

1 **Modeling the effects of litter stoichiometry and soil mineral N**
2 **availability on soil organic matter formation using CENTURY-CUE**
3 **(v1.0)**

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18 **Abstract**

19 Microbial decomposition of plant litter is a crucial process for the land
20 carbon (C) cycle, as it directly controls the partitioning of litter-C between CO₂
21 released to the atmosphere versus the formation of new soil organic matter (SOM).
22 Land surface models used to study the C cycle rarely considered flexibility in the
23 decomposer C use efficiency (CUE_d) defined by the fraction of decomposed litter-C
24 that is retained as SOM (as opposed to be respired). In this study, we adapted a
25 conceptual formulation of CUE_d based on assumption that litter decomposers
26 optimally adjust their CUE_d as a function of litter substrate C to nitrogen (N)
27 stoichiometry to maximize their growth rates. This formulation was incorporated into
28 the widely used CENTURY soil biogeochemical model and evaluated based on data
29 from laboratory litter incubation experiments. Results indicated that the CENTURY
30 model with new CUE_d formulation was able to reproduce differences in respiration
31 rate of litter with contrasting C:N ratios and under different levels of mineral N
32 availability, whereas the default model with fixed CUE_d could not. Using the model
33 with flexible CUE_d, we also illustrated that litter quality affected the long-term SOM
34 formation. Litter with a small C:N ratio tended to form a larger SOM pool than litter
35 with larger C:N ratios, as it could be more efficiently incorporated into SOM by
36 microorganisms. This study provided a simple but effective formulation to quantify
37 the effect of varying litter quality (N content) on SOM formation across temporal
38 scales. Optimality theory appears to be suitable to predict complex processes of litter
39 decomposition into soil C, and to quantify how plant residues and manure can be
40 harnessed to improve soil C sequestration for climate mitigation.

41

42 *Keywords:* microbial carbon use efficiency, litter decomposition, litter stoichiometry,
43 soil organic matter, litter decay model, nitrogen

44

45 **1 Introduction**

46 Plant litter decomposition plays a key role in global carbon (C) cycle, thus
47 needs to be well represented in land surface models. The decomposition and
48 transformation processes of plant litter control the formation of soil organic matter
49 (SOM) (Prescott, 2010; Schmidt *et al.*, 2011; Walela *et al.*, 2014; Cotrufo *et al.*, 2015)
50 and associate immobilization and mineralization of essential plant nutrients
51 (Moorhead and Sinsabaugh, 2006; Parton *et al.*, 2007; Manzoni *et al.*, 2008; Manzoni
52 and Porporato, 2009). Hence, a reliable litter decay model is necessary for estimating
53 soil C balance and turnover of ecosystem C (Allison, 2012; Bonan *et al.*, 2013;
54 Wieder *et al.*, 2013; Campbell and Paustian, 2015). In particular, a realistic
55 representation of litter decomposition in land surface models is helpful to decrease the
56 uncertainties in predicted effects of climate change and anthropogenic management
57 on ecosystems (Gholz *et al.*, 2000; Campbell and Paustian, 2015; Luo *et al.*, 2016). As
58 litter decomposition is a very complex process determined by climate (e.g. temperature
59 and moisture), litter quality (e.g. nitrogen (N) concentration), soil nutrients and the
60 physiological characteristics of microorganisms (Lekkerkerk *et al.*, 1990; Prescott,
61 2010; Manzoni *et al.*, 2012; Frey *et al.*, 2013; Sinsabaugh *et al.*, 2013; Garc ía-Palacios
62 *et al.*, 2016), there remain large uncertainties in existing litter decay models (Zhang *et*
63 *al.*, 2008; Bonan *et al.*, 2013; Campbell and Paustian, 2015). Many litter decay
64 models, especially those incorporated in global land surface models, have ignored
65 stoichiometric constraints to microbial processes (Bonan *et al.*, 2013; Cotrufo *et al.*,
66 2013; Wieder *et al.*, 2013; Wieder *et al.*, 2014).

67 Microbial carbon use efficiency (CUE), defined as the ratio of microbial
68 biomass production to material uptake from substrates (Lekkerkerk *et al.*, 1990;
69 Manzoni *et al.*, 2012), is an important parameter constraining litter decay, but it has
70 rarely been represented as a flexible quantity in land surface models. During litter
71 decomposition, only a part of the decomposed litter-C is being transferred into SOM,
72 while the remaining C is being released as CO₂ to the atmosphere by microbial
73 respiration (Campbell and Paustian, 2015; Cotrufo *et al.*, 2015). While CUE is a

74 physiological property of each decomposer community, it also determines the
75 ecosystem-level efficiency at which litter C is transferred into SOM a step further from
76 simple microbial incorporation. We denote this efficiency as carbon use efficiency of
77 litter decomposition (CUE_d). With higher CUE_d , more plant-produced litter is
78 transformed biologically into SOM, and soil C storage can reach higher values (Six *et*
79 *al.*, 2006; Sinsabaugh *et al.*, 2013). In most existing soil biogeochemical models,
80 CUE_d is assumed to be same as microbial CUE and considered as a fixed parameter.
81 The Verberne model (Verberne *et al.*, 1990) assumes for instance $CUE_d \approx 0.25$. In the
82 Yasso model (Liski *et al.*, 2005), the CUE_d is set to 0.2. The CENTURY model sets
83 the CUE_d for decomposition of surface and belowground metabolic litter to 0.55 and
84 0.45, respectively (Parton *et al.*, 1988). In Daisy (Hansen *et al.*, 1991), NCSOIL
85 (Molina *et al.*, 1983) and ICBM (Kätterer and Andr n, 2001), $CUE_d = 0.6$ for the
86 labile litter pools and takes a lower value for recalcitrant substrates. Only a few
87 models account for variable CUE, letting it vary in response to substrate
88 stoichiometry (Schimel and Weintraub, 2003) or temperature (Allison *et al.*, 2010).

89 The increasing evidence for a variable microbial CUE leads to a conceptual
90 CUE model which can explain trends in CUE of microorganisms along stoichiometric
91 gradients (Manzoni *et al.*, 2017). The values of CUE_d used in existing litter decay
92 models are mostly derived from CUE obtained in laboratory studies on microbial
93 physiology or limited observations in certain ecosystems, thus show large variations
94 without a dynamic link to environmental conditions (Parton *et al.*, 1988; Verberne *et*
95 *al.*, 1990; Hansen *et al.*, 1991; Liski *et al.*, 2005; Manzoni *et al.*, 2012). Recent studies
96 (Manzoni *et al.*, 2008, 2012) suggested that the microbial CUE in terrestrial
97 ecosystems ranges from less than 0.1 for wood decomposers to about 0.5 for
98 decomposition of N-rich and high-quality litter. To explain those differences, Manzoni
99 *et al.* (2017) proposed a conceptual model of microbial CUE based on the assumption
100 that decomposers adapt their metabolism (and hence CUE) to maximize their growth
101 rate. This model based on optimality theory links CUE to substrate and decomposers
102 stoichiometry, where the optimal CUE decreases with increasing substrate
103 C-to-nutrient ratio, and increases with soil nutrient availability. The predictions of this

104 theoretical model have been verified by empirical evidence from CUE estimates for
105 different microorganisms in both aquatic and terrestrial ecosystems (Manzoni *et al.*,
106 2017).

107 Besides variable CUE_d , many previous studies have also indicated the
108 necessity for litter decomposition models to consider soil mineral N availability as a
109 driver of litter decomposition rates, in particular under low N availability (Wieder *et*
110 *al.*, 2015; Luo *et al.*, 2016; Averill and Waring, 2018). Microbial biomass is nearly
111 homeostatic (Cleveland and Liptzin, 2007; Franklin *et al.*, 2011; Allison, 2012). When
112 the supply of N from substrates is lower than the demand of microbes to fulfill their
113 specific stoichiometric C:N ratio, microbes will utilize mineral N (immobilization)
114 (Manzoni *et al.*, 2012). Thus, low availability of mineral N can limit microbial activity,
115 and in turn litter decay rate (Manzoni and Porporato 2009; Fujita *et al.*, 2014).

116 Although there are fertilization experiments reporting insignificant or even negative
117 impacts of added N on litter decay rate (Fog, 1988; Hobbie and Vitousek, 2000; Finn
118 *et al.*, 2015), many incubation experiments showed a significant decrease of litter
119 decomposition rate with declining mineral N availability (Recous *et al.*, 1995; Hobbie
120 and Vitousek, 2000; Guenet *et al.*, 2010). Moreover, recent modeling studies have
121 indicated that including the limiting effect of low mineral N on decomposition
122 improved predictions of C and N fluxes (Bonan *et al.*, 2013; Fujita *et al.*, 2014).
123 Therefore, soil mineral N can alter litter C flux by affecting both the litter decay rate
124 and the partition of decayed litter-C (via flexible CUE_d).

125 Some detailed microbial decomposition models actually have included
126 variable microbial CUE and the limitation of low mineral N availability on litter
127 decay rate (Ingwersen *et al.*, 2008; Pagel *et al.*, 2013; Campbell *et al.*, 2016; Huang *et*
128 *al.*, 2018); however, the parameterization and evaluation of these models pose
129 significant challenges due to their complexity and limited verification data (Wieder *et*
130 *al.*, 2014; Campbell and Paustian, 2015). There is still scope for implementing the
131 effects of litter stoichiometry and soil mineral N availability on litter decomposition in
132 litter decay models with more generalizable structure. In particular, it is important to
133 test the role of these effects in models that have already been incorporated into land

134 surface model for long-term and large-scale applications (e.g. CENTURY, Parton *et*
135 *al.*, 1988). In this study, we incorporated flexible CUE_d based on substrate C:N ratios
136 and mineral N limitations into a soil biogeochemical model based on the CENTURY
137 equations to simulate the decomposition and transfer processes of litter-C. The study
138 was organized as follows. First, the new model was calibrated and tested against data
139 from laboratory litter incubation experiments for its ability to capture the effect of
140 variable litter quality and soil mineral N on litter respiration rates (short-term
141 simulations). Second, the model parameterized assuming flexible CUE_d and mineral
142 N limitations was used to explore the consequences of such stoichiometric constraints
143 on the production of soil organic carbon (SOC) (long-term simulations). With these
144 two modeling analyses, we aimed at linking stoichiometric constraints acting on
145 short-term (months to years) decomposition dynamics to their consequences on SOC
146 accumulation occurring at decadal to centennial time scales.

147

148 **2 Materials and methods**

149 2.1 The CENTURY decomposition model

150 The basis of the litter decay model used in this study is the CENTURY model
151 (Fig. 1), a first-order decay model that describes decomposition as a function of
152 substrate availability and quality, clay content, soil moisture and soil temperature
153 (Parton *et al.*, 1988). Most land surface models (e.g. Kucharik *et al.*, 2000; Sitch *et*
154 *al.*, 2003; Krinner *et al.*, 2005) adopted a similar structure to simulate the litter and
155 soil biogeochemical processes. Dead organic matter in CENTURY is separated into
156 structural and metabolic litter and three SOM pools (active, slow, passive) with
157 different turnover times. There is no explicit representation of microbial biomass in
158 CENTURY, instead the biomass of microbes is assumed to be in equilibrium with
159 labile SOM and thus implicitly included in the active SOM pool. When C is being
160 decomposed, a fraction of the decomposed C is respired to the atmosphere and the
161 remaining fraction (CUE_d conceptually equal to microbial CUE) enters the acceptor
162 SOM pool. Three of such fractions are defined to characterize the transfer of C from

163 litter to SOM: CUE_{ma} for transfer of the metabolic litter to the active SOM pool, and
 164 CUE_{sa} and CUE_{ss} for transfer of structural litter to active and slow SOM pools,
 165 respectively (Fig. 1). These fractions are set to be time invariant in the original version
 166 of CENTURY, so that a fixed fraction of decomposed C is retained in the acceptor
 167 pool regardless of environmental conditions and changes in the quality of the donor
 168 pool. The N flows in CENTURY follow the C flows and are equal to the product of C
 169 flow by the N:C ratio of the acceptor SOM pool. N mineralization is defined as the
 170 difference between N obtained from the donor pools and N stoichiometric demand of
 171 the acceptor pool (Parton et al., 1988; Metherell et al., 1993). In this way, net N
 172 mineralization occurs when the donor pool has low C:N ratio, but N is immobilized
 173 (taken up by microbes) when the donor pool has high C:N ratio.

174

175 2.2 Optimal CUE

176 To quantify how microbial CUE varies along gradients of nutrient
 177 availability, it can be hypothesized that microorganisms maximize their growth rate,
 178 and hence their ecological competitiveness, by adapting resource (C and nutrients) use
 179 efficiencies. This follows the growth maximization hypothesis (Mooshammer *et al.*,
 180 2014; Manzoni *et al.*, 2017). Based on this hypothesis, Manzoni *et al.* (2017)
 181 formulated a theoretical model expressing microbial CUE as a function of the
 182 stoichiometric difference between decomposers and their substrate. The CUE for
 183 which growth rate is maximized is the optimal CUE (CUE_{opt}) given by:

$$184 \quad CUE_{opt} = CUE_{max} \times \min \left[1, \frac{CN_D}{CUE_{max}} \times \left(\frac{1}{CN_S} + \frac{I_N}{U_0} \right) \right] \quad (1)$$

185 where CUE_{max} is the maximum microbial CUE (dimensionless) when growth is
 186 limited by C from the organic substrate. CN_D and CN_S are the C:N ratio (in mass,
 187 dimensionless) of decomposer and their substrate, respectively. Although Manzoni *et al.*
 188 *et al.* (2017) indicated that mineral phosphorus (P) could also affect optimal CUE we
 189 only considered N as a limiting nutrient. I_N (g N kg⁻¹ soil) is the maximum rate at
 190 which mineral N can be taken up by microbes, and U_0 (g C kg⁻¹ soil) is the C-limited
 191 uptake rate (corresponding to the decomposition rate at optimal mineral N

192 concentration). When litter C:N is low or soil mineral N is in excess, the second term
 193 in the minimum function (Eq.(1)) is higher than one, and $CUE_{opt} = CUE_{max}$ (C limited
 194 conditions, as in nutrient-rich litter). In contrast, when mineral N is scarce, CUE_{opt}
 195 decreases with increasing substrate C:N ratio (N limited conditions, N-poor litter).
 196 Lack of N in the organic substrates can be compensated by mineral N being
 197 immobilized by microorganisms from the soil solution. Immobilization meets the
 198 nutrient demands as long as it is lower than the maximum supply rate I_N , at which
 199 point microbial CUE starts being down regulated. Thus, for any given C:N ratio in the
 200 substrate, CUE_{opt} increases with inorganic N concentration in the soil solution until
 201 CUE_{max} is reached. It should also be noted that Eq. (1) is interpreted at the microbial
 202 community scale, not for individual organisms.

