Takeda, S., and Yasuda, I.: Subpolar marginal seas fuel the North Pacific through the intermediate water at the termination of the global ocean circulation, *Proceedings of the National Academy of Sciences*, 117, 12 665–12 673, <https://doi.org/10.1073/pnas.2000658117>, <https://www.pnas.org/content/117/23/12665>, 2020.

840 Olsson, J., Stipp, S., Makovicky, E., and Gislason, S.: Metal scavenging by calcium carbonate at the Eyjafjallajökull volcano: A carbon capture and storage analogue, *Chemical Geology*, 384, 135 – 148, <https://doi.org/https://doi.org/10.1016/j.chemgeo.2014.06.025>, <http://www.sciencedirect.com/science/article/pii/S0009254114003076>, 2014.

Oeschies, A., Duteil, O., Getzlaff, J., Koeve, W., Landolfi, A., and Schmidtke, S.: Patterns of deoxygenation: sensitivity to natural and anthropogenic drivers, *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 375, 20160325, <https://doi.org/10.1098/rsta.2016.0325>, <https://royalsocietypublishing.org/doi/abs/10.1098/rsta.2016.0325>, 2017.

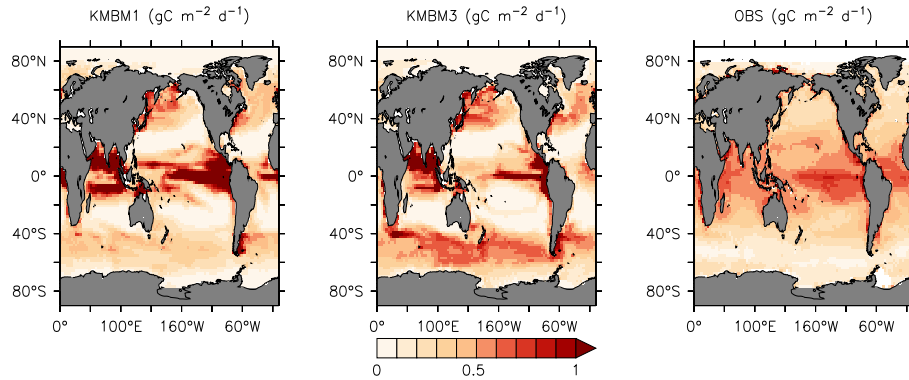
845

- O'Brien, C. J.: Global distributions of coccolithophores abundance and biomass - Gridded data product (NetCDF) - Contribution to the MAREDAT World Ocean Atlas of Plankton Functional Types, <https://doi.org/10.1594/PANGAEA.785092>, <https://doi.org/10.1594/PANGAEA.785092>, 2012.
- 850 Paasche, E.: A review of the coccolithophorid *Emiliania huxleyi* (Prymnesiophyceae), with particular reference to growth, coccolith formation, and calcification-photosynthesis interactions, *Phycologia*, 40, 503–529, 2001.
- Partanen, A., Keller, D. P., Korhonen, H., and Matthews, H. D.: Impacts of sea spray geoengineering on ocean biogeochemistry, *Geophysical Research Letters*, 43, 7600–7608, <https://doi.org/10.1002/2016GL070111>, <https://doi.org/10.1002/2016GL070111>, 2016.
- Pasquier, B. and Holzer, M.: Inverse-model estimates of the ocean's coupled phosphorus, silicon, and iron cycles, *Biogeosciences*, 14, 4125–
- 855 4159, <https://doi.org/10.5194/bg-14-4125-2017>, <https://www.biogeosciences.net/14/4125/2017/>, 2017.
- Renaudie, J.: Quantifying the Cenozoic marine diatom deposition history: links to the C and Si cycles, *Biogeosciences*, 13, 6003–6014, <https://doi.org/10.5194/bg-13-6003-2016>, <https://www.biogeosciences.net/13/6003/2016/>, 2016.
- Riahi, K., Gruebler, A., and Nakicenovic, N.: Scenarios of long-term socio-economic and environmental development under climate stabilization, *Technological Forecasting and Social Change*, 74, 887–935, <https://doi.org/10.1016/j.techfore.2006.05.026>, 2007.
- 860 Ridgwell, A., Watson, A., and Archer, D.: Modeling the response of the oceanic Si inventory to perturbation, and consequences for atmospheric CO<sub>2</sub>, *GLOBAL BIOGEOCHEMICAL CYCLES*, 16, <https://doi.org/10.1029/2002GB001877>, 2002.
- Rousseaux, C. S. and Gregg, W. W.: Recent decadal trends in global phytoplankton composition, *Global Biogeochemical Cycles*, 29, 1674–1688, <https://doi.org/10.1002/2015GB005139>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1002/2015GB005139>, 2015.
- 865 Sarmiento, J. and Gruber, N.: *Ocean Biogeochemical Dynamics*, Princeton University Press, Princeton and Oxford, 2006.
- Schmidtko, S., Stramma, L., and Visbeck, M.: Decline in global oceanic oxygen content during the past five decades, *Nature*, 542, 335–339, <https://doi.org/10.1038/nature21399>, <https://doi.org/10.1038/nature21399>, 2017.
- Schmittner, A., Oschlies, A., Giraud, X., Eby, M., and Simmons, H.: A global model of the marine ecosystem for long-term simulations: Sensitivity to ocean mixing, buoyancy forcing, particle sinking, and dissolved organic matter cycling, *Global Biogeochemical Cycles*, 19,
- 870 <https://doi.org/10.1029/2004GB002283>, 2005.
- Schmittner, A., Oschlies, A., Matthews, H. D., and Galbraith, E. D.: Future changes in climate, ocean circulation, ecosystems, and biogeochemical cycling simulated for a business-as-usual CO<sub>2</sub> emission scenario until year 4000 AD, *Global Biogeochemical Cycles*, 22, <https://doi.org/10.1029/2007GB002953>, 2008.
- Somes, C. J., Oschlies, A., and Schmittner, A.: Isotopic constraints on the pre-industrial oceanic nitrogen budget, *Biogeosciences*, 10, 5889–
- 875 5910, <https://doi.org/10.5194/bg-10-5889-2013>, <https://www.biogeosciences.net/10/5889/2013/>, 2013.
- Stramma, L., Schmidtko, S., Bograd, S. J., Ono, T., Ross, T., Sasano, D., and Whitney, F. A.: Trends and decadal oscillations of oxygen and nutrients at 50 to 300 m depth in the equatorial and North Pacific, *Biogeosciences*, 17, 813–831, <https://doi.org/10.5194/bg-17-813-2020>, <https://bg.copernicus.org/articles/17/813/2020/>, 2020.
- Tréguer, P., Bowler, C., Moriceau, B., Dutkiewicz, S., Gehlen, M., Aumont, O., Bittner, L., Dugdale, R., Finkel, Z., Iudicone, D., Jahn, O.,
- 880 Guidi, L., Lasbleiz, M., Leblanc, K., Levy, M., and Pondaven, P.: Influence of diatom diversity on the ocean biological carbon pump, *Nature Geoscience*, 11, 27–37, <https://doi.org/10.1038/s41561-017-0028-x>, <https://doi.org/10.1038/s41561-017-0028-x>, 2018.
- Tréguer, P. J. and De La Rocha, C. L.: The World Ocean Silica Cycle, *Annual Review of Marine Science*, 5, 477–501, <https://doi.org/10.1146/annurev-marine-121211-172346>, <https://doi.org/10.1146/annurev-marine-121211-172346>, PMID: 22809182, 2013.

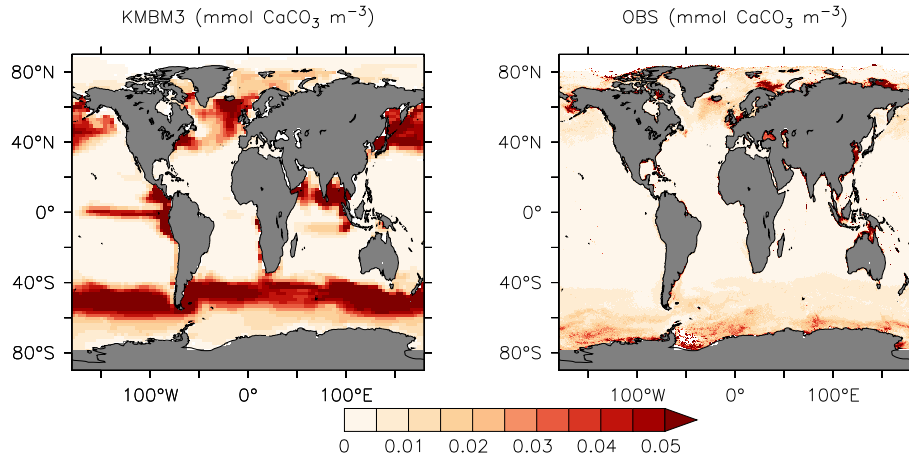


**Figure 1.** The latest biogeochemical model structure for the KMBM3. Previously unpublished features are shown in orange.

