

Interactive comment on “The DeepMIP contribution to PMIP4: methodologies for selection, compilation and analysis of latest Paleocene and early Eocene climate proxy data, incorporating version 0.1 of the DeepMIP database” by Christopher J. Hollis et al.

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Hollis et al. 2019. The DeepMIP contribution to PMIP4: methodologies for selection, compilation and analysis of latest Paleocene and early Eocene climate proxy data, incorporating version 0.1 of the DeepMIP database. Geosci. Model Dev. Discuss.

1. General comments.

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The paper by Hollis et al. provides a well written summary of the proxy methods used to derive past climate states and greenhouse gas levels from the geological record – specifically for the latest Paleocene (LP), Paleocene-Eocene thermal maximum (PETM) and early Eocene climatic optimum (EECO). The focus on this geological interval is appropriate - as stated by the authors – as at around 56 Ma the world experienced a geologically short (~220 ky) and strong warming, the PETM, that was followed by a series of additional hyperthermals (e.g., ETM-2) and the longer duration EECO from ~53 to ~50 Ma. The PETM in particular offers a possible analog for current and forecast climate warming due to its short duration and the range of pCO₂ reconstructed for the PETM spanning current to projected greenhouse gas levels under different projections.

The authors provide a comprehensive yet succinct review of both paleontological proxies as well as geochemical proxies of temperature of waters and of the air, past CO₂ levels, as well as a range of paleoclimate parameters including annual and seasonal precipitation based primarily on terrestrially derived proxies. Further, their review highlights the challenges – strengths and weaknesses – posed by such a wide range of methods and approaches, ranging from concerns over precision versus accuracy, alternative methods of analysis and interpretation, taphonomy, limited geographic coverage, and the issues around compiling such disparate data into a database that can be used by the paleoclimate modelling community (model-model and model-data comparisons). As such, the article largely succeeds, offering a set of guidelines – as the article states – for the selection, compilation and analysis of such proxy data.

The article closes with a proposal for an “atlas” of climate conditions arrayed on an agreed paleogeography for the three selected time intervals to constrain and provide insight into the mechanisms controlling past hyperthermals via ‘database ver. 0.1’. The article draws on the expertise of a large set of authors – the DeepMIP team – whose research foci span these topics. The manuscript therefore represents a substantial contribution to modelling science and is appropriate for the journal. I have few

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concerns, and these mainly reflect topics dealt with in less depth than I would consider necessary, or restricting discussion to particular research teams, as detailed in the next section. In my specific comments I have focused on my area of expertise; paleobotanical reconstruction of past pCO₂ and climate, so “5 Terrestrial proxies for air temperature.”

2. Specific comments

Section 5 in its title only states ‘Terrestrial proxies for air temperature’, when this section also considers precipitation (e.g., p. 31 line 24, in section 5.2 or in section 5.3 at line 15 on p. 32), and should be re-titled as ‘Terrestrial proxies for climate’. Further, as leaf physiognomic methods as well as NLR approaches have been applied to reconstructing latest Paleocene to EECO precipitation (e.g., Greenwood et al. 2010; Pross et al. 2012; Eldrett et al. 2014; West et al. 2015; Suan et al. 2017; Hyland et al. 2018), the leaf-based and NLR proxies for precipitation should be included, e.g., CLAMP estimates growing season precipitation as well as that of the wettest 3 months and the driest 3 months, but these are not differentiated on p. 31 line 24, nor discussed at any point in section 5.2.

5.2 Leaf morphology-based approaches. This section is very focused on the univariate leaf margin analysis (LMA) and the multivariate CLAMP, and ignores other methods such as digital leaf physiognomy (DiLP; Peppe et al. 2011), and also (as noted above and below) barely mentions the use of leaf physiognomy for reconstructing precipitation (annual and seasonal values) in PETM and EECO studies (e.g., Leaf Area Analysis [LAA] and DiLP). Ignoring DiLP is unfortunate, but perhaps forgivable as DiLP has been applied to very few Paleogene floras to date. However, LAA (Wilf et al. 1998; Peppe et al. 2011) has been applied to a wider set of floras, including LP to EECO floras (e.g., Greenwood et al. 2010; Sunderlin et al. 2011; Smith et al. 2012; West et al. 2015; Lowe et al. 2018) and – as noted – yields data on precipitation that is critical to the DeepMIP project. The discussion of weaknesses and strengths in this section is also out of date as it misses current and recent literature on LMA and CLAMP. Citing

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Carpenter et al. (2012) at line 22, p. 30 is idiosyncratic as an example as this paper proposed a method that has been never used in another study, whereas LAA (Wilf et al. 1998) is cited, but this univariate method’s application to estimate Paleogene annual precipitation is not mentioned, despite LAA being used in multiple studies of the LP to EECO interval.

5.3 Nearest Living Relative Analysis. I disagree that the ‘most widely used method’ is the Coexistence Approach (CA), particularly when applied to the LP to EECO interval of concern. By focusing on a method that has been widely criticized – and further is not the most widely used NLR-proxy used in existing studies of the PETM and EECO – the authors create problems rather than solve them. My recommendation (as argued below) is that CA should not be the focus of recommendations.