203

204 2.3 Adaption of the optimal CUE model in the CENTURY model

205 CUE of decomposition (CUE_d) is also assumed to be equivalent to microbial
 206 CUE in this study. Then we followed the theory from Manzoni *et al.* (2017) (Eq. (1))
 207 to parameterize CUE_d during litter decomposition into CENTURY (Fig.1). Due to the
 208 implicit representation of microbial growth in CENTURY, we replaced the original
 209 optimality CUE model (Eq. (1)) by a simpler equation that involves the C:N ratios of
 210 the donor and acceptor pools, rather than microbial C:N ratios:

$$211 \quad CUE_{opt} = CUE_{max} \times \min \left[1, \left(\frac{CN_{lit}}{CN_{SOM}} \right)^a \right] \quad (2)$$

212 where CN_{lit} and CN_{SOM} are the C:N ratio (dimensionless) of litter (metabolic or
 213 structural) and SOM pools (active, slow or passive), respectively. The C:N ratio of
 214 SOM (around 9:1 on a mass basis in CENTURY) is representative of the decomposer
 215 biomass, its value being between the average C:N ratio of soil microbial communities
 216 including fungi and bacteria (7.4:1 in Cleveland and Liptzin, 2007) and the C:N ratio
 217 of soil fungi (13.4:1 in Zhang and Elser, 2017), which are probably largely
 218 responsible for fresh litter decomposition. CUE_{max} (dimensionless) is the maximum
 219 CUE_d achieved when nutrients are not limiting, and it is set to 0.8 based on a
 220 synthesis of observed CUE of soil microbes (Manzoni *et al.*, 2012). The exponent a (g

221 N kg⁻¹ soil) captures the effect of mineral N uptake by microbes on CUE_d. Because
 222 CUE_d is expected to increase with mineral N availability (Eq. (1)), a is assumed to be
 223 a linear function of the mineral N concentration (N_{min} , g N kg⁻¹ soil):

$$224 \quad a = m_1 \times (N_{min} - n_1) \quad (3)$$

225 where m_1 (kg g⁻¹ N) and n_1 (g N kg⁻¹ soil) are two coefficients that need to be
 226 calibrated. Eqs. (2) and (3) modulate the decrease in CUE_d with decreasing litter
 227 quality when mineral N availability changes– the exponent a increases with
 228 increasing mineral N availability, causing an increase in CUE_d at any given litter C:N
 229 ratio. Hence, increasing a mimics an increase in I_N in Eq. (1). Fig. 2a illustrates how
 230 CUE_d from Eq. (2) varies as a function of mineral N concentration, for different
 231 values of litter C:N.

232 Eqs. (2) and (3) were implemented in CENTURY to modify the originally
 233 fixed CUE_d (Fig. 1). With this change, the fractions of C from litter that remain in
 234 SOM are mediated by stoichiometric constraints and mineral N availability, at the
 235 expense of additional parameters to fit. The CUE_d for C transfers between SOC pools
 236 (active, slow and passive) are not modified.

237

238 2.4 Constraint of soil nutrient availability on litter decomposition rate

239 CENTURY is a first-order decay model in which decomposition rates of
 240 metabolic and structural litter are modulated by scaling factors of soil temperature
 241 ($f(tem)$) and moisture ($f(water)$) (Parton *et al.*, 1988). Here, we introduced an
 242 additional mineral N scaling factor ($f(N_{min})$, 0–1, dimensionless) to account for the
 243 limitation imposed by low mineral N availability on litter decay rate ($D(C_{lit})$).

$$244 \quad D(C_{lit}) = C_{lit} \times k \times f(tem) \times f(water) \times f(N_{min}) \quad (4)$$

245 where C_{lit} is the C (g C kg⁻¹ soil) in litter pool (metabolic or structural) and k is the
 246 potential maximum turnover rate (day⁻¹) at optimal soil temperature, moisture and
 247 nutrient conditions.

248 In this study, we assumed that the scaling factor of mineral N increases
 249 linearly with increasing soil mineral N concentration (N_{min} , Eq. (5)) below a threshold
 250 value of $1/m_2$ g N kg⁻¹ soil, where m_2 is a positive coefficient which needs to be

251 calibrated (Fig.2b). The inhibition effect of mineral N only occurs in case of
 252 immobilization ($1/CN_{lit} < CUE_{opt}/CN_{SOM}$). The specific function $f(N_{min})$ can be
 253 expressed as:

$$254 \quad f(N_{min}) = \begin{cases} \min(1, m_2 \times N_{min}), & \frac{CUE_{opt}}{CN_{SOM}} - \frac{1}{CN_{lit}} > 0 \\ 1 & , \frac{CUE_{opt}}{CN_{SOM}} - \frac{1}{CN_{lit}} \leq 0 \end{cases} \quad (5)$$

255 Existing studies have adopted approaches that differ from our definition to
 256 explicitly represent the N inhibition effects on microbial processes (Manzoni and
 257 Porporato, 2009; Bonan *et al.*, 2013; Fujita *et al.*, 2014; Averill and Waring, 2018). In
 258 these previous studies, $f(N_{min})$ was assumed equal to the ratio between immobilized
 259 mineral N and the N deficit for maintaining a stable C:N of decomposer biomass or
 260 other receiver pools. Using the notation of Section 2, this definition of $f(N_{min})$ can be
 261 expressed as:

$$262 \quad f(N_{min}) = \begin{cases} \min\left(1, \frac{m_3 \times N_{min}}{U_0 \times \left(\frac{CUE_{opt}}{CN_{SOM}} - \frac{1}{CN_{lit}}\right)}\right), & \frac{CUE_{opt}}{CN_{SOM}} - \frac{1}{CN_{lit}} > 0 \\ 1 & , \frac{CUE_{opt}}{CN_{SOM}} - \frac{1}{CN_{lit}} \leq 0 \end{cases} \quad (6)$$

263 where m_3 is a coefficient that needs to be optimized. U_0 ($\text{g C kg}^{-1} \text{ soil day}^{-1}$) is the C
 264 uptake rate (equivalent to the litter decomposition rate in absence of leaching) when
 265 soil mineral N is fully adequate for litter decay (i.e. $f(N_{min}) = 1$), and can be calculated
 266 from Eq. (7) as:

$$267 \quad U_0 = C_{lit} \times k \times f(tem) \times f(water) \quad (7)$$

268 In this study, we also tested this formulation in the CENTURY-based model, in
 269 addition to the Eq. (5) (see model M4 in Table A3).

270

271 2.5 Model parameterization and validation

272 To determine the respective impacts of including flexible CUE_d and N
 273 availability constraining decay rates, we built four conceptual litter decay models
 274 (Table 1). Model M0 corresponds to the default CENTURY parameterization of a
 275 fixed CUE_d and no constraints of N availability on litter decay rates ($f(N_{min}) = 1$).
 276 Model M1 accounts for flexibility in CUE from Eq. (2) and N constraints on decay

277 rates by Eq. (5). Model M2 has flexible CUE_d but no N constraints on decay rates
278 ($f(N_{min})= 1$). Model M3 has N constraints on decay rates but a fixed CUE_d (Table 1).
279 All of these four models are run at a daily time step. Finally, model M4 also accounts
280 for flexibility in CUE and N constraints on decays (Table A3), but it uses Eq. (6) to
281 represent the N constraints on decays rate rather than Eq. (2). Results from model M4
282 are presented in the main text, but only shown in the Appendix. This range of models
283 allows identifying which mechanisms are at play during decomposition – flexible
284 CUE_d only (M3), mineral N limitation only (M2), both mechanisms (M1, M4), or
285 none (M0).

286 For calibrating model parameters and evaluation of their results, we collected
287 data of laboratory litter incubation experiments from Recous *et al.* (1995) (5
288 experiments) and Guenet *et al.* (2010) (9 experiments, Table A2). The incubation
289 experiments of Recous *et al.* (1995) and Guenet *et al.* (2010) continued 80 and 124
290 days, respectively. Recous *et al.* (1995) used corn residues (C:N = 130) and Guenet *et*
291 *al.* (2010) used wheat straw (C:N = 44) in their incubation experiments. The C:N
292 ratios of those corn residue and wheat straw span the range of litter C:N ratios among
293 different ecosystems (Harmon *et al.*, 2009; Brovkin *et al.*, 2012; Manzoni *et al.*, 2010).
294 In the incubation experiments, plant litter was firstly cut into fine fragments before it
295 was mixed with mineral soil. Soil temperature and moisture condition were kept
296 constant during the experiment. Respired C from the incubated litter and SOC, as well
297 as the soil mineral N concentrations were measured continuously across the
298 incubation period. To distinguish the litter- and SOC-derived CO_2 flux, Guenet *et al.*
299 (2010) used straw from wheat grown under ^{13}C labeled CO_2 and they are therefore
300 able to track the CO_2 coming from litter and the CO_2 coming from soil. In the
301 experiments by Recous *et al.* (1995), litter-derived CO_2 flux is calculated as the
302 difference in CO_2 flux between the incubation samples with both soil and litter, and
303 the control samples without added litter. More detailed information about the
304 incubation experiments of Recous *et al.* (1995) and Guenet *et al.* (2010) can be found
305 in Table A2.

306 The initial C storage and C:N ratios of litter and SOM pool, as well as soil

307 temperature and moisture condition for decomposition in all of the five versions of the
308 model (M0-M4) were set based on observations (Table A2). Plant litter was firstly
309 separated into metabolic and structural litter pools based on its lignin to C ratio (LC_{lit} ,
310 dimensionless). The fraction of metabolic litter-C (f_m , 0-1, dimensionless) is
311 calculated by:

$$312 \quad f_m = f_{max} - m_4 \times LC_{lit} \quad (8)$$

313 where m_4 is a coefficient to be calibrated; $f_{max}=0.85$ is the maximum fraction of
314 metabolic litter (i.e., the default value in CENTURY; Parton *et al.*, 1988). The
315 fraction of structural litter-C is thus $1-f_m$. The C:N ratios of both metabolic and
316 structural pools are assumed to be equal to the C:N ratio of litter input.

317 In M1 and M3 models, the observed mineral N concentrations across the
318 incubation period were used to calculate the daily N inhibition effect (Eq. (5)). The
319 observed cumulative respired litter-C (g C kg^{-1} soil) measured in the incubation
320 experiments was used to calibrate the model parameter values. Moreover, to quantify
321 the simulated CO_2 flux derived from the litter, we also performed a set of control
322 simulations with only SOM (initial litter pools were set to 0 g kg^{-1} soil) using the four
323 model versions. The simulated litter-derived CO_2 flux is calculated as the difference
324 in CO_2 flux between the simulation with both litter and SOM inputs and the
325 simulation with only SOM input.

326 Parameter calibration was performed for each model with the shuffled
327 complex evolution (SCE) algorithm developed by Duan *et al.*, (1993). The SCE
328 algorithm relies on a synthesis of four concepts that have proved successful for global
329 optimization: combination of probabilistic and deterministic approaches; clustering;
330 systematic evolution of a complex of points spanning the space in the direction of
331 global improvement and competitive evolution(Duan *et al.*, 1993). A more detailed
332 description of this SCE optimization method can be found in Duan *et al.* (1993, 1994).
333 In this study, the RMSE (root mean square error, Eq.(9)) between simulated and
334 measured cumulative respired litter-C (%) on all observation days (Table A2) of each
335 incubation experiment was used as the objective function, and the parameters
336 minimizing RMSE between simulated and observed cumulative respired litter-C were

337 regarded as optimal parameter values.

$$338 \quad RMSE = \sqrt{\left(\frac{\sum_{i=1}^n (Sim_i - Obs_i)^2}{n}\right)} \quad (9)$$

339 where n is the number of observation days, Sim_i and Obs_i (%) are the simulated
340 and observed percent of cumulative litter-C flux on day i , respectively.

341 We used leave-one-out cross-validation (Kearns and Ron, 1997; Tramontana
342 *et al.*, 2016) to evaluate each of the four models (i.e. M0-M3), a cross validation
343 method used when data is scarce. The number of cross-validations corresponds to the
344 number of incubation experiments (14). Each time, one of the 14 incubation
345 experiments was left out as the validation sample, and the remaining 13 experiments
346 were used to train model parameters. In addition to RMSE, we also adopted the
347 Akaike Information Criterion (AIC, Bozdogan, 1987, Eq. (10)) to determine the
348 relative quality of the four version models on estimating cumulative respired litter-C.

$$349 \quad AIC = n \times \ln\left(\frac{\sum_{i=1}^n (Sim_i - Obs_i)^2}{n}\right) + 2n_p \quad (10)$$

350 where n_p is the number of model parameters. The evaluation of AIC is important here
351 because depending on the model version, different numbers of parameters have to be
352 determined (Table 1), requiring us to weigh both model accuracy and robustness.