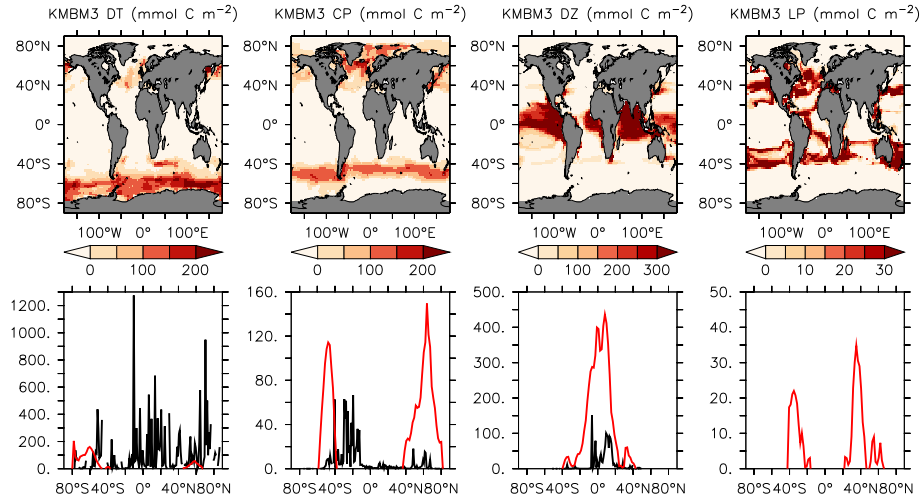
- 885 Weaver, A., Eby, M., Wiebe, E., Bitz, C., Duffy, P., Ewen, T., Fanning, A., Holland, M., MacFadyen, A., Matthews, H., Meissner, K., Saenko, O., Schmittner, A., Wang, H., and Yoshimori, M.: The UVic Earth System Climate Model: Model description, climatology, and applications to past, present and future climates, *Atmosphere-Ocean*, 39, 361–428, 2001.
- Weber, T. and Deutsch, C.: Oceanic nitrogen reservoir regulated by plankton diversity and ocean circulation, *Nature*, 489, 419, <https://doi.org/10.1038/nature11357><http://10.0.4.14/nature11357><https://www.nature.com/articles/nature11357{#}supplementary-information>, 2012.
- 890 Wernand, M. R., van der Woerd, H. J., and Gieskes, W. W. C.: Trends in Ocean Colour and Chlorophyll Concentration from 1889 to 2000, Worldwide, *PLOS ONE*, 8, 1–20, <https://doi.org/10.1371/journal.pone.0063766>, <https://doi.org/10.1371/journal.pone.0063766>, 2013.
- Westberry, T., Behrenfeld, M. J., Siegel, D. A., and Boss, E.: Carbon-based primary productivity modeling with vertically resolved photoacclimation, *Global Biogeochemical Cycles*, 22, <https://doi.org/10.1029/2007GB003078>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1029/2007GB003078>, 2008.
- 895 Yao, W., Kvale, K. F., Achterberg, E., Koeve, W., and Oschlies, A.: Hierarchy of calibrated global models reveals improved distributions and fluxes of biogeochemical tracers in models with explicit representation of iron, *Environmental Research Letters*, 14, 114 009, <https://doi.org/10.1088/1748-9326/ab4c52>, <https://doi.org/10.1088/1748-9326/ab4c52>, 2019.
- Yasunaka, S., Ono, T., Nojiri, Y., Whitney, F. A., Wada, C., Murata, A., Nakaoka, S., and Hosoda, S.: Long-term variability of surface nutrient concentrations in the North Pacific, *Geophysical Research Letters*, 43, 3389–3397, <https://doi.org/10.1002/2016GL068097>, <https://agupubs.onlinelibrary.wiley.com/doi/abs/10.1002/2016GL068097>, 2016.
- 900 Zhang, Y., Mahowald, N., Scanza, R. A., Journet, E., Desboeufs, K., Albani, S., Kok, J. F., Zhuang, G., Chen, Y., Cohen, D. D., Paytan, A., Patey, M. D., Achterberg, E. P., Engelbrecht, J. P., and Fomba, K. W.: Modeling the global emission, transport and deposition of trace elements associated with mineral dust, *Biogeosciences*, 12, 5771–5792, <https://doi.org/10.5194/bg-12-5771-2015>, <https://bg.copernicus.org/articles/12/5771/2015/>, 2015.



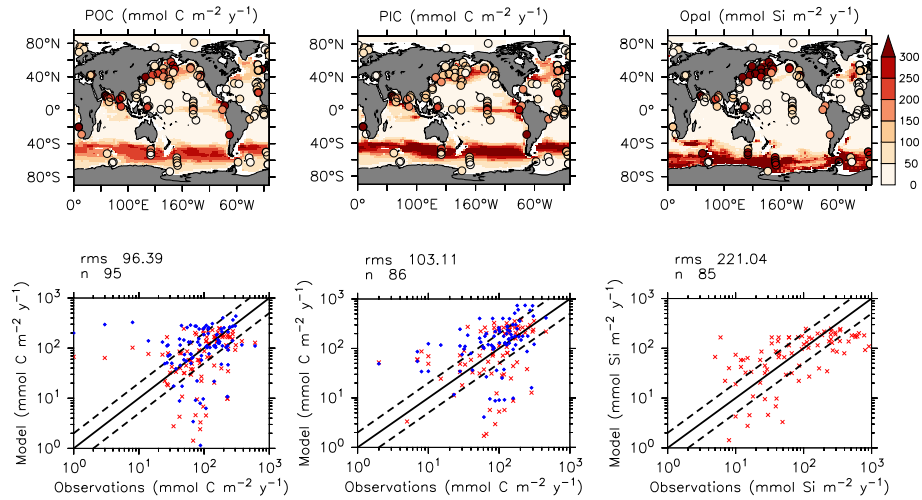
**Figure 2.** Comparison of model NPP output at 2014 (left panel; Keller et al. (2012), and middle panel; this model) and satellite-derived NPP climatology (NASA, 2018), 2012-2018 (right panel) in  $\text{gC m}^{-2} \text{ day}^{-1}$ .



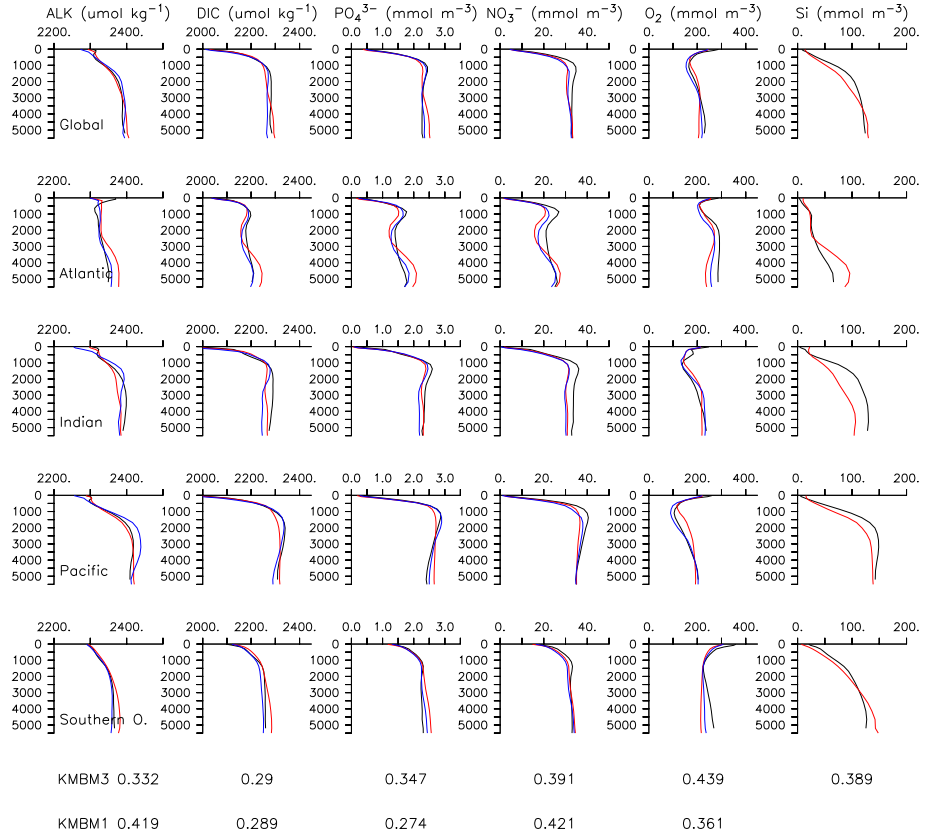
**Figure 3.** Comparison of model average 2004-2014 (left panel) and satellite  $\text{CaCO}_3$  climatology, 2002-2018 (right panel) in  $\text{mmol CaCO}_3 \text{ m}^{-3}$ . Data product is scaled by the model grid in the  $z$  direction and in both plots only the upper 20 meters are represented, where uniform coccolith concentration is assumed (Balch and Utgoff, 2009).



