Review1

Authors' Response: We thank the reviewer for their thorough review of the paper. We have addressed every review point, to the improvement of the paper. Individual responses to different review points follow:

Review: I have two main concerns with the paper: the first refers to the C3MP dataset used to derive the emulator response functions, and the second refers to how these response functions are going to be used. The C3MP dataset is a large set of 99 CTW sensitivity tests carried out by a number of site-based crop models covering a range of different crops at a total of 1135 sites, of which data from 575 sites are actually used in this study. The climate change signal used in these sensitivity tests is completely synthetic since it consists of applying a temporally uniform temperature offset or precipitation multiplier to a historical baseline weather timeseries. However, in reality, climate change is not constant over time. For example, precipitation might increase during part of the year, while decreasing during other times. I am not convinced that the constant CTW perturbation experiments are equivalent to using a more realistic climate timeseries, and I would suggest that the authors either show in the paper or at least point the reader to other literature showing that this is a valid approach. Otherwise, the authors risk that there is already a bias in the data used to train the emulator which would propagate to the emulated yields.

Response: We have clarified that we are looking at climatological mean TW changes during the growing season only, rather than changes to seasonality to our methods, as outlined in the two peer-reviewed citations detailing C3MP and its use. Analysis within Ruane et al., 2014 showed that the explicit modeling of future scenarios was quite consistent with aggregating the seasonal changes (see figure comparing simulated vs. emulated yields). Changes in seasonality will be reflected in the seasonal precipitation and temperature changes, but explicit action to adjust growing seasons to match new seasonality would require adaptation. Autonomous adaptation and technological gains are included in IAMs as part of the exogenous trend, so we focus on the primary climatological pressure, recognizing that we are excluding the secondary effects of seasonality. Climatological changes are considered here specifically because this is for use in GCAM, which solves for an equilibrium on 5 year timesteps during which subannual dynamics (such as the distribution of precipitation *during* the growing season) tend to average out. A user could investigate the impacts of different growing seasons by processing the climate data of interest from annual monthly to growing season average with an appropriate growing season mask for their intent. We believe such investigations are outside the scope of this paper. However, we have also included a subsection 2.1.1 explicitly addressing several known caveats of the C3MP data set.

Review: My second concern is that response functions for some of the 25 production groups are based on a very small number of sites, in the most extreme case only two sites. I cannot help but wonder how representative these results really are, keeping in mind that each production group represents one crop-irrigation-latitude combination, with latitudes only distinguished into extended tropics and mid-latitudes. In addition, these sites are not only used to derive a mean yield response, but also an estimate of response uncertainty. The high and low response functions are supposed to represent site responses at the mean plus/minus one standard deviation level but how meaningful are these estimates based on such a small sample

size? The emulator response functions are only derived for two regions (extended tropics and mid-latitudes) but I wonder at what level of spatial detail they will be used later.
Response: We have included a new section 3.1.1 to directly address model performance in the production groups with small sample size. We also have clarified language regarding spatial scales throughout the paper. As a brief summary, we acknowledge that some crops and regions are under-represented in the training data. However, for many crops and regions, there is no comparable dataset available. In Section 3.1.1, we discuss validation that our modeling framework does represent the underlying data well for the production groups with small sample sizes. We also distinguish between the two-region response function and the degree of spatial heterogeneity that still results when combined with gridded temperature and precipitation change projections (throughout the text, but particularly in Section 2.1.1 Known caveats of the C3MP data set).

Review: In Figure 8 and 9 of the paper, the authors take spatial patterns of climate change from the HadGEM2-ES model and use their response functions to derive corresponding patterns of yield change. These patterns are shown, but not evaluated in any way. I would suggest that the authors compare their derived patterns of yield change to simulations of yield change from global gridded crop models for the same climate data. After all, the intent of the emulator is to replace simulations by global gridded crop models. Such crop yield simulations for the HadGEM2-ES RCP8.5 scenario used in Figure 8 and 9 of the paper are, for example, available from a number of crop models and for a number of different crops within the ISIMIP data archive at https://esg.pik-potsdam.de/projects/isimip/ Such a comparison would help to address both of my concerns voiced above.
Response: We thank the reviewer for this helpful insight. We have added an extensive new section 4.1 directly comparing to the ISIMIP global gridded crop modeling results, as well as to other modeling efforts.

Review: Page 2, ll. 25 – 28: You state that previous emulators were restricted to emulating yield change under RCP scenarios. I am aware of at least two other global crop yield change emulators derived from crop model simulations that are applicable to any future climate scenario: Oyebamiji et al. (2015) and Ostberg et al. (2018). It might be useful to contrast them to the work presented in this paper.
Response: We apologize for this oversight and have added a discussion of these papers to our Introduction (paragraph beginning on P3L3).

Review: Page 4, starting with line 11, and Figure 1: You outline three use cases for the Persephone yield emulator, but none of this is actually done in this paper. So I'm not sure if the Methods section is the right place for this.
Response: We have streamlined this discussion of use cases and moved it from the Methods to the Introduction, as the intended use in GCAM has motivated many of our modeling choices.

Review: Page 6, section 2.2: The Copernicus guidelines request that datasets should be cited with a reference in the reference list. Is there a reference for the C3MP dataset?
Response: We have included language in Section 2.1 that the two peer-reviewed C3MP publications (Ruane et al, McDermid et al) include data availability information. The relevant data to this work is also included in the paper analysis archive.

Review: Page 6, ll. 8 – 14: This part does not refer to the setup of C3MP or to your processing of the C3MP data. Instead, it refers to how climate data is pre-processed before use with the finished emulator. It should probably be moved to section 4.'
Response: This has been done.

Review: Page 7, ll. 7 – 9: Unless I am mistaken, the global gridded crop models within AgMIP also conducted the 99 CTW sensitivity tests. They should offer a much better global coverage. Would it be worthwhile adding some discussion of why this paper used the site-based results instead of the global simulations?
Response: Unfortunately, the global gridded crop models did *not* participate in the 99 CTW tests that create the C3MP archive. The global gridded crop models have conducted their own, separate sensitivity tests and the data is not yet publicly available. Therefore, we developed this emulator using data we that was publicly available, but with the aim of a sufficiently flexible framework as to update to newer data sets (like the globally gridded crop models) if/when they become available. We have added language to the introduction and discussion highlighting that the Persephone framework described here can be updated in future versions, possibly with different explanatory variables, to include such gridded simulations as they become available. In the new section 4.1, comparing to the ISIMIP GGCM results, we now explicitly state that these models did not participate in the C3MP exercise

Review: Page 8, ll. 19 – 20: Are the 8 different functional forms documented anywhere? Are these the same as are used in the emulator?
Response: Thank you for highlighting this oversight. The functional forms used to estimate site-specific baseline Yields are now explicitly documented in the new Appendix B.

Review: Page 11, ll. 9 – 10: Is the value of b0 constrained between -0.02 and 0.02 or is it 0.02%? The following sentence suggests that it is 0.02%.
Response: We thank the reviewer for catching this typo. We have corrected this.

Review: Given the very small sample sizes of about 1/3 of the production groups the 84.135th and 15.865th percentiles do not seem very meaningful'
Response: This is addressed with our new section 3.1.1 directly evaluating the performance of the three functional forms for small sample size production groups.

Review: Section 3.2: It seems to me that this is rather a test whether the crop models used in training feature these important relationships. Or would you say it is possible that these relationships are present in the models, but missing in the emulator? Since the evaluation is

positive I guess it means that the relationships are present in the models and retained in the emulator.
Response: Yes, the relationships are present in the models and retained by the emulator. The final sentence is correct and we have clarified the text to reflect this point.

Review: Page 20, l. 4 – page 21, l. 1: Climate change can affect both the start and the length of the growing season. Is this accounted for or is the same growing season used under climate change as during the reference period?
Response: The same growing season was used by every model for their training runs. We have updated our methods and discussion to explicitly state this. We also note that we plan to use with GCAM and are focused on long term climatological changes, rather than specific changes to seasonality that may average out over 5 year timesteps.

Review: Page 21, ll. 8 – 10: On the one hand you talk about passing CTW changes for regions into the emulator, on the other hand you mention a gridded map of yield changes. So are the yield changes at the same spatial resolution as the climate data or is there a difference between resolutions (region versus grid)? Please clarify
Response: We have updated this language to be clearer (now occurs on P24).

Review: Page 21, ll. 17 – 18, and Figure 9: Looking at Figure 9, it seems to me that most regions show a positive yield change under the high response function, not just "a few regions". Also, I think that in the bottom row of Figure 9 the maps for mean and high response are swapped. The bottom right map (which should be the high response) looks identical to the map for Maize in Figure 8 (which shows the mean response).
Response: Thank you for catching this mistake, we have corrected the error in figure 9. Rather than swapping the mean and high response maps for Maize in figure 9, we inadvertently included the Wheat mean response map with the Maize high and low maps in the original figure. We have corrected the rainfed Maize mean figure, and clarified the language discussing Figure 9.

Review: Page 23, ll. 6 – 8: Here, you emphasize the rapid evaluation time of the response functions relative to a global gridded crop model, but I think you should really try to show that the emulator response is actually comparable to what you would get using a global gridded crop model. This step is missing in the paper.
Response: Thank you for this suggestion, we have added section 4.1, comparing many of our results to some of the ISIMIP GGCM results.

Review: Page 24, ll. 8 – 10: Given that the response functions are only derived for two latitudinal bands I would say that they cannot really be used to characterize the range of uncertainty within national or multi-national units (unless the respective unit is covered by C3MP sites).
Response: We have clarified the language in this section of the Conclusions and discussion that the response functions are able to characterize the range of C3MP response sites, and that this

is only a partial characterization of the response uncertainty within larger national land units. And that using more spatially complete data sets for training in the future will improve this characterization.

Review: Page 24: In the paragraph on caveats, I would suggest to add discussion of potential biases arising out of way the CTW experiments are set up. Another source of uncertainty results from the fact that crop model simulations generally omit adaptation options such as changing sowing dates or switching to different cultivars. This is done for simplicity and comparability but is not realistic considering that agriculture is a highly managed system, and potentially creates another bias in the crop model simulations of yield change that are used to train the emulator.
Response: We have added text to this effect.

Review: The greyscale lines in Figure 6, 8 and 9 are hard to see at all, let alone distinguish the different shades of grey.
Response: We have replaced the grayscale lines with black contours placed at values of 10%, 20%, etc and included explicit labeling of the black contours on the plots.

Review 2

Review:

The authors present a methodology, with an accompanying R package, to emulate changes in crop yield under global change scenarios. The functions produced by this framework can be introduced in other models such as GCAM, which can help to speedup different types of simulations. The manuscript is well written and presents important results that merit publication in GMD. However, I have one major comment to this work. Although I really liked the Bayesian approach proposed here, which is more robust than previous linear regression approaches, I had problems understanding the approach for modeling the standard deviation term. It seems that the approach yields negative values of σCTW , which is evident by the use of absolute values in equation 6. As far as I am concerned, standard deviation values can never be negative since theoretically they are the square root of the variance. The choice of prior distributions for modeling σCTW expressed in equation 5, explains the reason for the negative values. For the baseline case, $b0 \sim N(0, 0.001)$ yields a distribution of standard deviations centered around zero, which I find difficult to understand. Modeling prior distributions for the variance in Bayesian analysis is not trivial, and there are many analyses dealing with this problem (e.g. see papers by Andrew Gelman). Most controversies about this topic deal with the choice of the prior distribution for variance parameters and whether gamma, inverse gamma, or other distributions are appropriate choices. These distributions however, are always defined in the positive part of the real line R +. I suggest the authors to revise this part of the manuscript. If there is important information that I am missing regarding this issue, the authors should at least explain their choice of distribution and its interpretation.

Response:

We thank the reviewer for reading the work so closely. Indeed, the reviewer has correctly characterized our methodology, but highlighted that we did not communicate our methods clearly. We have clarified the text in our section on emulation, specifically addressing the reviewer's comments.

A crop yield change emulator for use in GCAM and similar models: Persephone v1.0

Abigail Snyder[1], Katherine V. Calvin[1], Meridel Phillips[2, 3], and Alex C. Ruane[3]

[1]Joint Global Change Research Institute, Pacific Northwest National Laboratory, College Park, MD 20740
[2]Columbia University Earth Institute Center for Climate Systems Research, New York, NY, USA
[3]NASA Goddard Institute for Space Studies, New York, NY, USA

Correspondence: Abigail Snyder (abigail.snyder@pnnl.gov)

Abstract. Future changes in Earth system state will impact agricultural yields and, through these changed yields, can have profound impacts on the global economy. Global gridded crop models estimate the influence of these Earth system changes on future crop yields, but are often too computationally intensive to dynamically couple into global multi-sector economic models, such as GCAM and other similar-in-scale models. Yet, generalizing a faster site-specific crop model's results to be used globally will introduce inaccuracies, and the question of which model to use is unclear given the wide variation in yield response across crop models. To examine the feedback loop among socioeconomics, Earth system changes, and crop yield changes, rapidly generated yield responses with some quantification of crop response uncertainty are desirable. The Persephone v1.0 response functions presented in this work are based on the Agricultural Model Intercomparison and Improvement Project (AgMIP) Coordinated Climate-Crop Modeling Project (C3MP) sensitivity test data set and are focused on providing the Global Change Assessment Model (GCAM) and similar models with a tractable number of rapid to evaluate, dynamic yield response functions corresponding to a range of the yield response sensitivities seen in the C3MP data set. With the Persephone response functions, a new variety of agricultural impact experiments will be open to GCAM and other economic models; for example, examining the economic impacts of a multi-year drought in a key agricultural region and how economic changes in response to the drought can, in turn, impact the drought.

1 Introduction

Agricultural yields are susceptible to changes in temperature, precipitation, growing season length, CO_2 concentrations, and other Earth system factors. While both the nature of the future climate and its impact on agricultural yields are uncertain (Rosenzweig et al., 2014; Pirttioja et al., 2015; Fronzek et al., 2018; Asseng et al., 2013, 2015; Martre et al., 2015; Lobell, 2013), it is clear that there is potential for identifying the important effects on agriculture and, in turn, the economic state of the world at large. The global multi-sector economic model Global Change Assessment Model (GCAM)[1] ~~(Kim et al., 2006; Clarke et al., 2007;~~

[1]Model and documentation available at https://github.com/JGCRI/gcam-core, http://jgcri.github.io/gcam-doc/toc.html

(Kyle et al., 2011; Wise et al., 2014; Calvin et al., 2019; Hartin et al., 2015) and other similar-in-scale models (Nelson et al., 2014) are ideal for understanding the far reaching impacts of this climate-agriculture-economic cycle, but rely on external projections of agricultural yields to quantify these effects (Figure 1, panel A). This asynchronous process results in inconsistencies between the economic and biophysical world, and overlooks feedbacks and unintended consequences as the future shifts (Ruane et al., 2017).

Several modeling groups, including the GCAM model development team, are interested in explicitly modeling and understanding bidirectional feedbacks between the Earth and the human systems ~~.~~ (e.g. Figure 1, panel C). Agriculture is one important pathway (of many) through which these systems directly interact. A prime example would be to study the impacts of a multi-year drought in a key agricultural region. The drought would affect yields, which would affect the agricultural supply to the global economic market. In a model like GCAM, this would lead to price changes and shifting land to more profitable crops. The new spatial distribution of agricultural land would change land related emissions, which will in turn affect climate and therefore yields moving forward. Being able to model each component of this process and the interactions among them is key to considering important questions like this one.

Currently, GCAM operates on a five year time step and is coupled with a physical Earth system emulator, Hector (Hartin et al., 2015) (as in Figure 1, panels A and B), to explore global change questions in rapid enough evaluation times to allow for large numbers of simulations to be analyzed as part of a wide range of experiments. GCAM is a recursive dynamic partial equilibrium model that is calibrated to a historical base year of 2010 and used to simulate forward in time by incorporating changes in quantities such as population, GDP, and technology to produce outputs that include land, water, and energy use as well as emissions and commodity prices. For agricultural production in GCAM, yield change trends representing generally positive change assumptions over time due to *non-climate* factors (changes in management, new seed genetics, new technologies, use of chemicals/fertilizers, adaptation, etc.) are used to calculate the profitability of a crop-irrigation-fertilizer combination in each of 384 GCAM land units at each time step based on the global crop price. This profitability determines land allocated to each crop, and the combination of exogenous yields and land allocation gives production of each crop-irrigation-fertilizer combination such that global supply and global demand are met on each timestep. The details of this allocation are provided in Kyle et al. (2011); Wise et al. (2014); Calvin et al. (2019). Shifting land allocation among different crop-irrigation-fertilizer combinations leads to a degree of endogenous yield intensification within GCAM.

Past agricultural impacts studies using GCAM ~~(Calvin et al., 2013)~~ (Calvin and Fisher-Vanden, 2017) have focused on using outputs of global gridded crop model (GGCM) studies (e.g., Rosenzweig et al., 2014; Elliott et al., 2014; Müller et al., 2017) in a strictly feed-forward way (Figure 1, panel A). Direct coupling of a GGCM to GCAM ~~is prohibitively expensive in the computational resources required to run the~~ would result in a computationally expensive modeling framework, limiting the number of simulations that could be performed. Yet, large ensembles of simulations are necessary to explore and understand future response options, so there is great need for a computationally efficient model that could explore the uncertainty space. While GCAM is already coupled to a simple climate model, Hector (Hartin et al., 2015), this coupling is one-way: emissions are passed to the climate model, but to date dynamic ~~, bidirectional~~ feedbacks between climate and humans ~~on~~ at each timestep are missing. In this paper, we describe the first version of Persephone (v1.0), a simple representation of mean agricultural response and uncertainty to future climate that can be incorporated into GCAM and similar models. Further detail of the desired studies this yield change emulator would be used for are given in Section 2.1 and discussed at length in Ruane et al. (2017).

An ideal solution to the computational expense of coupling a GGCM to GCAM is a yield response emulator, which uses past crop yield model runs to predict what the model *would* have done under different conditions, had it been run. However, previous work in this area has been restricted to either emulating ~~GGCM results under~~ crop model results under fixed [CO_2]-temperature pathways such as the various RCPs ~~(Blanc, 2017)~~ (Oyebamiji et al., 2015; Blanc, 2017; Ostberg et al., 2018) or building statistical models from empirical and historical data (Lobell, 2013; Moore et al., 2017; Mistry, 2017; Mistry et al., 2017)~~, neither of which span a wide range~~. While an emulator trained on RCP-driven scenarios can be used to estimate yield change in any future climate, the RCPs only span a subset of possible future ~~climate. These approaches then~~ climates. In particular, should one want to consider the impacts of [CO_2]-temperature pathways that substantially differ from the RCPs, these emulators would face the difficult ~~problem of extrapolating into the future,~~ task of predicting yield changes outside of the conditions of the training data. Statistical models of empirical and historical data also must predict yield changes in response to future climate outside of the conditions of the training data, ~~to serve the~~ especially in response to large [CO_2] increases. Substantial departure from the RCPs and historical values of [CO_2] is very possible in the bidirectional coupled human-earth system applications outlined above and an emulator equipped to handle that is desirable. Finally, many of these past studies have lacked a way to capture aspects of uncertainty that would be useful for the GCAM bidirectional feedback experiments described in Section 2.1.

The Agricultural Model Intercomparison and Improvement Project (AgMIP) (Rosenzweig et al., 2013) took steps to begin addressing these issues with the Coordinated Climate-Crop Modeling Project (C3MP), a modeling study specifically designed to, among other things, provide the data necessary to develop a flexible and dynamic crop yield emulator (Ruane et al., 2014; McDermid et al., 2015). C3MP invited point-based crop modelers from across the AgMIP community to simulate their calibrated agricultural system's response to 99 sensitivity tests in which 1980-2009 baseline climate data were modified to synthesize changes in mean carbon dioxide concentration ([CO_2]), temperature, and precipitation. The 99 Carbon-Temperature-Precipitation (denoted CTW, W for Water rather than P for Precipitation) tests that make up the C3MP protocol were selected using a Latin hypercube to ensure that future scenarios through the end of the 21st ~~centure~~century, including all RCPs, fall within the training model simulation data over the vast ~~marjority~~ majority of agricultural lands (Ruane et al., 2014). The full space of CTW changes that these 99 tests represent is: 330-900 ppm global [CO_2], -1°C to +8°C from local baseline temperature, and -50% to +50% from local baseline precipitation (applied as a multiplicative factor). A particular CTW perturbation could be associated with a specific time slice, for example the 2050s climate changes from a given Earth System Model (ESM) RCP4.5 projection, or from a climate condition generated within GCAM as a result of interactions between socioeconomic development and the natural environment. Finally, the C3MP study featured broad spatial coverage (albeit not uniform) of a wider variety of crop models, crops, and management practices than has been incorporated into past GGCM or emulator work. More than 50 participating crop modelers helped C3MP record yield response simulation results from a total of 1135 sites, differing by location, crop species, cultivars, crop model, farm management, etc.

The Persephone ~~response functions~~ framework presented in this work ~~are designed to~~ is designed to develop yield response functions to CTW changes from a given data set. The Persephone V1.0 response functions, based on the C3MP data set, provide a computationally inexpensive estimate of the change in agricultural yield due to a change in the Earth system, and make use of the promising data relating yield changes to CTW changes collected in C3MP. Specifically, we present biologically reasonable response functions that are rapid-to-evaluate and more dynamic than past options for incorporating crop responses into models like GCAM. ~~The response functions also represent the large uncertainty in yield response across crop models to a given change in local Earth system state.~~ We strictly considered responses to long term Earth system changes. The C3MP results or other appropriate data sets could be further used to examine the effect of inter-annual variability on yields in ~~the future~~Persephone V2.0 and beyond, although this would require additional complexities in seasonal yield variations that are largely averaged out in long-term trends.