353 Note that the turnover times of SOM pools (active, slow and passive) used in
354 this study are obtained from the ORCHIDEE-MICT (v8.4.1, Guimberteau *et al.*,
355 2018). The turnover times of litter pools (metabolic and structural), as well as the
356 coefficient m_d in Eq. (8) were optimized against the observed cumulative respired
357 litter-C from all of the 14 incubation experiments using the M0 and M1 models (Table
358 A3). A previous study has shown that litter decomposability is negatively correlated to
359 its physical size (for example, Tuomi *et al.*, 2011). Therefore, the turnover times of
360 the fine litter fragments used in the incubation experiments of Recous *et al.* (1995)
361 and Guenet *et al.* (2010) are expected to be shorter than the values set in
362 ORCHIDEE-MICT, which are representative of the turnover times of natural plant
363 residues. In addition, the mixing of soil and litter particle in the incubation experiment
364 likely enhances decomposition as spatial disconnection of decomposer and substrate,
365 which can occur under natural soil conditions (Barnes *et al.*, 2012; Hewins *et al.*,

366 2013), is prevented. The calibrated turnover times of the metabolic and structural
367 pools and the value of m_4 in Eq. (8) are 3.5 and 30 days and 0.5, respectively.

368

369 2.6 Impacts of litter stoichiometry and mineral N availability on SOM accumulation

370 We used the model M1, with flexible CUE_d and decomposition rate function
371 of available N to study the impacts of litter stoichiometry (C:N ratio) and soil mineral
372 N availability on the formation and accumulation of SOM. In total, 24 idealized
373 simulation experiments with different values of litter C:N ratios and soil mineral N
374 availabilities were conducted (Table A4). The assumed litter C:N ratios (CN_{lit}) of 10,
375 15, 30, 60, 120 and 200 span the variation among most natural substrates and soil
376 amendments from organic matter input in agriculture (Harmon et al., 2009; Brovkin *et*
377 *al.*, 2012; Manzoni *et al.*, 2010). The assumed range of mineral N availability (N_{min})
378 of 0.001, 0.005, 0.01 and 0.05 g N kg⁻¹ soil span the observed concentrations of soil
379 mineral N in major terrestrial ecosystems (Metherall *et al.*, 1993).

380 In each simulation experiment, model M1 was run for 5000 years to bring the
381 litter and SOM pools in equilibrium with the prescribed litter input flux. The daily
382 input rate of plant litter was set to 0.006 g C kg⁻¹ soil day⁻¹, and the initial C stock of
383 litter and SOM pools were all set to be 0 g C kg⁻¹ soil. During the simulation, soil
384 temperature and soil water content were assumed to be 25 °C and 60% of water
385 holding capacity, respectively. We emphasized that our goal with this simplified
386 scenario was to single out the effects of stoichiometric constraints, not to simulate the
387 effects of a realistic climatic regime. Parameter values for M1 (with $m_1 = 0.54$, $n_1 =$
388 0.50 and $m_2 = 296.8$) used here were optimized based on all of the 14 incubation
389 experiments from Recous *et al.* (1995) and Guenet *et al.* (2010) (see above). More
390 detailed information about the specific settings of our simulation experiments can be
391 found in Table A4.

392

393 **3 Results**

394 3.1 Evaluation of different models

395 Results of leave-one-out cross-validation suggest that model M1 provides
396 more accurate prediction of cumulative respired litter-C than other models (Fig. 3).
397 The differences between simulated and observed cumulative respired litter-C from
398 M1 are mostly less than 6% for over 93% of the data (Fig. A1b). The average RMSE
399 of predicted cumulative respired litter-C from M1 (3.0%) is lower than that of model
400 M0 (4.1%). Models M2 and M3 have slightly lower RMSE values than M0 (3.7% and
401 3.8%, respectively) but perform worse than M1 (Fig.4). However, the average AIC of
402 all the models are comparable, suggesting that models with more fitted parameters do
403 not over-fit the observations (Fig. 4).

404 Model M1 captures the differences in respiration rates due to different C:N
405 ratios of substrate and varying levels of mineral N availability across the 14
406 incubation experiments (Fig.5). While model M3 can reproduce the observed effect of
407 soil mineral N availability on litter respirations rates (Fig.5d), it underestimates the
408 cumulative respired CO₂ from low quality litter ($CN_{lit} = 130$) at high mineral N
409 concentrations ($> 0.04 \text{ g N kg}^{-1} \text{ soil}$). Models M0 and M2 cannot represent the effects
410 of soil mineral N on litter respiration rate (Figs.5a, c), and their predictions are more
411 biased from the observed values compared to M1. The Model M4, which uses the
412 alternative formulation for N constraints on litter decay (Eq. (6)), reproduces the
413 different respiration rates of substrates with contrasting C:N ratios and at different
414 levels of mineral N availability (Fig. A2), but with a slightly higher average RMSE of
415 cumulative respired litter-C than model M1.

416 The predicted CUE_d and the limitation effects of soil mineral N availability
417 on litter decay rate ($f(N_{min})$ function Eq. (5)) are different among the four tested
418 models (Fig. A3). In models M0 and M3, which used a fixed CUE_d , the fitted values
419 of CUE_d calculated with optimized parameters during the incubation period are about
420 0.57 and 0.54, respectively (Figs. A3a, d). In models M1 and M2, the CUE_d varies
421 with the C:N ratios of plant litter, and is only slightly affected by soil mineral N
422 concentrations (Figs. A3b, c). For very low quality litter with a C:N ratio of 130, the
423 CUE_d in models M1 and M2 are 0.40 and 0.44, respectively, which are lower than for
424 better quality litter with C:N ratio of 44 (approximately 0.55 and 0.56 in M1 and M2,

425 respectively). Models M0 and M2 do not include the N inhibition effects on litter
426 decay rate, thus the $f(N_{min})$ in these two models is always 1 (Figs. A3e, g). In M1 and
427 M3, the N inhibition effect changes with both the litter C:N ratio and the mineral N
428 availability (Figs. A3f, h).

429 CUE_d from Eq. (2) calibrated with the data of the two incubation
430 experiments, decreases with increasing CN_{lit}/CN_{SOM} (Fig. 6). The average CUE_d value
431 is larger than the average of data compiled for microbial CUE of litter decomposition
432 in terrestrial ecosystems by Manzoni *et al.* (2017). This is shown by the gray circles in
433 Fig. 6. Our optimized values of CUE_d for a given C:N ratio are more comparable with
434 microbial CUE observed in incubations of soil mixed with litter (Gilmour and
435 Gilmour, 1985; Devêre and Horváth, 2000; Thiet *et al.*, 2006), shown as black
436 squares in Fig. 6.

437

438 3.2 The effect of litter quality vs quantity on equilibrium SOM stocks

439 Model M1 predicts that the size of the SOM pool at equilibrium is mainly
440 determined by litter stoichiometry, with a minor effect of soil mineral N (Fig. 7). The
441 lower C:N ratio of litter is, the higher equilibrium SOC stock. For litter with a specific
442 C:N ratio, high soil mineral N concentration (e.g. above 0.05 g N kg⁻¹ soil) generally
443 produces a slightly larger equilibrium SOC stock than a low mineral N concentration
444 (Fig. 7). Further analysis suggests that the SOC at equilibrium increases with
445 decreasing litter C:N because the SOC pool is positively related to the CUE_d ; however
446 the limitation of soil mineral N on litter decomposition rate almost shows no impact
447 on SOC (Fig. A4).

448

449 **4 Discussion**

450 We hypothesized that stoichiometric constraints (flexible CUE_d or inhibition
451 of decomposition under N limited conditions) played a role in shaping the trajectory
452 of litter decomposition, with potential consequences on predicted SOC stocks. Our
453 results suggest that flexible CUE_d and inhibition effects of soil mineral N on litter

454 decay rate improve prediction of litter decomposition when using a modified version
455 of CENTURY model (denoted as M1). Evaluation of the model M1 using data from
456 incubation experiments indicate that this modified model captures the effects of both
457 variable litter quality (stoichiometry) and mineral N availability on respiration rates
458 (Fig. 5), without strongly inflating the complexity of CENTURY (Table 1). As the
459 stoichiometric constraints are implemented in the generalizable and widely used
460 structure of CENTURY and require only three parameters to be calibrated, they can
461 also be easily implemented into land surface models for large spatial scale
462 applications.

463 Accurately representing N control of microbial processes during litter
464 decomposition has been suggested to be important for modeling the connection
465 between the litter inputs, CUE_d , and soil C dynamics (Gerber *et al.*, 2010; Manzoni *et al.*,
466 2012; Cotrufo *et al.*, 2013; Sinsabaugh *et al.*, 2013). In model M1, soil mineral N
467 affects the litter-C flux via two mutually different pathways: (1) mineral N availability
468 affects the litter decay rate and (2) flexible CUE_d determining the partition of
469 decomposed C into SOC products and respired CO_2 (Fig. 1). Therefore, an increase in
470 soil mineral N concentration enhances litter decay rates, which alone will increase the
471 flux of litter-derived CO_2 (Eq. (5) and Fig. A5). However, as higher N concentration
472 also results in a higher CUE_d (Eq. (2)), more C is transferred to SOC and less C is
473 respired. In this way, SOC is predicted to accumulate with increasing mineral N
474 availability when using model M1 (Fig. 7). In addition, the model M4, which is
475 comparable to model M1 but uses an alternative formulation for N effects on the
476 decomposition rate (Eq. (6)), performed slightly worse than model M1 (Fig. A2).
477 Arguably, Eq. (6) represents the underlying mechanisms of N inhibition effects
478 (Manzoni *et al.*, 2009; Bonan *et al.*, 2013; Fujita *et al.*, 2014; Averill and Waring,
479 2018) better than Eq. (5) and due to the minor differences in RMSE and AIC (Figure
480 A2b) between these formulations it can serve as an alternative to M1.

481 Our results indicate that the observed diversity of responses of litter
482 respiration rate to mineral N additions (Hobbie and Vitousek, 2000; Guenet *et al.*,
483 2010; Janssens *et al.*, 2010) is likely due to the combined effects of changes in litter

484 decay rate and CUE_d (Fig. A5). Thus, N addition effects can differ among fertilization
485 experiments if litter quality and background N availability vary. In addition to altering
486 litter decay rate and CUE_d , mineral N addition can induce abiotic formation of
487 compounds that resist microbial attack, inhibit oxidative enzymes involved in lignin
488 degradation, stimulate microbial biomass production early in decomposition, or lead
489 to the accumulation of microbial residues that are resistant to decay (Fog, 1988;
490 Hobbie, 2015). All these effects might decrease litter respiration rate by inhibiting the
491 decomposition process, but have not been considered in our current model.

492 This study provides insights on processes leading to increased SOM
493 sequestration. Enhancing the efficiency at which plant residuals are transformed into
494 stable SOM has been suggested as an effective strategy to sequester C in soil (Prescott,
495 2010; Cotrufo *et al.*, 2013). Simulation results from our model suggest a positive
496 linear relationship between equilibrium SOC stock and CUE of decomposed litter
497 (Fig. A4), in line with the earlier findings with a similar model (for example Frey *et al.*
498 2013). In fact, with linear models such as CENTURY it can be shown that the steady
499 state SOC scales linearly with CUE, different from nonlinear models predicting that
500 higher CUE can trigger SOC loss (Allison *et al.*, 2010). Our model goes beyond
501 earlier attempts (Bonan *et al.*, 2013; Fujita *et al.*, 2014; Averill and Waring, 2018) by
502 adapting the optimal metabolic regulation hypothesis of Manzoni *et al.* (2017) to link
503 CUE, litter quality and SOM formation in a process-oriented way.

504 The importance of litter quality for SOM formation as found here is in line
505 with recent experiments (Bahri *et al.*, 2008; Rubino *et al.*, 2010; Walela *et al.*, 2014)
506 and modeling studies (Grandy and Neff, 2008; Cotrufo *et al.*, 2013). SOM is mainly
507 formed through the partial decomposition of plant debris by microorganisms (Paul,
508 2007; Knicker, 2011; Cotrufo *et al.*, 2013). The conceptual model developed by
509 Cotrufo *et al.* (2013) suggested that although labile litter was decomposed faster than
510 recalcitrant litter, a higher fraction of this labile litter-C would be incorporated into
511 microbial biomass and subsequently incorporated into SOM pool (corresponding to a
512 higher CUE_d). Therefore, labile litter inputs tend to form a larger SOM pool than the
513 poor-quality (high C:N ratio) litter that is generally used by microbes at lower

514 efficiency. Our simulations of decomposition process of plant litter with different C:N
515 ratios also suggest that litter of good quality (with low C:N ratio) can induce a larger
516 SOM pool than the poor-quality litter (Fig. 7). CUE_d plays a more important role than
517 the inhibition effect of low mineral N concentration in determining the size of the
518 stable SOM pool (Fig. A4).

519 The predictions from Cotrufo *et al.* (2013) and this study contrasts with the
520 conventional hypothesis whereby the poor-quality litter with low decay rate and small
521 CUE_d are preferential to be accumulated in SOM (Berg and Mcclaugherty, 2008;
522 Walela *et al.*, 2014). This view of SOM stabilization, however, seems to apply to
523 N-limited systems with high C:N litter and where microbial remains are recalcitrant to
524 decomposition (e.g., boreal forests) – in these systems SOC does accumulate despite
525 its low quality (Kyaschenko *et al.* 2017). Moreover, one could argue that higher CUE_d
526 implies larger microbial biomass, allowing faster decomposition (Allison *et al.*, 2010).
527 These feedbacks between microbial biomass and decomposition rate were not
528 implemented in the current model, but could offer additional flexibility – again at the
529 expense of more difficult model parameterization.