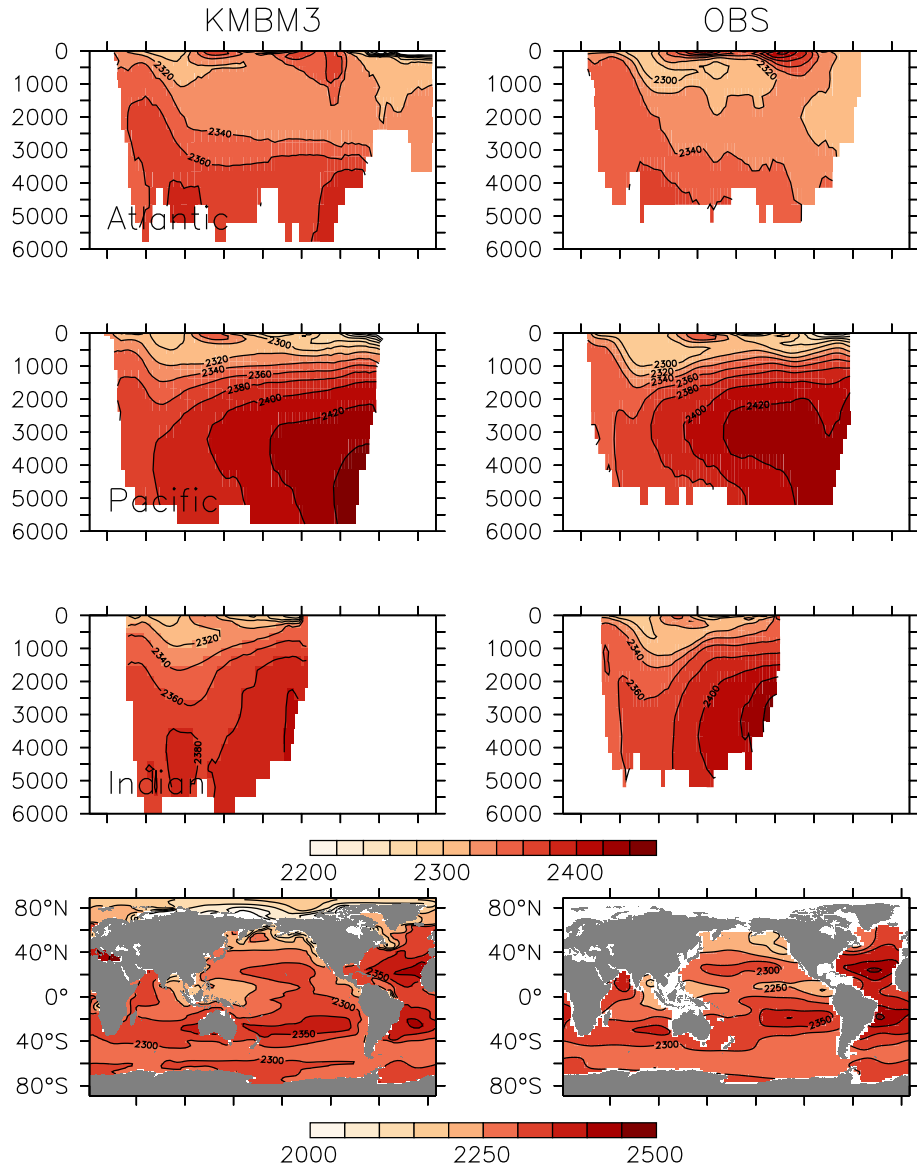
**Figure 4.** Annually-averaged and depth-integrated model biomass at year 2014 (top row). Zonally-averaged and depth-integrated model biomass plotted with the equivalent MAREDAT compilations (bottom row; Leblanc et al., 2012; O'Brien, 2012; Luo et al., 2012, no data compilation of “mixed” phytoplankton, a model-specific category). All units are  $\text{mmol C m}^{-2}$ . Black lines represent MAREDAT, red lines are the model output.



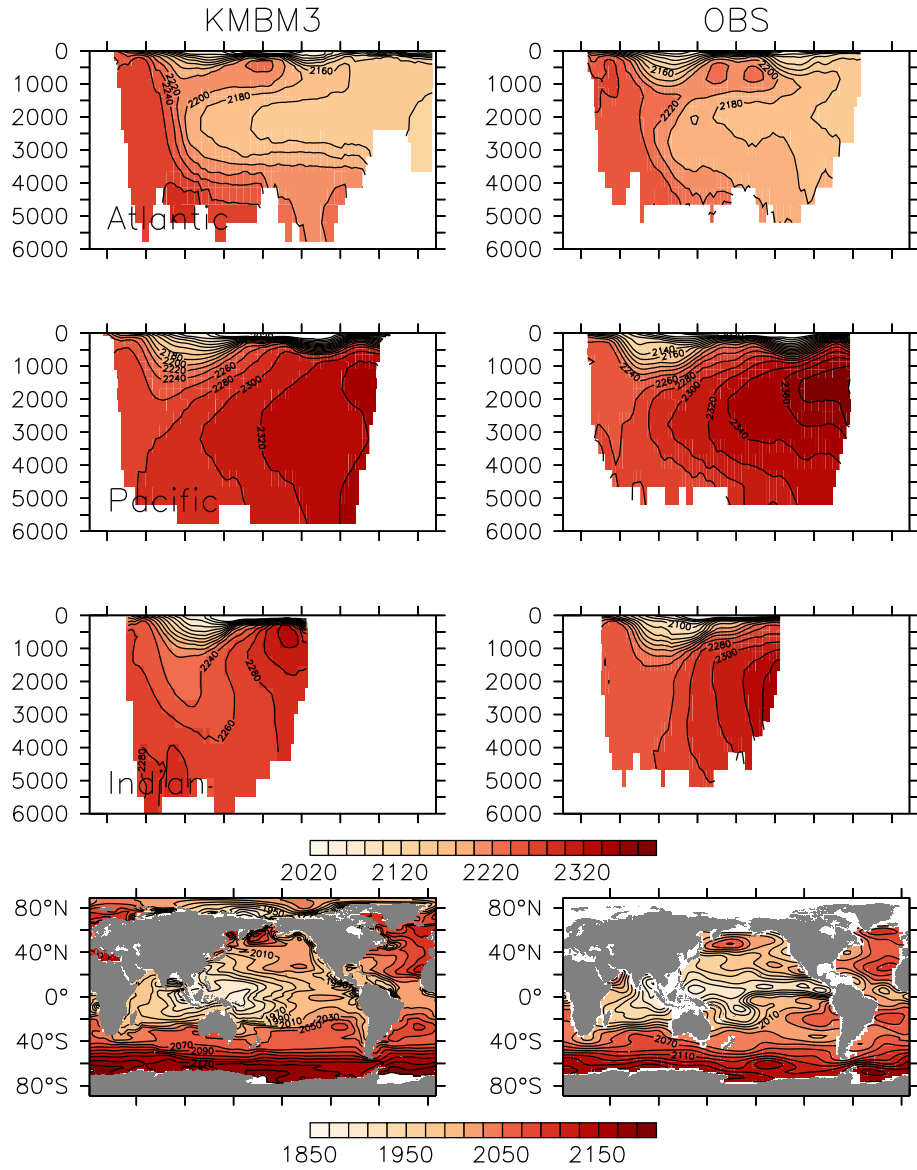
**Figure 5.** Comparison of model POC (left panels),  $\text{CaCO}_3$  (middle panels), and opal (right panels) fluxes at year 2004 and 2 km depth with Honjo et al. (2008) data. Red points are this model (KMBM3), blue points are the Keller et al. (2012) model (KMBM1). Statistics are given for KMBM3 only.



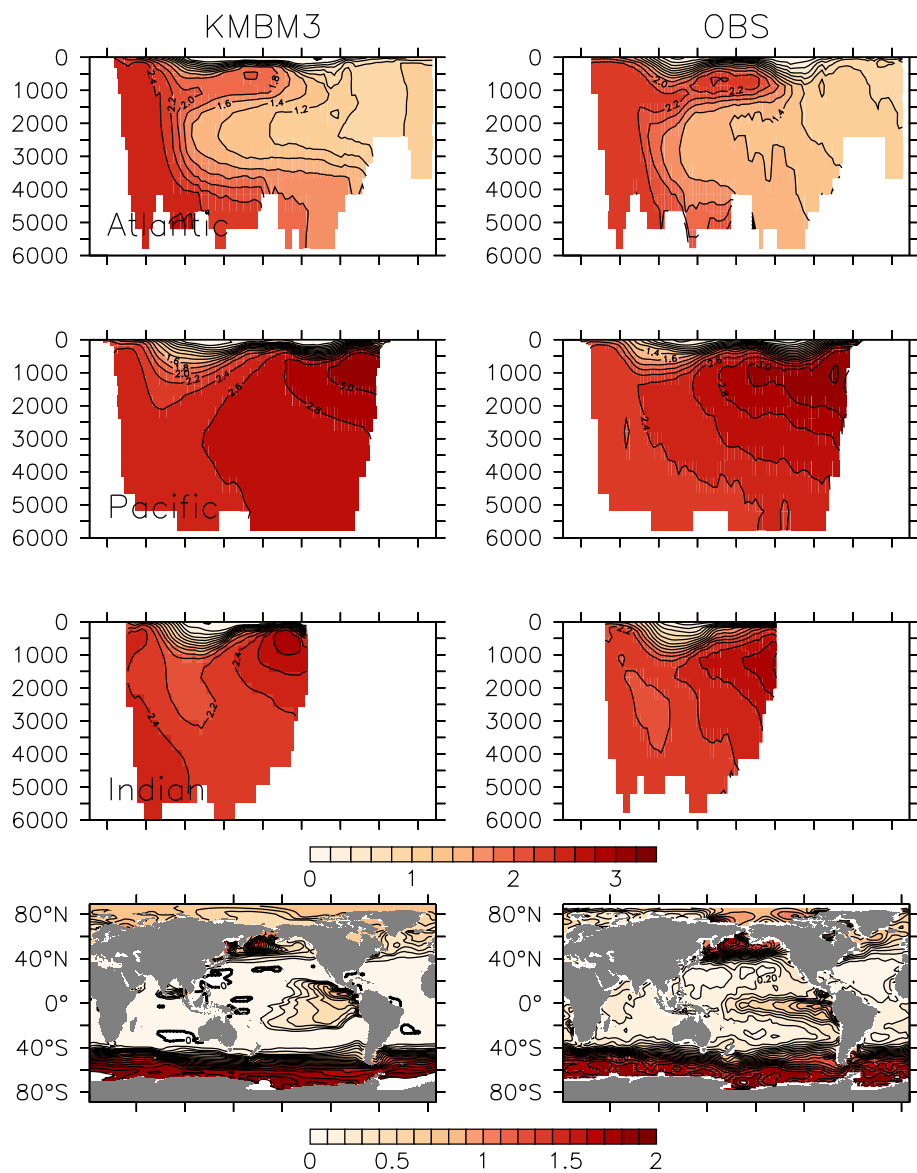
**Figure 6.** Model global and basin average nutrient and carbon depth profiles at year 2014 (KMBM3; red lines), compared to model output from the Keller et al. (2012) version (KMBM1; blue lines) and observational data (black lines; Garcia et al., 2014a, b; Key et al., 2015; Lauvset et al., 2016). Global root mean square error is given below each column.