2 ~~Methods~~

1.1 ~~GCAM background and experimental goals~~

~~The Persephone yield response functions are developed for use with models that couple energy, economy, agriculture and land-use, such as GCAM. GCAM operates on a five year time step and is coupled with a physical Earth system emulator, Hector (as in Figure 1, panels A and B), to explore global change questions in rapid enough evaluation times to allow for large numbers of simulations to be analyzed as part of a wide range of experiments.~~

~~GCAM is a recursive dynamic partial equlibrium model that is calibrated to a historical base year of 2010 and used to simulate forward in time by incorporating changes in quantities such as population, GDP, and technology to produce outputs that include land, water, and energy use as well as emissions and commodity prices. For agricultural production in GCAM, yield change trends representing (generally positive) change assumptions over time due to *non-climate* factors (changes in management, new seed genetics, new technologies, use of chemicals/fertilizers, adaptation, etc.) are used to calculate the profitability of a crop-irrigation-fertilizer combination in each of 384 GCAM land units at each time step based on the global crop price. This profitability determines land allocated to each crop, and the combination of exogenous yields and land allocation gives production of each crop-irrigation-fertilizer combination such that global supply and global demand are met on each timestep. The details of this allocation are provided in Kyle et al. (2011); Wise et al. (2014). Shifting land allocation among different crop-irrigation-fertilizer combinations leads to a degree of endogenous yield intensification within GCAM[2].~~

~~To date, the only method for using GCAM to explore the far reaching impacts of agricultural yield changes due to future climate has been to draw predetermined scenarios undertaken by the GGCMs, such as crop yield under select emissions pathways and ESM combinations, from public archives. These predetermined crop yield data sets are converted to exogneous multipliers which are applied to GCAM's exogenous technological yield change assumptions. Using this new yield change assumption set, GCAM is re-run (Figure 1, panel A).~~

[2] ~~Note that this is a new feature from GCAM 5.0 and onward.~~

~~The Persephone yield response functions were developed~~ The response functions also represent the uncertainty in yield response across crop models in the C3MP data set to a given change in local Earth system state, for use in three ~~new~~ types of agricultural impacts studies with GCAM:

1. A partially coupled, feed forward study (Figure 1, Panel B) similar to methodology in Ruane et al. (2018). A future climate time series of interest (a non-traditional RCP, climate stabilization level, or hypothetical drought, for instance) is input to the yield response functions, returning yield changes. These yield changes are applied as multipliers to GCAM input files and GCAM is run forward for the entire time period of interest in order to trace the broad impacts on energy, water, and land use of the future climate time series. In this type of study, we only capture the implications of climate for human systems.

2. A fully coupled feedback loop that updates on every model timestep to understand how societal pressures drive environmental impacts which in turn create or reduce societal pressures (Figure 1, Panel C). In this case, the yield changes must be calculated very quickly in order to evaluate on each step and interact with GCAM. In this type of study, we can capture the effects of humans on climate and climate on humans, simultaneously.

3. Joint climate-crop uncertainty studies of the above two experiments. For tractability, the GCAM development team specifically seeks a mean response function as well as two additional response functions that represent a range of yield response uncertainty. Persephone also stores the full predictive distributions of yield changes for any given CTW change that these three response functions span. If a user desires a different representation of uncertainty, the distribution may be sampled.

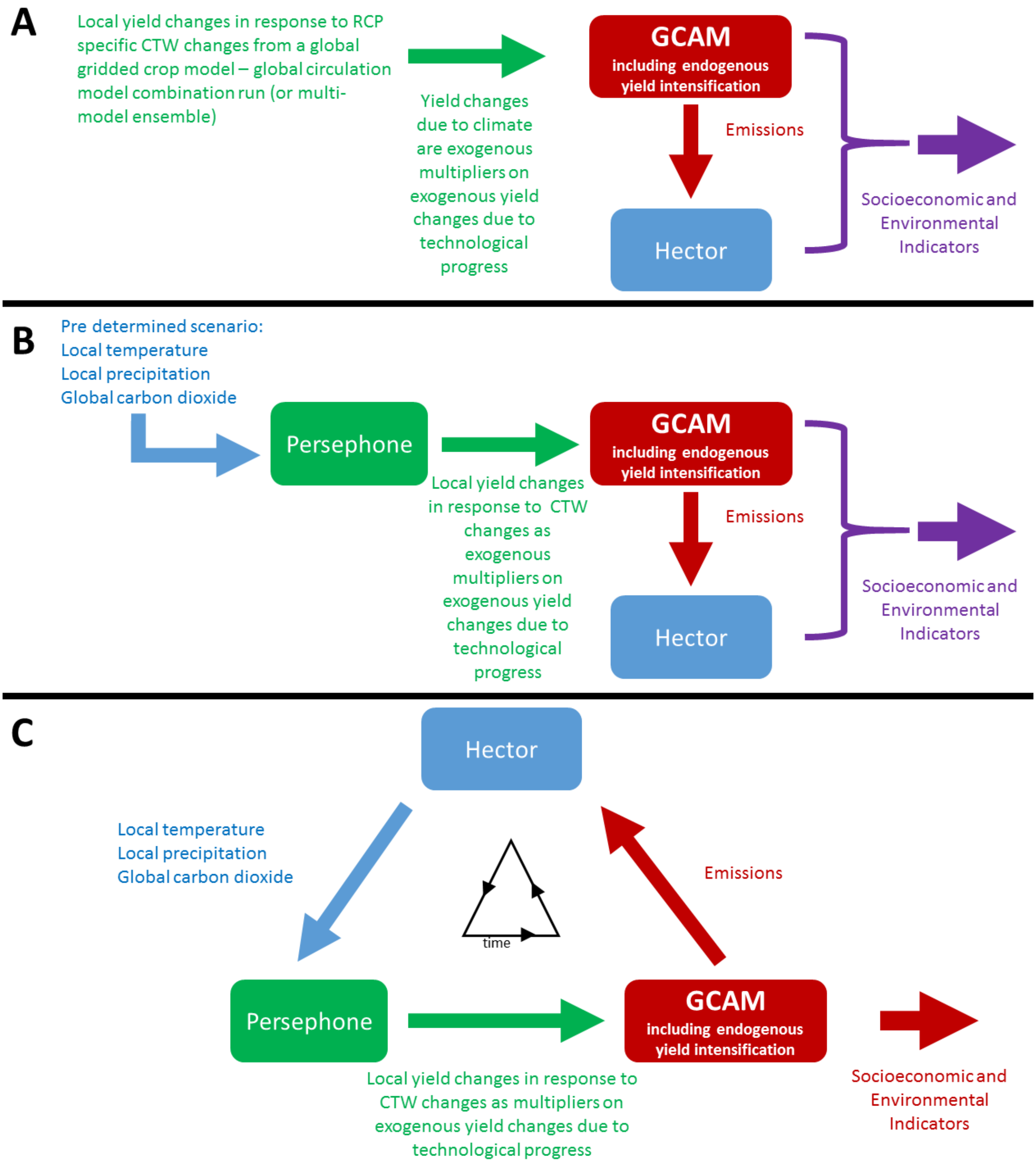


Figure 1. The current method for incorporating agricultural impacts into GCAM and two experimental designs for using Persephone v1.0 with GCAM. Panel A: The current method for incorporating yield changes from a global gridded crop model into GCAM. Panel B: A partially coupled, feed forward study incorporating yield changes from a predetermined climate scenario into GCAM. Panel C: A fully coupled feedback loop that iteratively updates agricultural yield impacts.

2 Methods

2.1 C3MP dataset

Full details of the C3MP protocols, design, and the location output archive can be found in Ruane et al. (2014); McDermid et al. (2015). Here, we highlight some of the key features of the data set and outline our processing of C3MP data for ~~use in training response functions~~ using the Persephone framework to train V1.0 response functions with the Persephone framework.

C3MP recorded yield response simulation results from a total of 1135 sites (differing by location, crops, crop model, management, etc) for each of 99 CTW sensitivity tests designed to cover a range of CTW changes that most future climates would fall into. For each site, each CTW test is applied to change a local ~~timeseries~~ time series of weather data from 1980-2009 and then the crop model is run to produce 30 years of impacted yields for the CTW test, which are then averaged. ~~In a typical RCP 8.5 scenario, there are sometimes a few grid cells with local precipitation changes that are out of sample. We convert these out of sample points to the extreme of our sample so that we avoid extrapolation (eg a 74% local increase in precipitation gets the response of 50% increase in precipitation - the maximum response to increased precipitation). Note that many of these large percentage changes in precipitation are actually the symptoms of ESM biases or small precipitation changes in arid regions that are unlikely to have agriculture. Holding to 50% precipitation change likely improves the fidelity of these estimates (Ruane et al., 2014).~~

The C3MP design resulted in a wider range of crops than had been previously sampled in a coordinated agricultural modeling study. We separate the C3MP data into 25 different production groups for ~~this analysis~~training in the Persephone framework to create V1.0 response functions. Twenty-four of the 25 groups for this paper are collections of sites corresponding to different crop-irrigation-latitude combinations: irrigated and rainfed versions of six key crops (~~Maize, Rice, Wheat, Soybeans~~maize, rice, wheat, soybeans, a C3-photosynthesis average, and a C4-photosynthesis average), based on sites at the extended tropics (30°S to 30°N) and the mid-latitudes (30- 70°S, 30- 70°N) ~~. The choice of breaking up groups by latitude zone was a rough way to account for baseline local temperature (which is important *in addition* to the change from local temperature)without having to eliminate the many valid C3MP sites that could not report local weather data due to data gaps or local government restrictions~~(see Section 2.1.1 for more details on spatial scales). It is also noteworthy that the majority of C3MP sites had high rates of fertilizer application, even in the extended tropics. These six crop groups were chosen because most IAMs already have experience incorporating such impacts from previous AgMIP exercises (e.g., Ruane et al. (2017); Calvin and Fisher-Vanden (2017); Nelson et al. (2014); Wiebe et al. (2015); Ruane et al. (2018)), they cover the major agricultural commodities globally, and they offer additional benchmarks for evaluating emulator success. In particular, the C3-photosynthesis production groups represent an average response of a very wide range of C3 crops, including ~~Wheat, Rice, and Soybeans~~wheat, rice, and soybeans. The C4-photosynthesis average is similarly defined, with sugarcane considered separately. The 25th production group is rainfed sugarcane in the extended tropics: no sugarcane sites outside of 30°S to 30°N were submitted to C3MP and only one irrigated sugarcane site was submitted.

We cull the 1135 contributed C3MP output datasets according to a range of criteria:

1. Sites simulated with notably older versions of crop models are eliminated. We thus eliminated uses of the DSSAT crop model v3 (and prior), given that important updates in crop physiology were added in version 4 (Jones et al., 2003).

2. Site simulations that exclude CO_2 fertilization responses, a fundamental variable examined here, were eliminated. We thus eliminated the SarraH-Hv32 crop model (primarily millet and sorghum sites in West Africa).

3. When C3MP modelers provided simulation sets that were identical other than the use of local weather data or AgMERRA climate forcing data (Ruane et al., 2015)), we used only the local dataset to avoid double counting. AgMERRA was provided for all datasets given frequent data gaps and governmental restrictions (Ruane et al., 2014).

These steps together eliminate more than 550 of the C3MP sites. Finally, for each production group, outliers are statistically identified and eliminated (Davies and Gather, 1993; Bond-Lamberty et al., 2014), in addition to those previously identified by the C3MP steering team. A total of 575 unique sites remain after culling, maps of which are included in Figure 2. These remaining sites cover 43 countries, 85 models, and 17 crop species. More than half of the C3MP sites have been eliminated, but this still results in a larger number of diverse sites, models, and crop species performing coordinated sensitivity tests than in any previous study (Asseng et al., 2013; Pirttioja et al., 2015; Fronzek et al., 2018). Since C3MP, the AgMIP-Wheat team has conducted an extensive analysis of temperature response at 30 wheat sites with 30 models (Asseng et al., 2015), but this only captures one of the CTW dimensions. ~~While~~

2.1.1 Known caveats of the C3MP data set

Additional discussion of the C3MP data set in the context of other AgMIP modeling efforts is presented in Ruane et al. (2017). One relevant point to this work is that, while C3MP spatial coverage is not ~~uniform~~ spatially uniform or production-weighted for any of the crops under consideration, sites for many of the major production regions are represented for each crop ~~.~~ (Figure 2). A major advantage of using site-specific crop models run voluntarily by experts is that the individual baseline runs at each site have been configured against local information in the historical period. However, the application of crop yield response from these sites to estimate response in any given grid cell with temperature and precipitation data is imperfect by its methodological nature. Yet this extension is necessary for use with GCAM: gridded yield changes for a subset of crops must be aggregated and converted to yield impact multipliers for each GCAM commodity in each land unit, defined as water basins in GCAM (Calvin et al., 2019).

Given the size and details of the C3MP data set, production groups were formed based on two latitude zones as a way to account for baseline local temperature (which is important in addition to the change from local temperature) without having to eliminate the many valid C3MP sites that could not report local weather data due to data gaps or local government restrictions. As this breakdown already results in some production groups with small sample sizes (see Table 1 and Section 3.1.1), further spatial disaggregation of production group is unjustified in this data set. While this means there will be limited spatial granularity in yield response *functions*, there can still be appreciable spatial granularity in yield *changes* due to variation in the gridded fields of temperature and precipitation changes. Future data sets with more comprehensive spatial coverage than the C3MP data may be used rather to create V2.0 response functions.

The site-specific percent change in yield from the 1980-2009 baseline yield is the dependent variable used to train our emulator (~~next section). While the output yields reported to the C3MP archive~~ see Section 2.2). Baseline yields differ widely across ~~sites for any given CTW combination,~~ the C3MP archive due to regional and system differences, however the percent change in yield from baseline is more consistent across sites for each CTW. Further, by training on change in yield rather than yield, we are able to introduce additional, scientifically grounded constraints to the functional forms we fit (Equations (4) - (6)). However, no baseline simulation was requested under the C3MP protocols. Therefore, for each individual set of output yields corresponding to each of the 575 simulation sites, we ~~estimate baseline yield so that we may calculate change in yield for training the emulator. For each simulation site, we~~ perform ordinary least squares ~~estimation for 8~~ regression for eight different functional forms relating the site-specific output yield to the input CTW values ~~. The form with the smallest root mean square error across the 99 tests for the site is the one used to provide a best estimate of baseline yield . This best estimate of baseline yield is used to convert~~ and select the best performing regression to estimate baseline yield (details in Appendix B, Equations (B1)-(B8)).

It is also worth noting that the C3MP ~~output yields at the site to percent changes in yield from baseline for emulator training~~experimental protocols (Ruane et al., 2014; McDermid et al., 2015) do not account for changing growing seasons, either through changes of within season distribution of temperature and rainfall or in the possible autonomous adaptation of farmers to shift planting and harvest dates. Ruane et al. (2014) showed that within season distribution changes had a small effect and the possible shift in planting and harvest dates are a topic of adaptation. Modeling autonomous adaptation behaviors is a challenging area for coordinated agricultural efforts and is only beginning to be addressed in coordinated sensitivity intercomparison studies as a scenario option, with no publicly available data sets at this time.

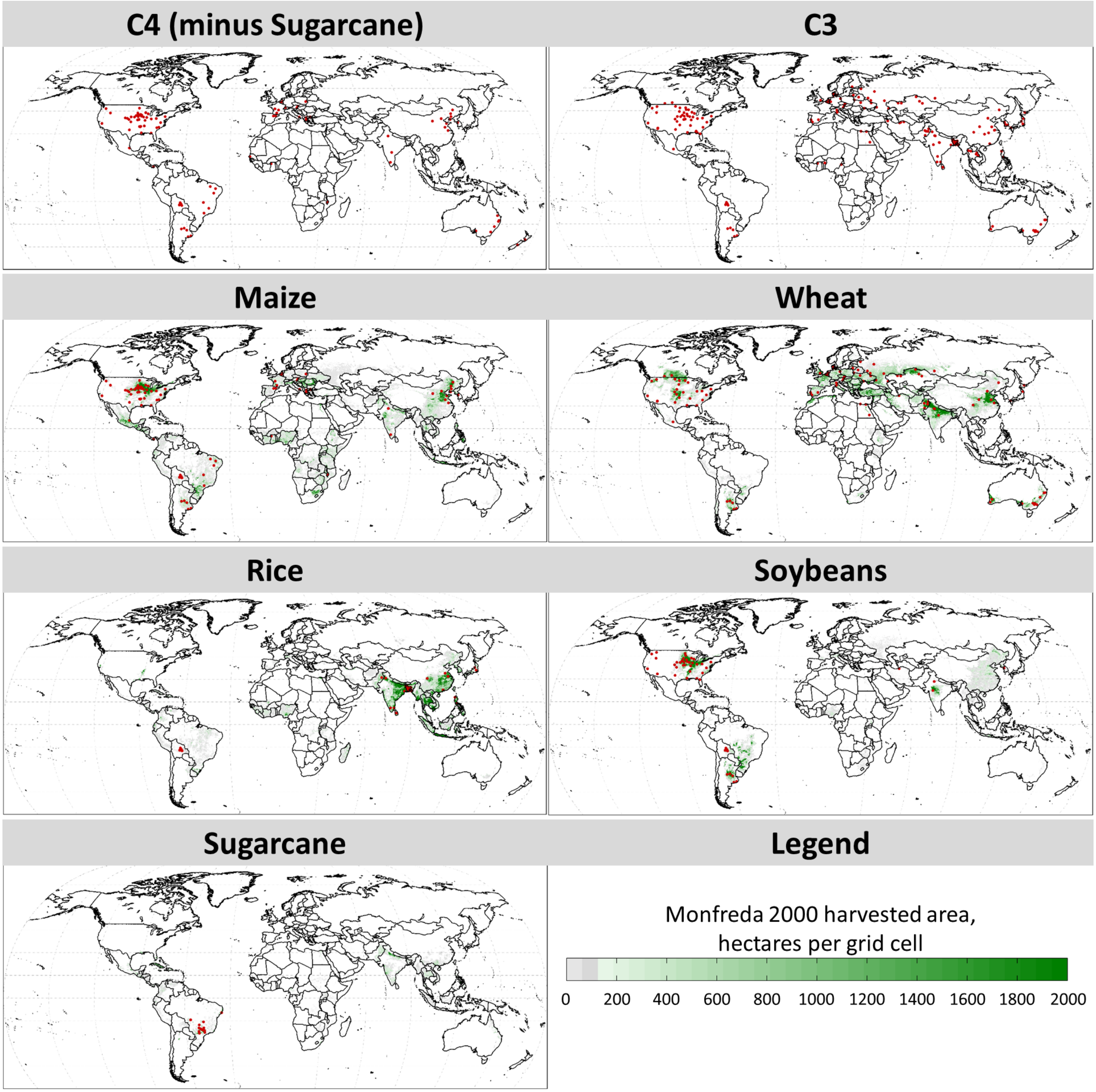


Figure 2. Maps of the C3MP data set culled sites. Each site represents a site-specific model of a single crop, with differing management practices. The sites are overlaid on Monfreda et al. (2008) harvested area data, except for the C3 and C4 averages.

2.2 Emulation

The majority of past agricultural yield emulator work has used ordinary least squares regression to estimate coefficients of functional forms. Given a set of predictors, $\mathbf{x}$, and given a particular value of the predictors $\mathbf{x}_i$ with corresponding training data y_i, an emulator would be some linear-in-parameters function $f(\mathbf{x})$ that returns an emulated value $f(\mathbf{x}_i)$ for comparison with y_i. Ordinary least squares regression requires that residuals $r_i = y_i - f(\mathbf{x}_i) \sim N(0,\sigma^2)$ for all i (e.g., Williams and Rasmussen, 2006, Section 2.1.1). A key requirement is that σ is a constant value across all i.

Figure 3 displays the spread of yield responses across sites for each CTW test for one production group, rainfed soybeans between 30- 70°S, 30- 70°N (the mid-latitudes). A successful emulator will produce the mean response (Figure 3, black dots) across sites for ~~eah~~ each CTW. Therefore examining the spread of the individual site yield changes about the mean yield gives some sense of the behavior of residuals in the most successful emulation case.The spread of yield change across sites relative to the mean response is different for each CTW test and appears to change in a systematic way - larger magnitude changes in yield are correlated with greater spread across sites. In light of this, a classic, ordinary least squares regression is not an appropriate approach for this emulator. We also desire more than just the mean response: we desire a measure of how this variation of site responses changes with CTW. With these considerations in mind, we take a slightly different approach to creating the Persephone ~~response functions~~V1.0 response functions, working from texts such as Gelman et al. (2013); Sivia and Skilling (2006); McElre
.

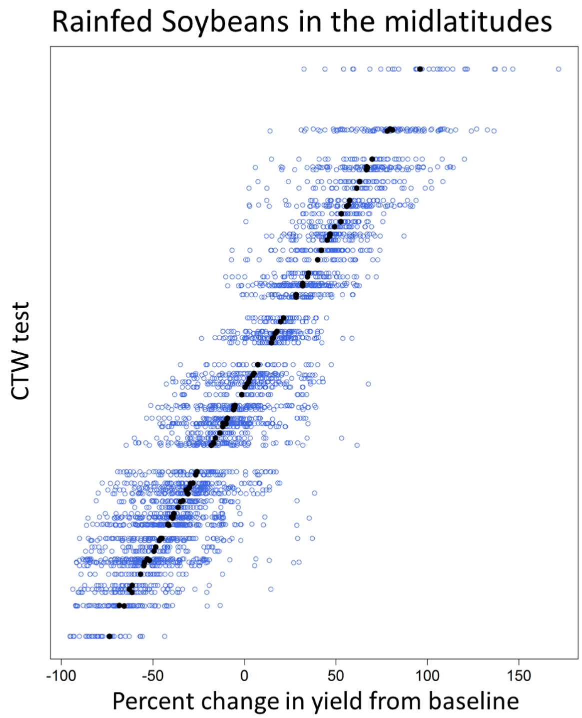


Figure 3. A plot of the percent yield change at each ~~Rainfed Soybeans~~ rainfed soybeans in the mid-latitudes site (blue points) for each CTW test (each horizontal line of points is a different test). The black dot for each test represents the mean response across the sites for that test.

