<u>Complete responses FINAL</u>

Anonymous Referee #1

1. Comment:

Model formulation - The model assumes no overlap of mineralization reactions with different terminal electron acceptors, and assumes that secondary redox reactions can be collapsed onto the interfaces between different mineralization zones (p.9). This is probably ok in environments typically encountered at greater water depth, but there is ample evidence of 'overlapping' mineralization pathways in surficial sediments, in particular in permeable or bioturbated settings.

Response:

We agree with the reviewer that different biogeochemical zones can overlap. However, as stated in the text, OMEN-SED is designed for the coupling to ESMs and its formulation is thus first and foremost guided by achieving numerical efficiency while retaining biogeochemical reality. As summarized in the manuscript, there are essentially two approaches that can be used to describe biogeochemical processes in models. The first approach solves the general diagenetic equation numerically on a regular or irregular grid and biogeochemical zonation emerges in response to inhibition terms allowing a certain degree of overlap between biogeochemical zones. This approach is highly flexible and thus preferable. Yet, its excessive computational demand unfortunately renders its application within a three-dimensional Earth System Model framework impossible. On the other hand, analytical models that subdivide the sediment into distinct biogeochemical zones are computationally efficient and thus ideally suited to describe diagenetic dynamics in ESM.

By their very nature, analytical models do not allow for overlapping biogeochemical zones. As stated in the manuscript, this is a simplification. However, we disagree with the reviewer that this simplification would *per-se* prevent the application of such analytical approaches in shallower aquatic environments. In fact, OMEN-SED builds on a number of analytical models that were developed to investigate local, coupled nutrient and oxygen cycles in coastal sediments (e.g. Billen, 1982; Goloway and Bender, 1982; Jahnke et al., 1982; Slomp et al. 1996). Similar approaches were later successfully applied from oxic to anoxic sediments and at the regional coastal ocean scale (e.g. Ruardij and Van Raaphorst, 1995; Tromp et al., 1995; Gypens et al., 2008). In particular, Gypens et al., (2008) points out that accounting for secondary redox process in the boundary condition induces little error as: "Using a numerical model, Soetaert et al. (1996) showed that this re-oxidation mainly occurs at the oxic-anoxic transition interface."

Finally, the good agreement between OMEN-SED and the results obtained with a fully formulated numerical RTM (compare Section 3.3, allowing for overlapping TEA use) shows that this is not a critical limitation of OMEN-SED - even for shallow sediments.

We included a little paragraph on this in the limitation section (pg 54 lines 3-8):
"Furthermore, by their very nature, analytical models do not allow for overlapping biogeochemical zones or depth dependent porosity, which introduces a certain error to simulation results. However, the energy yield dependent sequence of oxidants is generally valid (e.g. Hensen et al., 2006) and the good agreement between OMEN-SED and the results obtained with a fully formulated numerical RTM (allowing for overlapping TEA use and depth dependent porosity, Section 3.3) shows that these are not critical limitations of OMEN-SED - even for shallow sediments."

2. Comment:

In the denitrification layer, all N goes to N2. However, in the suite of processes involved in the breakdown of organic matter, ammonium is produced as well, even if nitrate serves as TEA. Ammonium produced in the denitrification zone would need to be accounted for at the transition to the oxic zone as well, which may further reduced the O2 penetration depth.

Also, processes such as DNRA or anammox are not included, even though literature surveys (e.g. Dalsgaard et al. 2005) indicate that anammox is relevant at shallower water depth.

Response:

Anammox is implicitly included in the model. The organic nitrogen released during the denitrification process is assumed to be directly oxidized with nitrite to N_2 through a coupling between denitrification and anaerobic ammonium oxidation.

However, we would like to stress again that OMEN-SED is a benthic model designed for the coupling to ESMs. Most ESMs do not even explicitly resolve N-dynamics. In addition, OMEN-SED is a system/global scale model that aims to resolve the most pertinent biogeochemical dynamics on a global scale (including a paleoenvironmental context) and estimate the main SWI-fluxes and not a model that aims at resolving specific local scale dynamics. Even most local scale RTM applications do not resolve DNRA and anammox explicitly. However, OMEN-SED could be easily adapted to explicitly resolve these processes if the specific application requires their representation (e.g. coastal ocean).