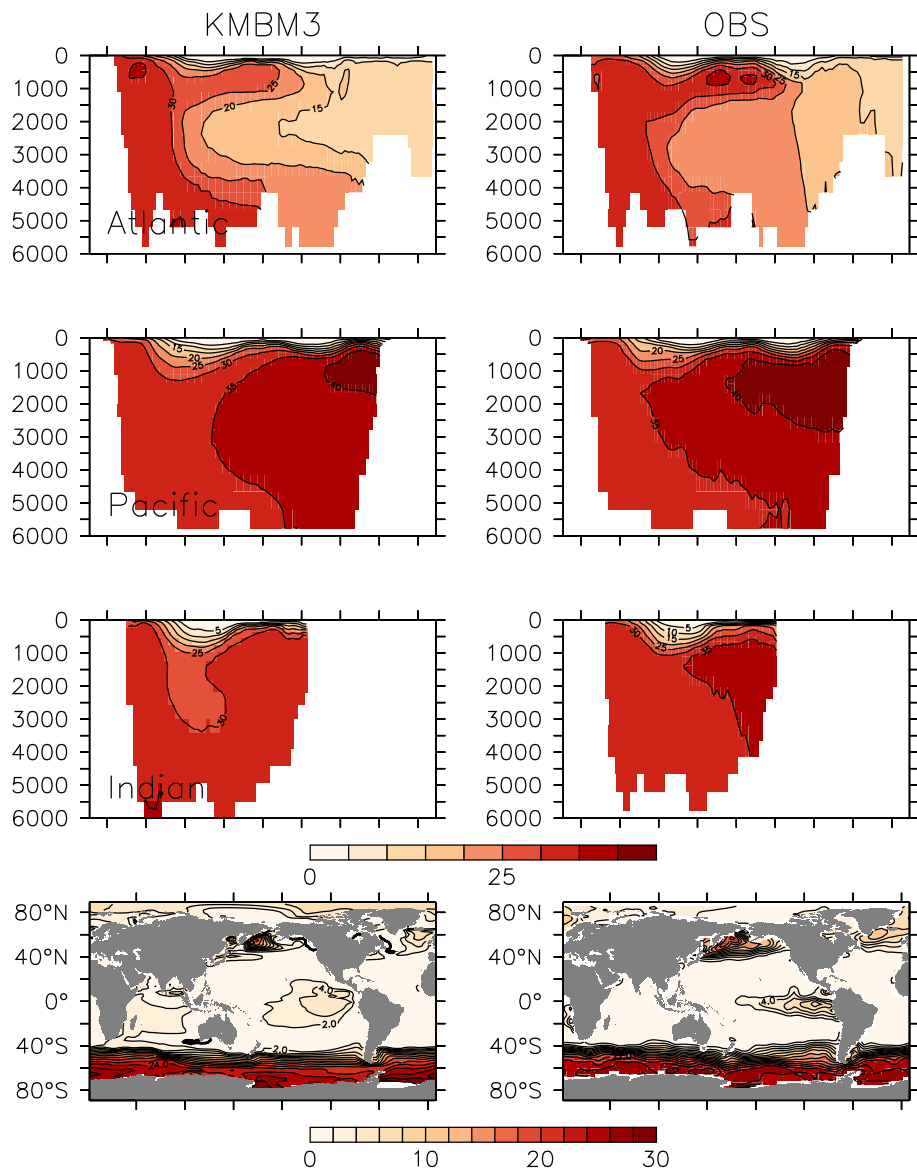
**Figure 7.** Model (left column) year 2014 alkalinity ( $\mu\text{mol kg}^{-1}$ ) averaged by basin compared to GLODAP (right column; Key et al., 2015; Lauvset et al., 2016).



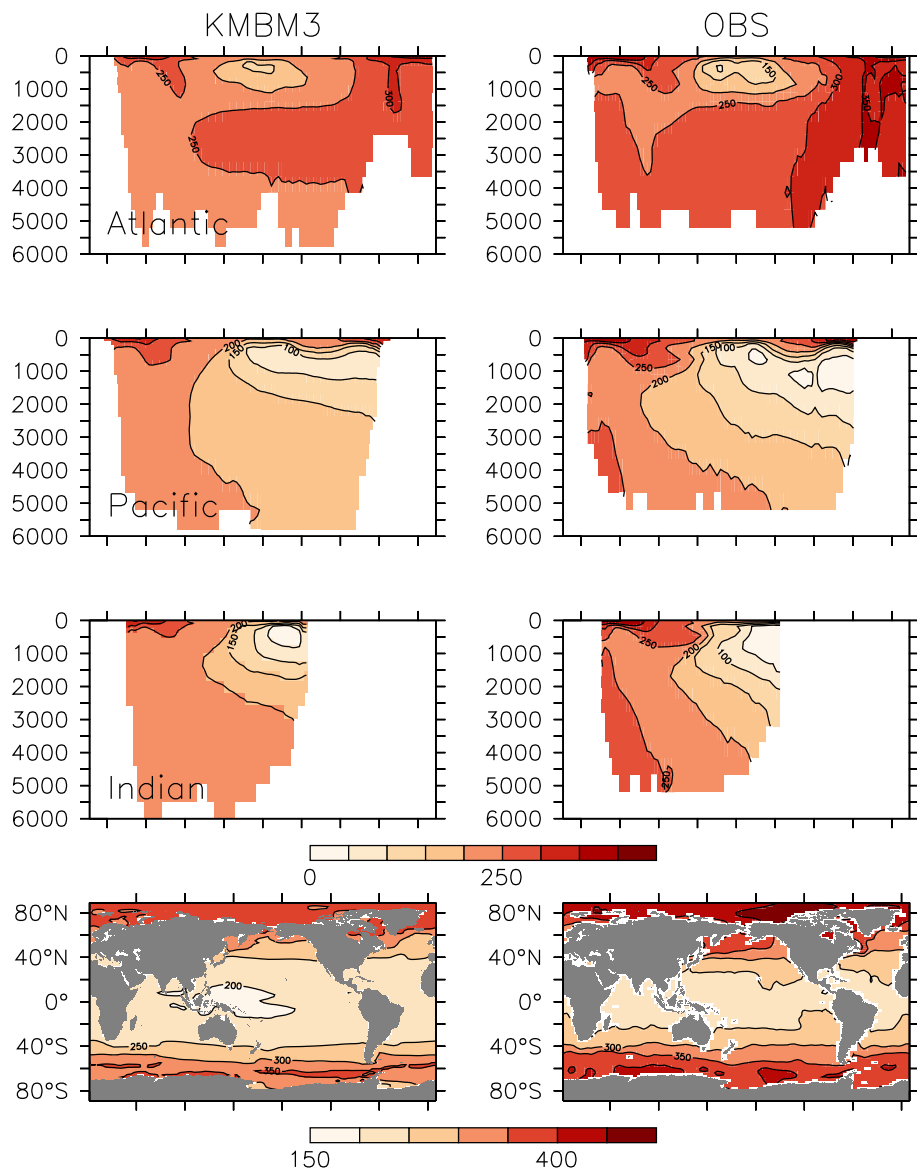
**Figure 8.** Model (left column) year 2014 DIC ( $\mu\text{mol kg}^{-1}$ ) averaged by basin compared to GLODAP (right column; Key et al., 2015; Lauvset et al., 2016).