We create the Persephone V1.0 response functions to emulate the mean yield response and two additional yield response scenarios spanning a range of individual site responses~~for use in GCAM~~. For a given production group (crop - irrigation - latitude zone combination), we collect the data for the 99 CTW tests for each of K C3MP simulation sets drawn from the culled-down archive. In other words, for each of 99 CTW combinations, there exist K 30-year average yield percent changes from the baseline (no changes in CTW) for a group. This ensemble of $99K$ yield changes is used to calculate the posterior densities for every parameter of μ_{CTW} and σ_{CTW} in the model defined by Equations (1) - (7) according to Bayes' theorem ($posterior \propto likelihood \times prior$). From the posteriors, the *maximum a posteriori* (MAP) estimates of parameters, the most plausible value for each parameter given both the model being used and the training data, is returned.

We define our likelihood as a normal distribution with mean μ_{CTW} and variance σ^2_{CTW}:

$$\Delta Y^{emulated}_{CTW} \sim N(\mu_{CTW}, \Sigma\sigma^2{}_{CTW}) \tag{1}$$

For a production group with site-specific yield responses that are normally distributed for each CTW value, μ_{CTW} is the mean response across sites for that CTW value (the black points in Figure 3), and ~~Σ_{CTW}~~ σ^2_{CTW} is a measure of agreement (or disagreement) of responses across sites for that CTW value. We present results for our most broadly optimal ~~μ_{CTW} and Σ_{CTW}~~ mean and variance functional form combination in this paper, and present the details of our selection criteria among the different functional forms in the ~~AppendixI~~Appendix.

To have unitless coefficients in our emulator, all predictor variables are standardized. Defining the collection of 99 T changes sampled by C3MP as T_{C3MP}, the collection of precipitation changes as W_{C3MP}, and the collection of CO_2 concentrations as C_{C3MP}, we have:

$$
\begin{aligned}
\Delta T &= \frac{T - T_{baseline}}{sd(T_{C3MP})} \\
\Delta W &= \frac{W - W_{baseline}}{sd(W_{C3MP})} \\
\Delta C &= \frac{C - C_{baseline}}{sd(C_{C3MP})}
\end{aligned}
\tag{2}
$$

$T_{baseline}$ is a change of 0° C from baseline, $W_{baseline}$ is a 0% change in precipitation from baseline, and $C_{baseline}$ is 360ppm. Plugging these baseline values into Equation (2) returns $\Delta T_{baseline} = \Delta W_{baseline} = \Delta C_{baseline} = 0$, as one would expect.

We exploit the fact that we are emulating change in yield (and not yield) and the fact that $\Delta T_{baseline} = \Delta W_{baseline} = \Delta C_{baseline} = 0$ in constructing Equations (4)-(7), which relate the mean and standard deviation of the likelihood in Equation (1) to our unitless predictor values $\Delta C, \Delta T, \Delta W$. By definition, percentage change in yield in response to no change in CTW is 0% at baseline for *every* individual C3MP site. This implies that ~~$\mu_{baseline} = \Sigma_{baseline} = 0$~~ both mean and variance at baseline are 0 for all production groups, and we must construct the Persephone response ~~funtions~~ functions to reflect this, independent of the estimated baseline yield at each site~~.~~ :

$$
\begin{aligned}
\mu_{baseline} &= 0 \\
\sigma^2_{baseline} &= 0
\end{aligned}
\tag{3}
$$

~~This~~ Implementing this constraint for the mean is straightforward. Any functional form representation of μ_{CTW} that does not include a constant parameter a_0 ~~and so at baseline,~~ will force $\mu_{baseline} = 0\%$ yield change ~~, as desired.~~ precisely because $\Delta T_{baseline} = \Delta W_{baseline} = \Delta C_{baseline} = 0$.

~~$\Sigma_{CTW} = |\sigma_{CTW}|$ where~~

$$
\begin{aligned}
\mu_{CTW} = {} & a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\
& + a_{10}\Delta T\Delta W\Delta C + a_{11}(\Delta T)^2\Delta W + a_{12}(\Delta T)^2\Delta C + a_{13}\Delta T(\Delta W)^2 + a_{14}\Delta T(\Delta C)^2 + a_{15}(\Delta W)^2\Delta C \\
& + a_{16}\Delta W(\Delta C)^2 + a_{17}(\Delta T)^3 + a_{18}(\Delta W)^3 + a_{19}(\Delta C)^3
\end{aligned}
\tag{4}
$$

Constraining the variance to be 0 at baseline as in Equation (3) should be equally easy by simply not considering any functional form that includes a constant parameter. However, this approach leads to numerical stability issues when estimating parameters. Therefore, we estimate the variance using the following functional form:

$$
\sigma^2_{CTW} = \Big(b_0 + b_1\Delta T + b_2(\Delta T)^2 + b_3\Delta W + b_4(\Delta W)^2 + b_5\Delta C + b_6(\Delta C)^2 + b_7\Delta T\Delta W + b_8\Delta T\Delta C + b_9\Delta W\Delta C\Big)^2 \tag{5}
$$

This results in the following functional form representation for the standard deviation:

$$\begin{aligned}\sigma_{CTW} &= +\sqrt{\sigma^2_{CTW}} \\ &= |b_0 + b_1\Delta T + b_2(\Delta T)^2 + b_3\Delta W + b_4(\Delta W)^2 + b_5\Delta C + b_6(\Delta C)^2 + b_7\Delta T\Delta W + b_8\Delta T\Delta C + b_9\Delta W\Delta C|\end{aligned} \tag{6}$$

This functional form estimates parameters that may individually be negative but which together result in a non-negative standard deviation for any CTW value being considered. At baseline, this functional form representation has ~~$\sigma_{baseline} = b_0$~~ standard deviation $\sigma_{baseline} = |b_0|$ as opposed to the required $\sigma_{baseline} = 0$ in Equation (3). This is done for numerical reasons and is ~~addresed~~ addressed with the prior for ~~$b_0 \sim N(0, 0.01)$~~ $b_0 \sim N(0, 0.01^2)$. This constrains the value of b_0 to be between -0.02% and 0.02% with 95.45% probability, reflecting that b_0 should be as close to 0 as possible without causing numerical solver issues. ~~We consider it acceptable even if a scenario results in $\Delta Y^{emulated}_{baseline} = 0.02\%$ because such a ΔY will be incorporated~~ This results in $\sigma_{baseline}$ values between 0% and 0.02% and therefore $0\% \leq \Delta Y^{emulated}_{baseline} \leq 0.02\%$, which we judge acceptable for incorporating into GCAM as a ~~multipler~~multiplier. All other parameters have very broad priors:

$$\begin{aligned} &b_0 \sim N(0, 0.01^2) \\ &a_i, b_i \sim Uniform(-300, 300)\ \forall a_i, b_i, i \neq 0 \end{aligned} \tag{7}$$

The functional form for μ_{CTW} is equivalent to estimating the coefficients of a third order Taylor polynomial, which can approximate a wide variety of functions fairly well. Similarly, the functional form for σ_{CTW} is ~~equivalent~~ conceptually related to estimating the coefficients of a second order Taylor polynomial. Because of the C3MP ~~experiemental~~ experimental design, emulating yield changes throughout the 21st century using Equations (1)-(7) does not require extending beyond the range of mean growing season CTW values used to train the Persephone V1.0 response functions. These functional forms are an evolution from C3MP's hybrid polynomial (Ruane et al., 2014). An exploration of other functional forms to address potential overfitting is included in Appendix A. Ruane et al. (2014) also reviews previous emulator forms across the literature, including discussion of the potential to look at non-linear terms such as killing degree days used in Schlenker and Roberts (2009), for example.

From the model defined by Equations (1)-(7), we construct the three Persephone v1.0 response functions for each production group~~, for use in GCAM and similar models~~:

$$\begin{aligned} &\text{Mean response: } \Delta Y^{emulated}_{CTW} = \mu_{CTW};\ \Delta Y^{emulated}_{baseline} = \mu_{baseline} = 0\% \\ &\text{High response: } \Delta Y^{emulated}_{CTW} = \mu_{CTW} + |\sigma_{CTW}|;\ \Delta Y^{emulated}_{baseline} \in (-0.02\%, 0.02\%) \text{ with } 95.45\% \text{probability} \\ &\text{Low response: } \Delta Y^{emulated}_{CTW} = \mu_{CTW} - |\sigma_{CTW}|\ \Delta Y^{emulated}_{baseline} \in (-0.02\%, 0.02\%) \text{ with } 95.45\% \text{probability} \end{aligned} \tag{8}$$

The default high and low responses are at one standard deviation of the production group yield responses (as opposed to two or three) because we are interested in scenarios that capture a range of the simulated site responses, but not the most extreme simulated site response. This does not affect how μ and σ are fit in Persephone v1.0, only how they are used. The Persephone v1.0 code is written flexibly enough that a user more interested in capturing the most extreme simulated site response could certainly add a multiplicative factor (e.g. $\mu + 2|\sigma|$) when using μ and σ without having to spend the computational time refitting.

3 Evaluation

We primarily present figures and analysis using the model and response functions defined by Equations (1)-(8) because we found these functional forms to be the most broadly optimal of those considered. ~~We~~ To investigate overfitting, we also examined nine other possible functional form combinations of μ_{CTW} and σ_{CTW} for each production group, defined in Equations (A1)-(A7). Details of the cross-validation experiments used as a method of functional form selection are in the Appendix. Briefly, because we are interested in the ability of any given response function to accurately predict yield changes in response to CTW values *not* used for training, we perform leave-one (CTW test)-out cross-validation experiments for each production group. The best performing functional form at the cross-validation experiments is then the selected functional form. This can be done to find the most broadly optimal functional form (using the same functional form for all production groups, Figure A1) or to find the best functional form for each production group (if a user wishes to vary the functional form for each production group, Table A10). This choice does not introduce additional fitting, or computational time. It is changed only by the calls to each function in the Persephone R package by the user.

Here, we quantitatively evaluate the performance of the Persephone V1.0 response functions (Equation (8)) trained on the full span of CTW values that the 99 tests represent for each production group (Section 3.1). We also present heuristic evaluations of mean response function performance (Section 3.2).

Files with the point estimate, as well as the standard deviation of the posterior distribution, for each coefficient in μ and σ for all 10 functional form combinations for all production groups are available (archived at https://doi.org/10.5281/zenodo.1414423) and as part of the Persephone v1.0 R package (https://github.com/JGCRI/persephone).

3.1 Quantitative

We categorize the performance of the Persephone V1.0 response functions trained on the full span of CTW values (mean, high, and low response, Equation (8)) for each production group based on comparing the 99 emulated yields output from the response functions to the 99 corresponding values from the C3MP simulation data: the in sample measurement of error. These are the actual response functions an end user would have and it is important to have a performance measure for them, although this is not the performance measure used to select functional forms.

The categorization is based on the normalized root mean square error (NRMS) and the comparison for each response function is as follows:

- The 99 emulated yields returned by the mean response function are compared to the mean yield response across the production group C3MP sites for each of the 99 ~~senstivity~~ sensitivity tests (what we call the ~~simualted~~ simulated mean yields).

- The 99 emulated yields returned by the high response function are compared to the 84.135^{th} percentile of yield responses across C3MP sites for each of the 99 ~~senstivity~~ sensitivity tests (what we call the simulated high yields). This corresponds to matching C3MP site responses at the mean plus one standard deviation level for each of the 99 sensitivity tests when the production group C3MP site responses were normally distributed for each sensitivity test.

– The 99 emulated yields returned by the low response function are compared to the 15.865th percentile of yield responses across C3MP sites for each of the 99 senstivity tests (what we call the simulated low yields). This corresponds to matching C3MP site responses at the mean minus one standard deviation level for each of the 99 sensitivity tests when the production group C3MP site responses were normally distributed for each sensitivity test.

As noted in Willmott (1984); Legates and McCabe (1999); Snyder et al. (2017), NRMS < 1 is one benchmark for adequate model performance, NRMS< 0.5 is a benchmark for good model performance, and NRMS = RMSE = 0 is perfect model performance. We further subdivide these categories and define excellent in-sample performance as NRMS≤ 0.25 for all three response functions; good performance to be $0.25 < NRMS \leq 0.5$ for at least one response function and NRMS≤ 0.25 for at least one response function; adequate performance to be all three response functions having $NRMS < 1$ but at least one response function with $0.5 < NRMS < 1$; and finally poor performance occurs when any one of the three response functions has $NRMS \geq 1$.

The mean response function performs excellently for all of our production groups~~. Non-excellent in-sample performance is driven by~~ , although the performance of the high and low response functions differs. These measures are presented in Table 1 for the response functions defined using cubic μ_{CTW} (Equation (4)) and quadratic σ_{CTW} (Equation (6)) for all production groups. The excellent performance of the mean response function holds across all functional form combinations explored (Table A1-A9). In the event that a user is only concerned with a mean response scenario, a shared functional form for all production groups is acceptable. A user interested in the high and low response functions may wish to use the production group specific functional form combinations listed in ~~Tabel~~ Table A10, which includes the in-sample performance metric for the optimal functional form for each production group. The majority of production groups (17/25) feature excellent in-sample performance while the remaining 8 production groups feature good overall performance. For more detail than the summary tables presented here, files of results for the leave-one-out cross validation exercises for all functional form combinations for all production groups are available in the paper analysis archive.

We also present a dashboard of quantitative evaluation plots for four of our 25 production groups in Figures 5 and 4 to provide a visual interpretation of the four in-sample performance categories. Each dashboard is organized to address the following questions:

– Top Left: For a given group, do the three representative responses span the range of sites? In this plot, individual site yield changes for each test (blue dots), are overlaid with the emulated mean, high, and low response functions evaluated for each test (black dots). Each horizontal line of points represents one of the 99 CTW sensitivity tests.

– Top Right: For a given group, how does the emulated mean for each of the 99 tests compare to the simulated mean for each test?

– Bottom Left: For a given group, how does the emulated high response for each of the 99 tests compare to the simulated high yield for each test?

Table 1. Persephone v1.0 response function performance for all production groups, for cubic μ_{CTW} (Equation (4)), quadratic σ_{CTW} (Equation (6))

Production group[1]	Num. C3MP sites	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c4 IRR mid	47	0.010	0.148	0.112	Excellent
Maize IRR mid	45	0.010	0.164	0.116	Excellent
Rice RFD mid	4	0.044	0.150	0.195	Excellent
Rice RFD tropic	41	0.020	0.199	0.146	Excellent
Soybeans IRR mid	32	0.017	0.230	0.176	Excellent
Soybeans IRR tropic	2	0.039	0.150	0.170	Excellent
Soybeans RFD mid	35	0.016	0.151	0.145	Excellent
Soybeans RFD tropic	9	0.043	0.198	0.160	Excellent
c3 RFD mid	165	0.010	0.316	0.270	Good
c4 RFD mid	74	0.016	0.319	0.241	Good
c4 RFD tropic	25	0.019	0.365	0.177	Good
Maize IRR tropic	7	0.012	0.345	0.118	Good
Maize RFD mid	66	0.018	0.293	0.230	Good
Maize RFD tropic	20	0.022	0.407	0.170	Good
Rice IRR tropic	53	0.088	0.339	0.261	Good
Wheat IRR mid	61	0.024	0.372	0.380	Good
Wheat IRR tropic	8	0.076	0.382	0.329	Good
Wheat RFD mid	103	0.021	0.302	0.280	Good
Wheat RFD tropic	4	0.093	0.364	0.311	Good
c3 RFD tropic	63	0.024	0.757	0.546	Adequate
c4 IRR tropic	14	0.012	0.998	0.214	Adequate
Rice IRR mid	6	0.029	0.656	0.427	Adequate
c3 IRR mid	103	0.012	1.038	0.701	Poor
c3 IRR tropic	67	0.072	1.662	0.790	Poor
Sugarcane RFD tropic	12	0.047	1.382	1.162	Poor

1. “IRR" = irrigated, “RFD" = rainfed, “mid" = mid-latitudes (30- 70°S, 30- 70°N), “tropic" = 30°S to 30°N. 2. Note that the mean response function performs “excellent" for all production groups.

– Bottom Right: For a given group, how does the emulated low response for each of the 99 tests compare to the simulated low yield for each test?

Figure 4 displays one performance dashboard from each in-sample performance category for the broadly optimal, shared functional form cubic μ_{CTW} and quadratic σ_{CTW} (Equations (4)-(6)), to aid interpretation of Table 1 (and Tables A1-A9).

As indicated in Table A10, any production group can be fit to result in response functions with an in-sample performance of good or excellent, if a user is willing to vary the functional forms used for each production group. Figure 5, left, presents the dashboard for one of the production groups that featured poor performance when the common functional form cubic μ_{CTW} and quadratic σ_{CTW} (Equations (4)-(6)) was used for all production groups: rainfed sugarcane in the extended tropics. Figure 5, right, presents the dashboard when the response functions are based on the production group specific functional forms selected by cross-validation (Table A10): C3MP μ_{CTW} (Equation (A2)) and cubic σ_{CTW} (Equation (A7)). The high and low response functions perform better in the latter case, though it is at the cost of a slightly worse (but still excellent) mean response function performance. Examination of the sugarcane entry in Tables 1, A1-A9 indicates that a cubic description of σ_{CTW} (Equation (A7)) leads to better high and low response function performance than a quadratic representation (Equation (A6)), regardless of functional form used for μ_{CTW} (Equations (A1)-(A5)). In other words, the uncertainty across C3MP site responses for each CTW test requires a more detailed Taylor series approximation to describe. This is also generally the case for the other production groups that rated adequate or poor in-sample performance in Table 1: sometimes the C3MP individual site yield responses are distributed in such a way for each CTW test that a more flexible fit for σ_{CTW} is necessary. Perhaps unsurprisingly, this usually occurs for either very broad production groups (such as those based on C3-photosynthesis), or for production groups with very few C3MP site outputs (irrigated rice in the mid-latitudes) rather than due to a discernible biophysical trend or requirement.

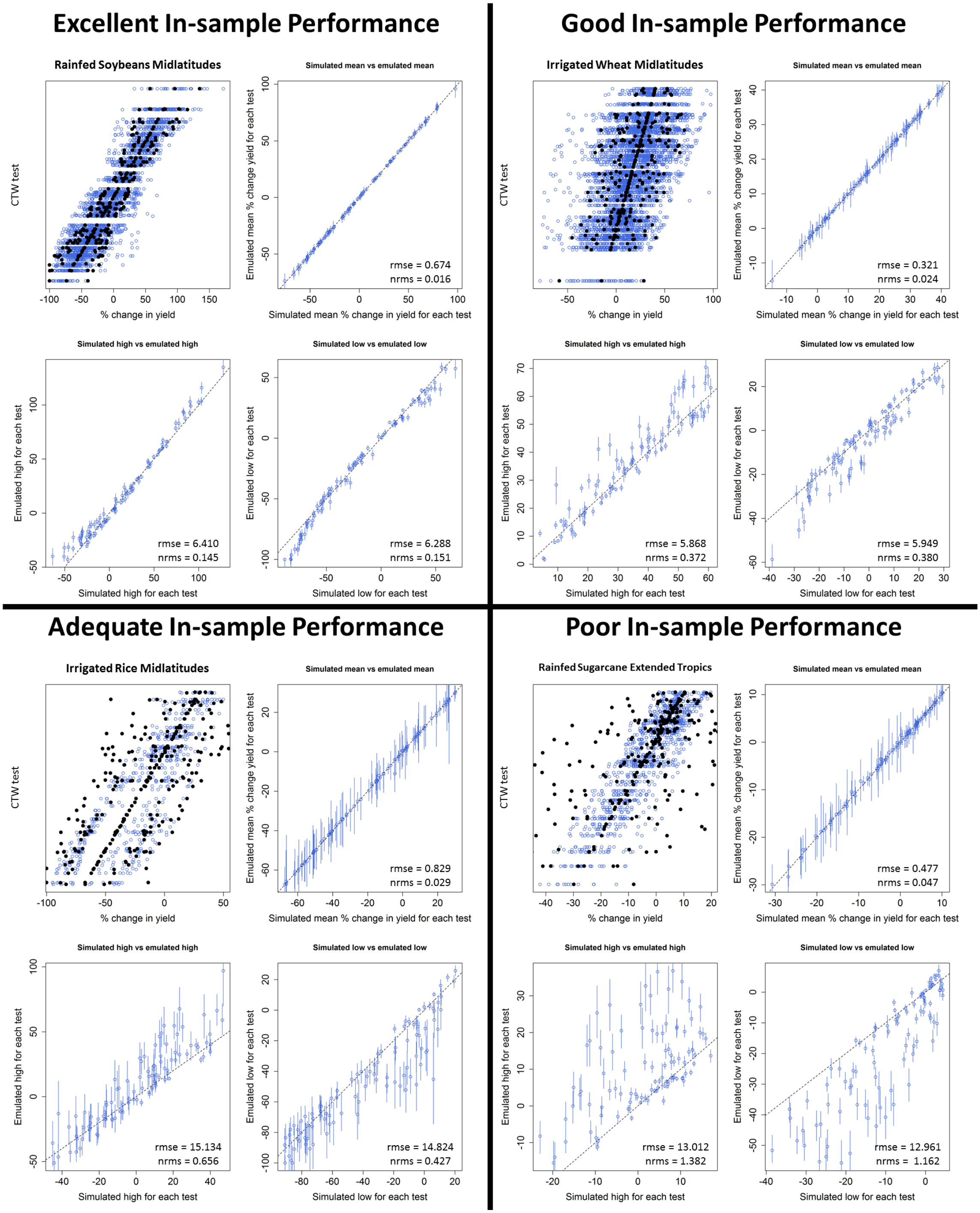


Figure 4. Top left: Rainfed ~~Soybeans~~ soybeans in the ~~Mid-latitudes~~mid-latitudes, an example of the excellent in-sample performance category. Top right: Irrigated ~~Wheat~~ wheat in the mid-latitudes, an example of the good in-sample performance category. Bottom left: Irrigated ~~Rice~~ rice in the mid-latitudes, an example of the adequate in-sample performance category. Bottom right: Rainfed ~~Sugarcane~~ sugarcane in the extended tropics, an example of the poor in-sample performance category (also seen in Figure 5, left). Vertical error bars indicate 95% credible interval for each of mean, high, low emulated responses.