We included a sentence on this in the Section 2.2.3 "Nitrate and Ammonium" (pg. 12, lines 22-24):
"*Anaerobic ammonium oxidation (anammox) is implicitly included in the model. The organic nitrogen released during denitrification is assumed to be directly oxidized with nitrite to N2 through a coupling between denitrification and anammox.*"

3. Comment:

Methane oxidation: All methane is assumed to be oxidized anaerobically. Is this done for simplicity? Is there no leakage term (gamma_CH4; comparable to incomplete sulfide and ammonium oxidation) because methane escaping from the anoxic zone is assumed to be removed by aerobic methane oxidation?

Response:

First, there is a leakage term γ_{CH4} comparable to incomplete sulfide and ammonium oxidation (see Table 10, SO_4 boundary conditions 5 and 8.2; H2S boundary conditions 5 and 9). Second, it can be safely assume that almost all CH4 is oxidized anaerobically in the sediments (e.g. Reeburgh (2007) and Hinrichs & Boetius (2002) estimated > 90%) - except for active (very localized) sites and slope failure, which can, in theory, be accounted for through the gamma term.

We included this sentence in the Section 2.2.4 "Sulfate and Sulfide" (pg. 14 line 10 – pg 15 line 1):
"*It can be safely assumed that almost all CH4 is oxidized anaerobically in the sediments (e.g. Reeburgh (2007) suggests up to 90%)- except for active (very localized) sites and slope failure, which can, in theory, be accounted for through the γ_{CH4} term.*"

4. Comment:

Methanogenesis: the 1/2 methane to DIC ratio seems to imply acetoclastic methanogenesis. What evidence is there to ignore hydrogenotrophic methanogenesis?

Response:
We thank the reviewer for this valid point. However, we are not aware of an approach that would allow splitting the two pathways in quantitative terms in a global context. Following the approach of Jourabchi et al. (2005), we assume a ½ methane to DIC ratio (i.e. the acetoclastic pathway). However, OMEN-SED could be easily adapted to represent hydrogenotrophic methanogenesis globally (i.e. by adapting the stoichiometric factor by a value depending on the specific pathway).

5. Comment:
For a globally applicable model, the lack of CaCO3 dissolution is an obvious issue. Thus, can you expand on what problems the modeling of CaCO3 dissolution would cause (page 18)? Is this linked to the calculation of pH? Why can pH (and then carbonate) not be estimated from DIC and alkalinity?

Response:
In general, the coupling between multiple reaction-transport equations (i.e. strongly coupled biogeochemical dynamics) make finding an analytical solution difficult or even impossible. Calcium carbonate dissolution is kinetically controlled by the amount of $CaCO_3$ and thermodynamically controlled by the porewater concentrations of calcium ion, Ca^{2+}, and carbonate ions, CO_3^{2-}. The reaction terms of the $CaCO_3$, (the Ca^{2+}), the DIC and balance equations would thus depend on the concentrations of $CaCO_3$, Ca^{2+} and CO_3^{2-} (DIC, pH/Alk). This set of strongly coupled equations does not have a simple analytical solution. Finding a solution to the problem requires some creativity and a number of assumptions that need to be independently tested and validated. This task thus requires some additional work and model development that go beyond the scope of this first model description paper centered on organic carbon and nutrients. However as pointed out in Section 5 "already planned future extensions of OMEN-SED include an explicit description of carbonate dissolution and iron."

6. Comment:
Is there no P sorption on iron oxides below the oxic zone? If so, why?

Response:
We here follow the approaches of Slomp et al. (1996) and Gypens et al. (2008) who both assume that HPO_4^{2-} is released due to the reduction of Fe oxides throughout the reduced sediment (i.e. starting at zox).

7. Comment:
Why does some of the ammonium created below the oxic zone escape oxidation, but oxidation of ammonium to nitrate is complete in the oxic zone?

Response:
This is a misunderstanding: There is also leakage term (γ_{NH4}) for ammonium to nitrate oxidation in the oxic zone (see Eqs. 12, 15 and 16).

8. Comment:
A fraction of the sulfide produced is assumed to escape complete oxidation. Does this mimicking the effect of precipitation with iron, rather than escape from the sediment?