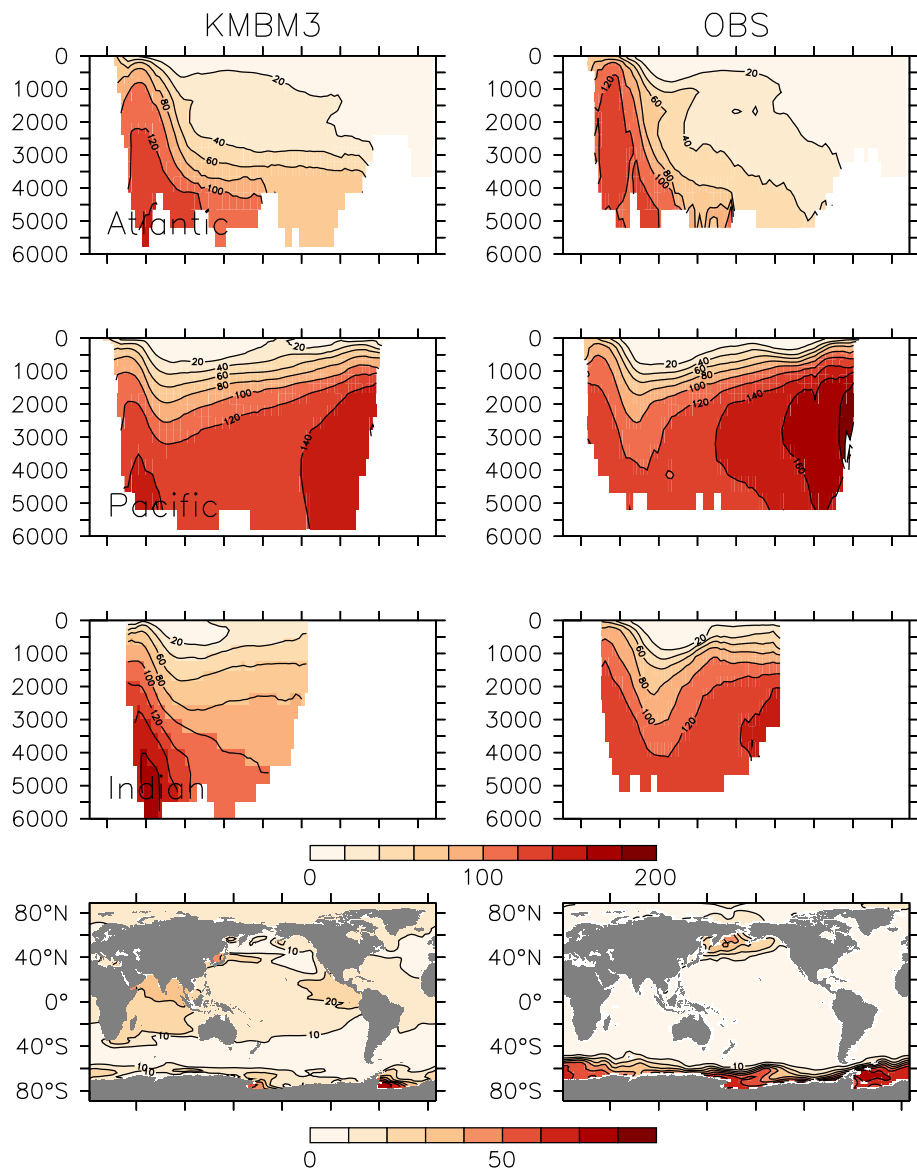
**Figure 9.** Model (left column) year 2014  $\text{PO}_4^{3-}$  (mmol  $\text{m}^{-3}$ ) averaged by basin compared to WOA (right column; Garcia et al., 2014b).



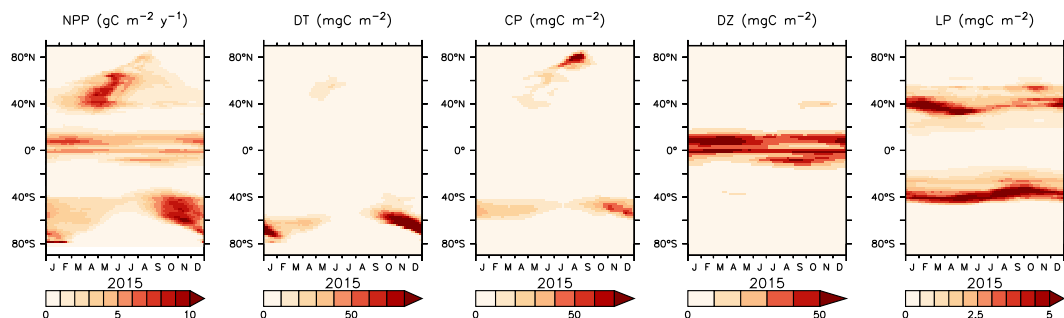
**Figure 10.** Model (left column) year 2014  $\text{NO}_3^-$  (mmol  $\text{m}^{-3}$ ) averaged by basin compared to WOA (right column; Garcia et al., 2014b).



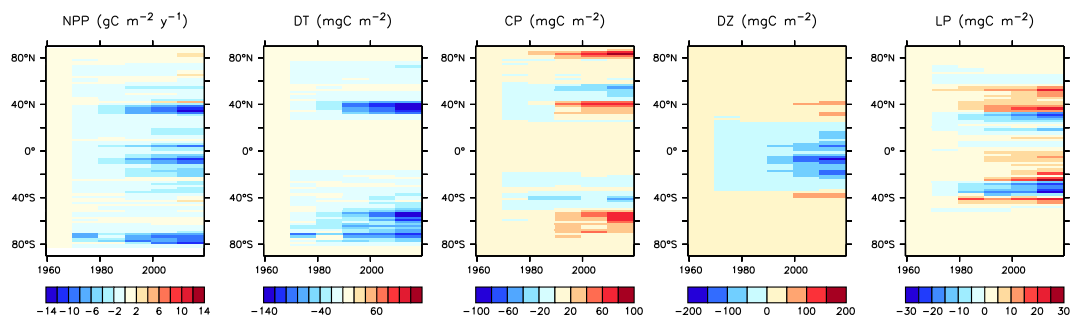
**Figure 11.** Model (left column) year 2014  $\text{O}_2$  ( $\text{mmol m}^{-3}$ ) averaged by basin compared to WOA (right column; Garcia et al., 2014a).



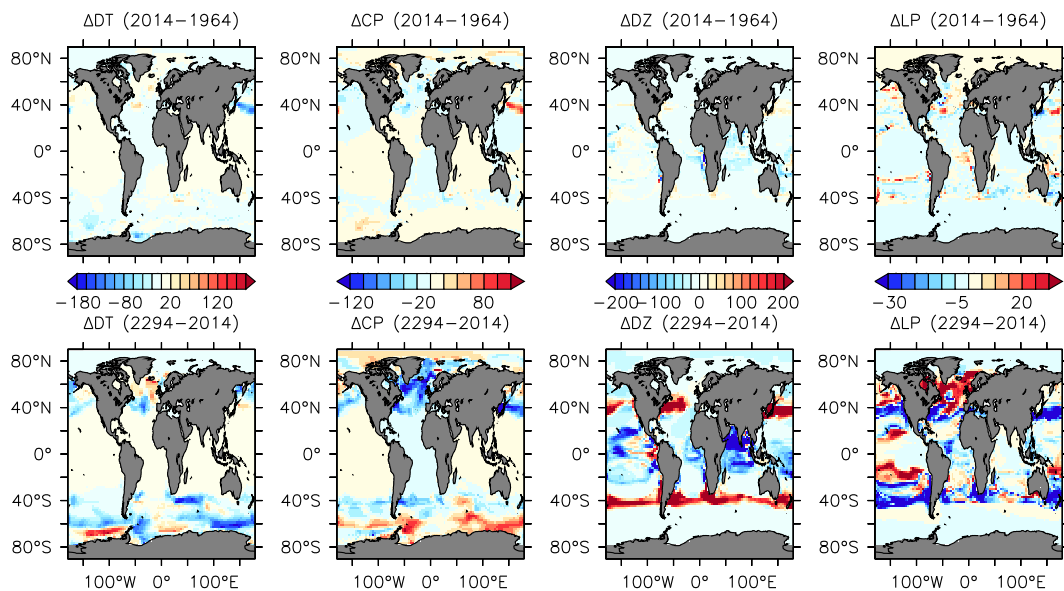
**Figure 12.** Model (left column) year 2014 Si (mmol m<sup>-3</sup>) averaged by basin compared to WOA (right column; Garcia et al., 2014b).



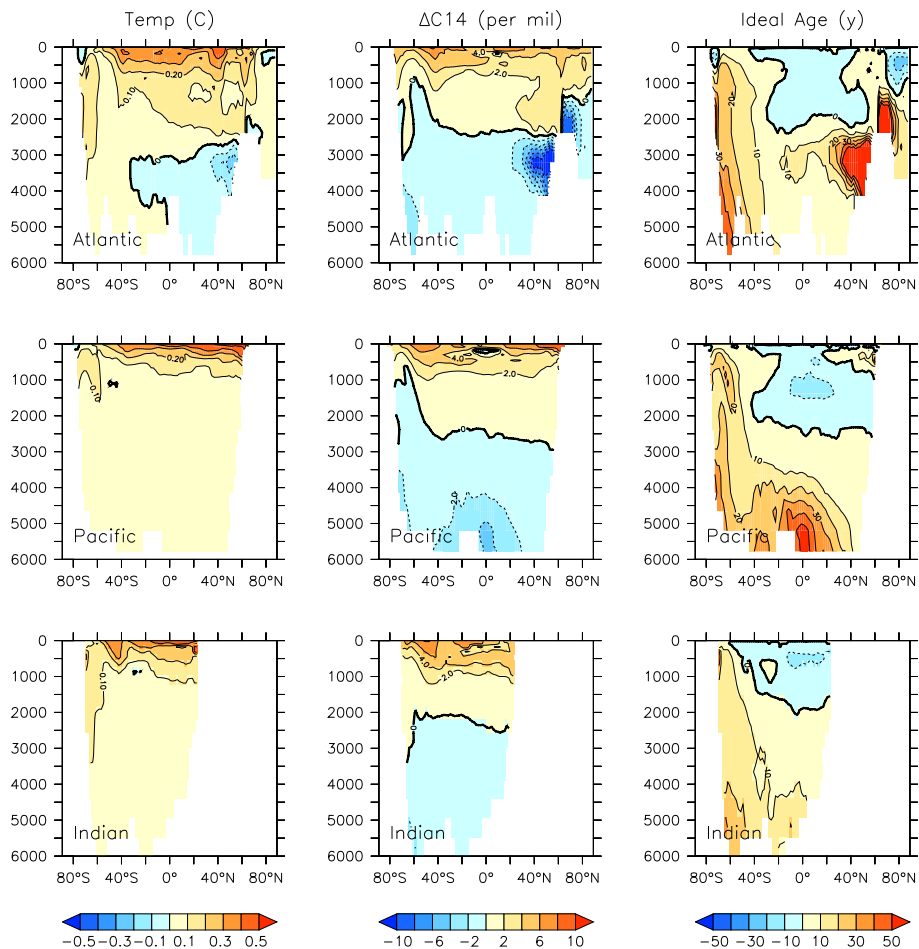
**Figure 13.** Model year 2015 zonally averaged surface NPP and phytoplankton biomass.