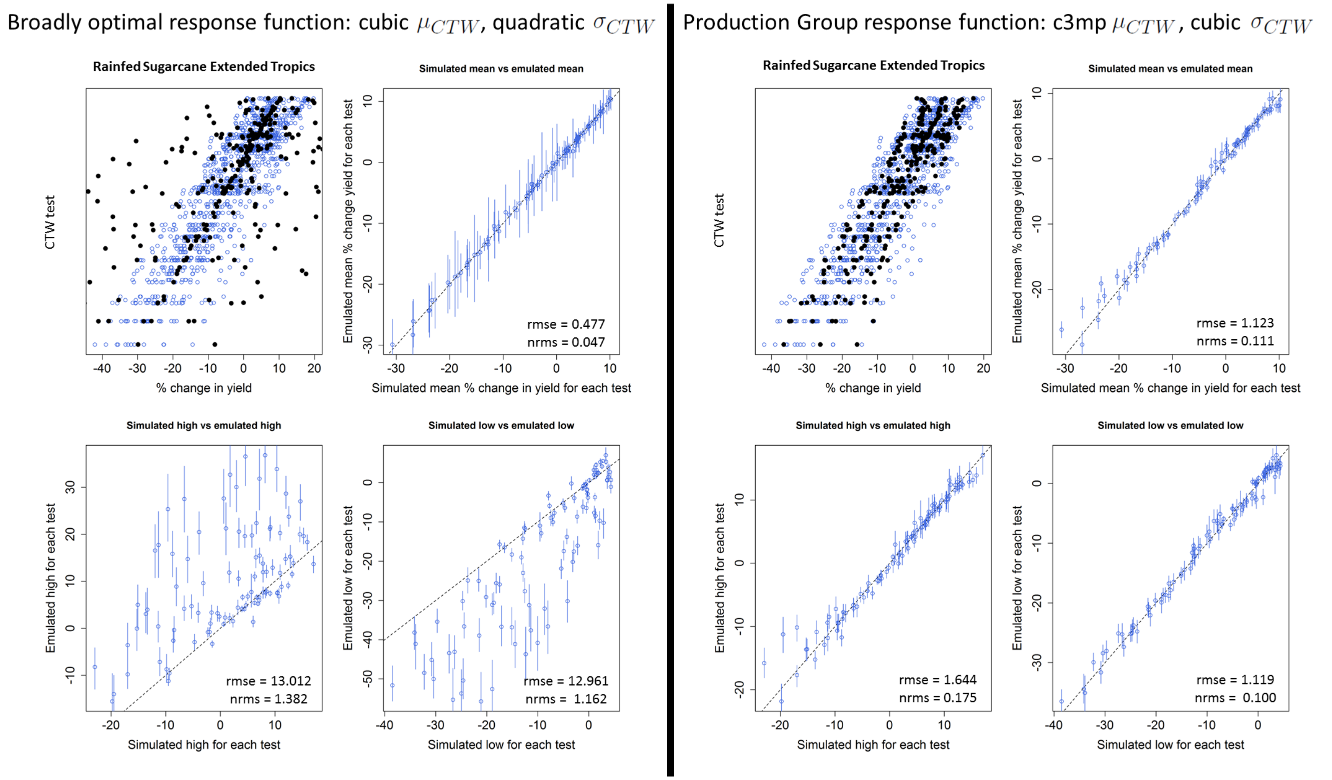


Figure 5. Rainfed ~~Sugarcane~~ sugarcane in the extended tropics. Left: The performance dashboard for the most broadly optimal functional form representations (i.e. if we want to use the same functional form combination for all production groups), and for which the high and low response functions poorly reproduce the simulated high and low yields for each of the 99 tests. Right: The performance dashboard for the production group specific functional forms (i.e. if we want the functional form to vary by production group). Vertical error bars indicate 95% credible interval for each of mean, high, low emulated responses.

3.1.1 Production groups with small sample size

3.2 ~~Heuristic~~

It is worth noting that 7 of the 25 production groups considered here are characterized by fewer than 10 C3MP sites (Table 1). For all of these groups, it is possible to fit high and low response functions that capture the spread of the group's C3MP site responses well (Figures 6 and 7). For many of these groups, the spread in response is relatively small. The Persephone framework does not fail, rather the data upon which the V1.0 response functions are trained is imperfect and would be improved by greater density in spatial sampling. Had the spatial disaggregation used in forming production groups resulted in small sample size groups with more significant spread in site response, the Persephone framework is unlikely to represent the full spread of the sample. As this is not the case, it is left to an eventual user to judge whether such responses serve their purpose.

Figure 6 highlights this fact for the production group with smallest sample size, irrigated soybeans in the Extended tropics. The spread of C3MP sites as well as the performance dashboards for the shared optimal functional form (as from Table 1) and for the group-specific optimal functional form (Table A10). While the shared optimal functional form (middle panel) overestimates the small spread between the two C3MP sites, the group-specific optimal functional form (right panel) captures the spread well.

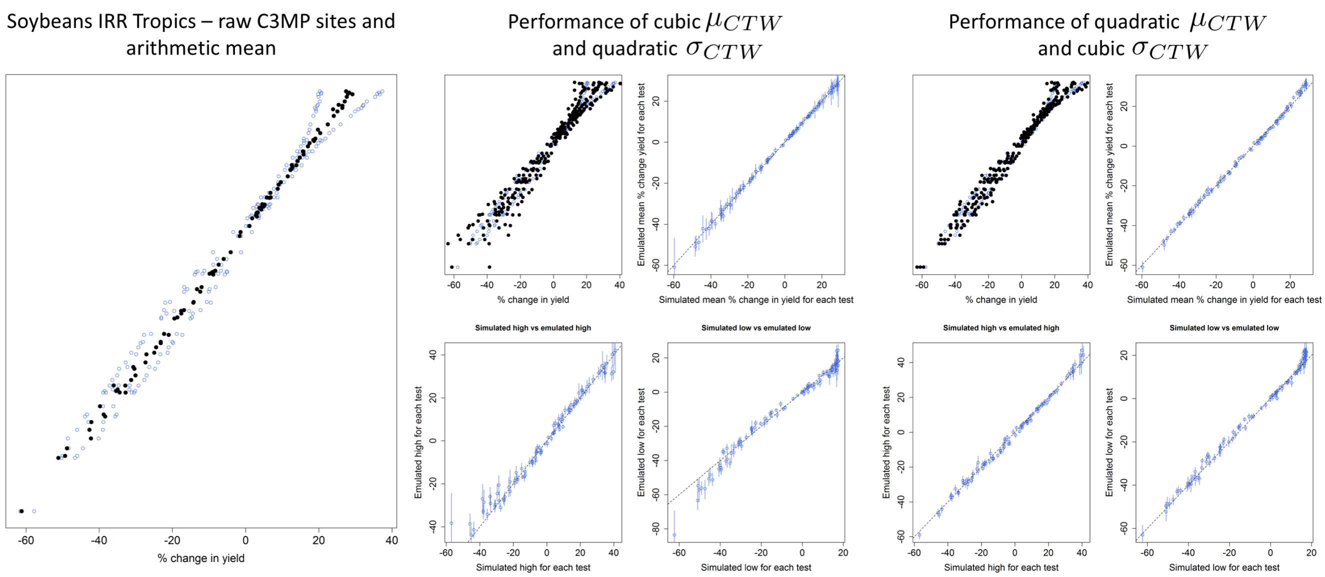


Figure 6. Left:The spread of yield responses for the two C3MP sites making forming the irrigated soybeans in the extended tropics production group. Middle: The performance dashboard of the shared optimal functional from (Table 1) for this production group. Right: The performance dashboard of the group-specific optimal functional form (Table A10) for this production group.

Figure 7 repeats this analysis for the next three smallest sample size groups: rainfed wheat in the extended tropics (Top), rainfed rice in the mid-latitudes (Middle), and irrigated ice in the mid-latitudes (Bottom). In all three cases the group-specific optimal functional form represents the spread of the data well. This is also the case for the two remaining production groups with fewer than 10 C3MP training sites: irrigated wheat in the extended tropics and rainfed soybeans in the extended tropics (not shown).

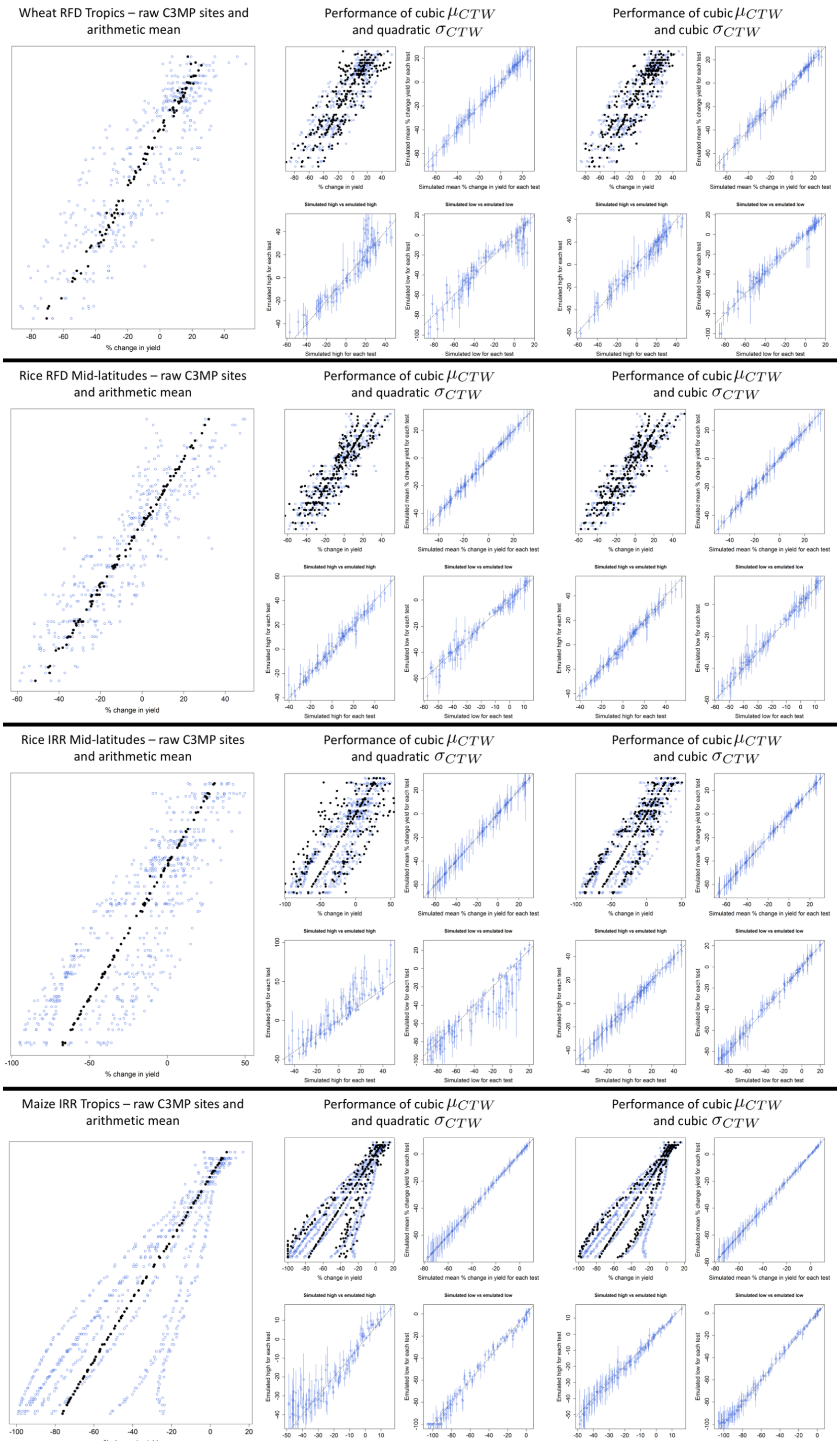


Figure 7. The same arrangement of figures as in Figure 6, for the rainfed wheat in the extended tropics (Top row), rainfed rice in the mid-latitudes (Second row), irrigated rice in the mid-latitudes (Third row), and irrigated maize in the extended tropics (bottom row) production groups.

3.2 Qualitative

One motivation for the 25 production groups based on [~~Corn, Wheat, Rice, Soybeans~~maize, wheat, rice, soybeans, C3, C4 (minus sugarcane), and sugarcane] X [~~irrgated~~ irrigated or rainfed] X [extended tropics or mid-latitudes] is to evaluate emulator performance beyond the quantitative. Given that some GCAM users will only be interested in the mean response functions, it is particularly important to validate that these functions capture key biological features of each crop, beyond the quantitative agreement for the 99 C3MP tests measured by the in-sample performance metric in Section 3.1. In particular, these are features motivated by biophysical intuition and present in most of the C3MP sites. Therefore we verify that these features are retained in the emulator.

We use impact response surfaces to visualize these features, examples of which are given in Figures 8 and 9. The three-dimensional CTW space is most easily examined by looking at cross sections where one of the CTW dimensions is kept constant while the other two vary. The brown to blue ~~colorbar~~ color bar in each of these figures depicts contours for the value of the mean yield response (μ_{CTW}) while the overlaid ~~grayscale~~ labeled black lines depict contours representing uncertainty (σ_{CTW}, used to create the high and low response functions).

We first identify three important relationships we would expect a successful emulation of C3MP mean responses (brown to blue ~~colorbar~~color bar) to obey:

- C3 crops respond strongly and positively to increases in global CO_2 concentrations; C4 crops have noticeably less benefit from CO_2 increases.

- Agriculture in the tropics tends to response more negatively/less positively to changes in temperature than agriculture in the higher latitudes as the extended tropics correspond to a higher baseline temperature.

- Irrigated crops have almost no response to changes in precipitation, whereas rainfed crops do.

These benchmarks are met: Figure 8 features impact response surfaces that highlight the C3-photosynthesis and C4-photosynthesis difference, the rainfed and irrigated difference, and the latitude difference. The full collection of impact response surfaces for all production groups are included in the paper analysis archive. These benchmarks for the mean response are met in those as well. When there are exceptions, we have investigated to find that the mean response function is faithfully representing the underlying C3MP data and that it is the sampling of C3MP sites making up the production group responsible for the discrepancy. Note that, in Figure 8, uncertainty is greatest in the CO_2-precipitation and CO_2-temperature slices, and increases with larger changes from the baseline condition. This follows with current practices for the process-based crop models forming the C3MP data set: CO_2 is clearly related to yields but the details of this relationship are highly uncertain and implemented differently across process-based, site specific crop models.

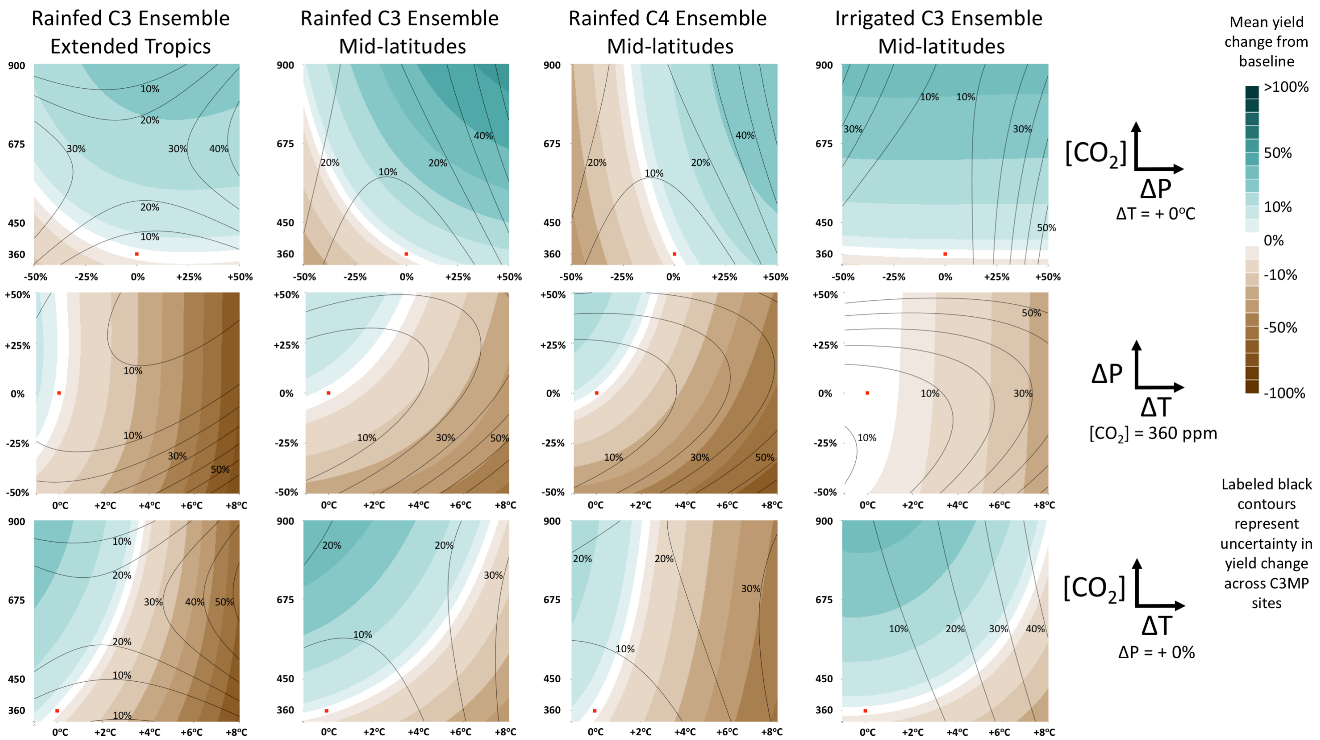


Figure 8. Select impact response surfaces - a collection of 2-parameter slices of our 3-parameter space (not a visualization of the full space). The color represents the yield change for a given local CTW perturbation as a % of baseline yields (1980-2009 planting year average, position shown as red square). ~~The grayscale lines~~ Labeled black contours are uncertainty across the submitted site specific crop models.

The *pattern* of yield response to CTW changes appears to be more qualitatively consistent across C3MP sites than the quantitative differences across sites (for example, Figure 3). Figure 9 displays this pattern for one cross-section of CTW space for 12 of 66 rainfed maize sites in the mid-latitudes, and for the emulated mean response. While the actual numerical values of the response surfaces differ at each site, the pattern of response seen at most sites (increasing yield with high CO_2 and low temperature changes in the upper left, decreasing yields elsewhere) is consistent and shared by the emulated mean response. The high and low response functions are able to capture much of the quantitative spread in site responses, though, as noted in Section 2.3, not the most extreme sites. We specifically included the sites at Ames, IA, Naousa, Greece, and Lublin Poland because they feature the most qualitatively different patterns. The pattern at the 54 sites not displayed closely resemble the other 9 sites in Figure 9. This pattern is seen in the broader impact response surfaces literature (Ruane et al., 2014; Pirttioja et al., 2015; Fronzek et al., 2018) as well, further improving confidence in the emulated mean response. All individual site impact response surfaces are included in the paper analysis archive.