530 The CUE_d formulation from Eq. (2) with parameters calibrated from the two
531 sets of incubation experiments might underestimate the impacts of litter quality on
532 microbial CUE under natural conditions, in particular in case of SOM decomposition.
533 In both incubation experiments, litter is firstly cut into fine fragments and then fully
534 mixed with mineral soil (Recous *et al.*, 1995; Guenet *et al.*, 2010). Thus, the nutrient
535 accessibility, air permeability and some other environmental factors (e.g. pH) of
536 incubated litter are different from those of decaying litter in more natural,
537 heterogeneous soil conditions. Those different decomposition conditions might be
538 responsible for the differences observed in Fig. 6 between our CUE estimates and
539 previously reported values. We speculate that more heterogeneous conditions reduce
540 nutrient availability and thus might cause lower CUE. Similarly, CUE of surface litter
541 decomposers may be lower than we estimated (Fig. 6), because litter not mixed with
542 soil is probably subject to stronger nutrient limitation.

543 Further validation and development of our model are still necessary to

544 decrease the model uncertainties. Soil mineral N which affects both litter decay rate
545 and CUE of decayed litter is seldom monitored in litter incubation experiments (e.g.
546 Walela *et al.*, 2014; Stewart *et al.*, 2015) and field litter decay experiments (e.g. Gholz
547 *et al.*, 2000; Harmon *et al.*, 2009), with few exceptions (Recous *et al.*, 1995; Guenet *et*
548 *al.*, 2010). An increasing number of land surface models (Wang *et al.*, 2010; Zaehle *et*
549 *al.*, 2014; Goll *et al.*, 2017) have representations of the terrestrial N cycle. By
550 incorporating our newly developed formulations of CUE_d and $f(N_{min})$ in these land
551 surface models that simulate the dynamics of soil mineral N concentration, it will be
552 possible to test and validate our developments with more extensive data from
553 laboratory and field experiments. Moreover, similar to N, P has also been suggested as
554 another important factor for litter decomposition and SOM formation (Güsewell and
555 Verhoeven, 2006; Talkner *et al.*, 2009; Manzoni *et al.*, 2010; Prescott, 2010),
556 especially in regions with highly weathered soil (Goll *et al.*, 2012, 2017; Yang *et al.*,
557 2014). So it might be necessary to include the effects of P on litter decay rate and
558 CUE_d into our model for further decrease the simulation uncertainties.

559

560 **5 Conclusions**

561 By adapting the hypothesis of optimal microbial CUE proposed by Manzoni
562 *et al.* (2017) for use in a CENTURY-based model and also introducing a N scaling
563 function to represent the limits of mineral N availability on litter decay rate, we
564 developed a simple but effective litter decomposition model that accounts for key
565 stoichiometric constraints during decomposition. Validation using observation data
566 obtained from laboratory incubation experiments indicated that our model could well
567 predict the respiration rates of litter in different qualities at various levels of mineral N
568 availability. Idealized simulations using our model revealed that the quality of litter
569 inputs plays an important role in determining the soil C stock at equilibrium.
570 High-quality litter (i.e. with low C:N ratio) tends to form a larger SOM pool as it can
571 be more efficiently utilized by microorganisms than recalcitrant litter (e.g. high C:N
572 ratio). Overall, the developed model captures the microbial mechanisms mediating

573 litter stoichiometry and soil mineral N effects on litter decomposition and SOM
574 formation. Due to the simple and generalizable structure of our model, it can be
575 incorporated into existing land surface models for further long-term and large spatial
576 scale applications.
577

578 **Code and data availability**

579 The CENTURY-based model used here is programmed in MATLAB
580 language. The source code is available online
581 (https://github.com/hchzhang/CENYUTY_CUE/tree/v1.0, DOI:
582 10.5281/zenodo.1307384). All the data used in this study can be obtained from
583 published literatures. Specific references of these data can be found in section 2.5.

584

585 **Competing interests**

586 The authors declare that they have no conflict of interest.

587

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594 **References**

- 595 Allison, S. D., Wallenstein, M. D., and Bradford, M. A.: Soil-carbon response to warming
596 dependent on microbial physiology, *Nature Geoscience*, 3, 336, 10.1038/ngeo846, 2010.
- 597 Allison, S. D.: A trait-based approach for modelling microbial litter decomposition, *Ecology*
598 *letters*, 15, 1058-1070, 10.1111/j.1461-0248.2012.01807.x, 2012.
- 599 Averill, C., and Waring, B.: Nitrogen limitation of decomposition and decay: How can it occur?,
600 *Glob Chang Biol*, 24, 1417-1427, 10.1111/gcb.13980, 2018.
- 601 Bahri, H., Rasse, D.P., Rumpel, C., Dignac, M.F., Bardoux, G., Mariotti, A. (2008) Lignin
602 degradation during a laboratory incubation followed by ¹³C isotope analysis. *Soil Biology*
603 *and Biochemistry*, 40, 1916-1922.
- 604 Barnes, P. W., Throop, H. L., Hewins, D. B., Abbene, M. L. Archer, S. R.: Soil coverage reduces
605 photodegradation and promotes the development of soil microbial films on dryland leaf litter,
606 *Ecosystems*, 15, 311-321, 2012.
- 607 Berg, B., and Mcclaugherty, C.: *Plant Litter. Decomposition, Humus Formation, Carbon*
608 *Sequestration*, Springer Verlag, Heidelberg, 2008.
- 609 Bonan, G. B., Hartman, M. D., Parton, W. J., and Wieder, W. R.: Evaluating litter decomposition
610 in earth system models with long-term litterbag experiments: an example using the
611 *Community Land Model version 4 (CLM4)*, *Glob Chang Biol*, 19, 957-974.
- 612 Bozdogan, H.: Model selection and Akaike's Information Criterion (AIC): The general theory and
613 its analytical extensions, *Psychometrika*, 52, 345-370, 1987.
- 614 Brovkin, V., Bodegom, P. M. V., Kleinen, T. and Wirth, C.: Plant-driven variation in
615 decomposition rates improves projections of global litter stock distribution. *Biogeosciences*
616 *Discussions*, 8, 565-576, 2011.
- 617 Campbell, E. E., and Paustian, K.: Current developments in soil organic matter modeling and the
618 expansion of model applications: a review, *Environmental Research Letters*, 10, 123004,
619 10.1088/1748-9326/10/12/123004, 2015.
- 620 Campbell, E. E., Parton, W. J., Soong, J. L., Paustian, K., Hobbs, N. T., and Cotrufo, M. F.: Using
621 litter chemistry controls on microbial processes to partition litter carbon fluxes with the Litter
622 *Decomposition and Leaching (LIDEL) model*, *Soil Biology and Biochemistry*, 100, 160-174,

623 10.1016/j.soilbio.2016.06.007, 2016.

624 Cleveland, C. C., and Liptzin, D.: C:N:P stoichiometry in soil: is there a “Redfield ratio” for the
625 microbial biomass?, *Biogeochemistry*, 85, 235-252, 10.1007/s10533-007-9132-0, 2007.

626 Cotrufo, M. F., Wallenstein, M. D., Boot, C. M., Deneff, K., and Paul, E.: The Microbial
627 Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition
628 with soil organic matter stabilization: do labile plant inputs form stable soil organic matter?,
629 *Glob Chang Biol*, 19, 988-995, 10.1111/gcb.12113, 2013.

630 Cotrufo, M. F., Soong, J. L., Horton, A. J., Campbell, E. E., Haddix, Michelle L., Wall, D. H., and
631 Parton, W. J.: Formation of soil organic matter via biochemical and physical pathways of
632 litter mass loss, *Nature Geoscience*, 8, 776-779, 10.1038/ngeo2520, 2015.

633 Devêre, O. C., and Horwáth, W. R.: Decomposition of rice straw and microbial carbon use
634 efficiency under different soil temperatures and moistures, *Soil Biology & Biochemistry*, 32,
635 1773-1785, 2000.

636 Duan, Q., Sorooshian, S., and Gupta, V. K.: Optimal use of the SCE-UA global optimization
637 method for calibrating watershed models, *Journal of Hydrology*, 158, 265-284, 1994.

638 Duan, Q. Y., Gupta, V. K., and Sorooshian, S.: Shuffled complex evolution approach for effective
639 and efficient global minimization, *Plenum Press*, 501-521 pp., 1993.

640 Finn, D., Page, K., Catton, K., Strounina, E., Kienzle, M., Robertson, F., Armstrong, R., and Dalal,
641 R.: Effect of added nitrogen on plant litter decomposition depends on initial soil carbon and
642 nitrogen stoichiometry, *Soil Biology and Biochemistry*, 91, 160-168,
643 10.1016/j.soilbio.2015.09.001, 2015.

644 Fog, K.: The effect of added nitrogen on the rate of decomposition of organic matter, *Biological*
645 *Reviews*, 63, 433-462, 1988.

646 Franklin, O., Hall, E. K., Kaiser, C., Battin, T. J., and Richter, A.: Optimization of Biomass
647 Composition Explains Microbial Growth-Stoichiometry Relationships, *American Naturalist*,
648 177, E29, 2011.

649 Frey, S. D., Lee, J., Melillo, J. M., and Six, J.: The temperature response of soil microbial
650 efficiency and its feedback to climate, *Nature Climate Change*, 3, 395-398,
651 10.1038/nclimate1796, 2013.

652 Fujita, Y., Witte, J.-P. M., and van Bodegom, P. M.: Incorporating microbial ecology concepts into

653 global soil mineralization models to improve predictions of carbon and nitrogen fluxes,
654 *Global Biogeochemical Cycles*, 28, 223-238, 2014.

655 Garc ía-Palacios, P., McKie, B. G., Handa, I. T., Frainer, A., H ätenschwiler, S., and Jones, H.: The
656 importance of litter traits and decomposers for litter decomposition: a comparison of aquatic
657 and terrestrial ecosystems within and across biomes, *Functional Ecology*, 30, 819-829,
658 10.1111/1365-2435.12589, 2016.

659 Gerber, S., Hedin, L. O., Oppenheimer, M., Pacala, S. W., and Shevliakova, E.: Nitrogen cycling
660 and feedbacks in a global dynamic land model, GB1001, *Global Biogeochemical Cycles*,
661 2010.

662 Gholz, H. L., Wedin, D. A., Smitherman, S. M., Harmon, M. E., and Parton, W. J.: Long-term
663 dynamics of pine and hardwood litter in contrasting environments: toward a global model of
664 decomposition, *Global Change Biology*, 6, 751-765, 2000.

665 Gilmour, C. M., and Gilmour, J. T.: Assimilation of carbon by the soil biomass, *Plant & Soil*, 86,
666 101-112, 1985.

667 Goll, D. S., Brovkin, V., Parida, B. R., and Reick, C. H.: Nutrient limitation reduces land carbon
668 uptake in simulations with a model of combined carbon, nitrogen and phosphorus cycling,
669 *Biogeosciences Discussions*, 9, 3547-3569, 2012.

670 Goll, D. S., Vuichard, N., Maignan, F., Jornet-Puig, A., Sardans, J., Violette, A., Peng, S., Sun, Y.,
671 Kvakic, M., and Guimberteau, M.: A representation of the phosphorus cycle for ORCHIDEE
672 (revision 4520), *Geoscientific Model Development*, 10, 3745-3770, 2017.

673 Grandy, A. S., and Neff, J. C.: Molecular C dynamics downstream: the biochemical decomposition
674 sequence and its impact on soil organic matter structure and function, *Science of the Total
675 Environment*, 404, 297-307, 2008.

676 Guenet, B., Neill, C., Bardoux, G., and Abbadie, L.: Is there a linear relationship between priming
677 effect intensity and the amount of organic matter input?, *Applied Soil Ecology*, 46, 436-442,
678 10.1016/j.apsoil.2010.09.006, 2010.

679 Guimberteau, M., Zhu, D., Maignan, F., Huang, Y., Yue, C., Dantec-N édec, S., Ottl é C.,
680 Jornet-Puig, A., Bastos, A., Laurent, P., Goll, D., Bowring, S., Chang, J., Guenet, B., Tifafi,
681 M., Peng, S., Krinner, G., Ducharne, A., Wang, F., Wang, T., Wang, X., Wang, Y., Yin, Z.,
682 Lauerwald, R., Joetzjer, E., Qiu, C., Kim, H. & Ciais, P. (2018) ORCHIDEE-MICT (v8.4.1),

683 a land surface model for the high-latitudes: model description and validation. *Geoscientific*
684 *Model Development*, 11, 121-163.

685 Güsewell, S., and Verhoeven, J. T. A.: Litter N:P ratios indicate whether N or P limits the
686 decomposability of graminoid leaf litter, *Plant & Soil*, 287, 131-143, 2006.

687 Hansen, S., Jensen, H. E., Nielsen, N. E., and Svendsen, H.: Simulation of nitrogen dynamics and
688 biomass production in winter wheat using the Danish simulation model DAISY, *Fertilizer*
689 *Research*, 27, 245-259, 1991.

690 Harmon, M. E., Silver, W. L., Fasth, B., Chen, H. U. A., Burke, I. C., Parton, W. J., Hart, S. C.,
691 and Currie, W. S.: Long-term patterns of mass loss during the decomposition of leaf and fine
692 root litter: an intersite comparison, *Global Change Biology*, 15, 1320-1338,
693 10.1111/j.1365-2486.2008.01837.x, 2009.

694 Hewins, D. B., Archer, S. R., Okin, G. S., McCulley R. L., Throop, H. L.: Soil-litter mixing
695 accelerates decomposition in a Chihuahuan Desert grassland, *Ecosystems*, 16, 183-195, 2013.