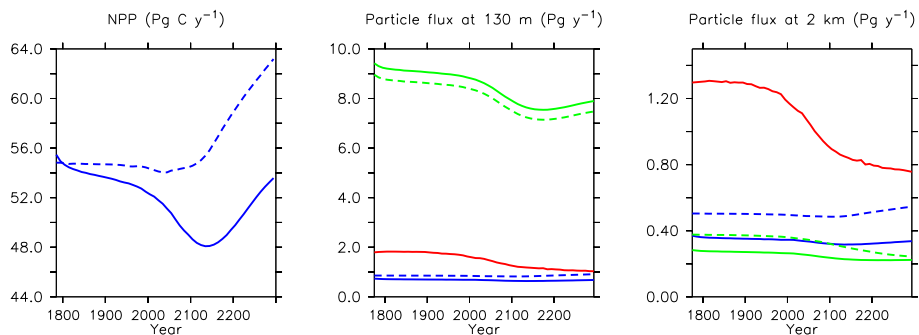
**Figure 14.** Modelled historical changes in zonally averaged and depth integrated NPP, and phytoplankton biomass, from 1964 to 2014.



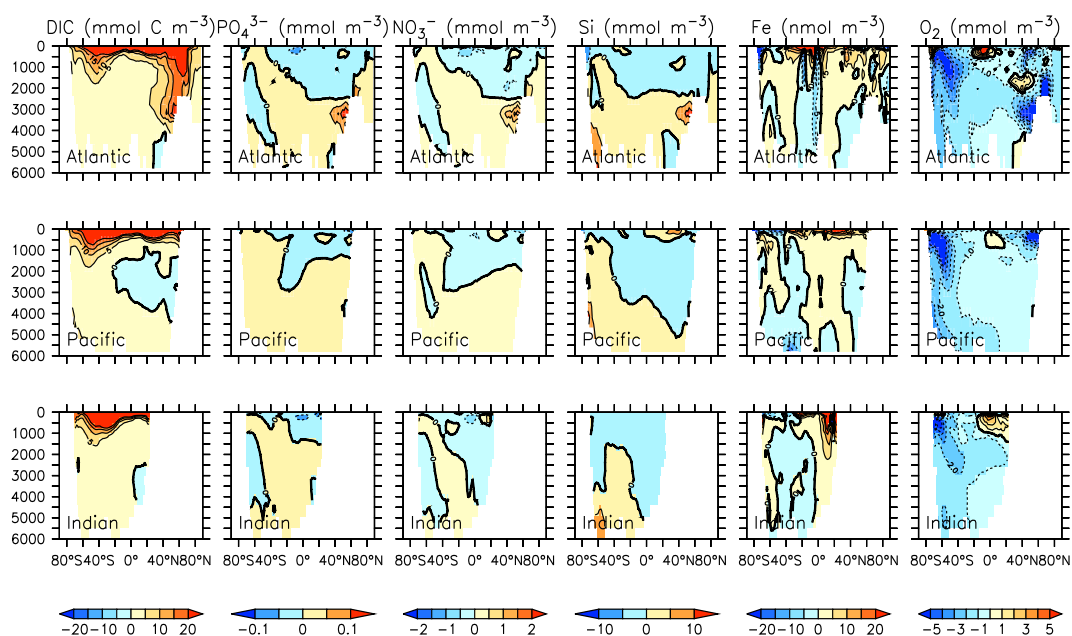
**Figure 15.** Modelled changes in phytoplankton biomass ( $\text{mmol C m}^{-3}$ ), from 1964 to 2014 (top row), and from 2014 to 2294 (bottom row).



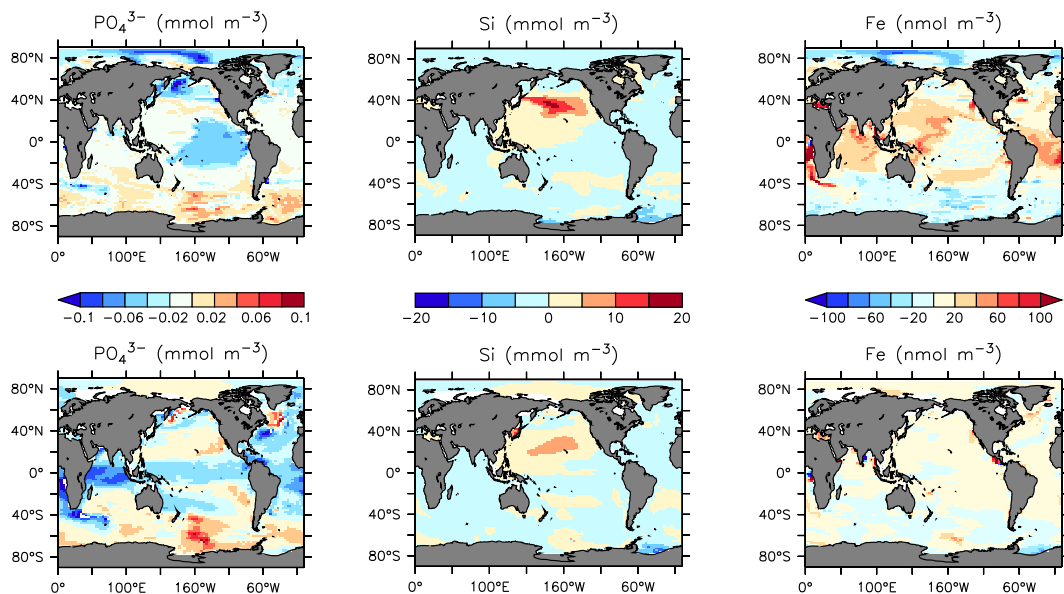
**Figure 16.** Modelled change in temperature, radiocarbon, and ideal age profiles by major ocean basin, from 1964 to 2144.



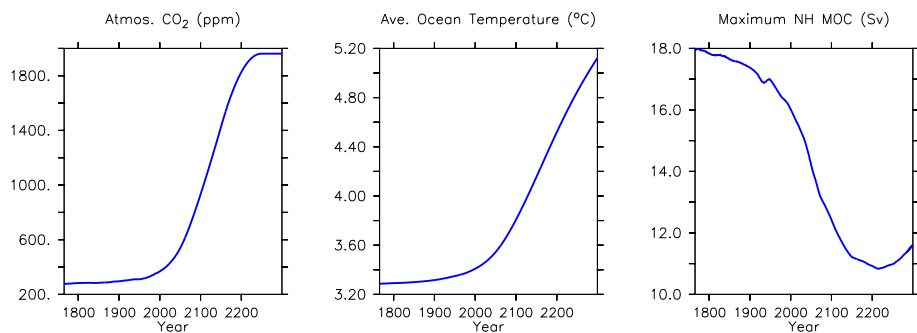
**Figure 17.** Major biogeochemical fluxes in the model, 1776 to 2300. Left panel is total net primary production, middle panel is total particle fluxes at 130 m depth, right panel is total particle fluxes at 2 km depth. Blue lines in the middle and right panels are PIC, green lines are POC, red lines are opal. Solid lines are from this model (KMBM3), dashed lines from the Keller et al. (2012) version (KMBM1).



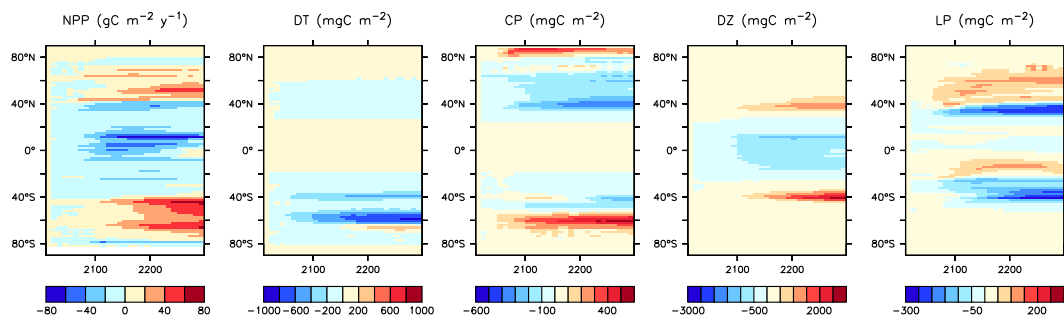
**Figure 18.** Modelled change in carbon and nutrient profiles by major ocean basin, from 1964 to 2014.