Response:
In the reviewed version of OMEN-SED, this fraction mimicked the escape from the sediment. However, in response to the first critical comment of reviewer 3 (K. Wallmann) we made the following changes to OMEN-SED:
When coupled to an ESM the γ_{H2S} value (fraction of H2S that is oxidised) becomes dependent on the bottom water oxygenation state. That is, $\gamma_{H2S} = 1.0$ for oxic bottom waters and a user defined value γ_{H2S} <1.0 for anoxic bottom waters.
In addition, we introduce another parameter (γ_{FeS}) representing the fraction of sulfide that is precipitated as pyrite (i.e. $0.0 <= \gamma_{FeS} < 1 - \gamma_{H2S}$) in the sulfate reduction zone. γ_{FeS} is an auxiliary parameter used as a fix until iron is explicitly represented. We thus assume that a user-defined fraction of the produced H2S precipitates as FeS(FeS2) in the sulfate reduction zone. If a user does not want to make any assumptions about FeS precipitation – it can be set it to 0.

The text, tables and equations (for O_2, SO_4, H_2S and alkalinity) are changed accordingly. The presented results have not been changed and we note that $\gamma_{FeS} = 0.0$ for all simulations, as we do not want to make any assumptions. About FeS precipitation.

Changes made in:
2.2.2 Oxygen: Table 3 boundary condition 4.2 + Text pg. 12 lines 14-16:
" *It is assumed that respective fractions (γNH4 and γH2S) are directly reoxidised at the oxic/anoxic interface and the remaining fraction escapes reoxidation **(or is precipitated as pyrite, γFeS)**.*"

2.2.4 Sulfate and Sulfide: Equations 23 + Table 5 boundary condition 5) + (pg. 14 lines 9-10):
"*In the sulfidic zone a defined fraction of sulfide, γFeS , can be precipitated as pyrite (in the presented simulations γFeS = 0.0 as we do not want to make any assumptions about pyrite precipitation).*"

2.2.7 Alkalinity: Table 8 boundary condition 5)
Text (pg. 20 lines 5-7):
" *In addition, the effect of secondary redox reactions, such as nitrification, sulfide and methane oxidation**, as well as pyrite precipitation,** are implicitly accounted for in the boundary conditions.* "

Text (pg. 20 lines 21-23):
"*The decrease of alkalinity due to oxidation of reduced species produced in the anoxic zones **and due to the precipitation of pyrite** (with stoichiometry ALK^{NIT} , ALK^{H2S} and **ALK^{FeS}**) is implicitly taken into account through the flux boundary condition at zox (Table 8 Eq. 5).* "

2.4.2 Stoichiometries and reaction parameters: (pg. 27 lines 19-23):
"*However, when coupled to an ESM γH2S becomes dependent on the bottom water oxygenation state. That is, γH2 S = 1.0 for oxic bottom waters and a user defined value γH2S < 1.0 for anoxic bottom waters. The parameter γFeS represents the fraction of sulfide that is precipitated as pyrite in the sulfidic zone. The majority of H2S produced by sulfate reduction is reoxidised, but it is estimated that ∼ 10 − 25% is eventually buried as pyrite (Bottrell and Newton, 2006). However, this fraction can vary significantly over geological timescales (Berner, 1984). If a user does not want to make any assumptions about pyrite precipitation – it can be set to 0 (as in the results presented here).*"

We also added pyrite precipiation to Table 1 and A1 in the Appendix.

9. Comment:
Iron cycling is not represented explicitly but some of the effect of iron cycling is parameterised. With its effect on sulphur cycling, P sorption and C mineralization (metal reduction can be the main mineralization pathway, see e.g. Canfield et al. 1993), I don't fully understand the reason for doing so (apart from the added complexity when dealing with another solid phase).

Response:
OMEN-SED will be mainly applied on a system/global scale, coupled to an ESM, where iron reduction has been shown to play just a minor role (i.e. about 3% of the global carbon mineralization rate Thullner et al. (2009); or compare Archer et al. (2002) Fig. 9D). Also, Fe-dynamics are generally not explicitly resolved in ESMs. However, as stated in Section 5, "already planned future extensions of OMEN-SED include an explicit description of iron."

10. Comment:
transport processes: A 1D diffusion/bioturbation model clearly faces major challenges in the coastal ocean, where sediments one predominantly are permeable. And setting fir=1, implying no bioirrigation, is also a very strong assumption.