696 Hobbie, S.E.: Plant species effects on nutrient cycling: revisiting litter feedbacks. *Trends Ecol*
697 *Evol*, 30, 357-363, 2015.

698 Hobbie, S. E., and Vitousek, P. M.: Nutrient limitation of decomposition in Hawaiian forests,
699 *Ecology*, 81, 1867-1877, 2000.

700 Huang, Y., Guenet, B., Ciais, P., Janssens, I. A., Soong, J. L., Wang, Y., Goll, D., Blagodatskaya,
701 E., and Huang, Y.: ORCHIMIC (v1.0), a microbe-driven model for soil organic matter
702 decomposition designed for large-scale applications, *Geoscientific Model Development*
703 *Discussions*, 1-48, 10.5194/gmd-2017-325, 2018.

704 Ingwersen, J., Poll, C., Streck, T., and Kandeler, E.: Micro-scale modelling of carbon turnover
705 driven by microbial succession at a biogeochemical interface, *Soil Biology and Biochemistry*,
706 40, 864-878, 10.1016/j.soilbio.2007.10.018, 2008.

707 Janssens, I. A., Dieleman, W., Luysaert, S., Subke, J. A., Reichstein, M., Ceulemans, R., Ciais, P.,
708 Dolman, A. J., Grace, J., Matteucci, G., Papale, D., Piao, S. L., Schulze, E. D., Tang, J., and
709 Law, B. E.: Reduction of forest soil respiration in response to nitrogen deposition, *Nature*
710 *Geoscience*, 3, 315-322, 10.1038/ngeo844, 2010.

711 Käterer, T., and Andr n, O.: The ICBM family of analytically solved models of soil carbon,
712 nitrogen and microbial biomass dynamics — descriptions and application examples,

713 Ecological Modelling, 136, 191-207, 2001.

714 Kearns, M. & Ron, D. (1997) Algorithmic stability and sanity-check bounds for leave-one-out
715 cross-validation. *Neural Computation*, 11, 1427-1453.

716 Knicker, H.: Soil organic N - An under-rated player for C sequestration in soils?, *Soil Biology &*
717 *Biochemistry*, 43, 1118-1129, 2011.

718 Krinner, G., Viovy, N., de Noblet-Ducoudré N., Ogée, J., Polcher, J., Friedlingstein, P., Ciais, P.,
719 Sitch, S., and Prentice, I. C.: A dynamic global vegetation model for studies of the coupled
720 atmosphere-biosphere system, *Global Biogeochemical Cycles*, 19,
721 doi:10.1029/2003GB002199, 10.1029/2003gb002199, 2005.

722 Kucharik, C. J., Foley, J. A., Delire, C., Fisher, V. A., Coe, M. T., Lenters, J. D., Young-Molling,
723 C., Ramankutty, N., Norman, J. M., and Gower, S. T.: Testing the performance of a dynamic
724 global ecosystem model: Water balance, carbon balance, and vegetation structure, *Global*
725 *Biogeochemical Cycles*, 14, 795-825, 10.1029/1999gb001138, 2000.

726 Kyaschenko, J., Clemmensen, K. E., Karlton, E., and Lindahl, B. D.: Below-ground organic
727 matter accumulation along a boreal forest fertility gradient relates to guild interaction within
728 fungal communities, *Ecology letters*, 20, 1546-1555, 10.1111/ele.12862, 2017.

729 Lekkerkerk, L., Lundkvist, H., Ågren, G. I., Ekbohm, G., and Bosatta, E.: Decomposition of
730 heterogeneous substrates; An experimental investigation of a hypothesis on substrate and
731 microbial properties, *Soil Biology & Biochemistry*, 22, 161-167, 1990.

732 Liski, J., Palosuo, T., Peltoniemi, M., and Sievänen, R.: Carbon and decomposition model Yasso
733 for forest soils, *Ecological Modelling*, 189, 168-182, 10.1016/j.ecolmodel.2005.03.005, 2005.

734 Luo, Y., Ahlström, A., Allison, S. D., Batjes, N. H., Brovkin, V., Carvalhais, N., Chappell, A.,
735 Ciais, P., Davidson, E. A., and Finzi, A.: Toward more realistic projections of soil carbon
736 dynamics by Earth system models, *Global Biogeochemical Cycles*, 30, n/a-n/a, 2016.

737 Manzoni, S., Jackson, R. B., Trofymow, J. A., and Porporato, A.: The global stoichiometry of litter
738 nitrogen mineralization, *Science*, 321, 684-686, 2008.

739 Manzoni, S., and Porporato, A.: Soil carbon and nitrogen mineralization: Theory and models
740 across scales, *Soil Biology and Biochemistry*, 41, 1355-1379, 10.1016/j.soilbio.2009.02.031,
741 2009.

742 Manzoni, S., Trofymow, J. A., Jackson, R. B., and Porporato, A.: Stoichiometric controls on

743 carbon, nitrogen, and phosphorus dynamics in decomposing litter, *Ecological Monographs*,
744 80, 89-106, 2010.

745 Manzoni, S., Taylor, P., Richter, A., Porporato, A., and Agren, G. I.: Environmental and
746 stoichiometric controls on microbial carbon-use efficiency in soils, *The New phytologist*, 196,
747 79-91, 10.1111/j.1469-8137.2012.04225.x, 2012.

748 Manzoni, S., Capek, P., Mooshammer, M., Lindahl, B. D., Richter, A., and Santruckova, H.:
749 Optimal metabolic regulation along resource stoichiometry gradients, *Ecology letters*, 20,
750 1182-1191, 10.1111/ele.12815, 2017.

751 Metherall, A. K., Harding, L. A., Cole, C. V., and Parton, W. J.: CENTURY Soil Organic Matter
752 Model Environment Technical Documentation, Agroecosystem Version 4.0, Great Plains
753 System Research Unit, Technical Report No. 4. USDA-ARS, Ft. Collins., 1993.

754 Molina, J. A. E., Clapp, C. E., Shaffer, M. J., Chichester, F. W., and Larson, W. E.: NCSOIL, A
755 Model of Nitrogen and Carbon Transformations in Soil: Description, Calibration, and
756 Behavior1, *Soil Science Society of America Journal*, 47, 85-91, 1983.

757 Moorhead, D. L., and Sinsabaugh, R. L.: A Theoretical Model of Litter Decay and Microbial
758 Interaction, *Ecological Monographs*, 76, 151-174, 2006.

759 Mooshammer, M., Wanek, W., Hammerle, I., Fuchslueger, L., Hofhansl, F., Knoltsch, A.,
760 Schneckner, J., Takriti, M., Watzka, M., Wild, B., Keiblinger, K. M., Zechmeister-Boltenstern,
761 S., and Richter, A.: Adjustment of microbial nitrogen use efficiency to carbon:nitrogen
762 imbalances regulates soil nitrogen cycling, *Nature communications*, 5, 3694,
763 10.1038/ncomms4694, 2014.

764 Pagel, H., Ingwersen, J., Poll, C., Kandeler, E., and Streck, T.: Micro-scale modeling of pesticide
765 degradation coupled to carbon turnover in the detritusphere: model description and sensitivity
766 analysis, *Biogeochemistry*, 117, 185-204, 10.1007/s10533-013-9851-3, 2013.

767 Parton, W., Silver, W. L., Burke, I. C., Grassens, L., Harmon, M. E., Currie, W. S., King, J. Y.,
768 Adair, E. C., Brandt, L. A., Hart, S. C., and Fasth, B.: Global-scale similarities in nitrogen
769 release patterns during long-term decomposition, *Science*, 315, 361-364,
770 10.1126/science.1134853, 2007.

771 Parton, W. J., Stewart, J. W. B., and Cole, C. V.: Dynamics of C, N, P and S in grassland soils: a
772 model, *Biogeochemistry*, 5, 109-131, 1988.

773 Paul, E. A.: *Soil Microbiology, Ecology and Biogeochemistry*, Academic Press, San Diego, CA,
774 USA., 2007.

775 Prescott, C. E.: Litter decomposition: what controls it and how can we alter it to sequester more
776 carbon in forest soils?, *Biogeochemistry*, 101, 133-149, 10.1007/s10533-010-9439-0, 2010.

777 Recous, S., Robin, D., Darwis, D., and Mary, B.: Soil inorganic N availability: Effect on maize
778 residue decomposition, *Soil Biology & Biochemistry*, 27, 1529-1538, 1995.

779 Rubino, M., Dungait, J. A. J., Evershed, R. P., Bertolini, T., Angelis, P. D., D'Onofrio, A.,
780 Lagomarsino, A., Lubritto, C., Merola, A., and Terrasi, F.: Carbon input belowground is the
781 major C flux contributing to leaf litter mass loss: Evidences from a ¹³C labelled-leaf litter
782 experiment, *Soil Biology & Biochemistry*, 42, 1009-1016, 2010.

783 Schimel, J. P., and Weintraub, M. N.: The implications of exoenzyme activity on microbial carbon
784 and nitrogen limitation in soil: a theoretical model, *Soil Biology & Biochemistry*, 35,
785 549-563, 2003.

786 Schmidt, M. W., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., Kleber, M.,
787 Kogel-Knabner, I., Lehmann, J., Manning, D. A., Nannipieri, P., Rasse, D. P., Weiner, S., and
788 Trumbore, S. E.: Persistence of soil organic matter as an ecosystem property, *Nature*, 478,
789 49-56, 10.1038/nature10386, 2011.

790 Sinsabaugh, R. L., Manzoni, S., Moorhead, D. L., and Richter, A.: Carbon use efficiency of
791 microbial communities: stoichiometry, methodology and modelling, *Ecology letters*, 16,
792 930-939, 10.1111/ele.12113, 2013.

793 Sitch, S., Smith, B., Prentice, I. C., Arneth, A., Bondeau, A., Cramer, W., Kaplan, J. O., Levis, S.,
794 Lucht, W., Sykes, M. T., Thonicke, K., and Venevsky, S.: Evaluation of ecosystem dynamics,
795 plant geography and terrestrial carbon cycling in the LPJ dynamic global vegetation model,
796 *Global Change Biology*, 9, 161-185, 10.1046/j.1365-2486.2003.00569.x, 2003.

797 Six, J., Frey, S. D., Thiet, R. K., and Batten, K. M.: Bacterial and Fungal Contributions to Carbon
798 Sequestration in Agroecosystems, *Soil Science Society of America Journal*, 70, 555--569,
799 2006.

800 Stewart, C. E., Moturi, P., Follett, R. F., and Halvorson, A. D.: Lignin biochemistry and soil N
801 determine crop residue decomposition and soil priming, *Biogeochemistry*, 124, 335-351,
802 10.1007/s10533-015-0101-8, 2015.

803 Talkner, U., Jansen, M., and Beese, F. O.: Soil phosphorus status and turnover in central-European
804 beech forest ecosystems with differing tree species diversity, *European Journal of Soil*
805 *Science*, 60, 338-346, 2010.

806 Thiet, R. K., Frey, S. D., and Six, J.: Do growth yield efficiencies differ between soil microbial
807 communities differing in fungal:bacterial ratios? Reality check and methodological issues,
808 *Soil Biology and Biochemistry*, 38, 837-844, 10.1016/j.soilbio.2005.07.010, 2006.

809 Tramontana, G., Jung, M., Schwalm, C. R., Ichii, K., Campsvalls, G., R aluly, B., Reichstein, M.,
810 Altaf Arain, M., Cescatti, A., and Kiely, G.: Predicting carbon dioxide and energy fluxes
811 across global FLUXNET sites with regression algorithms, *Biogeosciences Discussions*, 13,
812 1-33, 2016.

813 Tuomi, M., Laiho, R., Repo, A., Liski, J.: Wood decomposition model for boreal forests,
814 *Ecological Modelling*, 222, 709-718, 2011.

815 Verberne, E. L. J., Hassink, J., Willigen, P. D., Groot, J. J. R., and Veen, J. A. V.: Modelling
816 organic matter dynamics in different soils, *Netherlands Journal of Agricultural Science Issued*
817 *by the Royal Netherlands Society for Agricultural Science*, 38, 221-238, 1990.

818 Walela, C., Daniel, H., Wilson, B., Lockwood, P., Cowie, A., and Harden, S.: The initial
819 lignin:nitrogen ratio of litter from above and below ground sources strongly and negatively
820 influenced decay rates of slowly decomposing litter carbon pools, *Soil Biology and*
821 *Biochemistry*, 77, 268-275, 10.1016/j.soilbio.2014.06.013, 2014.

822 Wang, Y.P., Law, R.M. and Pak, B.: A global model of carbon, nitrogen and phosphorus cycles for
823 the terrestrial biosphere, *Biogeosciences*, 7, 7, 9891-9944, 2010.

824 Wieder, W. R., Bonan, G. B., and Allison, S. D.: Global soil carbon projections are improved by
825 modelling microbial processes, *Nature Climate Change*, 3, 909-912, 10.1038/nclimate1951,
826 2013.

827 Wieder, W. R., Cleveland, C. C., Smith, W. K., and Todd-Brown, K.: Future productivity and
828 carbon storage limited by terrestrial nutrient availability, *Nature Geoscience*, 8, 441-444,
829 2015.

830 Yang, X., Post, W. M., Thornton, P. E., and Ricciuto, D. M.: The role of phosphorus dynamics in
831 tropical forests - a modeling study using CLM-CNP, *Biogeosciences*, 11, 14439-14473, 2014.