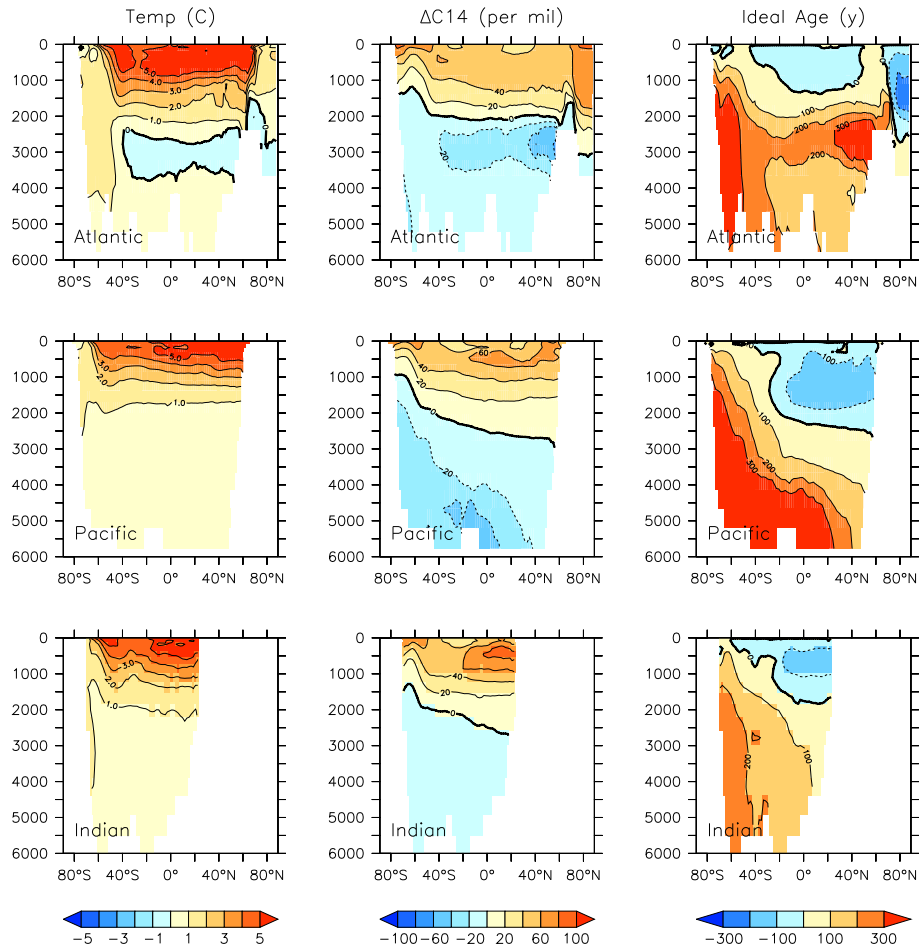
**Figure 19.** Modelled change in nutrients at the surface (top row) and 300 m depth (bottom row), from 1964 to 2014.



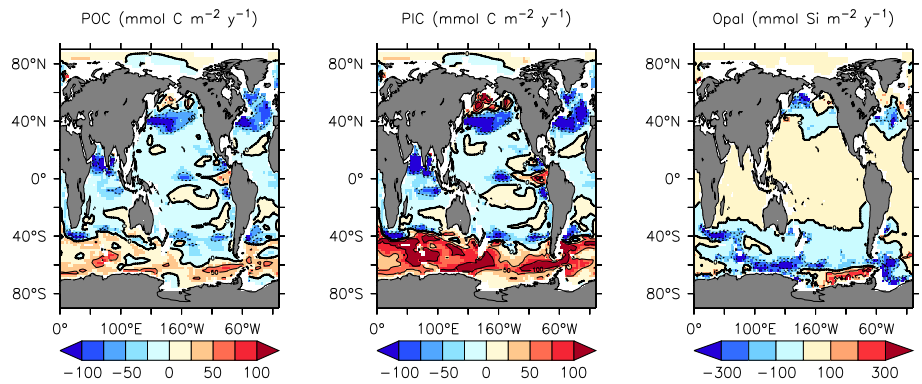
**Figure 20.** Major physical changes in the model, 1776 to 2300.



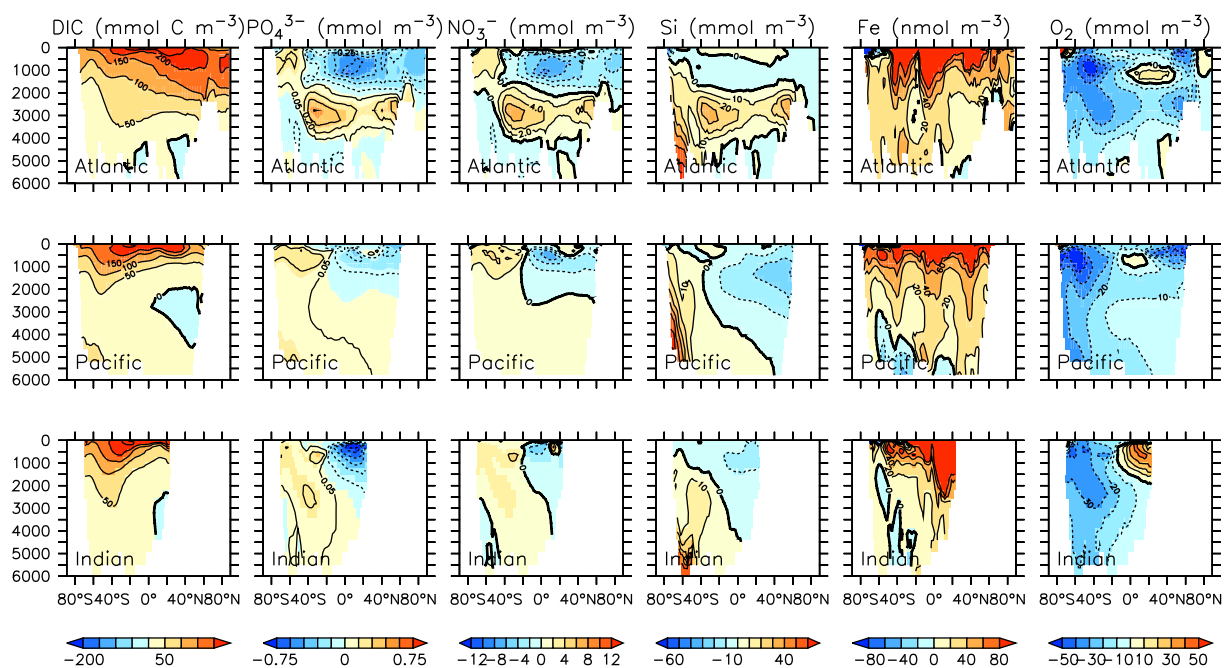
**Figure 21.** Modelled changes in zonally averaged and depth integrated NPP, and phytoplankton biomass, from 2014 to 2294.



**Figure 22.** Modelled change in temperature, radiocarbon, and ideal age profiles by major ocean basin, from 2014 to 2296.



**Figure 23.** Modelled change in particle fluxes at 2 km depth, from 2014 to 2294.



**Figure 24.** Modelled change in carbon and nutrient profiles by major ocean basin, from 2014 to 2294.

**Table 1.** KMBM3 state variables.

| Variable                   | Subscript Symbol              | Units                  |
|----------------------------|-------------------------------|------------------------|
| Silica                     | Si                            | mol Si m <sup>-3</sup> |
| Iron                       | Fe                            | mol Fe m <sup>-3</sup> |
| Phosphate                  | PO <sub>4</sub> <sup>3-</sup> | mol P m <sup>-3</sup>  |
| Nitrate                    | NO <sub>3</sub> <sup>-</sup>  | mol N m <sup>-3</sup>  |
| Dissolved inorganic carbon | DIC                           | mol C m <sup>-3</sup>  |
| Calcite                    | CaCO <sub>3</sub>             | mol C m <sup>-3</sup>  |
| Living calcite             | CaCO <sub>3liv</sub>          | mol C m <sup>-3</sup>  |
| Alkalinity                 | Alk                           | mol C m <sup>-3</sup>  |
| Oxygen                     | O <sub>2</sub>                | mol O m <sup>-3</sup>  |
| Diazotrophs                | DZ                            | mol N m <sup>-3</sup>  |
| Diatoms                    | DT                            | mol N m <sup>-3</sup>  |
| Low latitude phytoplankton | LP                            | mol N m <sup>-3</sup>  |
| Calcifiers                 | CP                            | mol N m <sup>-3</sup>  |
| Zooplankton                | Z                             | mol N m <sup>-3</sup>  |
| Free detritus              | Detr <sub>free</sub>          | mol N m <sup>-3</sup>  |
| Ballasted detritus         | Detr <sub>bal</sub>           | mol N m <sup>-3</sup>  |
| Detrital iron              | Detr <sub>Fe</sub>            | mol Fe m <sup>-3</sup> |

**Table 2.** Miscellaneous KMBM3 parameters.

| Parameter                              | Symbol                          | Units                                   | Value    |
|--|---------------------------------|---|----------|
| E-folding temperature                  | $T_b$                           | °C                                      | 15.65    |
| Molar organic P:N ratio                | R <sub>P:N</sub>                | unitless                                | 0.0625   |
| Molar organic C:N ratio                | R <sub>C:N</sub>                | unitless                                | 6.625    |
| Molar organic O:N ratio                | R <sub>O:N</sub>                | unitless                                | 8.46     |
| Molar sedimentary Fe:P ratio           | R <sub>Fe:P<sub>sed</sub></sub> | unitless                                | 0.004    |
| Molar organic Fe:N ratio               | R <sub>Fe:N</sub>               | unitless                                | 6.625E-6 |
| Molar mass of carbon                   | M <sub>C</sub>                  | g M <sup>-1</sup>                       | 12.011   |
| Molar mass of carbonate                | M <sub>CaCO<sub>3</sub></sub>   | g M <sup>-1</sup>                       | 60.01    |
| Light attenuation by phytoplankton     | $k_c$                           | (m mmol m <sup>-3</sup> ) <sup>-1</sup> | 0.07     |
| Light attenuation by CaCO <sub>3</sub> | $k_{CaCO_3}$                    | (m mmol m <sup>-3</sup> ) <sup>-1</sup> | 0.2      |
| Light attenuation by ice               | $k_i$                           | m <sup>-1</sup>                         | 5.0      |
| Light attenuation by water             | $k_w$                           | m <sup>-1</sup>                         | 0.04     |