Response:
Most sediments on the globe are non-sandy, therefore we had decided to neglect sandy sediments. However, the bioirrigation coefficient has been changed and is now represented by the empirical relationship with seafloor depth derived by Soetaert et al. (1996): fir = Min{1; $15.9 \cdot z-0.43$ }. We also added to the limitations section that our approach might not be appropriate to simulate non-accumulating permeable sands.
The text has been changed to (pg. 25 lines 21-23):
"*Soetaert et al. (1996) derived an empirical relationship between fir and seafloor depth (fir = Min{1; 15.9 · z −0.43 }) based on observations from Archer and Devol (1992) and Devol and Christensen (1993) which is used in OMEN-SED*"
And in the limitations section (pg. 54 lines 17-18):
"*Also note that our 1-D diffusion/bioturbation model might not be appropriate to simulate non-accumulating permeable sands of the coastal ocean.*"

11. Comment:
The 2G model is a sensible choice. Model parameterisation is a general concern (page 3, line 30), but I suggest to cite some promising new approaches to address this issues such as presented by https://www.biogeosciences-discuss.net/bg-2017-397

Response:
We added the suggested reference to the manuscript (Section 4.2 "Parameterising the OM degradation rate constants in a global model"). However, the suggested approach is based on empirical relationships derived from modern ocean data, as well as strong assumptions. Its applicability to past and future oceans is thus questionable and the problem of parameterizing organic matter reactivity remains for these applications.

Added sentence (page 45 lines 19-22):
"*Stolpovsky et al. (2015, 2017) suggested empirically derived approaches to constrain degradation rate constants in a 2G model on a global scale. These approaches are derived from present-day observations and might help constrain parameters for present-day applications. However, the problem*

of constraining 2G degradation model parameters remains for largely different environmental conditions encountered in the past that could also prevail in the future (Arndt et al., 2013)."

12. Comment:
site comparison case studies: - Table 13: fix the depths for the SB and IM sites (SB is the 585m site ...).

Response:
Changed.

13. Comment:
site comparison case studies: - Why are the stoichiometric factors set to default values when Epping et al. provide the C/N ratio of the surface sediment?

Response:
We intended to do as little site tuning as possible in order to test how the default model performs for these sites and to be able to evaluate the performance of the model in data poor areas.

14. Comment:
site comparison case studies: - If a non-local exchange mechanism resulting from bioirrigation is invoked for the Canyon site, what is the source of the high ammonium (and DIC) leading to the observed increase in concentration at depth?

Response:
As suggested by the papers describing the study area (van Weering et al., 2002; Epping et al., 2002), we assume it is a result of degradation of organic matter which has been delivered from the shelf to the ocean interior.

15. Comment:
- global transect case study: I don't see the value of the 5% reoxidation case. This is simply unrealistic for the conditions considered here. A possible explanation why lower gammas are giving better match to the data in the shallow sites is that under those conditions, the conceptual model of a vertical separation of reaction zones is more and more violated, so the gamma become a 'fudge-factor' to account for this (see also above comments about the coastal ocean).

Response:
We agree with the reviewer, that gamma is a fudge factor. It accounts for all the processes that may enhance escape from re-oxidation but are not explicitly resolved. However, we would reject the comment that this is a problem with assuming strict zonation. We would argue that this reflects the more intense dynamics in shallow ocean regions and that the increased escape is due to enhanced macrofaunal activity.
We also want to stress again, that similar analytical approaches (with distinct redox zones) have given good results for coastal/estuarine sediments (e.g. Billen, 1982; Goloway and Bender, 1982; Jahnke et al., 1982; Ruardij and Van Raaphorst, 1995; Slomp et al., 1996; Gypens et al., 2008).

16. Comment:
- link to cGENIE: - on page 43, it says tha if fPOC is computed to be > 1 (more than 100% is preserved), then this is discarded and all POC is remineralized. Imposing constraints is fine, but what is the rationale for jumping from >100 to 0 % preservation?

Response:
The result fPOC > 1.0 does not imply that 100% preservation is a realistic result. It just means that the OMEN-SED solution does not provide sensitive values. Therefore, its results are discarded and a reflective boundary is assumed (a reflective boundary is a better choice than the conservative as in most cases the majority of OM is degraded during early diagenesis). However, this is just a safety measure and has not occurred in our experiments so far.

17. Comment:
- link to cGENIE. Page 44 discusses the challenges in applying the model in such a setting. In addition, deposition fluxes may change over time. At what point is the steady state assumption on the POC profile still valid under such settings? This is addressed summarily at of the bottom of page 54. However, I think it is important to lead with this, before interpreting the data-model comparison.