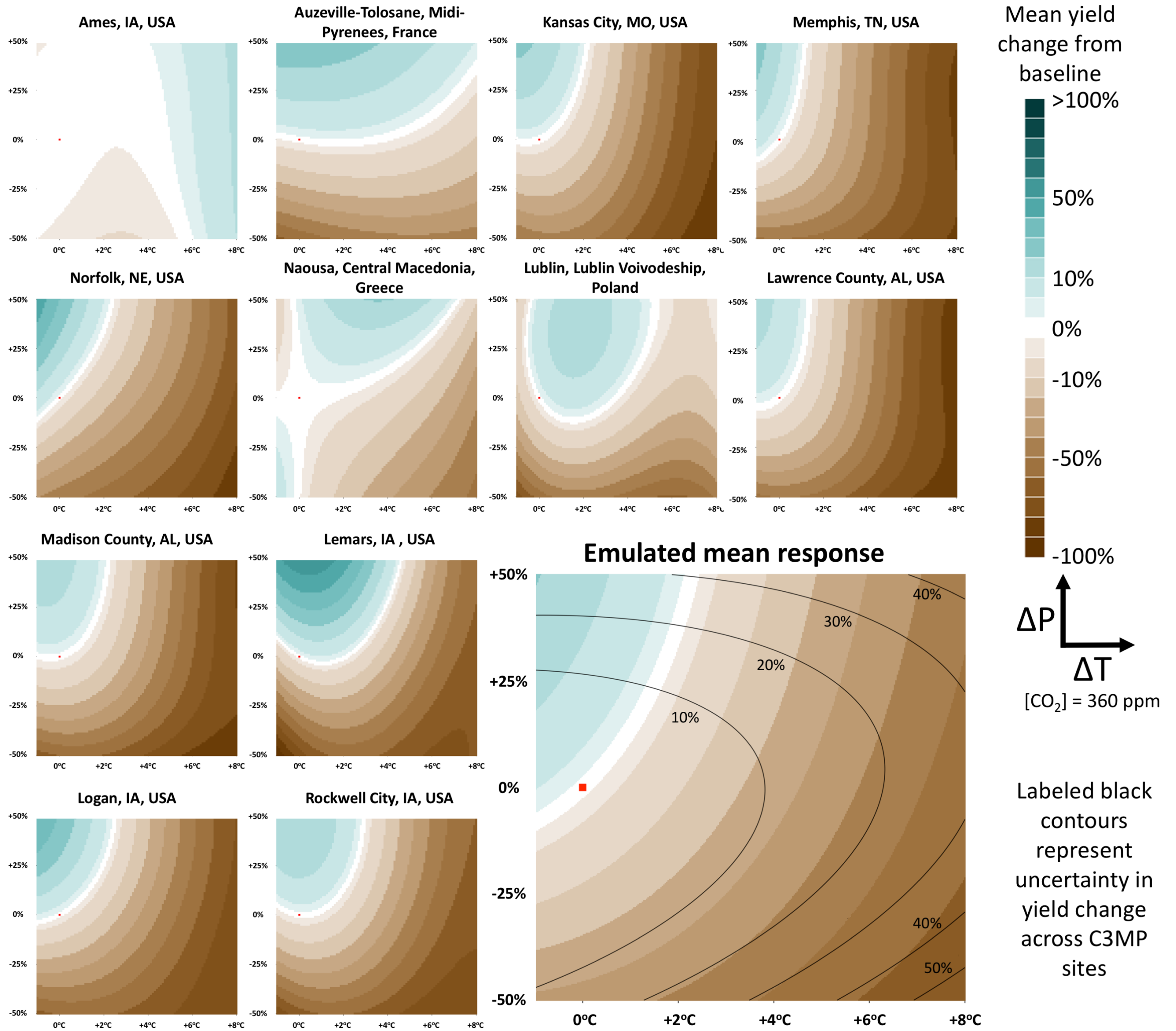


Figure 9. Yield responses to changes in temperature and precipitation with fixed $[CO_2]$ = 360 ppm for 12 (of 66 total) rainfed ~~Maize~~ maize sites located in the mid-latitudes, as well as the emulated mean response for use in GCAM.

4 Applications

Figure 10 demonstrates the basic procedure followed in using Persephone within GCAM (using the average of 2071-2100 HadGEM2-ES RCP 8.5 projections as an example). The first requirement is a global gridded file of local precipitation and local temperature drawn from climate projections, along with a global CO_2 concentration level. Temperature and precipitation changes ~~should be~~ are calculated only for the relevant local growing season months in comparison to a 1980-2009 baseline value. The different maps of local temperature and precipitation changes on the left side of Figure 10 reflect that there are differences in the dates of the local growing season for rainfed maize and wheat. Note that this includes a global CO_2 concentration of 812 ppm, compared to the baseline level of 360 ppm. The CO_2 change alone leads to increased yields for rainfed wheat mid-latitude even in the absence of changes in temperature and precipitation. Indeed, the higher CO_2 elevates yields (compared to the baseline) across all but the most extreme hot and dry conditions. Conversely, the yield response for rainfed tropical maize is barely helped by elevated CO_2.

In a typical RCP 8.5 scenario, there are sometimes a few grid cells with local precipitation changes that are out of sample. We convert these out of sample points to the extreme of our sample so that we avoid extrapolation (eg a 74% local increase in precipitation gets the response of 50% increase in precipitation - the maximum response to increased precipitation). Note that many of these large percentage changes in precipitation are actually the symptoms of ESM biases or small precipitation changes in arid regions that are unlikely to have agriculture. Holding to 50% precipitation change likely improves the fidelity of these estimates (Ruane et al., 2014).

The second step in using Persephone for GCAM is that CTW changes for each ~~agricultural region~~ grid cell with climate data are passed into the Persephone V1.0 response functions (depending on species/management/latitude zone) to create the desired global gridded map of yield changes that would represent the likely agricultural response. The abrupt change in behavior across 30°N and 30°S (particularly noticeable for wheat in Southern Asia) are due to our division of training data into mid-latitudes and extended tropics production groups. Those abrupt changes will soften as these impacts are aggregated to the larger GCAM land region level before being applied as multipliers in the experiments ~~detailed in Section 2.1.~~ outlined in Figure 1.

Figure 11 presents the rainfed maize impact response surfaces and yield change maps for the bias-corrected ISIMIP entry of HadGEM2-ES RCP 8.5 (Warszawski et al., 2014) 2071-2100 average CTW changes (displayed in Figure 10) for the low (left), mean (center) and high (right) response functions. The high and low response surfaces result from adding or subtracting the gray uncertainty contours to the brown-blue mean yield response contours in the mean response surfaces (Equation (8)). Note that under the high response function, there are a few regions that experience increased yields due to large increases in precipitation offsetting temperature increases. The differences in these three response functions will allow the boundaries of crop response uncertainty to be run through GCAM, resulting in a spread of socioeconomic and environmental impacts in response to a particular future climate.

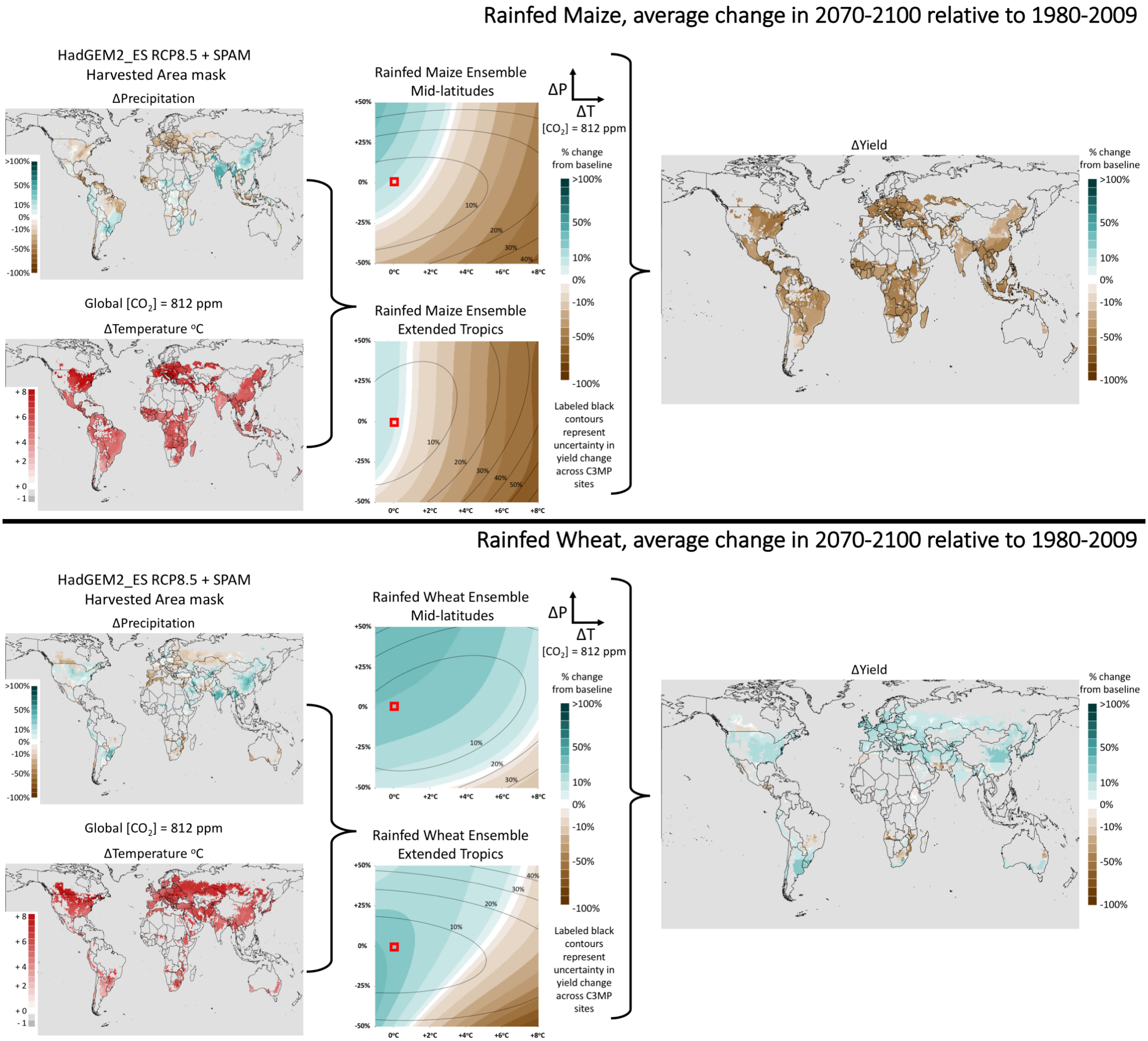


Figure 10. Tracing the path from gridded local growing season temperature and precipitation changes and global CO_2 = 812 ppm concentration under HadGEM2-ES RCP 8.5 for 2071-2100 compared to 1980-2009, through the relevant yield response functions (represented here as ~~Impact Response Surfaces~~impact response surfaces) to generate mean yield change maps for Rainfed ~~Maize~~ maize (top) and ~~Rainfed Wheat~~ rainfed wheat (bottom). The open red square is placed at no change in temperature and precipitation for each ~~Impact Response Surfaces~~impact response surface. For plotting clarity, we use a harvested area mask of grid cell harvested area > 10 hectares in the SPAM ~~2010~~ 2005 data set (You et al., 2014)

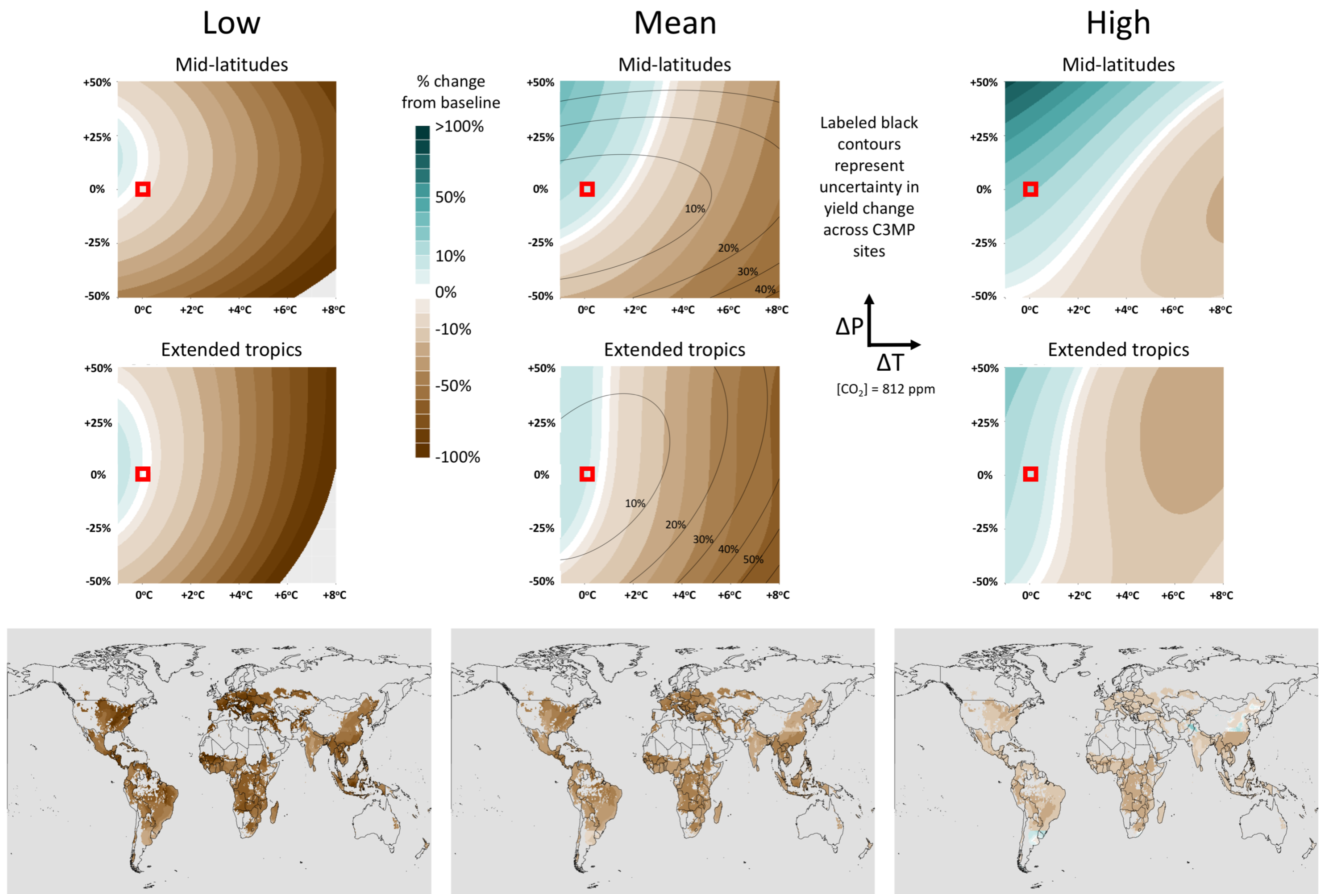


Figure 11. Low, mean, and high response surfaces for the mid-latitudes (top row) and extended tropics (middle row) for ~~Rainfed Maize~~rainfed maize, as well as the resulting maps of yield changes under the same HadGEM2-ES RCP 8.5 2071-2100 CTW changes as Figure 10.

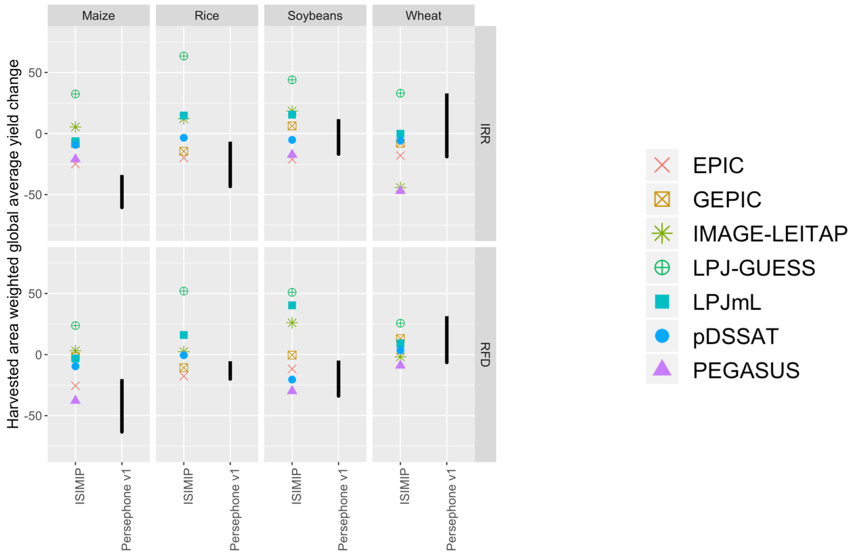
4.1 Comparison to other crop modeling results

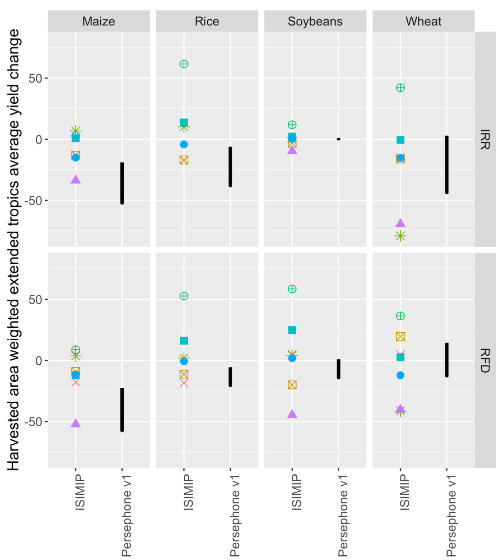
We further examine the Persephone V1.0 response functions driven by HadGEM2-ES RCP 8.5 CTW changes by comparing our results with previous AgMIP global gridded crop model (GGCM) yield change data released under ISIMIP (Rosenzweig et al., 2014; Warsz . In order to compare the best possible emulation of the C3MP data set to the range of AgMIP/ISIMIP GGCM results, the production group-specific optimal functional forms provided in Table A10 are used here. To have the most direct comparison possible, the ISIMIP GGCM yield time series were converted from actual yield values to percent changes from 1980-2009 baseline yields. It is important to note that the GGCMs were driven by historical climate data from 1980-2004. 2005-2009 yield data for each GGCM was driven by HadGEM2-ES RCP 8.5, given that this was considered a "future" simulation according to the GCM projections from 2005 forward. The results from the GGCMs which include model-specific [CO_2] effects were used. Both Persephone V1.0 and the ISIMIP GGCM yield change data are compared only on grid cells with harvested area > 10 hectares in the SPAM 2005 data set (You et al., 2014).

As the ISIMIP GGCMs did not directly participate in the C3MP exercise, no version of these GGCMs was used in the training data that produced the Persephone V1.0 response functions, and there is no *a priori* reason to expect the Persephone V1.0 range of yield changes to match the ISIMIP range. The site-specific simulations using various versions of DSSAT submitted to the C3MP exercise feature different configurations and model versions than the ISIMIP GGCM pDSSAT (a global gridded implementation of DSSAT). Given this fact, and that the C3MP archive includes results from non-DSSAT site-specific crop models, there is again no expectation of replicating pDSSAT results even though the fundamental crop responses are similar. Finally, it is also worth noting is that the 1980-2009 historical/RCP8.5 HadGEM2-ES simulation is not the same as the historical, site specific and AgMERRA data used by modelers submitting to C3MP. This combination of different responses and different baselines across C3MP and the ISIMIP GGCMs means there could be considerable differences in interannual variability and mean yields, which may be a reason that the Persephone V1.0 response functions may predict different yield changes from the ISIMIP range for some crops.

However, it is still worth evaluating our results against the GGCM data. Figure 12 compares the range of aggregated (via MIRCA2000 harvested area Portmann et al., 2010), time averaged 2071-2100 yield changes from Persephone V1.0 response functions to the range of ISIMIP yield changes at the global level (top), in the extended tropics latitude band (bottom left) and in the mid-latitudes band (bottom right) for both irrigated and rainfed maize, rice, soybeans, and wheat. For context, in the time since the AgMIP/ISIMIP results were published, the IMAGE-LEITAP model has been largely abandoned. Further, IMAGE-LEITAP, LPJ-GUESS, and LPJmL feature relatively unlimited nutrient constraints, resulting in frequent yield increases given an unconstrained CO2 response. For many of the production groups, the range of Persephone V1.0 yield changes lies at least partially within the ISIMIP range, suggesting that the response functions for those production groups result in yield changes consistent with ISIMIP. Those production groups that differ substantially from the ISIMIP yield range are due to underlying differences in the C3MP data set versus those produced from the ISIMIP GGCMs. That is, while the Persephone framework emulates the C3MP data well, response functions based on a different data set may behave more consistently with the ISIMIP GGCMs given differences between the model selection and local farm system configurations of the C3MP and GGCM ensembles.

Of the production groups with yield ranges much smaller than the range of ISIMIP yield changes, several (irrigated and rainfed soybeans in the extended tropics and irrigated and rainfed rice in the mid-latitudes) are small sample size groups (Table 1, Section 3.1.1). Future coordinated sensitivity studies of site-specific crop models would ideally include more participation in a broader range of regions, but this is a current limitation of the Persephone V1.0 response functions. This adds additional support to the call for a designed network of site-based crop models, intended to cover all regions and systems, to participate in coordinated sensitivity studies raised in Ruane et al. (2017).





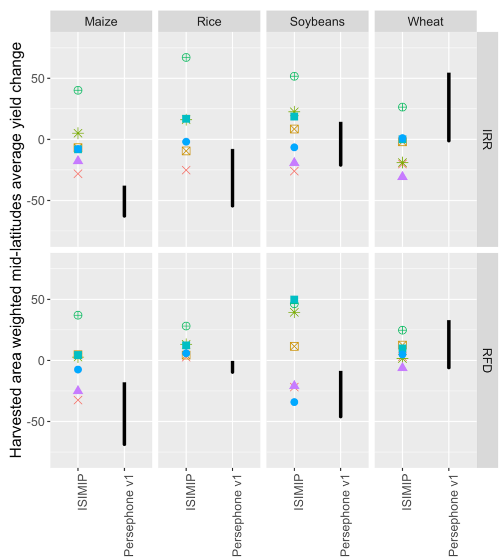


Figure 12. Aggregated (via MIRCA2000 harvested area (Portmann et al., 2010)), time averaged 2071-2100 yield changes for Persephone V1.0 response functions and the ISIMIP GGCM range of results for multiple production groups. Top: Comparison of global average yield changes. Bottom left: Comparison of average yield changes in the extended latitude band. Bottom right: Comparison of average yield changes in the mid-latitude band.