**Table 3.** KMBM3 phytoplankton production and mortality parameters.

| Parameter                    | Symbol                       | Units                                  | Value                |
|------------------------------|------------------------------|--|----------------------|
| Growth rate                  | $a_{LP}$                     | $\text{day}^{-1}$                      | 0.4                  |
|                              | $a_{CP}$                     |  | 0.6                  |
|                              | $a_{DT}$                     |  | 0.7                  |
|                              | $a_{DZ}$                     |  | $0.81 \times a_{LP}$ |
| Half-saturation constant N   | $k_{NLP}$                    | $\text{mmol m}^{-3}$                   | 0.1                  |
|                              | $k_{NCP}$                    |  | 0.2                  |
|                              | $k_{NDT}$                    |  | 0.5                  |
| Half-saturation constant P   | $k_{PDZ}$                    | $\text{mmol m}^{-3}$                   | $k_{NLP}/16$         |
| Half-saturation constant Fe  | $k_{FeLP}$                   | $\text{nmol m}^{-3}$                   | 0.08                 |
|                              | $k_{FeCP}$                   |  | 0.34                 |
|                              | $k_{FeDT}$                   |  | 0.78                 |
|                              | $k_{FeDZ}$                   |  | 0.08                 |
| Half-saturation constant Si  | $k_{Si}$                     | $\text{mmol m}^{-3}$                   | variable             |
| Half-saturation Si scaling   | $k_{Si}^*$                   | $\text{mol Si m}^{-3}$                 | 0.03                 |
| Initial slope of P-I curve   | $\alpha_{\min}^{\text{chl}}$ | $(\text{W m}^{-2})^{-1} \text{d}^{-1}$ | 1.59                 |
|                              | $\alpha_{\max}^{\text{chl}}$ |  | 6.36                 |
| Chl:C ratio                  | $\theta_{\min}$              | unitless                               | 0.01                 |
|                              | $\theta_{\max}$              | unitless                               | 0.04                 |
| Phytoplankton mortality rate | $m_{LP}$                     | $\text{day}^{-1}$                      | 0.03                 |
|                              | $m_{CP}$                     |  | 0.03                 |
|                              | $m_{DT}$                     |  | 0.03                 |
| Microbial fast recycling     | $\mu_{0LP}^*$                | $\text{day}^{-1}$                      | 0.015                |
|                              | $\mu_{0CP}^*$                |  | 0.015                |
|                              | $\mu_{0DT}^*$                |  | 0.015                |
|                              | $\mu_{0DZ}^*$                |  | 0.015                |

**Table 4.** KMBM3 zooplankton parameters. Temperature-dependent parameter values are given for 0°C.

| Parameter                       | Symbol         | Units                | Value |
|---------------------------------|----------------|----------------------|-------|
| Maximum grazing rate            | $\mu_Z^\theta$ | day <sup>-1</sup>    | 0.571 |
| Maximum grazing rate parameters | $b$            | unitless             | 1.066 |
|                                 | $c$            | °C <sup>-1</sup>     | 1.0   |
| Food preferences                | $\psi_{LP}$    | unitless             | 0.2   |
|                                 | $\psi_{CP}$    |                      | 0.25  |
|                                 | $\psi_Z$       |                      | 0.2   |
|                                 | $\psi_{DZ}$    |                      | 0.05  |
|                                 | $\psi_{DT}$    |                      | 0.25  |
|                                 | $\psi_{Det}$   |                      | 0.05  |
| Half saturation constant        | $k_z$          | mmol m <sup>-3</sup> | 0.15  |
| Growth efficiency constant      | $\varpi$       | unitless             | 0.4   |
| Food assimilation efficiency    | $\gamma$       | unitless             | 0.7   |
| Mortality rate                  | $m_z$          | day <sup>-1</sup>    | 0.06  |

**Table 5.** KMBM3 particle export-production parameters.

| Parameter   | Symbol           | Units  | Value |
|---|------------------|--|-------|
| Detrital remineralisation rate                            | $\mu_{D,0}$      | day <sup>-1</sup>  | 0.07  |
| Detrital sinking speed at surface                         | $w_{D,0}$        | m day <sup>-1</sup>  | 12.28 |
| Detrital increase in sinking speed                        | $wdd$            | m day <sup>-1</sup>  | 5.8   |
| CaCO <sub>3</sub> ballast:total detrital production ratio | $R_{bal:tot}$    | unitless   | 0.05  |
| CaCO <sub>3</sub> :POC production ratio                   | $R_{CaCO_3:POC}$ | unitless   | 0.07  |
| CaCO <sub>3</sub> sinking speed at surface                | $w_{C,0}$        | m day <sup>-1</sup>  | 22.43 |
| CaCO <sub>3</sub> increase in sinking speed               | $wdc$            | m day <sup>-1</sup>  | 1.8   |
| Base opal:POC production ratio                            | $R_{Opal:POC,0}$ | unitless   | 0.5   |
| Opal dissolution rate constant                            | $\lambda_{Opal}$ | day <sup>-1</sup>  | 0.03  |
| Opal sinking rate   | $w_{Opal}$       | m day <sup>-1</sup>  | 75.0  |
| Organic particle iron scavenging rate                     | $k_{Fe_{org}}$   | (m <sup>3</sup> (gC d) <sup>-1</sup> ) <sup>0.58</sup>                 | 0.45  |
| Calcite iron scavenging rate                              | $k_{Fe_{ca}}$    | (m <sup>3</sup> (gCaCO <sub>3</sub> d) <sup>-1</sup> ) <sup>0.58</sup> | 0.45  |

**Table 6.** Globally integrated diagnosed biogeochemical properties at year 2004. Corresponding values are also given using the model of Keller et al. (2012).

| Property  | KMBM3 | KMBM1 | Independent Estimate   |
|---|-------|-------|------------------------|
| Primary Production (Pg C y <sup>-1</sup> )              | 52.27 | 54.33 | 44–78 <sup>a</sup>     |
| Calcite Production (Pg C y <sup>-1</sup> )              | 0.84  | 0.86  | 1.08–1.60 <sup>b</sup> |
| Opal Production (Pg Si y <sup>-1</sup> )                | 7.68  | -     | 14.4 <sup>c</sup>      |
| Nitrogen Fixation (Pg N y <sup>-1</sup> )               | 0.37  | 0.16  | 0.71–1.54 <sup>d</sup> |
| POC flux at 130 m (Pg C y <sup>-1</sup> )               | 8.80  | 7.04  | 5.73 <sup>e</sup>      |
| POC flux at 2 km (Pg C y <sup>-1</sup> )                | 0.26  | 0.36  | 0.43 ± 0.05            |
| CaCO <sub>3</sub> flux at 130 m (Pg C y <sup>-1</sup> ) | 0.69  | 0.84  | 1.1 ± 0.3              |
| CaCO <sub>3</sub> flux at 2 km (Pg C y <sup>-1</sup> )  | 0.35  | 0.49  | 0.41 ± 0.05            |
| CaCO <sub>3</sub> dissolution (Pg C y <sup>-1</sup> )   | 0.37  | -     | 0.5 ± 0.2 <sup>f</sup> |
| Opal flux at 130 m (Pg Si y <sup>-1</sup> )             | 1.60  | -     | 6.31 <sup>g</sup>      |
| Opal flux at 2 km (Pg Si y <sup>-1</sup> )              | 1.17  | -     | 2.07 ± 0.16            |
| Opal dissolution (Pg Si y <sup>-1</sup> )               | 3.87  | -     | 9.68 <sup>h</sup>      |
| Net Silica seafloor flux (Pg Si y <sup>-1</sup> )       | 1.32  | -     | 0.38                   |
| Silica river input (Pg Si y <sup>-1</sup> )             | 0.09  | -     | 0.44                   |
| Total Phytoplankton (Pg C)                              | 0.80  | 0.53  | 0.5–2.4 <sup>i</sup>   |
| Phytoplankton LP (Pg C)                                 | 0.07  | 0.51  | 0.28–0.64 <sup>j</sup> |
| Phytoplankton CP (Pg C)                                 | 0.12  | -     | 0.001–0.03             |
| Phytoplankton DT (Pg C)                                 | 0.12  | -     | 0.1–0.94               |
| Phytoplankton DZ (Pg C)                                 | 0.49  | 0.02  | 0.008–0.12             |
| Zooplankton (Pg C)                                      | 0.39  | 0.52  | 0.03–0.67 <sup>k</sup> |

<sup>a</sup>Carr et al. (2006), Jin et al. (2006)

<sup>b</sup>Balch (2018)

<sup>c</sup>Tréguer and De La Rocha (2013)

<sup>d</sup>Luo et al. (2014)

<sup>e</sup>all particle fluxes from Honjo et al. (2008) unless noted

<sup>f</sup>Luo et al. (2014)

<sup>g</sup>Tréguer and De La Rocha (2013)

<sup>h</sup>all subsequent silica fluxes from Tréguer and De La Rocha (2013)

<sup>i</sup>all biomass estimates from Buitenhuis et al. (2013)

<sup>j</sup>picophytoplankton

<sup>k</sup>pteropods