Response:
As suggested by the reviewer the steady-state assumption is addressed earlier in the text.
We added the following sentence to Section "4.2 Parameterising the OM degradation rate constants in a global model" (pg 45, lines 12-13):
"Furthermore, by assuming steady-state in OMEN-SED we assume that deposition fluxes of OM are constant over the characteristic timescales of the reaction-transport processes."

18. Comment:
On p.54 it says "In theory, its scope of applicability thus ranges from the regional to the global and from the seasonal to the millennial time-scale. " - in the following para-graph they recognize that "This steady-state assumption is only valid if the variability in boundary conditions and fluxes is generally longer than the characteristic timescales of the reaction-transport processes. "
I recommend to be a little more cautious in the application of the model, since I am not convinced that violations of assumptions underlying the conceptual model and non-steady state effects can be ignored. The model clearly requires substantial tuning. It is clear that the authors are aware of the shortcomings, and they discuss that the model may not be adequate to assess seasonal patterns (and one can think of additional set-tings, where fluxes vary over timescales intrinsic to the POC profile in the top 50cm of sediment modeled here). My concern is that they largely ignore them in their appli-cation, before acknowledging them in the discussion.

Response:
As stated in the manuscript, OMEN-SED is first and foremost designed for the coupling to ESMs.
More specific tuning/adaptation is needed if OMEN-SED is used for specific, regional environments, e.g coastal environments.
However, we would like to re-emphasize that the current version of OMEN-SED performs well across different depositional environments ranging from the coastal to the deep ocean as evidenced by the

model-data and model-model comparison. As outlined in the "Scope of applicability and model limitations" section additional developments, such as adapting pseudo-transient dynamics will further facilitate the application of OMEN-SED to more dynamic environments. A number of benthic models specifically designed for coastal/estuarine environments (e.g. ERSEM Ruardij and Rapphorst et al., 1997; Arndt and Regnier, 2007) have successfully applied such an approach. We therefore maintain our point of view that, in theory, the scope of applicability of OMEN-SED also includes coupling to system-scale estuarine and/or coastal ocean models..

19. Comment:
I also suggest to tone down the finding that "A comparison between simulated OM contents and observations indicates that depth dependent k-f relationships provide the best fit (Section 4.2.2), confirming more theoretical considerations regarding the different time and reactivity scales that need to be considered (see Section 4.2). " The age-reactivity relationship is pretty well established, without confirmation by this modeling effort.

Response:
This is a misunderstanding. We do not argue that model results "confirm" the reactivity-age link. We wanted to emphasize these results confirm that reducing the continuous distribution of organic matter reactivities into two distinct reactivity classes (2G Model) requires different k-f values for shallow vs deep ocean sediments because of the largely different reaction timescales involved (also see Fig. 10).

To clarify, we rephrased the sentence (pg 54 lines 22-25):
"A comparison between simulated OM contents and observations indicates that a depth dependent k-f relationship provides the best fit (Section 4.2.2). These results confirm that reducing the continuous distribution of organic matter reactivities into two distinct reactivity classes (2G Model) requires different k-f values for shallow vs deep ocean sediments because of the largely different reaction timescales involved (also see Fig. 10)."

OMEN-SED – cGENIE coupling
20. Comment:
The stated purpose of section 4 is …
… The validation of the coupled model requires more work, and I wonder whether this was not better done in a separate paper, in which the coupling to cGENIE and the parameterization of POC mineralization was explored in more detail.

Response:
As stated in the manuscript (page 45, lines 23-26):
"Our objective is not to perform and discuss a detailed calibration of the coupled models as this is beyond the scope of this sediment model development paper. Rather we want to showcase the feasibility of the model coupling, illustrate the range of results and thus information that can be generated with OMEN-SED and verify that model results capture the main observed global benthic biogeochemical features."

We think that demonstrating how OMEN-SED can be coupled to an ESM and illustrating the type of output/information generated by OMEN-SED within such a coupling is a central aspect of the model description paper.
However, we agree trimming down this section (as in the re-submitted version). We will discuss an

improved model-data analysis (also using observations of SWI-fluxes) in a follow-up publication (as also suggested by reviewer #3 K. Wallmann).
Specifically, the sensitivity analysis for the spatially uniform degradation rate constants (Figure 12) and it's discussion has been removed (compare pages 47-50).

21. Comment:
Figures 12 - 14: I gather the R2 values are for the bin averages. I don't see much value of that, as over- and underpredictions cancel each other out in the averaging. Why not compute statistics for the actual model results with the Seiter data directly?