The Persephone V1.0 range of yield changes for irrigated maize also noticeably departs from the ISIMIP range of yield changes in the mid-latitudes (Figure 12, bottom right), which in turn drives the disagreement at the global level (Figure 12, top). This is not due to an error in emulation or due to small sample sizes (Table 1, Figure 7, bottom), but rather due to a fundamental disagreement in the predicted maize response among the site-specific crop models of C3MP and the ISIMIP

GGCMs. It is worth noting that yield changes predicted by Persephone V1.0 response functions are consistent with work examining maize site data. Namely, a 2014 site-specific model comparison study by the AgMIP-Maize team found irrigated and rainfed maize yield changes in response to a local temperature +6°C of similar values to the Persephone V1.0 range of responses (see Figure 3 of Bassu et al., 2014). The HadGEM2-ES RCP 8.5 local growing season temperature 2071-2100 change map for irrigated maize used to drive the Persephone V1.0 response functions is shown in Figure 13 and it is worth noting that many major producers of maize see temperature increases of at least +6°C. Further, recent analysis of FACE experiments and crop model results suggest that maize primarily benefits from high [CO_2] during drought, indicating that models of the effects of [CO_2] fertilization on irrigated maize (and rainfed maize during non-drought periods) may be overly beneficial (Durand et al., 2018). This suggests that the more pessimistic irrigated maize yield changes predicted by the C3MP sites and therefore Persephone V1.0 are more consistent with site-specific crop models and FACE experiments than they are with the ISIMIP GGCM range of results. While it would be ideal to have GGCM results from more GGCMs and more recent model versions for comparison, such results are not yet public. This discrepancy between the results of site-specific crop models and FACE experiments versus GGGMs supports the call in Leakey et al. (2012) for further investigation to understand regional and system-specific variation in [CO_2] response.

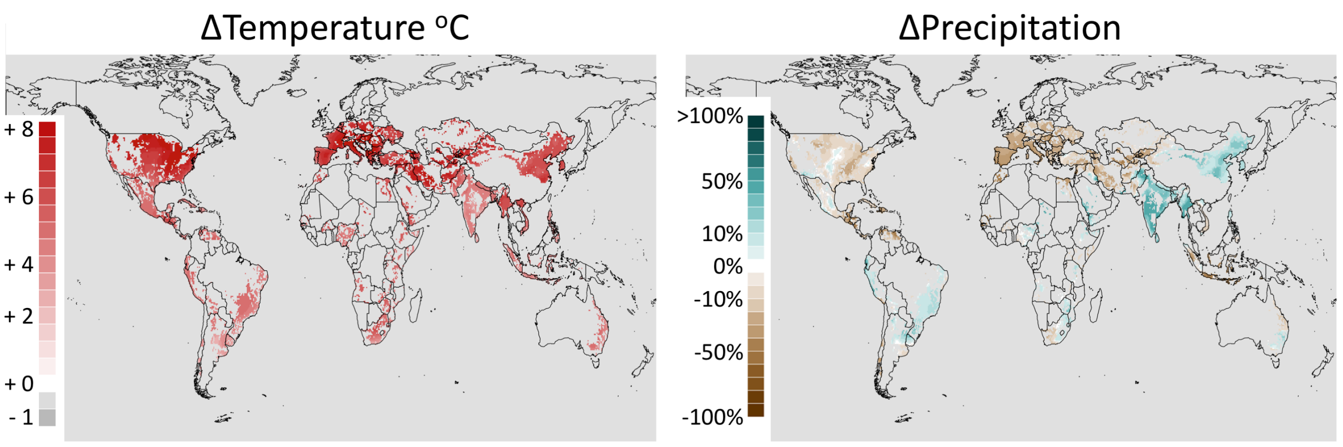


Figure 13. Gridded local growing season temperature and precipitation changes for irrigated maize under HadGEM2-ES RCP 8.5 for 2071-2100 compared to 1980-2009, global CO_2 = 812 ppm concentration.

Figure 14 includes a spatial comparison of the Persephone V1.0 low, mean, and high yield changes for irrigated maize (analogous to Figure 11) with the range of ISIMIP responses in each grid cell. Specifically, maps of the minimum, median, and maximum irrigated maize yield change across the ISIMIP GGCMs are plotted in each grid cell; no individual GGCM would produce any of these maps. As noted above, the Persephone range of yield changes in each grid cell is generally more pessimistic than the ISIMIP range, but there does appear to be spatial consistency in terms of response strength in several regions between the Persephone V1.0 range and the ISIMIP range. C3MP, and therefore the Persephone V1.0 projections, capture a strong temperature dependence and a lesser response to precipitation (particularly for irrigated crops). Because warmer temperatures are nearly universal in the HadGEM2-ES RCP 8.5 projection (Figure 13), there is limited irrigated crop response to precipitation changes, and [CO_2] response for maize is small among the mechanistic models that are more prominent in C3MP than in the GGCMs, there is nothing but yield decreases in the Persephone projection. In the ISIMIP range of GGCMs, there are models that are more positively responsive to precipitation and [CO_2] in the C4-photosynthesis maize crop, so wetter conditions and/or higher [CO_2] are much more beneficial in the ISIMIP maximum map (Rosenzweig et al., 2014) .

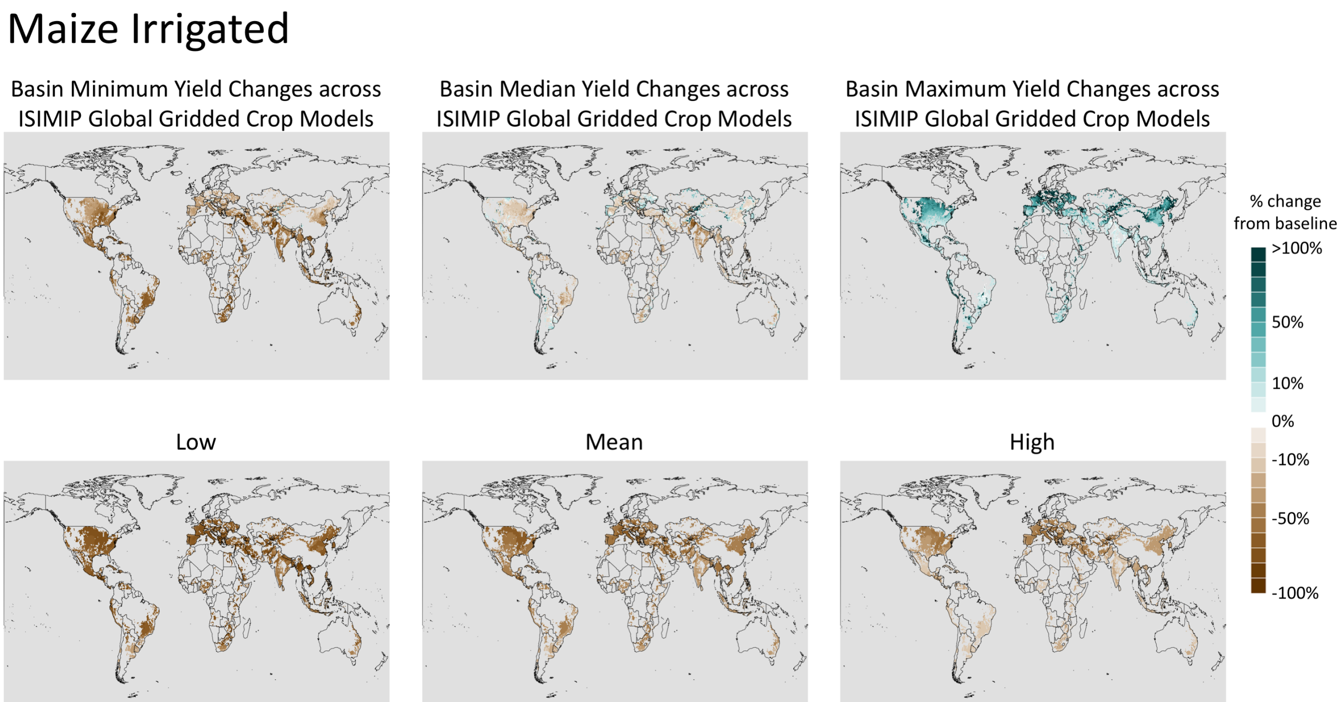


Figure 14. Top: grid cell specific minimum (Left), median (Middle), and maximum (Right) yield change across the ISIMIP GGCMs for irrigated maize. Bottom: the low, mean, and high Persephone V1.0 yield changes in each grid cell for irrigated maize.

Figure 15 presents the same analysis for a production group that Persephone V1.0 matches the ISIMIP global average range well: rainfed wheat. For reference, the HadGEM2-ES RCP 8.5 local growing season temperature and precipitation projections for rainfed wheat are included in Figure 10, bottom. Again, there is noticeable spatial consistency in response strength between the Persephone V1.0 range and the ISIMIP range. For wheat, the C3MP models and therefore the Persephone V1.0 projections are in closer agreement with the ISIMIP GGCMs on C3-photosynthesis [CO_2] response and water limitations in many regions. Additionally, the harvested area mask used for rainfed wheat include many more regions that are limited by cool baseline temperatures and thus stand to gain from warmer conditions than the regions considered for irrigated maize. Put together, these observations indicate that both Persephone V1.0 and ISIMIP are capable of the large gains in the optimistic maximum model response scenario. Together with Figure 14, this suggests that the Persephone V1.0 response functions are spatially consistent with the ISIMIP range of yield changes even when the global average ranges may disagree.

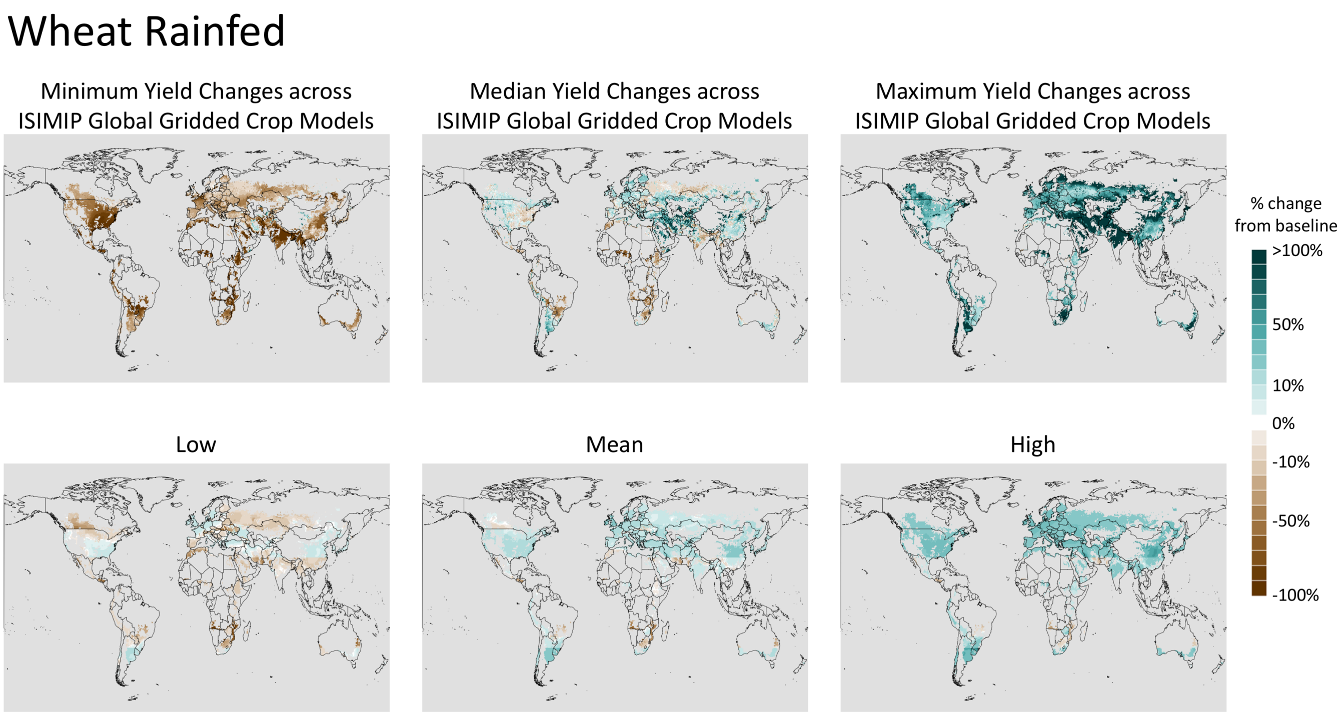


Figure 15. Top: grid cell specific minimum (Left), median (Middle), and maximum (Right) yield change across the ISIMIP GGCMs for rainfed wheat. Bottom: the low, mean, and high Persephone V1.0 yield changes in each grid cell for rainfed wheat.

5 Conclusions and discussion

We have presented ~~an~~ the Persephone emulator framework that results in ~~the three Persephone v1~~three V1.0 response functions to emulate a range of crop yield changes in response to future CTW changes for 25 production groups. The response functions are inexpensive to evaluate, open doors to new feedback loops between society and the natural environment (Figure 1), and represent multiple models and farming systems. The Persephone V1.0 response functions agree well with the underlying C3MP training data and are rapid to evaluate, with in-sample performance metrics being particularly strong for the mean response in each production group. The rapid evaluation time of the response functions, relative to a global gridded crop model, is extremely ~~important~~ imporant given that models such as GCAM are designed to be run rapidly to trace the impacts of future scenarios (at most hours per scenario). The GCAM model development team prioritizes staying on this order of computation time, even for the planned experiments outlined in Section 2.1, because it results in a nimble, flexible model that allows multiple iterations for probability, uncertainty, and process understanding. In addition to the good quantitative agreement of our response functions with all C3MP crop-irrigation-latitude ensembles, we further evaluated our mean response function heuristically, finding that the mean responds to changes in CTW as one would expect for comparisons across C3/C4 photosynthesis mechanisms, rainfed versus irrigated management, and latitude zones. Finally, the range of V1.0 yield changes were evaluated against a variety of past global gridded crop modeling, site specific crop modeling, and/or empirical studies for many of the production groups and found to be consistent.

As a result of the culling methods outlined in Section 2.2, 575 C3MP sites are used for training the Persephone functions. These sites account for many major crops where they are typically grown, as well as a wider variety of crops than has been examined in past studies. One key observation is that, if one were only concerned with capturing the mean response, any of the functional forms examined for μ_{CTW} (Equations (A1) - (A5)) in the Appendix would be excellent, with all five forms featuring in-sample NRMS < 0.2 for all production groups (Table 1). The challenge is in defining a pair of response functions, μ and σ, to characterize a range of uncertainty across C3MP site responses to each CTW change~~for use in~~. It should also be noted that such a range of uncertainty will capture only a portion of the uncertainty in response in national and multi-national GCAM units. The Persephone framework may be used with future more spatially dense data sets to characterize this uncertainty more fully.

The modeling choices made in this study introduce a variety of caveats. ~~GCAM, and many similar models, operates on 5-10~~ Foremost, it is likely that future versions of Persephone response functions, trained on different data sets, will almost certainly result in different response functions. Yet this work has shown that the Persephone framework is well-suited to this kind of problem, and that the V1.0 response functions developed from the C3MP data emulate that data well. They also perform reasonably well on heuristic metrics and in comparison to other crop modeling efforts. Another important caveat is that GCAM operates on five year timesteps. Therefore, the response functions in this work only characterize yield responses to long-term, local Earth system state changes. Capturing interannual variability and responses to abrupt weather shocks is an area ~~will~~ that may form future phases of this research. We note that this is a more difficult task, given that year-to-year variability depends on many ~~more~~ factors that tend to average out over longer terms (e.g. intra-seasonal variability such as heat waves or dry spells). ~~Using GCAM to examine the broad impacts of a sustained drought, hypothetical or emergent from the feedback loop sketched in Figure 1, would be an excellent application of this yield change emulator.~~ Additionally, this work did not account for differing nitrogen application rates across different C3MP sites. Nitrogen data is included in the C3MP archive, but the sites are heavily biased to high nitrogen application (this is likely a function of the most commonly simulated sites also being systems with higher input investment). There are also a number of sites with no recorded nitrogen information, which were kept for this study. With so few sites featuring low nitrogen application rates, we considered examining the nitrogen dimension of yield responses to be its own intellectual challenge reserved for future work, the methods of which will likely be determined by the desired use. ~~Simiarly~~Similarly, exploration of forming production groups based on different crop groups, different latitudinal zones, Koppen-Geiger or temperature zones would require trivial changes, limited only by the number of sites available to sort into different production groups. Finally, it is worth noting that any emulator is only as good as the data upon which it is trained. If crop modeling studies that provide data to an emulator do not account for real-world behaviors, the emulator will not capture such behaviors either.

For clear analysis in this paper, we have presented results for the functional form combination that performed best at the cross-validation ~~experiements~~ experiments described in the Appendix for the most production groups. Therefore one remedy to the presence of ensembles with poorer emulator performance on in-sample metrics (Table 1) would be to use different functional forms for each production group to create a more globally optimal set of response functions. These are laid out for each production group in Table A10, along with the in-sample performance of the group-specific optimal functional forms. Some analysis with these production group-specific optimal models is included in Section 3.1.1 and Section 4.1. The data processing, emulator fitting, and analysis techniques presented in this paper are agnostic of the actual functional forms used for μ_{CTW} and σ_{CTW} as long as they are linear-in-parameters. Varying functional form by production group will only require different inputs to the Persephone R functions, not refitting of any parameters.

The most immediate future work involving Persephone v1.0 will be to fully implement the feedback loop sketched in Figure 1. Specifically, using GCAM to examine the broad impacts of a sustained drought, hypothetical or emergent from the feedback loop sketched in Figure 1, would be an excellent application of this yield change emulator. Once the illustrated links have been implemented and full runs of the loop have been timed, future development may take place. In addition to the exploration of the nitrogen dimension of yield response and allowing response functional form to differ by production group, Persephone version 2 may incorporate other predictors as data is available~~and~~, explore more dynamic feature selection algorithms for functional form selection for μ_{CTW} and σ_{CTW} such as L1-regularization (which favors sparse models), and/or be trained with data sets that may be released in the future. Which of these is explored next will depend on the outcomes of the initial full feedback loop studies with GCAM. This study represents the first vital, necessary step in better identifying a pathway in which society can develop with balanced consideration of the natural environment and managed environments like agriculture through connecting Persephone and GCAM.

Code and data availability. Software implementing this technique is available as an R package released under the GNU General Public License. Full source can be found in the project's GitHub repository (https://github.com/JGCRI/persephone and https://doi.org/10.5281/zenodo.1415487). Release version 1.0.0 of the package was used for all of the work in this paper.

The data and analysis code for the results presented in this paper are archived at https://doi.org/10.5281/zenodo.1414423.

Appendix A: Model selection and performance

We fit the likelihood presented in Equation (1) with five different functional forms for μ_{CTW} (Equations (A1) - (A5)) and two different functional forms for σ_{CTW} (Equations (A6) and (A7)), resulting in data from a total of 10 emulator models (each with different likelihoods based on μ_{CTW}, σ_{CTW}) to compare to the C3MP data set.

The five functional forms for μ were selected intentionally. The first (Equation (A1)) is a second order Taylor polynomial approximation of mean yield response. Equation (A2) is the functional form for mean response used in Ruane et al. (2014), differing from the second order Taylor polynomial by only one third-order CTW interaction term, a_{10}. Equations (A3) and (A4) continue to build up from the second order Taylor polynomial, examining the impacts of adding third order CTW interaction terms and the impacts of adding pure third order CTW terms ~~respsectively~~respectively. Finally, Equation (A5) is the full third order Taylor polynomial, a flexible approximation for many complicated functions. The two functional forms for σ (Equations (A6) and (A7)) are simply the second and third order Taylor polynomial approximations of response spread across C3MP sites.

quadratic: $$\mu_{CTW} = a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \tag{A1}$$

C3MP: $$\begin{aligned}\mu_{CTW} &= a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ &+ a_{10}\Delta T\Delta W\Delta C\end{aligned} \tag{A2}$$

cross: $$\begin{aligned}\mu_{CTW} &= a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ &+ a_{10}\Delta T\Delta W\Delta C \\ &+ a_{11}(\Delta T)^2\Delta W + a_{12}(\Delta T)^2\Delta C + a_{13}\Delta T(\Delta W)^2 + a_{14}\Delta T(\Delta C)^2 + a_{15}(\Delta W)^2\Delta C + a_{16}\Delta W(\Delta C)^2\end{aligned} \tag{A3}$$

pure: $$\begin{aligned}\mu_{CTW} &= a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ &+ a_{10}(\Delta T)^3 + a_{11}(\Delta W)^3 + a_{12}(\Delta C)^3\end{aligned} \tag{A4}$$

cubic: $$\begin{aligned}\mu_{CTW} &= a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ &+ a_{10}\Delta T\Delta W\Delta C \\ &+ a_{11}(\Delta T)^2\Delta W + a_{12}(\Delta T)^2\Delta C + a_{13}\Delta T(\Delta W)^2 + a_{14}\Delta T(\Delta C)^2 + a_{15}(\Delta W)^2\Delta C + a_{16}\Delta W(\Delta C)^2 \\ &+ a_{17}(\Delta T)^3 + a_{18}(\Delta W)^3 + a_{19}(\Delta C)^3\end{aligned} \tag{A5}$$

quadratic: $$\begin{aligned}\sigma_{CTW} &= |b_0 + b_1\Delta T + b_2(\Delta T)^2 + b_3\Delta W + b_4(\Delta W)^2 + b_5\Delta C + b_6(\Delta C)^2 \\ &+ b_7\Delta T\Delta W + b_8\Delta T\Delta C + b_9\Delta W\Delta C|\end{aligned} \tag{A6}$$

$$\textbf{cubic: } \sigma_{CTW} = |b_0 + b_1\Delta T + b_2(\Delta T)^2 + b_3\Delta W + b_4(\Delta W)^2 + b_5\Delta C + b_6(\Delta C)^2 \\ + b_7\Delta T\Delta W + b_8\Delta T\Delta C + b_9\Delta W\Delta C + b_{10}\Delta T\Delta W\Delta C \\ + b_{11}(\Delta T)^2\Delta W + b_{12}(\Delta T)^2\Delta C + b_{13}\Delta T(\Delta W)^2 + b_{14}\Delta T(\Delta C)^2 \\ + b_{15}(\Delta W)^2\Delta C + b_{16}\Delta W(\Delta C)^2 + b_{17}(\Delta T)^3 + b_{18}(\Delta W)^3 + b_{19}(\Delta C)^3| \tag{A7}$$

We selected the model presented in the paper from the 10 combinations above based on leave-one (CTW test)-out cross-validation experiments to estimate out-of-sample prediction error for each production group. We do also include the in-sample performance metric defined in Section 3.1 for a more complete picture of model performance for all 10 functional form combinations for all 25 production groups (Tables A1-A9).