832 Zaehle, S., Medlyn, B.E., De Kauwe, M.G., Walker, A.P., Dietze, M.C., Hickler, T., Luo, Y., Wang,

833 Y.-P., El-Masri, B., Thornton, P., Jain, A., Wang, S., Warlind, D., Weng, E., Parton, W.,
834 Iversen, C.M., Gallet-Budynek, A., McCarthy, H., Finzi, A., Hanson, P.J., Prentice, I.C., Oren,
835 R. & Norby, R.J.: Evaluation of 11 terrestrial carbon–nitrogen cycle models against
836 observations from two temperate Free-Air CO₂ Enrichment studies, *The New Phytologist*, 202,
837 803-822, 2014.

838 Zhang, C. F., Meng, F. R., Bhatti, J. S., Trofymow, J. A., and Arp, P. A.: Modeling forest leaf-litter
839 decomposition and N mineralization in litterbags, placed across Canada: A 5-model
840 comparison, *Ecological Modelling*, 219, 342-360, 10.1016/j.ecolmodel.2008.07.014, 2008.

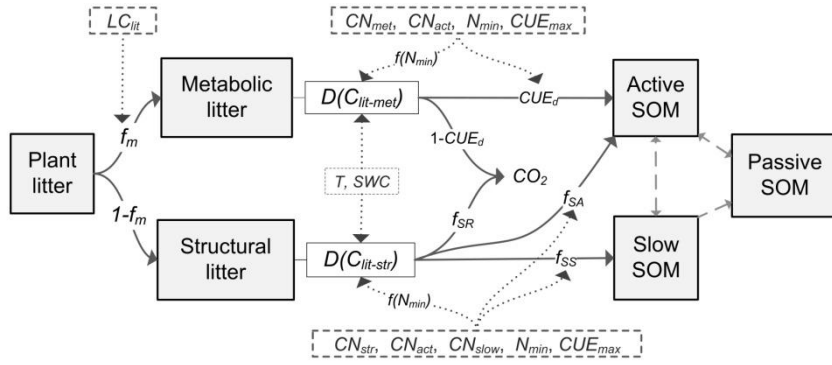
841 Zhang, J., and Elser, J. J.: Carbon:Nitrogen:Phosphorus Stoichiometry in Fungi: A Meta-Analysis,
842 *Frontiers in microbiology*, 8, 1281, 2017.

843

844 **Table 1** Optimized parameter values for the five versions of the litter decomposition
 845 model used in this study. cue_{fit} is the optimized value of CUE, m_1 and n_1 are the
 846 coefficients in Eq. (3), m_2 is the coefficient in Eq. (5), and m_3 is the coefficient in Eq.
 847 (6). Values in brackets following each parameter are the means (\pm standard deviations)
 848 of the fitted parameter values based on “leave-one-out” cross-validation (see Section
 849 2.5 for more details).

Version	CUE	$f(N_{min})$	Parameters
M0	Fixed	1	cue_{fit} (0.57 \pm 0.004)
M1	Eqs. (2), (3)	Eq. (5)	m_1 (0.61 \pm 0.34), n_1 (0.53 \pm 0.21), m_2 (297.4 \pm 38.0)
M2	Eqs. (2), (3)	1	m_1 (0.11 \pm 0.01), n_1 (1.96 \pm 0.13)
M3	Fixed	Eq. (5)	cue_{fit} (0.54 \pm 0.01), m_2 (396.9 \pm 23.6)
M4	Eqs.(2), (3)	Eq. (6)	m_1 (0.13 \pm 0.07), n_1 (1.91 \pm 0.37), m_3 (0.58 \pm 0.12)

850



851

852 **Figure 1** Schematic diagram of the C flows in the litter decay model used in this study.

853 f_m is the fraction of metabolic compounds in plant litter. $D(C_{lit-met})$ and $D(C_{lit-str})$ are

854 the decomposition rates ($\text{g C kg}^{-1} \text{ day}^{-1}$) of metabolic or structural litter, respectively.

855 LC_{lit} is the lignin:C ratio (on a mass basis) of plant litter; CN_{met} , CN_{str} , CN_{act} , and

856 CN_{slow} are the C:N ratio of metabolic litter pool, structural litter pool, active SOM

857 pool and slow SOM pool, respectively; N_{min} is the concentration of mineral N in

858 solution ($\text{g N kg}^{-1} \text{ soil}$); $f(N_{min})$ is a factor reducing litter decay rate when soil mineral

859 N availability is limiting; T ($^{\circ}\text{C}$) and SWC (%) are temperature and soil water content,

860 respectively; CUE_d is C use efficiency of the transformation from litter to soil organic

861 matter (SOM); $CUE_{max}=0.8$ is the maximum microbial CUE (dimensionless) when

862 growth is limited by C from the organic substrate; f_{SA} , f_{SS} and f_{SR} are the fractions of

863 decomposed structural litter-C that is transferred to active SOM pool, slow SOM pool

864 and released to atmosphere in forms of CO_2 , respectively. As in the algorithms in

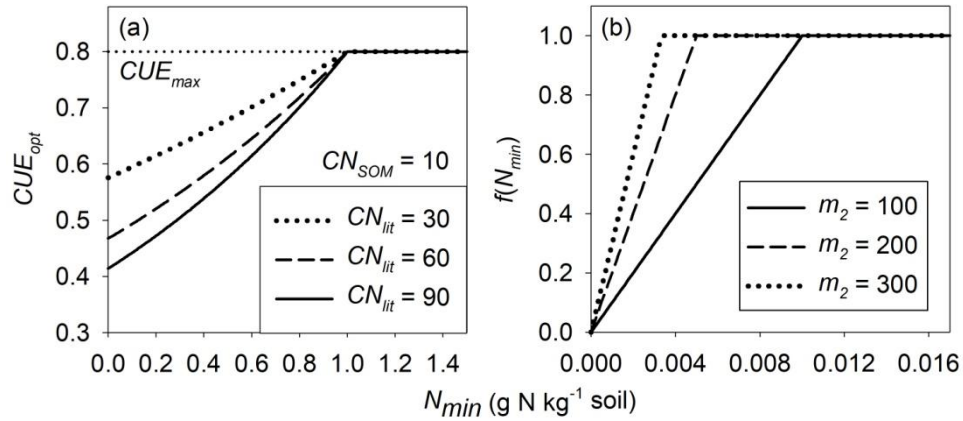
865 CENTURY model (Parton et al., 1988), here $f_{SA}=CUE_{d_SA} \times (1-f_{lig})$, $f_{SS}=CUE_{d_SS} \times f_{lig}$,

866 $f_{SR}=1-(f_{SA}+f_{SS})$, where f_{lig} is the lignin fraction (0–1, dimensionless) in the structural

867 litter pool, and CUE_{d_SA} and CUE_{d_SS} are the CUE of C transformation from structural

868 litter pool to active and slow SOM pool, respectively.

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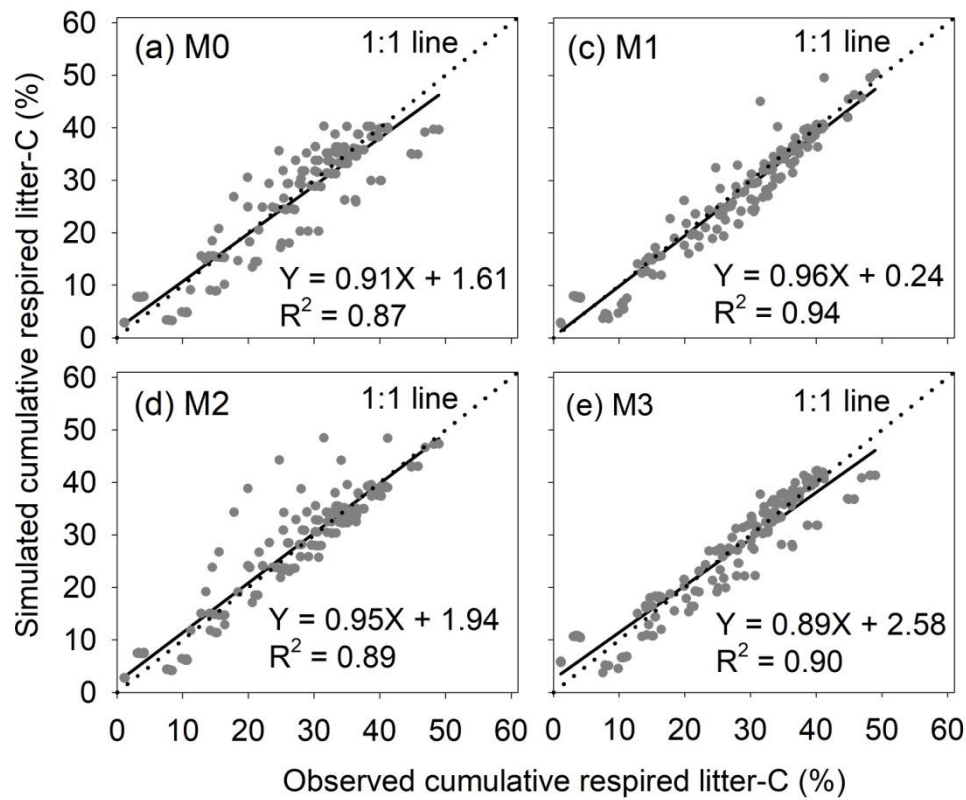


870

871 **Figure 2.** Schematic plot of (a) the optimal carbon use efficiency (CUE_{opt}) as a
 872 function of soil mineral nitrogen for different litter C:N ratios (from Eq. (2) in the
 873 main text with $m_l = 0.3$, $n_l = 1.0$) and (b) the N limitation function $f(N_{min})$ applied to
 874 litter decomposition rates (from Eq. (5) in the main text). CN_{lit} and CN_{SOM} are the C:N
 875 ratios of the litter and SOM pools, respectively. $CUE_{max} = 0.8$ is the maximum CUE
 876 under optimal nutrient condition (C limitation only). m_l and n_l are the parameters of
 877 Eq. (3) and m_2 is the parameter of Eq. (5).

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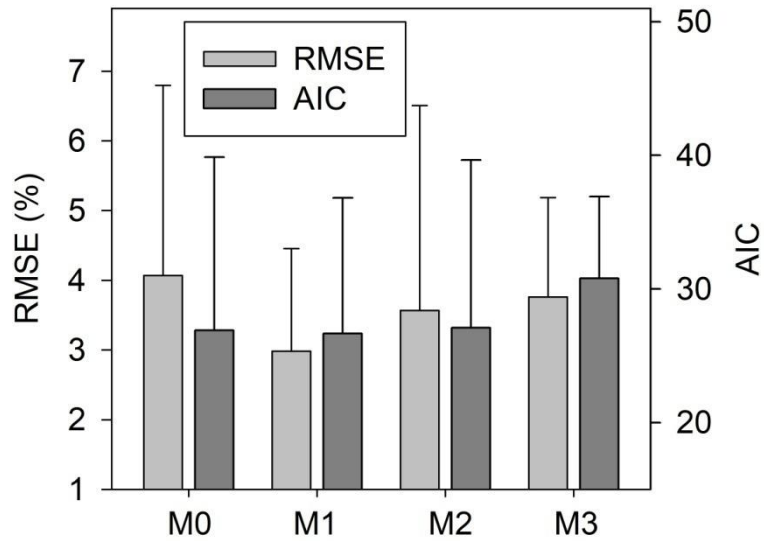
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881

882 **Figure 3** Comparison of the predicted cumulative respired litter-C to observed values
 883 at different times during litter decomposition process. Each dot denotes an
 884 observation of cumulative respired litter-C at a certain day. In total, there are 149
 885 points. M0-M3 are the four versions of litter decay model tested in this study (Table
 886 1).

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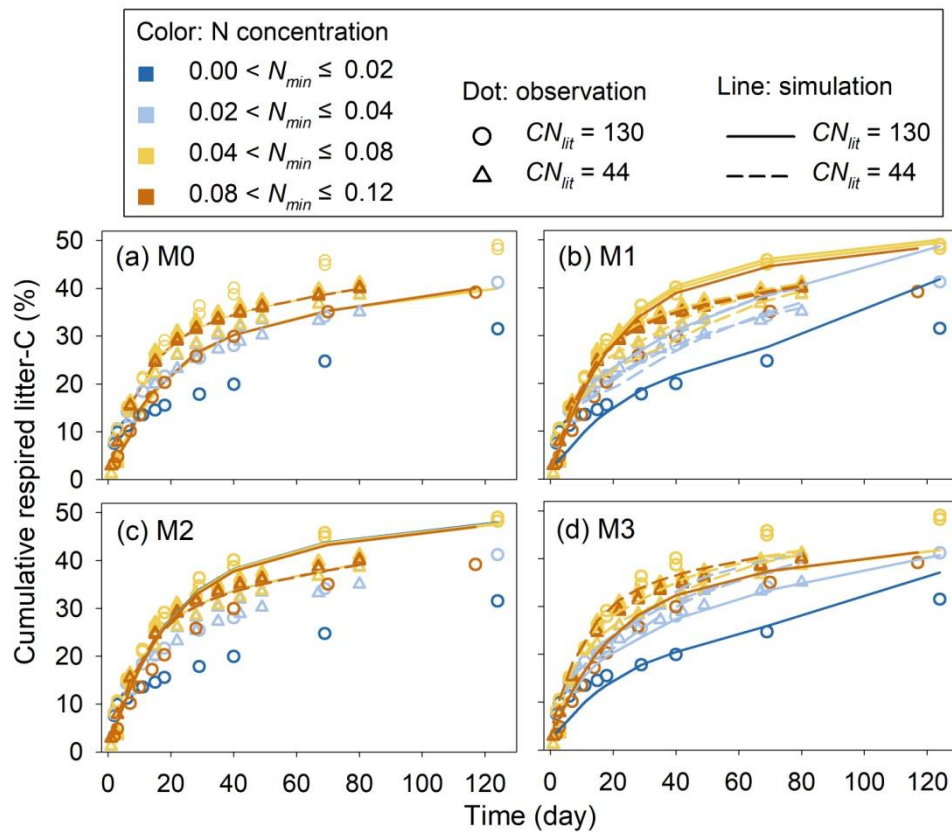


888

889 **Figure 4** The RMSE and AIC of the simulated cumulative respired litter-C from the
 890 four versions of litter decay model used in this study. Error bars denote the standard
 891 deviation of RMSE or AIC for different incubation experiments. M0-M3 denote the
 892 four models tested in this study (Table 1).