Response:
Figure 12 has been deleted from the manuscript. As the statistics for the actual data are not helpful/misleading (see also comment 4 of reviewer #2) we decided to remove the R2 values in Figures 13 and 14 (as well as their discussion in the text). Compare changes on pages 47-50.

Minor comments:

22. Comment:
page 8/ line 1: It is said that all parameters in Eq. 1 may vary with depth, but above it is stated that porosity and burial rates are constant with depth.
Response:
We thank the reviewer for highlighting this. This has been changed in the revised manuscript to (pg 8 lines 1-2):
*"All parameters in Eq. (1)**, apart from porosity and burial rate,** may vary with **sediment** depth and many reaction rate expressions depend on the concentration of other species. "*

23. Comment:
- the fraction of POC buried is defined as the POC at z=0 relative to the POC at depth. Why is it not defined as the flux at z=0 vs. the flux at depth (it seems Eq. 5 ignores the diffusion flux)?
Response:
We decided to calculate the fraction of POC preserved dependent on the concentrations of POC at z=0 and z=zinf mainly because this information is required by cGENIE. Also POC at z=0 is calculated on the basis of the flux provided by cGENIE, therefore does include advection-diffusion-reaction.

24. Comment:
Related to that, page 10, line 18 refers to a concentration/flux boundary conditions at the SWI. The following equations and Table 2 only show a known concentration, not a flux condition. However, the latter would be useful when connecting the sediment model to a model of the water column. On page 43, Eq. 51 this is addressed - make this clear earlier.
Response:
We thank the reviewer for highlighting this. The text has been changed and a reference to Eq. 51 has been added (pg 10 lines 17-20):
*"For organic matter, OMEN-SED applies a known **concentration** at the sediment-water interface and assumes continuity across the bottom of the bioturbated zone, zbio. **When OMEN-SED is coupled to an ESM, the POC depositional flux from the coupled ocean model is converted to a concentration by**

solving the flux divergence equation (51)."

25. Comment:
Can lines 21-23 on page 21 be deleted?
Response:
We refer to both equations later in the text (pg 22 line 14 and 22), therefore we would like to keep them in.

26. Comment:
page 31, line 2: specify NH4, SO4 and H2S FLUXES
Response:
Text has been corrected as suggested (pg 29 line 33).

27. Comment:
Figure 3 did not help me much. Does the green dashed vertical arrow indicate possible locations of zbio?
Response:
Yes, it does indicate locations of zbio. We added this to the caption of Figure 3. However, we strongly believe that Figure 3 illustrates the bioturbation boundary problem in an efficient way and also highlights the integration constants and ODE solutions for the different sediment layers.
New caption:
*"Schematic of the generic boundary condition matching (GBCM) problem. Showing the resulting integration constants (Ai , Bi) and ODE solutions (Ei , Fi, Gi) for the different sediment layers and the bioturbation boundary **(possible locations are indicated by the green vertical arrow)**."*

Anonymous Referee #2

1. Comment:
The model neglects the effect of sediment compaction "due to mathematical constraints". I understand the rational for this and accept a consistency of this assumption to near-seafloor (bioturbated) sediments; however, this might be a problem for deeper sediments discussed in the paper (down to 50 or 100cm). The authors should either define different porosity values for different depth-zones or to demonstrate that the results are not particularly sensitive to the value of this parameter.

Response:
Assuming constant porosity is required to solve the diagenetic equation analytically. It is a mathematical limitation and it will induce a certain error. However, the error is not very large and we have already shown this by comparing the performance of OMEN-SED against observed data (Section 3.2) and against model results from a fully formulated RTM with depth-varying porosity (Section 3.3). The comparison of OMEN-SED with the results of the numerically solved RTM (Section 3.3) allows evaluating to which extend simplifying assumptions (e.g. constant porosity, non-overlapping redox zones etc) affect simulation results and, thus, quantitatively test the performance of the computationally efficient OMEN-SED approach against the computationally expensive numerical approach.
We also want to reiterate that OMEN-SED is designed for the coupling to ESMs and thus for global scale applications (see responses to reviewer #1). The novel model represents a big advance compared to the description of benthic-pelagic exchange processes currently incorporated into ESMs (Hülse et al., 2017; also see comment by the K. Wallmann). Conservative and reflective boundaries, as well as simple box models are characterized by much stronger, simplifying assumptions and far bigger limitations than constant porosity.