First, to test each model's validity and robustness at predicting yield changes for CTW values not included in the training data for each group, we ran leave-one-out cross-validation experiments (Gelman et al., 2014) to analyze the performance of each model for each production group. For each group separately, one CTW test data was withheld and the model was fit on the remaining 98 CTW tests. Then the mean, high, and low response functions resulting from the model were evaluated on the C3MP site data for the withheld test. This process was repeated withholding each CTW test, and the results were averaged resulting in an RMSE measure of performance for each of the mean, high, and low response functions. Leave-on-out cross validation used in this way answers the question: For a particular production group and model, on average, how do the emulated [mean, high, low] yield changes compare against the C3MP [mean, high, low] yield changes for CTW values not in the training set?

The Latin Hypercube design of the C3MP sensitivity tests lends confidence to this leave-one-out exercise because the cross-validation has covered the full space of CTW combinations. The results are summarized in Figure A1: each row represents the average leave-one-out cross-validation RMSE measures for each functional form across all production groups for the high, low, or mean response function, and then the average across all three (total, bottom row Figure A1). We find that cubic μ_{CTW}, quadratic σ_{CTW} performs the best at this cross validation experiment for the highest number of ensembles across the three response functions we defined in Equation (8) (that is, the high, low, and mean response functions). We repeat these calculations for each production group separately (rather than averaging across production groups) to determine the group-specific optimal functional form, listed in Table A10 for each group.

Because cubic μ_{CTW}, quadratic σ_{CTW} performs the best at out-of-sample error measurements for the highest number of ensembles across mean, low, and high response functions, and is quite good (though not the best) at in-sample error measurements (Table 1), this is the form used throughtout the body of the paper as the most broadly optimal functional form combination. We particularly value performance on the cross-validation (out-of-sample error) experiments because most CTW changes that may arise in application are likely to differ from the 99 C3MP tests.

We also repeat the in-sample measurement of error presented in Section 3.1 for all functional form combinations. These results are summarized in Tables A1 to A9, and we find that, purely based on the in-sample measurements, cubic μ_{CTW}, cubic σ_{CTW} (Table A9) is the best functional form for the most production groups. Specifically, it only performs poorly for one crop, rainfed ~~Wheat~~ wheat in the mid-latitudes. However, it is very poor for that important ensemble. The in-sample performance information from these tables is included in Table A10 for each production-group specific optimal functional form combination.

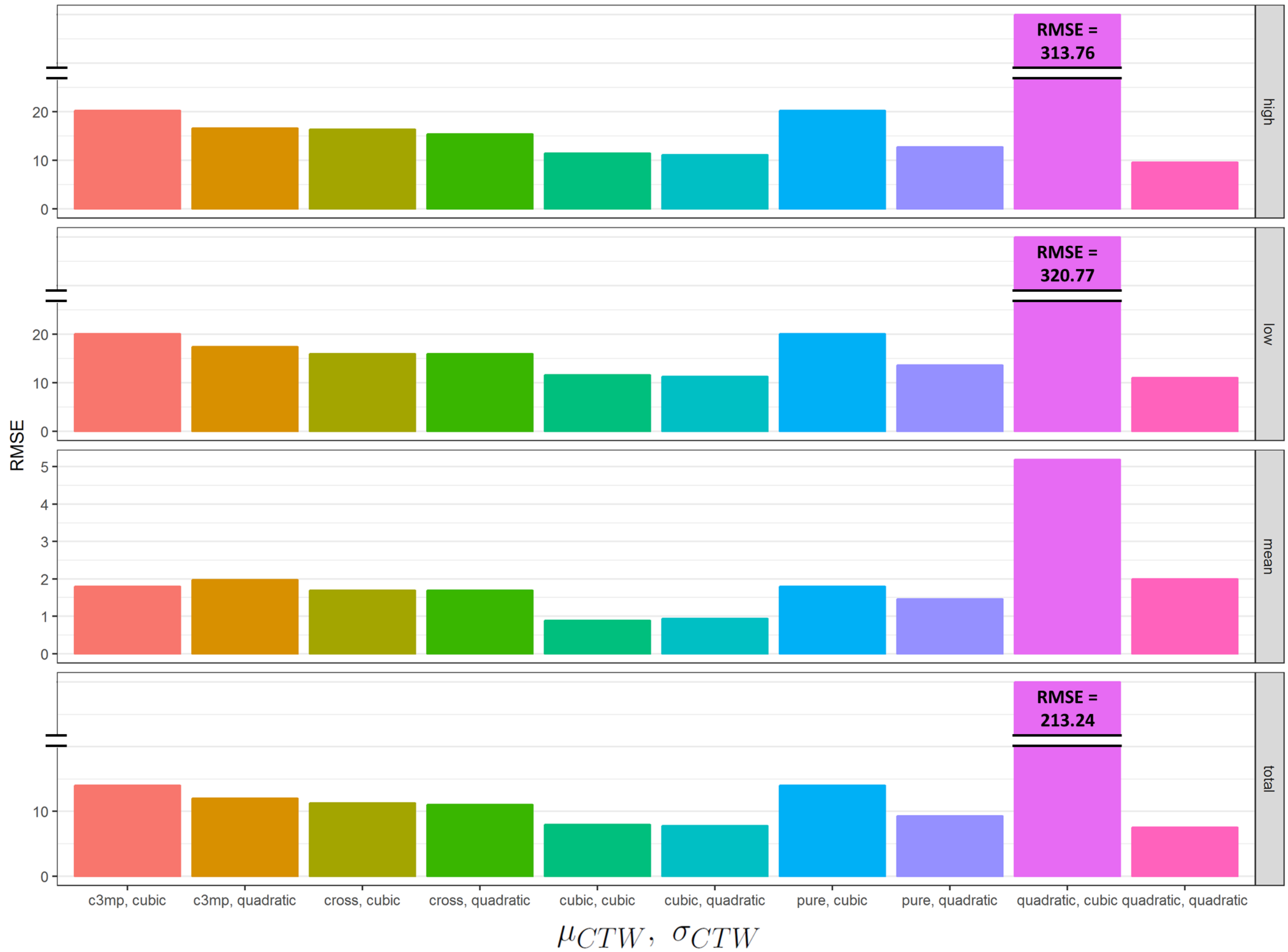


Figure A1. Comparison of leave-one-out cross-validation average RMSE measures for each functional form across all ensembles. Each functional form is labeled as μ_{CTW}, σ_{CTW}. Note the broken scales to capture the performance of quadratic μ_{CTW}, cubic $sigma_{CTW}$.

Table A1. Persephone v1.0 response function performance for all production groups, for quadratic μ_{CTW} (Equation (A1)), quadratic σ_{CTW} (Equation (A6))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.043	0.445	0.279	Good
c3 IRR tropic	0.074	0.270	0.188	Good
c3 RFD mid	0.044	0.301	0.280	Good
c3 RFD tropic	0.046	0.178	0.170	Excellent
c4 IRR mid	0.028	0.137	0.125	Excellent
c4 IRR tropic	0.041	1.662	0.440	Poor
c4 RFD mid	0.049	0.295	0.258	Good
c4 RFD tropic	0.094	0.280	0.209	Good
Maize IRR mid	0.028	0.150	0.130	Excellent
Maize IRR tropic	0.102	0.755	0.331	Adequate
Maize RFD mid	0.045	0.266	0.251	Good
Maize RFD tropic	0.108	0.318	0.188	Good
Rice IRR mid	0.069	0.259	0.181	Good
Rice IRR tropic	0.095	0.296	0.203	Good
Rice RFD mid	0.180	1.429	1.601	Poor
Rice RFD tropic	0.047	0.116	0.144	Excellent
Soybeans IRR mid	0.080	0.245	0.192	Excellent
Soybeans IRR tropic	0.068	0.119	0.175	Excellent
Soybeans RFD mid	0.069	0.139	0.178	Excellent
Soybeans RFD tropic	0.101	0.183	0.179	Excellent
Sugarcane RFD tropic	0.125	0.448	0.479	Good
Wheat IRR mid	0.069	0.408	0.351	Good
Wheat IRR tropic	0.085	0.309	0.267	Good
Wheat RFD mid	0.041	0.298	0.286	Good
Wheat RFD tropic	0.199	0.807	0.833	Adequate

1. "IRR" = irrigated, "RFD" = rainfed, "mid" = mid-latitudes (30- 70°S, 30- 70°N), "tropic" = 30°S to 30°N. 2. Note that the mean response function performs "excellent" for all production groups.

Table A2. Persephone v1.0 response function performance for all production groups, for quadratic μ_{CTW} (Equation (A1)), cubic σ_{CTW} (Equation (A7))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.036	0.142	0.110	Excellent
c3 IRR tropic	0.074	0.661	0.525	Adequate
c3 RFD mid	0.044	0.899	0.706	Adequate
c3 RFD tropic	0.052	0.817	0.571	Adequate
c4 IRR mid	0.028	2.169	0.863	Poor
c4 IRR tropic	0.035	0.300	0.063	Good
c4 RFD mid	0.042	0.125	0.080	Excellent
c4 RFD tropic	0.084	1.022	0.511	Poor
Maize IRR mid	0.035	0.763	0.471	Adequate
Maize IRR tropic	0.083	0.193	0.066	Excellent
Maize RFD mid	0.039	0.112	0.075	Excellent
Maize RFD tropic	0.086	0.390	0.147	Good
Rice IRR mid	0.064	0.159	0.098	Excellent
Rice IRR tropic	0.095	1.029	0.672	Poor
Rice RFD mid	0.153	0.166	0.187	Excellent
Rice RFD tropic	0.047	0.077	0.063	Excellent
Soybeans IRR mid	0.073	0.123	0.088	Excellent
Soybeans IRR tropic	0.057	0.078	0.089	Excellent
Soybeans RFD mid	0.075	1.137	0.893	Poor
Soybeans RFD tropic	0.084	0.355	0.303	Excellent
Sugarcane RFD tropic	0.114	2.163	1.469	Poor
Wheat IRR mid	0.060	0.149	0.197	Excellent
Wheat IRR tropic	0.082	0.206	0.204	Excellent
Wheat RFD mid	0.038	0.117	0.102	Excellent
Wheat RFD tropic	0.175	0.769	0.587	Adequate

1. “IRR” = irrigated, “RFD" = rainfed, “mid” = mid-latitudes (30- 70° S, 30- 70° N), “tropic" = 30° S to 30° N. 2. Note that the mean response function performs “excellent" for all production groups.

Table A3. Persephone v1.0 response function performance for all production groups, for C3MP μ_{CTW} (Equation (A2)), quadratic σ_{CTW} (Equation (A6))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.037	1.039	0.7	Poor
c3 IRR tropic	0.074	1.675	0.792	Poor
c3 RFD mid	0.046	0.303	0.276	Good
c3 RFD tropic	0.057	1.116	0.78	Poor
c4 IRR mid	0.027	0.139	0.123	Excellent
c4 IRR tropic	0.049	0.894	0.224	Adequate
c4 RFD mid	0.046	0.303	0.248	Good
c4 RFD tropic	0.093	0.3	0.199	Good
Maize IRR mid	0.027	0.152	0.129	Excellent
Maize IRR tropic	0.111	1.091	0.273	Poor
Maize RFD mid	0.042	0.272	0.242	Good
Maize RFD tropic	0.106	0.341	0.182	Good
Rice IRR mid	0.081	0.725	0.402	Adequate
Rice IRR tropic	0.093	0.287	0.209	Good
Rice RFD mid	0.115	1.055	1.08	Poor
Rice RFD tropic	0.047	0.18	0.164	Excellent
Soybeans IRR mid	0.08	0.248	0.191	Excellent
Soybeans IRR tropic	0.11	0.726	0.724 Adequate	
Soybeans RFD mid	0.066	0.149	0.157	Excellent
Soybeans RFD tropic	0.084	0.444	0.354	Good
Sugarcane RFD tropic	0.144	2.42	2.066	Poor
Wheat IRR mid	0.061	0.391	0.365	Good
Wheat IRR tropic	0.082	0.72	0.548	Adequate
Wheat RFD mid	0.041	0.298	0.287	Good
Wheat RFD tropic	0.147	0.297	0.376	Good

1. “IRR" = irrigated, “RFD" = rainfed, “mid" = mid-latitudes (30- 70°S, 30- 70°N), “tropic" = 30°S to 30°N. 2. Note that the mean response function performs “excellent" for all production groups.

Table A4. Persephone v1.0 response function performance for all production groups, for C3MP μ_{CTW} (Equation (A2)), cubic σ_{CTW} (Equation (A7))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.032	0.356	0.240	Good
c3 IRR tropic	0.073	0.113	0.113	Excellent
c3 RFD mid	0.039	0.121	0.094	Excellent
c3 RFD tropic	0.041	0.087	0.057	Excellent
c4 IRR mid	0.026	0.166	0.108	Excellent
c4 IRR tropic	0.037	0.296	0.064	Good
c4 RFD mid	0.038	0.449	0.358	Good
c4 RFD tropic	0.073	0.335	0.168	Good
Maize IRR mid	0.025	0.073	0.044	Excellent
Maize IRR tropic	0.082	0.244	0.082	Good
Maize RFD mid	0.036	0.109	0.076	Excellent
Maize RFD tropic	0.096	0.729	0.272	Adequate
Rice IRR mid	0.064	0.282	0.175	Good
Rice IRR tropic	0.094	0.120	0.143	Excellent
Rice RFD mid	0.134	0.175	0.178	Excellent
Rice RFD tropic	0.046	0.079	0.060	Excellent
Soybeans IRR mid	0.073	0.123	0.088	Excellent
Soybeans IRR tropic	0.075	0.213	0.194	Excellent
Soybeans RFD mid	0.060	0.080	0.068	Excellent
Soybeans RFD tropic	0.086	0.145	0.169	Excellent
Sugarcane RFD tropic	0.111	0.175	0.100	Excellent
Wheat IRR mid	0.061	0.961	1.039	Poor
Wheat IRR tropic	0.088	2.522	1.231	Poor
Wheat RFD mid	0.058	7.604	2.233	Poor
Wheat RFD tropic	0.164	0.934	0.924	Adequate

1. “IRR” = irrigated, “RFD” = rainfed, “mid” = mid-latitudes (30- 70°S, 30- 70°N), “tropic” = 30°S to 30°N. 2. Note that the mean response function performs “excellent” for all production groups.

Table A5. Persephone v1.0 response function performance for all production groups, for cross μ_{CTW} (Equation (A3)), quadratic σ_{CTW} (Equation (A6))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.022	1.038	0.701	Poor
c3 IRR tropic	0.073	1.671	0.792	Poor
c3 RFD mid	0.021	0.314	0.272	Good
c3 RFD tropic	0.030	1.201	0.634	Poor
c4 IRR mid	0.024	0.140	0.121	Excellent
c4 IRR tropic	0.041	0.928	0.220	Poor
c4 RFD mid	0.033	0.312	0.247	Good
c4 RFD tropic	0.069	0.340	0.187	Good
Maize IRR mid	0.025	0.152	0.128	Excellent
Maize IRR tropic	0.107	1.926	0.450	Poor
Maize RFD mid	0.030	0.286	0.236	Good
Maize RFD tropic	0.083	0.379	0.175	Good
Rice IRR mid	0.070	0.627	0.445	Poor
Rice IRR tropic	0.092	0.347	0.258	Good
Rice RFD mid	0.092	0.306	0.342	Good
Rice RFD tropic	0.020	0.210	0.141	Excellent
Soybeans IRR mid	0.090	1.595	1.103	Poor
Soybeans IRR tropic	0.051	0.203	0.161	Excellent
Soybeans RFD mid	0.036	0.150	0.148	Excellent
Soybeans RFD tropic	0.081	0.318	0.219	Good
Sugarcane RFD tropic	0.147	5.574	3.954	Poor
Wheat IRR mid	0.056	0.392	0.364	Good
Wheat IRR tropic	0.078	1.256	0.815	Poor
Wheat RFD mid	0.034	0.306	0.279	Good
Wheat RFD tropic	0.114	0.332	0.347	Good

1. "IRR" = irrigated, "RFD" = rainfed, "mid" = mid-latitudes (30- 70°S, 30- 70°N), "tropic" = 30°S to 30°N. 2. Note that the mean response function performs "excellent" for all production groups.

Table A6. Persephone v1.0 response function performance for all production groups, for cross μ_{CTW} (Equation (A3)), cubic σ_{CTW} (Equation (A7))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.019	0.303	0.196	Good
c3 IRR tropic	0.071	0.112	0.111	Excellent
c3 RFD mid	0.022	0.674	0.602	Adequate
c3 RFD tropic	0.025	0.071	0.056	Excellent
c4 IRR mid	0.024	0.168	0.114	Excellent
c4 IRR tropic	0.032	0.303	0.076	Good
c4 RFD mid	0.037	1.544	0.623	Poor
c4 RFD tropic	0.062	0.156	0.060	Excellent
Maize IRR mid	0.022	0.071	0.044	Excellent
Maize IRR tropic	0.074	0.179	0.063	Excellent
Maize RFD mid	0.028	0.097	0.081	Excellent
Maize RFD tropic	0.073	0.305	0.129	Good
Rice IRR mid	0.063	0.278	0.176	Good
Rice IRR tropic	0.092	0.120	0.141	Excellent
Rice RFD mid	0.096	0.237	0.219	Good
Rice RFD tropic	0.019	0.057	0.051	Excellent
Soybeans IRR mid	0.058	0.120	0.073	Excellent
Soybeans IRR tropic	0.063	0.120	0.212	Excellent
Soybeans RFD mid	0.034	0.054	0.054	Excellent
Soybeans RFD tropic	0.053	0.111	0.094	Excellent
Sugarcane RFD tropic	0.078	0.241	0.229	Excellent
Wheat IRR mid	0.044	0.721	0.748	Adequate
Wheat IRR tropic	0.084	0.185	0.219	Excellent
Wheat RFD mid	0.050	3.658	2.116	Poor
Wheat RFD tropic	0.111	0.212	0.179	Excellent

1. "IRR" = irrigated, "RFD" = rainfed, "mid" = mid-latitudes (30- 70° S, 30- 70° N), "tropic" = 30° S to 30° N. 2. Note that the mean response function performs "excellent" for all production groups.

Table A7. Persephone v1.0 response function performance for all production groups, for pure μ_{CTW} (Equation (A4)), quadratic σ_{CTW} (Equation (A6))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.031	1.045	0.697	Poor
c3 IRR tropic	0.071	1.660	0.791	Poor
c3 RFD mid	0.039	0.301	0.280	Good
c3 RFD tropic	0.052	0.662	0.498	Poor
c4 IRR mid	0.012	0.149	0.111	Excellent
c4 IRR tropic	0.018	0.985	0.216	Poor
c4 RFD mid	0.035	0.307	0.248	Good
c4 RFD tropic	0.045	0.334	0.189	Good
Maize IRR mid	0.012	0.165	0.117	Excellent
Maize IRR tropic	0.016	1.039	0.340	Poor
Maize RFD mid	0.035	0.277	0.242	Good
Maize RFD tropic	0.044	0.376	0.179	Good
Rice IRR mid	0.038	0.346	0.197	Good
Rice IRR tropic	0.091	0.343	0.260	Good
Rice RFD mid	0.124	0.123	0.275	Good
Rice RFD tropic	0.053	0.161	0.171	Excellent
Soybeans IRR mid	0.033	0.221	0.185	Excellent
Soybeans IRR tropic	0.066	0.072	0.172	Excellent
Soybeans RFD mid	0.056	0.137	0.170	Excellent
Soybeans RFD tropic	0.083	0.171	0.173	Excellent
Sugarcane RFD tropic	0.085	1.504	1.307	Poor
Wheat IRR mid	0.045	0.378	0.377	Good
Wheat IRR tropic	0.080	0.710	0.550	Good
Wheat RFD mid	0.034	0.294	0.289	Good
Wheat RFD tropic	0.175	0.371	0.341	Good

1. "IRR" = irrigated, "RFD" = rainfed, "mid" = mid-latitudes (30- 70° S, 30- 70° N), "tropic" = 30° S to 30° N. 2. Note that the mean response function performs "excellent" for all production groups.