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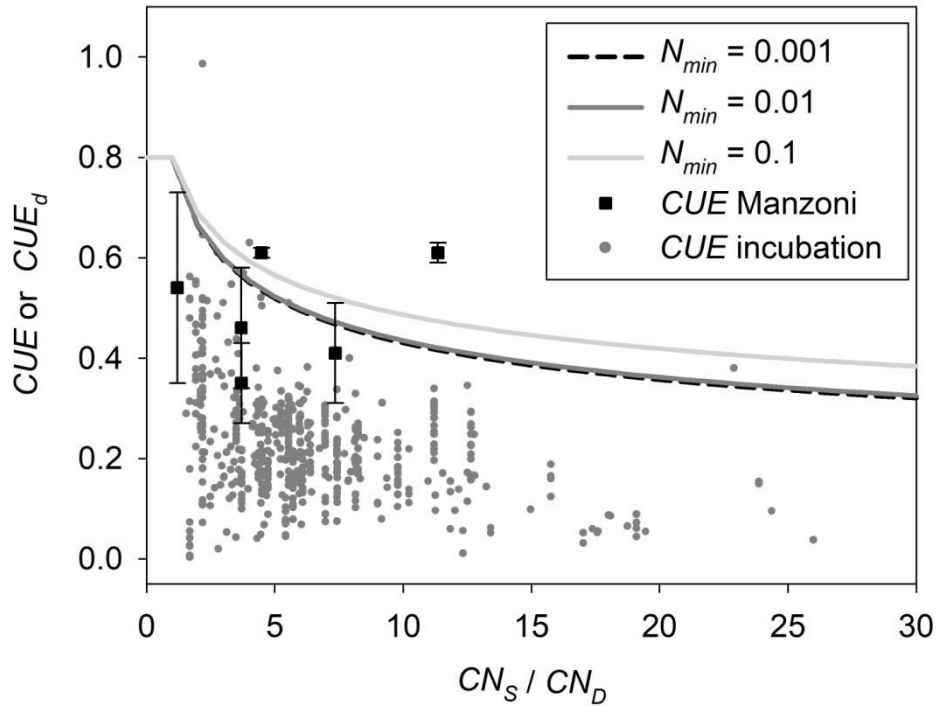
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895

896 **Figure 5** Time series of the simulated (lines) and observed (dots) cumulative respired
 897 litter-C (% of initial litter-C) at four different levels of soil mineral N availability (N_{min} ,
 898 g N kg^{-1} soil). CN_{lit} is the C:N ratio of plant litter. M0-M3 denote the four models
 899 tested in this study (Table 1). Here the simulation results of each model were
 900 calculated with parameters optimized based on all of the 14 samples of incubation
 901 experiments (Table A2).

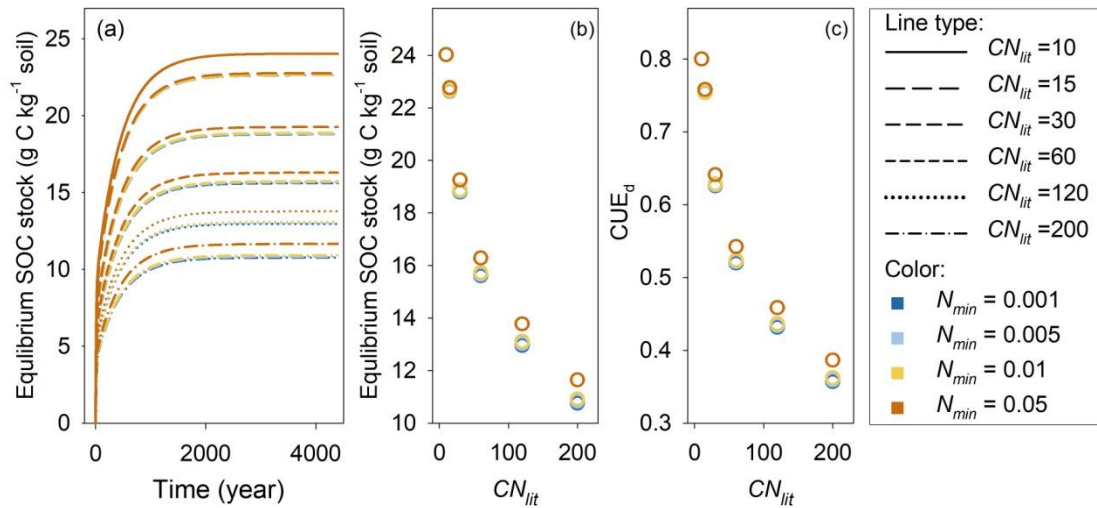
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903

904 **Figure 6** Comparison of CUE_d (lines) predicted by Eq. (2) with parameter values (m_2
 905 $= 0.54$, $n_1 = 0.50$) calibrated based on the incubation experiments (Table A2) of
 906 Recous et al. (1995) and Guenet et al. (2010) to observed CUE of terrestrial
 907 microorganisms along a gradient of CN_S/CN_D . For observed CUE (dots), CN_D and
 908 CN_S are the C:N ratio of decomposers and their substrates, respectively. For simulated
 909 CUE (lines), CN_S and CN_D correspond to the C:N ratio of donor (litter pool) and
 910 acceptor (the active SOM pool of the CENTURY), respectively. Gray dots are the
 911 estimated microbial CUE of litter decomposition in natural terrestrial ecosystems
 912 from Manzoni *et al.* (2017). Black squares are the microbial CUE measured via
 913 laboratory incubation experiments of Gilmour and Gilmour, (1985), Devêre and
 914 Horwáth (2000) and Thiet *et al.* (2006). Error bars represent the standard deviations.
 915 N_{min} (g N kg^{-1} soil) is the concentration of soil mineral N.

916



917

918 **Figure 7** (a) Accumulation of soil organic carbon (SOC) for constant substrates input
 919 (plant litter) with different C:N ratios (CN_{lit}) at different levels of soil mineral N
 920 concentrations (N_{min} , g N kg⁻¹ soil), (b) Change trends of equilibrium SOC stock and
 921 carbon use efficiency of decomposed litter (CUE_d) with increasing litter C:N ratio.
 922

923 **Appendix:**

924 **Table A1** List of symbols used in this study; stoichiometric ratios are all expressed on
 925 a mass basis.

Symbol	Unit	Description
A	g N kg^{-1} soil	Exponent in Eq. (2)
AIC	dimensionless	The Akaike Information Criterion (Eq. (10))
CN_{act}	dimensionless	C to N ratio of active soil organic matter pool
CN_D	dimensionless	C to N ratio of decomposer (Eq. (1))
CN_{met}	dimensionless	C to N ratio of metabolic litter pool
CN_{slow}	dimensionless	C to N ratio of slow soil organic matter pool
CN_{str}	dimensionless	C to N ratio of structural litter pool
CN_S	dimensionless	C to N ratio of substrate (Eq. (1))
C_{lit}	g C kg^{-1} soil	C stock of litter pool (Eq. (4))
CN_{lit}	dimensionless	C to N ratio of litter pool (metabolic or structural, Eq. (2))
CN_{SOM}	dimensionless	C to N ratio of soil organic matter pool
CUE	dimensionless	Microbial carbon use efficiency
CUE_d	dimensionless	Carbon use efficiency of decomposition (C incorporated in SOC over litter C decomposed)
CUE_{fit}	dimensionless	Optimized value of fixed CUE in model M0 and M4
CUE_{max}	dimensionless	Maximum CUE_d (Eqs. (1) and (2))
CUE_{opt}	dimensionless	Optimal CUE_d (Eq. (1))
$CUE_{d,SA}$	dimensionless	CUE of the transformation from structural litter to active SOM pool
$CUE_{d,SS}$	dimensionless	CUE of the transformation from structural litter to slow SOM pool
$D(C_{lit-met})$	g C kg^{-1} soil day^{-1}	Decomposition rate of metabolic litter
$D(C_{lit-str})$	g C kg^{-1} soil day^{-1}	Decomposition rate of structural litter
$f(N_{min})$	dimensionless	Limit factor of soil mineral N on litter decomposition (Eqs. (4) and (5))
$f(tem)$	dimensionless	Limit factor of soil temperature on litter decomposition (Eq. (4))
$f(water)$	dimensionless	Limit factor of soil water content on litter decomposition (Eq. (4))
f_m	dimensionless	Fraction of metabolic plant litter
f_{SA}	dimensionless	Fractions of decomposed structural litter-C that is transferred to active SOM pool
f_{SR}	dimensionless	Fractions of decomposed structural litter-C that is released to atmosphere
f_{SS}	dimensionless	Fractions of decomposed structural litter-C that is transferred to slow SOM pool
I_N	g kg^{-1} soil	Maximum mineral N immobilization rate (Eq. (1))
K	day^{-1}	potential maximum turnover rate (Eqs. (4) and (7))
LC_{lit}	dimensionless	Lignin to C ratio of litter input
m_1	kg g^{-1} N	Coefficients in Eq. (3)
n_1	g N kg^{-1} soil	Coefficients in Eq. (3)
m_2	day^{-1}	Coefficients in Eq. (5)

m_3	kg g ⁻¹ N	Coefficients in Eq. (6)
N_{min}	g N kg ⁻¹ soil	Soil mineral N concentration (Eq. (5))
$RMSE$	%	Root mean square error (Eq.(9))
SOC	g C kg ⁻¹ soil	Soil organic carbon
SOM	g C kg ⁻¹ soil	Soil organic matter
U_0	g C kg ⁻¹ soil day ⁻¹	C uptake rate when soil mineral N is fully adequate for litter decay (Eqs. (1) and (8))

926

927 **Table A2** Information about the 14 samples of laboratory incubation experiment used in this study. CN_{lit} and LC_{lit} are the C to N ratio and lignin
928 to C ratio of plant litter, respectively. CN_{SOM} is the C to N ratio of SOM pool. N_{min} is the concentration of soil mineral N (NO_3^- -N + NH_4^+ -N).
929 For the incubation experiments of Guenet *et al.* (2010), cumulative respired litter-C was measured on days 1, 3, 7, 15, 22, 28, 35, 42, 49, 67 and
930 80, and N_{min} was measured on days 3, 7, 17, 28 and 80. For the incubation experiments of Recous *et al.* (1995), both cumulative respired litter-C
931 and N_{min} were mostly measured on days 2, 3, 6, 11, 15, 18, 29, 40, 69 and 124.

Sample	CN_{lit}	LC_{lit}	CN_{SOM}	Initial N_{min} (g N kg ⁻¹ soil)	Duration (day)	Temperature (°C)	Soil moisture (%, in volume)	Litter type	Reference	
1	44	0.26	11	0.035	80	20	50	Crop (wheat)	Guenet <i>et al.</i> , 2010	
2	44	0.26	11	0.051	80	20	50	Crop (wheat)		
3	44	0.26	11	0.055	80	20	50	Crop (wheat)		
4	44	0.26	11	0.033	80	20	50	Crop (wheat)		
5	44	0.26	11	0.049	80	20	50	Crop (wheat)		
6	44	0.26	11	0.067	80	20	50	Crop (wheat)		
7	44	0.26	11	0.033	80	20	50	Crop (wheat)		
8	44	0.26	11	0.048	80	20	50	Crop (wheat)		
9	44	0.26	11	0.079	80	20	50	Crop (wheat)		
10	130	0.23	9	0.010	124	15	42	Crop (Corn)		
11	130	0.23	9	0.030	124	15	42	Crop (Corn)		Recous <i>et al.</i> , 1995
12	130	0.23	9	0.060	124	15	42	Crop (Corn)		
13	130	0.23	9	0.080	124	15	42	Crop (Corn)		
14	130	0.23	9	0.100	124	15	42	Crop (Corn)		

932

933 **Table A3** List of parameters calibrated for two versions of the litter decomposition
 934 model (M0, M1): k_{lim} and k_{lits} are respectively the turnover rates of metabolic and
 935 structural litter pools, m_4 is the coefficient in Eq. (8), cue_{fit} is the optimized value of
 936 CUE, m_1 and n_1 are the coefficients in Eq. (3), and m_2 is the coefficients in Eq. (5).