Grimm and Denk (2012) and Grimm and Potts (2016) posed useful concerns about the analytical approach that underpins the ‘coexistence approach’ as employed by the PALAEOFLORA group. Principally CA’s reliance on single taxa to define upper and lower limits.

Grimm directs deep time climate researchers towards statistical methods such as CR-ACLE (Harbert and Nixon 2015) and an earlier iteration of my bioclimatic approach (Greenwood et al. 2005; see also Ballantyne et al. 2010) where we took a more objective probability based approach to either remove outliers, or to produce an estimate that reflected the highest probability solution for the suite of taxa present in a fossil assemblage. A nice succinct summary of these concerns and the more appropriate probability based approaches is given by Hyland et al. (2018).

In essence the concerns are that:

1) genera that are low species richness today (e.g., Ginkgo, taxodioid Cupressaceae such as *Glyptostrobus*, *Metasequoia* and *Sequoia*, etc) may be restricted today in their climate range for reasons other than climate, and may have occupied wider (or just different) climatic spaces in the Paleogene when they were clearly far more geograph-

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ically widespread and so potentially more ecologically varied than today – this argues against using single taxa such as Ginkgo or Metasequoia (both of which are commonly present in LP, PETM and EECO macrofloras) to define hard limits;

2) that climatic tolerance may have evolved in some lineages – e.g., palms are often used in Paleogene hyperthermal studies to constrain winter temperatures, however pre-Eocene palms likely didn't include the most cold-tolerant subtribes present today as they were a late Eocene or even Miocene radiation (Reichgelt et al. 2018 and references therein);

3) issues with identification of the NLRs – plant taxa may show the same leaf or pollen morphology in Eocene fossils as today, but come from plants that were phylogenetically sister taxa – i.e. not the same genus – with morphologically different flowers or other plant organs to the NLR, so the climate tolerance of the fossil's NLR may be a non-match – e.g., Platanaceae and Betulaceae leaves and pollen present in many North American floras may show 'Platanus' or 'Carpinus' leaf or pollen morphology, but are from plants with extinct taxon flowers and other organs (Macginitiea and Palaeocarpinus); methods like CA are inherently subjective because 'outliers' – e.g., those relictual taxa noted in point 1, or that 'sit outside' the climate range of the majority of taxa – are arbitrarily excluded from the CA, or arbitrarily used to set a limit for the range of possible estimates;

4) in the past there were concerns about where NLR climate range data were coming from – a lack of standardization of data sources and data quality control.

Due to all of the above concerns, a number of authors have adopted Grimm's recommendations or independently came to a similar solution (e.g., Ballantyne et al. 2010). For example, the mathematical approach developed in R by my colleague Tammo Reichgelt – a method with comparable assumptions to CRACLE – where we use probability density functions and Monte Carlo runs to select the highest probability climate estimate based on the whole suite of fossil taxa's NLRs, including taxa at different tax-

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onomic ranks (Family, tribe or genus) to allow for differing degrees of confidence of matching fossils to an extant NLR.

Furthermore, Greenwood et al. (2017), Hyland et al. (2018) and Reichgelt et al. (2018) and others derive the NLR climate range data from the international online portal GBIF.org which warehouses distribution data from most of the world's university, museum and government herbaria and equivalent, with clear policies on data quality and ownership, so verifiable data. The GBIF records are not without bias (geographical gaps or low data density for reasons of local and national politics or economics – rich countries lots of good data, poor countries not so much), but they do constitute the best available, i.e. best practice. Climate data is queried using either WorldClim (Hijmans et al. 2005; Fick and Hijmans 2017) or one of its comparable global climate surfaces (interpolated met station data and digital elevation model coupled with some atmospheric physics to fine tune effects of slope, aspect, continentality etc). I would recommend the authors consider advocating as a standard that NLR methods use GBIF for the source of their extant taxon distributions, with a mathematical climate surface such as WorldClim (Hijmans et al. 2005; Fick and Hijmans 2017) as the analytical procedure to derive the climate range data.

Finally, a suggestion. In Ballantyne et al. (2010), Greenwood et al. (2017) and Lowe et al. (2018) we employ what we call a consensus or an ensemble approach, where multiple terrestrial climate proxies – including both geochemical and paleobotanical methods in some instances – are combined and assessed using probability density analysis. We think this is the solution to the problem of competing proxies; query the data to see where they overlap and are most consistent. I would invite the authors to consider recommending such an approach.

3. Technical corrections.

p. 4, line 5, typo: Paleogene, not Paleogne

p. 4, line 15, omission: cite Greenwood and Wing (1995) as the LAT global N & S

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hemisphere latitudinal compilation predecessor of Huber and Caballero (2011).

p. 4, Line 32, typo? I would think 'end-member' rather than without the hyphen.

p. 5, line 24: cite also Eldrett et al. (2014) as well as Suan et al. (2017) as these authors document from terrestrial plant palynomorphs this same point for the PETM at high northern latitudes.

p. 22, lines 9-34: I would like to see mention here the concerns expressed by Eberle et al. (2010) on a possible seasonal bias in Arctic TEX86 reconstructions.

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