Table A8. Persephone v1.0 response function performance for all production groups, for pure μ_{CTW} (Equation (A4)), cubic σ_{CTW} (Equation (A7))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.030	0.766	0.524	Adequate
c3 IRR tropic	0.071	0.115	0.110	Excellent
c3 RFD mid	0.035	0.117	0.095	Excellent
c3 RFD tropic	0.040	0.082	0.061	Excellent
c4 IRR mid	0.012	0.153	0.116	Excellent
c4 IRR tropic	0.013	0.249	0.072	Excellent
c4 RFD mid	0.038	2.286	0.778	Poor
c4 RFD tropic	0.040	0.120	0.061	Excellent
Maize IRR mid	0.012	0.061	0.046	Excellent
Maize IRR tropic	0.016	0.162	0.073	Excellent
Maize RFD mid	0.031	0.104	0.077	Excellent
Maize RFD tropic	0.041	0.126	0.060	Excellent
Rice IRR mid	0.038	0.109	0.076	Excellent
Rice IRR tropic	0.092	0.123	0.139	Excellent
Rice RFD mid	0.122	0.178	0.213	Excellent
Rice RFD tropic	0.043	0.213	0.149	Excellent
Soybeans IRR mid	0.029	0.091	0.071	Excellent
Soybeans IRR tropic	0.065	0.125	0.141	Excellent
Soybeans RFD mid	0.052	0.072	0.061	Excellent
Soybeans RFD tropic	0.066	0.112	0.105	Excellent
Sugarcane RFD tropic	0.066	0.260	0.177	Good
Wheat IRR mid	0.033	0.691	0.705	Adequate
Wheat IRR tropic	0.078	0.185	0.215	Excellent
Wheat RFD mid	0.037	5.732	2.313	Poor
Wheat RFD tropic	0.173	0.368	0.204	Good

1. “IRR" = irrigated, “RFD" = rainfed, “mid" = mid-latitudes (30- 70° S, 30- 70° N), “tropic" = 30° S to 30° N. 2. Note that the mean response function performs “excellent" for all production groups.

Table A9. Persephone v1.0 response function performance for all production groups, for cubic μ_{CTW} (Equation (A5)), cubic σ_{CTW} (Equation (A7))

Production group[1]	NRMS mean[2]	NRMS high	NRMS low	In-sample Performance
c3 IRR mid	0.013	0.488	0.326	Good
c3 IRR tropic	0.069	0.113	0.109	Excellent
c3 RFD mid	0.009	0.106	0.095	Excellent
c3 RFD tropic	0.021	0.065	0.058	Excellent
c4 IRR mid	0.010	0.152	0.116	Excellent
c4 IRR tropic	0.010	0.313	0.092	Good
c4 RFD mid	0.016	0.705	0.370	Adequate
c4 RFD tropic	0.018	0.102	0.058	Excellent
Maize IRR mid	0.010	0.062	0.044	Excellent
Maize IRR tropic	0.011	0.116	0.066	Excellent
Maize RFD mid	0.016	0.091	0.079	Excellent
Maize RFD tropic	0.021	0.109	0.056	Excellent
Rice IRR mid	0.029	0.104	0.073	Excellent
Rice IRR tropic	0.089	0.123	0.137	Excellent
Rice RFD mid	0.043	0.098	0.123	Excellent
Rice RFD tropic	0.018	0.060	0.048	Excellent
Soybeans IRR mid	0.015	0.087	0.068	Excellent
Soybeans IRR tropic	0.034	0.063	0.085	Excellent
Soybeans RFD mid	0.015	0.042	0.046	Excellent
Soybeans RFD tropic	0.035	0.100	0.089	Excellent
Sugarcane RFD tropic	0.042	0.209	0.171	Excellent
Wheat IRR mid	0.022	0.681	0.675	Adequate
Wheat IRR tropic	0.078	0.171	0.221	Excellent
Wheat RFD mid	0.042	5.268	1.905	Poor
Wheat RFD tropic	0.091	0.196	0.165	Excellent

1. "IRR" = irrigated, "RFD" = rainfed, "mid" = mid-latitudes (30- 70° S, 30- 70° N), "tropic" = 30° S to 30° N. 2. Note that the mean response function performs "excellent" for all production groups.

Table A10. The best performing functional form combination for each production group at the task of leave-one-out cross-validation(out of sample performance) and the corresponding In-sample Performance measure.

Production group	μ_{CTW}	σ_{CTW}	In-sample Performance
c3 IRR mid	quadratic	quadratic	Good
c3 IRR tropic	cubic	cubic	Excellent
c3 RFD mid	c3mp	cubic	Excellent
c3 RFD tropic	cubic	cubic	Excellent
c4 IRR mid	cubic	cubic	Excellent
c4 IRR tropic	cubic	cubic	Good
c4 RFD mid	cubic	quadratic	Good
c4 RFD tropic	pure	quadratic	Good
Maize IRR mid	cubic	cubic	Excellent
Maize IRR tropic	cubic	cubic	Excellent
Maize RFD mid	cubic	cubic	Excellent
Maize RFD tropic	cubic	quadratic	Good
Rice IRR mid	cubic	cubic	Excellent
Rice IRR tropic	quadratic	quadratic	Good
Rice RFD mid	cubic	cubic	Excellent
Rice RFD tropic	cubic	cubic	Excellent
Soybeans IRR mid	cubic	cubic	Excellent
Soybeans IRR tropic	quadratic	cubic	Excellent
Soybeans RFD mid	cubic	cubic	Excellent
Soybeans RFD tropic	cubic	cubic	Excellent
Sugarcane RFD tropic	c3mp	cubic	Excellent
Wheat IRR mid	quadratic	cubic	Excellent
Wheat IRR tropic	quadratic	quadratic	Good
Wheat RFD mid	cubic	quadratic	Good
Wheat RFD tropic	cubic	cubic	Excellent

Appendix B: C3MP baseline yield estimate functional forms

As mentioned in Section 2.1.1, the 8 different functional forms used to relate site-specific output yield in response to input CTW values are presented here in Equations (B1)-(B8). Each functional form was used with each specific C3MP site's data in order to provide a best estimate of baseline yield for that site. The form with the smallest root mean square error across the 99 tests for the site is the one used to provide a best estimate of baseline yield. This best estimate of baseline yield is used to convert the C3MP output yields at the site to percent changes in yield from baseline for emulator training.

$$Y_{CTW}^{site} = a_0 + a_1\Delta T + a_2\Delta W + a_3\Delta C \tag{B1}$$

$$Y_{CTW}^{site} = a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 \tag{B2}$$

$$Y_{CTW}^{site} = a_0 + a_1\Delta T + a_2\Delta W + a_3\Delta C + a_4\Delta T\Delta W + a_5\Delta T\Delta C + a_6\Delta W\Delta C \tag{B3}$$

$$Y_{CTW}^{site} = a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \tag{B4}$$

$$\begin{aligned} Y_{CTW}^{site} = {} & a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ & + a_{10}\Delta T\Delta W\Delta C \end{aligned} \tag{B5}$$

$$\begin{aligned} Y_{CTW}^{site} = {} & a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ & + a_{10}\Delta T\Delta W\Delta C \\ & + a_{11}(\Delta T)^2\Delta W + a_{12}(\Delta T)^2\Delta C + a_{13}\Delta T(\Delta W)^2 + a_{14}\Delta T(\Delta C)^2 + a_{15}(\Delta W)^2\Delta C + a_{16}\Delta W(\Delta C)^2 \end{aligned} \tag{B6}$$

$$\begin{aligned} Y_{CTW}^{site} = {} & a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\ & + a_{10}(\Delta T)^3 + a_{11}(\Delta W)^3 + a_{12}(\Delta C)^3 \end{aligned} \tag{B7}$$

$$
\begin{aligned}
Y_{CTW}^{site} = {} & a_0 + a_1\Delta T + a_2(\Delta T)^2 + a_3\Delta W + a_4(\Delta W)^2 + a_5\Delta C + a_6(\Delta C)^2 + a_7\Delta T\Delta W + a_8\Delta T\Delta C + a_9\Delta W\Delta C \\
& + a_{10}\Delta T\Delta W\Delta C \\
& + a_{11}(\Delta T)^2\Delta W + a_{12}(\Delta T)^2\Delta C + a_{13}\Delta T(\Delta W)^2 + a_{14}\Delta T(\Delta C)^2 + a_{15}(\Delta W)^2\Delta C + a_{16}\Delta W(\Delta C)^2 \\
& + a_{17}(\Delta T)^3 + a_{18}(\Delta W)^3 + a_{19}(\Delta C)^3
\end{aligned}
\tag{B8}
$$

Author contributions. ACR and KC conceived the uses of this emulator. AS, MP, ACR, KC developed the emulator. AS wrote the manuscript with contributions from MP, KC, ACR.

Competing interests. No competing interests are present.

Acknowledgements. The authors appreciate discussions with Sonali McDermid and Theo Mavromatis around the evaluation and application of the C3MP ensemble. Abigail Snyder, Katherine Calvin, and Meridel Phillips were supported by the U.S. Department of Energy, Office of Science, as part of research in the Multi-Sector Dynamics, Earth and Environmental System Modeling Program Area. Alex Ruane's work was supported by the NASA Climate Impacts Group under the Modeling, Analysis, and Prediction Program.

References

Asseng, S., Ewert, F., Rosenzweig, C., Jones, J., Hatfield, J., Ruane, A., Boote, K. J., Thorburn, P. J., Rötter, R. P., Cammarano, D., et al.: Uncertainty in simulating wheat yields under climate change, Nature Climate Change, 3, 827, 2013.

Asseng, S., Ewert, F., Martre, P., Rötter, R. P., Lobell, D., Cammarano, D., Kimball, B., Ottman, M., Wall, G., White, J. W., et al.: Rising temperatures reduce global wheat production, Nature Climate Change, 5, 143, 2015.

Bassu, S., Brisson, N., Durand, J.-L., Boote, K., Lizaso, J., Jones, J. W., Rosenzweig, C., Ruane, A. C., Adam, M., Baron, C., et al.: How do various maize crop models vary in their responses to climate change factors?, Global change biology, 20, 2301–2320, 2014.

Blanc, É.: Statistical emulators of maize, rice, soybean and wheat yields from global gridded crop models, Agricultural and Forest Meteorology, 236, 145–161, 2017.

Bond-Lamberty, B., Calvin, K., Jones, A. D., Mao, J., Patel, P., Shi, X., Thomson, A., Thornton, P., and Zhou, Y.: On linking an Earth system model to the equilibrium carbon representation of an economically optimizing land use model, Geoscientific Model Development, 7, 2545, 2014.

Calvin, K. and Fisher-Vanden, K.: Quantifying the indirect impacts of climate on agriculture: an inter-method comparison, Environmental Research Letters, 12, 115 004, 2017.

Calvin, K., Clarke, L., Edmonds, J., Eom, J., Hejazi, M., Kim, S., Kyle, P., Link, R., Luckow, P., Patel, P., et al.: GCAM wiki documentation, Pacific Northwest National Laboratory, 2011.

Calvin, K., Wise, M., Clarke, L., Edmonds, J., Kyle, P., Luckow, P., and Thomson, A.: Implications of simultaneously mitigating and adapting to climate change: initial experiments using GCAM, Climatic Change, 117, 545–560, 2013.

Calvin, K., Patel, P., Clarke, L., Asrar, G., Bond-Lamberty, B., Di Vittorio, A., Edmonds, J., Hartin, C., Hejazi, M., Iyer, G., et al.: GCAM v5.1: Representing the linkages between energy, water, land, climate, and economic systems, Geosci. Model Dev. Discuss. Accepted for publication in Geoscientific Model Development, forthcoming, 2019.

Clarke, L., Lurz, J., Wise, M., Edmonds, J., Kim, S., Smith, S., and Pitcher, H.: Model documentation for the minicam climate change science program stabilization scenarios: Ccsp product 2.1 a, Pacific Northwest National Laboratory, PNNL-16735, 2007.

Davies, L. and Gather, U.: The identification of multiple outliers, Journal of the American Statistical Association, 88, 782–792, 1993.

Durand, J.-L., Delusca, K., Boote, K., Lizaso, J., Manderscheid, R., Weigel, H. J., Ruane, A. C., Rosenzweig, C., Jones, J., Ahuja, L., et al.: How accurately do maize crop models simulate the interactions of atmospheric CO2 concentration levels with limited water supply on water use and yield?, European journal of agronomy, 100, 67–75, 2018.

Elliott, J., Müller, C., Deryng, D., Chryssanthacopoulos, J., Boote, K., Büchner, M., Foster, I., Glotter, M., Heinke, J., Iizumi, T., et al.: The Global Gridded Crop Model intercomparison: data and modeling protocols for Phase 1 (v1. 0), Geoscientific Model Development Discussions, 7, 4383–4427, 2014.

Fronzek, S., Pirttioja, N., Carter, T. R., Bindi, M., Hoffmann, H., Palosuo, T., Ruiz-Ramos, M., Tao, F., Trnka, M., Acutis, M., et al.: Classifying multi-model wheat yield impact response surfaces showing sensitivity to temperature and precipitation change, Agricultural Systems, 159, 209–224, 2018.

Gelman, A., Stern, H. S., Carlin, J. B., Dunson, D. B., Vehtari, A., and Rubin, D. B.: Bayesian data analysis, Chapman and Hall/CRC, 2013.

Gelman, A., Hwang, J., and Vehtari, A.: Understanding predictive information criteria for Bayesian models, Statistics and computing, 24, 997–1016, 2014.

Hartin, C. A., Patel, P., Schwarber, A., Link, R. P., and Bond-Lamberty, B.: A simple object-oriented and open-source model for scientific and policy analyses of the global climate system–Hector v1. 0, Geoscientific Model Development, 8, 939, 2015.

Jones, J. W., Hoogenboom, G., Porter, C. H., Boote, K. J., Batchelor, W. D., Hunt, L., Wilkens, P. W., Singh, U., Gijsman, A. J., and Ritchie, J. T.: The DSSAT cropping system model, European journal of agronomy, 18, 235–265, 2003.

Kim, S. H., Edmonds, J., Lurz, J., Smith, S., and Wise, M.: The Object-oriented Energy Climate Technology Systems (ObjECTS) framework and hybrid modeling of transportation in the MiniCAM long-term, global integrated assessment model, Energy J, 27, 63–91, 2006.

Kyle, G. P., Luckow, P., Calvin, K. V., Emanuel, W. R., Nathan, M., and Zhou, Y.: GCAM 3.0 agriculture and land use: data sources and methods, Tech. rep., Pacific Northwest National Laboratory (PNNL), Richland, WA (US), 2011.

Leakey, A. D., Bishop, K. A., and Ainsworth, E. A.: A multi-biome gap in understanding of crop and ecosystem responses to elevated CO2, Current opinion in plant biology, 15, 228–236, 2012.

Legates, D. R. and McCabe, G. J.: Evaluating the use of "goodness-of-fit" measures in hydrologic and hydroclimatic model validation, Water resources research, 35, 233–241, 1999.

Lobell, D. B.: Errors in climate datasets and their effects on statistical crop models, Agricultural and Forest Meteorology, 170, 58–66, 2013.

Martre, P., Wallach, D., Asseng, S., Ewert, F., Jones, J. W., Rötter, R. P., Boote, K. J., Ruane, A. C., Thorburn, P. J., Cammarano, D., et al.: Multimodel ensembles of wheat growth: many models are better than one, Global change biology, 21, 911–925, 2015.

McDermid, S. P., Ruane, A. C., Rosenzweig, C., Hudson, N. I., Morales, M. D., Agalawatte, P., Ahmad, S., Ahuja, L., Amien, I., Anapalli, S. S., et al.: The AgMIP coordinated climate-crop modeling project (C3MP): methods and protocols, in: HANDBOOK OF CLIMATE CHANGE AND AGROECOSYSTEMS: The Agricultural Model Intercomparison and Improvement Project Integrated Crop and Economic Assessments, Part 1, pp. 191–220, World Scientific, 2015.

McElreath, R.: Statistical Rethinking: A Bayesian Course with Examples in R and Stan, vol. 122, CRC Press, 2016.

Mistry, M. N.: Impacts of climate change and variability on crop yields using emulators and empirical models, 2017.

Mistry, M. N., Wing, I. S., and De Cian, E.: Simulated vs. empirical weather responsiveness of crop yields: US evidence and implications for the agricultural impacts of climate change, Environmental Research Letters, 12, 075 007, 2017.

Monfreda, C., Ramankutty, N., and Foley, J. A.: Farming the planet: 2. Geographic distribution of crop areas, yields, physiological types, and net primary production in the year 2000, Global biogeochemical cycles, 22, 2008.

Moore, F. C., Baldos, U., Hertel, T., and Diaz, D.: New science of climate change impacts on agriculture implies higher social cost of carbon, Nature Communications, 8, 1607, 2017.

Müller, C., Elliott, J., Chryssanthacopoulos, J., Arneth, A., Balkovic, J., Ciais, P., Deryng, D., Folberth, C., Glotter, M., Hoek, S., et al.: Global gridded crop model evaluation: benchmarking, skills, deficiencies and implications, Geoscientific Model Development, 10, 1403, 2017.

Nelson, G. C., Valin, H., Sands, R. D., Havlík, P., Ahammad, H., Deryng, D., Elliott, J., Fujimori, S., Hasegawa, T., Heyhoe, E., et al.: Climate change effects on agriculture: Economic responses to biophysical shocks, Proceedings of the National Academy of Sciences, 111, 3274–3279, 2014.

Ostberg, S., Schewe, J., Childers, K., and Frieler, K.: Changes in crop yields and their variability at different levels of global warming, Earth System Dynamics, 9, 479–496, 2018.

Oyebamiji, O. K., Edwards, N. R., Holden, P. B., Garthwaite, P. H., Schaphoff, S., and Gerten, D.: Emulating global climate change impacts on crop yields, Statistical Modelling, 15, 499–525, 2015.

Pirttioja, N., Carter, T. R., Fronzek, S., Bindi, M., Hoffmann, H., Palosuo, T., Ruiz-Ramos, M., Tao, F., Trnka, M., Acutis, M., et al.: Temperature and precipitation effects on wheat yield across a European transect: a crop model ensemble analysis using impact response surfaces, 2015.

Portmann, F. T., Siebert, S., and Döll, P.: MIRCA2000—Global monthly irrigated and rainfed crop areas around the year 2000: A new high-resolution data set for agricultural and hydrological modeling, Global Biogeochemical Cycles, 24, 2010.

Rosenzweig, C., Jones, J. W., Hatfield, J. L., Ruane, A. C., Boote, K. J., Thorburn, P., Antle, J. M., Nelson, G. C., Porter, C., Janssen, S., et al.: The agricultural model intercomparison and improvement project (AgMIP): protocols and pilot studies, Agricultural and Forest Meteorology, 170, 166–182, 2013.

Rosenzweig, C., Elliott, J., Deryng, D., Ruane, A. C., Müller, C., Arneth, A., Boote, K. J., Folberth, C., Glotter, M., Khabarov, N., et al.: Assessing agricultural risks of climate change in the 21st century in a global gridded crop model intercomparison, Proceedings of the National Academy of Sciences, 111, 3268–3273, 2014.

Ruane, A. C., McDermid, S., Rosenzweig, C., Baigorria, G. A., Jones, J. W., Romero, C. C., and DeWayne Cecil, L.: Carbon–Temperature–Water change analysis for peanut production under climate change: a prototype for the AgMIP Coordinated Climate-Crop Modeling Project (C3MP), Global change biology, 20, 394–407, 2014.

Ruane, A. C., Goldberg, R., and Chryssanthacopoulos, J.: Climate forcing datasets for agricultural modeling: Merged products for gap-filling and historical climate series estimation, Agricultural and Forest Meteorology, 200, 233–248, 2015.

Ruane, A. C., Rosenzweig, C., Asseng, S., Boote, K. J., Elliott, J., Ewert, F., Jones, J. W., Martre, P., McDermid, S. P., Müller, C., et al.: An AgMIP framework for improved agricultural representation in integrated assessment models, Environmental Research Letters, 12, 125 003, 2017.

Ruane, A. C., Phillips, M. M., and Rosenzweig, C.: Climate shifts within major agricultural seasons for+ 1.5 and+ 2.0° C worlds: HAPPI projections and AgMIP modeling scenarios, Agricultural and Forest Meteorology, 259, 329–344, 2018.

Schlenker, W. and Roberts, M. J.: Nonlinear temperature effects indicate severe damages to US crop yields under climate change, Proceedings of the National Academy of sciences, 106, 15 594–15 598, 2009.

Sivia, D. and Skilling, J.: Data analysis: a Bayesian tutorial, OUP Oxford, 2006.

Snyder, A. C., Link, R. P., and Calvin, K. V.: Evaluation of integrated assessment model hindcast experiments: a case study of the GCAM 3.0 land use module, Geoscientific Model Development, 10, 4307, 2017.

Warszawski, L., Frieler, K., Huber, V., Piontek, F., Serdeczny, O., and Schewe, J.: The inter-sectoral impact model intercomparison project (ISI–MIP): project framework, Proceedings of the National Academy of Sciences, 111, 3228–3232, 2014.

Wiebe, K., Lotze-Campen, H., Sands, R., Tabeau, A., van der Mensbrugghe, D., Biewald, A., Bodirsky, B., Islam, S., Kavallari, A., Mason-D'Croz, D., et al.: Climate change impacts on agriculture in 2050 under a range of plausible socioeconomic and emissions scenarios, Environmental Research Letters, 10, 085 010, 2015.

Williams, C. K. and Rasmussen, C. E.: Gaussian processes for machine learning, the MIT Press, 2, 4, 2006.

Willmott, C. J.: On the evaluation of model performance in physical geography, in: Spatial statistics and models, pp. 443–460, Springer, 1984.

Wise, M., Calvin, K., Kyle, P., Luckow, P., and Edmonds, J.: Economic and physical modeling of land use in GCAM 3.0 and an application to agricultural productivity, land, and terrestrial carbon, Climate Change Economics, 5, 1450 003, 2014.

You, L., Wood-Sichra, U., Fritz, S., Guo, Z., See, L., and Koo, J.: Spatial Production Allocation Model (SPAM) 2005 v2.0, Available from http://mapspam.info, 2014.