Version	CUE	$f(N_{min})$	Parameters
M0	Fixed	1	$cue_{fit}, k_{lim}, k_{lits}, m_4$
M1	Eqs.(2), (3)	Eq. (5)	$m_1, n_1, m_2, k_{lim}, k_{lits}, m_4$

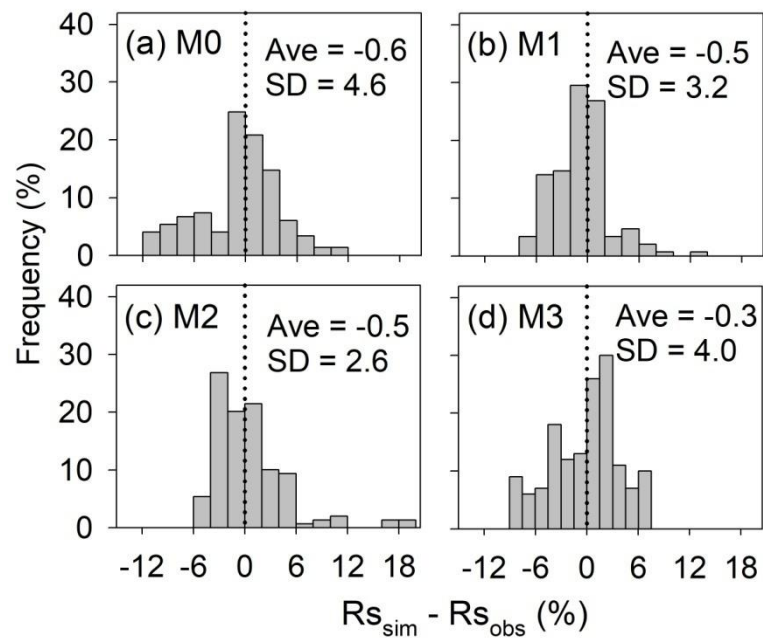
937

938 **Table A4** Specific setting of litter and SOM properties, and soil conditions in the 16
939 idealized simulations for exploring the impacts of litter stoichiometry (i.e. C:N ratio)
940 and soil mineral N on SOC accumulation. CN_{lit} and LC_{lit} are the C to N ratio and
941 lignin to C ratio of plant litter, respectively. Lit_{inp} (g C kg⁻¹ soil day⁻¹) is the daily input
942 rate of plant litter. CN_{SOM} is the C to N ratio of SOM pool. N_{min} (g N kg⁻¹ soil) is the
943 concentration of soil mineral N (NO₃⁻ -N + NH₄⁺ -N). Tem (°C) and SWC (%) are the
944 temperature and soil water content, respectively.

Experiment	CN_{lit}	LC_{lit}	Lit_{inp}	CN_{SOM}	N_{min}	Tem	SWC
1	15	0.2	0.006	12	0.001	25	60
2	30	0.2	0.006	12	0.005	25	60
3	60	0.2	0.006	12	0.01	25	60
4	120	0.2	0.006	12	0.05	25	60
5	15	0.2	0.006	12	0.001	25	60
6	30	0.2	0.006	12	0.005	25	60
7	60	0.2	0.006	12	0.01	25	60
8	120	0.2	0.006	12	0.05	25	60
9	15	0.2	0.006	12	0.001	25	60
10	30	0.2	0.006	12	0.005	25	60
11	60	0.2	0.006	12	0.01	25	60
12	120	0.2	0.006	12	0.05	25	60
13	15	0.2	0.006	12	0.001	25	60
14	30	0.2	0.006	12	0.005	25	60
15	60	0.2	0.006	12	0.01	25	60
16	120	0.2	0.006	12	0.05	25	60

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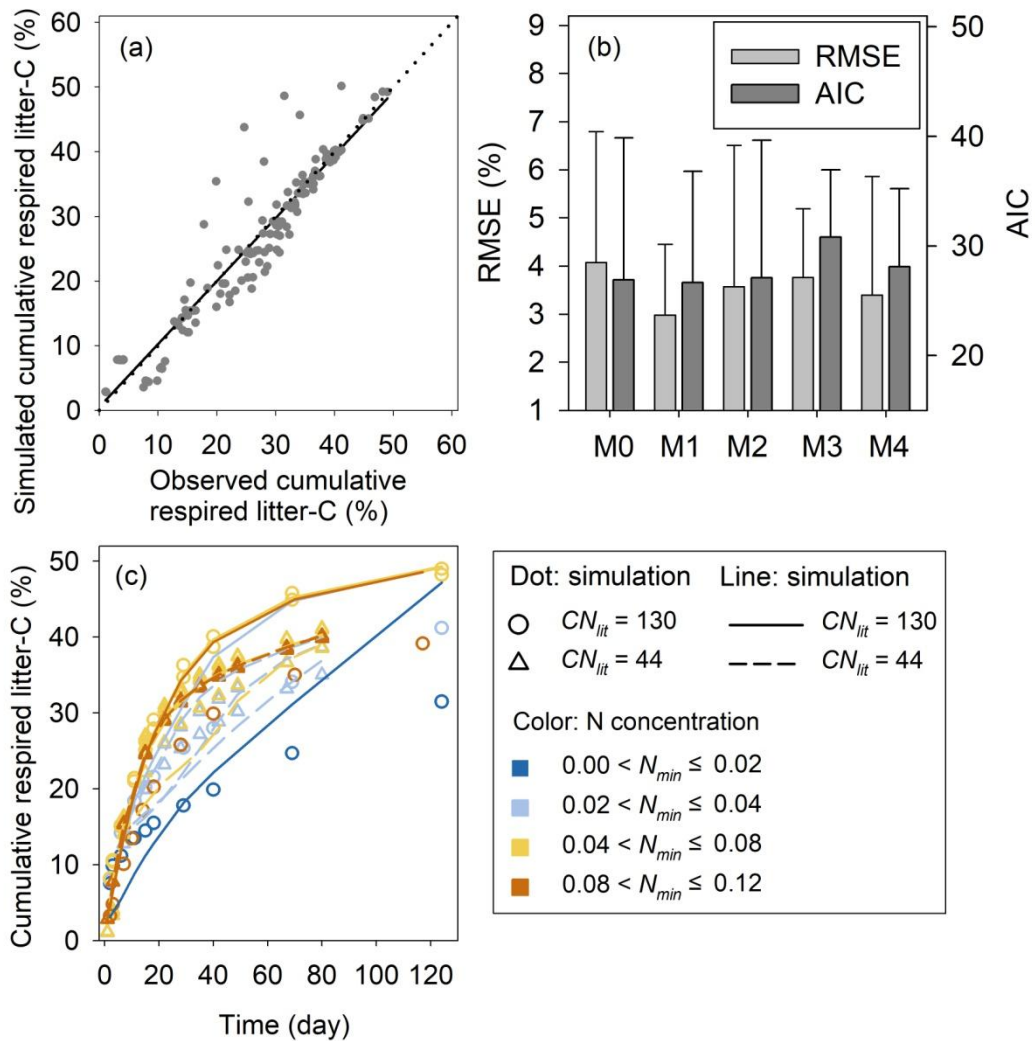
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947

948 **Figure A1** Distribution of the difference between the predicted cumulative respired
949 litter-C ($R_{s_{sim}}$, %) and the observed values ($R_{s_{obs}}$, %) for all experiments and points in
950 time. SD is standard deviation of the biases. M0-M3 denote the four models tested in
951 this study (Table 1).

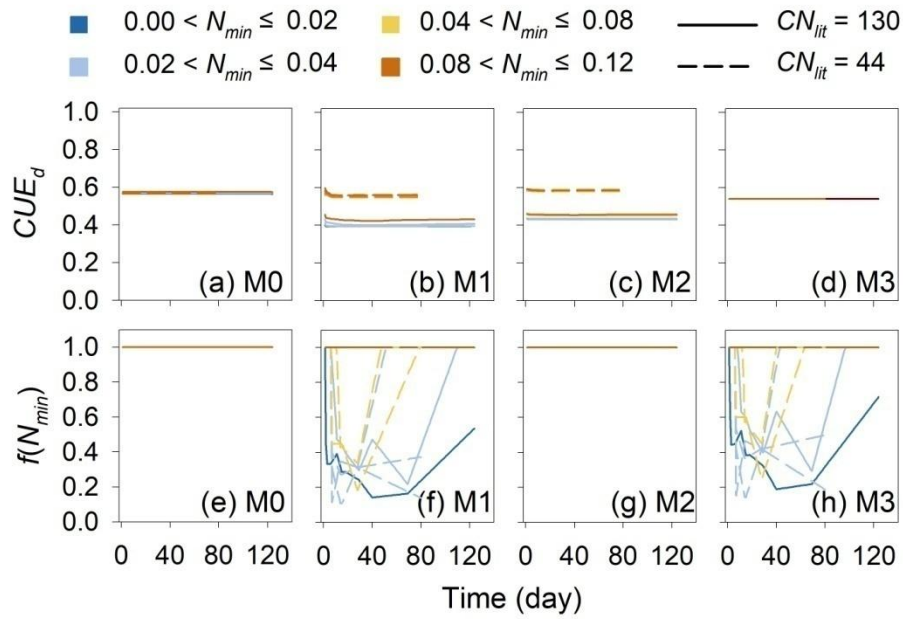
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953

954 **Figure A2** Comparison between simulated cumulative respired litter-C with $f(N_{min})$
 955 (inhibition effect of soil mineral N on litter decomposition) calculated by Eq. (6) and
 956 the observed results from incubation experiments. In figure (b), M0-M4 denote the
 957 five versions of litter decay model in Table 1. M4 denote the model which used Eq. (2)
 958 to calculate the dynamic CUE and Eq. (6) to calculate $f(N_{min})$.

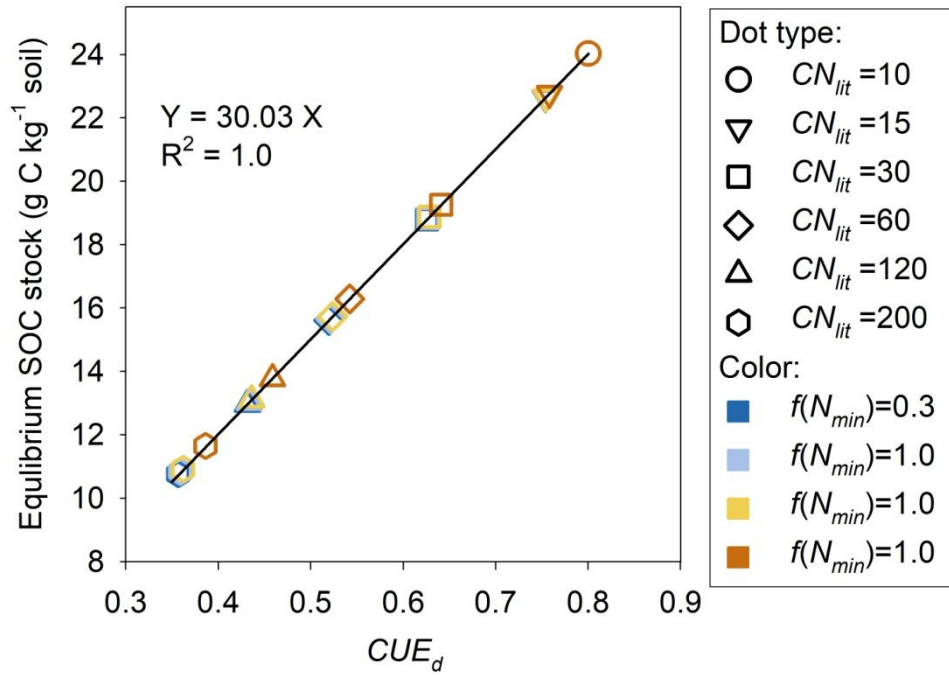
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960

961 **Figure A3** Dynamic of the simulated carbon use efficiency (CUE) and $f(N_{min})$ during
 962 the incubation experiments (Table A4). CN_{lit} is the C:N ratio of incubated litter, and
 963 N_{min} is the initial soil mineral N concentration (g N kg^{-1} soil). M0-M3 are the four
 964 models in Table 1. Here the simulation results of each model were calculated with
 965 parameters optimized based on all of the 14 samples of incubation experiments (Table
 966 A2).

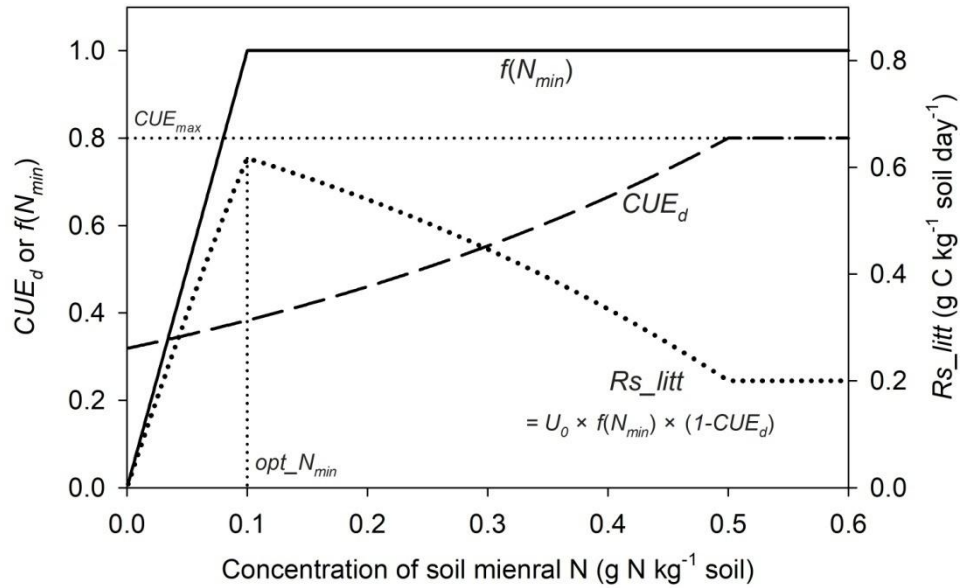
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968

969 **Figure A4** Relationship between C stock of the potentially equilibrated SOM pool
 970 and the carbon use efficiency of decomposed metabolic litter (CUE_d) at the dynamic
 971 equilibrium stage. $f(N_{min})$ denote the inhibition factor (0–1) of soil mineral N on litter
 972 decomposition.

973



974

975 **Figure A5** Schematic plot for change trends of $f(N_{min})$ (inhibition effect of mineral N,
 976 Eq. (6)), CUE_d (carbon use efficiency of decomposed litter, Eqs. (2), (3)) and Rs_{litt}
 977 (litter respiration rate) with increasing concentration of soil mineral N. CUE_{max} (= 0.8)
 978 is the maximum CUE set in this study. opt_N_{min} denotes the concentration of soil
 979 mineral N at which litter respiration is maximized. U_0 is the potential decomposition
 980 rate when mineral N is fully adequate for litter decay.