We included a little paragraph on this in the limitation section (pg 54 lines 3-8):
"Furthermore, by their very nature, analytical models do not allow for overlapping biogeochemical zones or depth dependent porosity, which introduces a certain error to simulation results. However, the energy yield dependent sequence of oxidants is generally valid (e.g. Hensen et al., 2006) and the good agreement between OMEN-SED and the results obtained with a fully formulated numerical RTM (allowing for overlapping TEA use and depth dependent porosity, Section 3.3) shows that these are not critical limitations of OMEN-SED - even for shallow sediments."

2. Comment:
Dividing the sediment column into functional zones in such a strict manner does not always represent reality well. Thus, "nitrogenous" zone may overlap with "oxic" zone. This assumption, as far as I understand, made it impossible to simulate nitrate SWI flux directed into the sediments in oxygenated environment, which is definitely not true. Validation of the model against measured benthic fluxes would probably demonstrate to some extent accordance of suggested method with real benthic system.

Response:
First, it is possible to simulate nitrate influx into the sediments in oxygenated environments with OMEN-SED (see e.g. Fig. 6C).

In the following we repeat the response given to the 1st comment of Reviewer #1:

We agree with the reviewer that different biogeochemical zones can overlap. However, as stated in the text, OMEN-SED is designed for the coupling to ESMs and its formulation is thus first and foremost guided by achieving numerical efficiency while retaining biogeochemical reality. As summarized in the manuscript, there are essentially two approaches that can be used to describe biogeochemical processes in models. The first approach solves the general diagenetic equation numerically on a regular or irregular grid and biogeochemical zonation emerges in response to inhibition terms allowing a certain degree of overlap between biogeochemical zones. This approach is highly flexible and thus preferable. Yet, its excessive computational demand unfortunately renders its application within a three-dimensional Earth System Model framework impossible. On the other hand, analytical models that subdivide the sediment into distinct biogeochemical zones are computationally efficient and thus ideally suited to describe diagenetic dynamics in ESM.

By their very nature, analytical models do not allow for overlapping biogeochemical zones. As stated in the manuscript, this is a simplification. However, we disagree with the reviewer that this simplification would *per-se* prevent the application of such analytical approaches in shallower aquatic environments. In fact, OMEN-SED builds on a number of analytical models that were developed to investigate local, coupled nutrient and oxygen cycles in coastal sediments (e.g. Billen, 1982; Goloway and Bender, 1982; Jahnke et al., 1982; Slomp et al. 1996). Similar approaches were later successfully applied from oxic to anoxic sediments and at the regional coastal ocean scale (e.g. Ruardij and Van Raaphorst, 1995; Tromp et al., 1995; Gypens et al., 2008). In particular, Gypens et al., (2008) points out that accounting for secondary redox process in the boundary condition induces little error as: "Using a numerical model, Soetaert et al. (1996) showed that this re-oxidation mainly occurs at the oxic-anoxic transition interface."

Finally, the good agreement between OMEN-SED and the results obtained with a fully formulated numerical RTM (compare Section 3.3, allowing for overlapping TEA use) shows that this is not a critical limitation of OMEN-SED - even for shallow sediments.

We again want to refer to the paragraph we added to the limitations section (see last comment, pg 54 lines 3-8).

3. Comment:
Nitrogen dynamics include "the metabolic production of ammonium, nitrification, denitrification as well as ammonium adsorption". Denitrification is considered as a single-step process ignoring NO2- production/consumption and anaerobic ammonium oxidation (Anammox) which is undoubtedly a significant component of the biogeochemical nitrogen cycle (Devol, 2015). In other words, nitrogen dynamics is somewhat simplified. This simplification should be quantified/discussed in more details.

Response:
In the following we repeat the response given to the 2nd comment of Reviewer #1:

Anammox is implicitly included in the model. The organic nitrogen released during the denitrification process is assumed to be directly oxidized with nitrite to N_2 through a coupling between denitrification and anaerobic ammonium oxidation.
However, we would like to stress again that OMEN-SED is a benthic model designed for the coupling to ESMs. Most ESMs do not even explicitly resolve N-dynamics. In addition, OMEN-SED is a system/global scale model that aims to resolve the most pertinent biogeochemical dynamics on a global scale (including a paleoenvironmental context) and estimate the main SWI-fluxes and not a model that

aims at resolving specific local scale dynamics. Even most local scale RTM applications do not resolve DNRA and anammox explicitly. However, OMEN-SED could be easily adapted to explicitly resolve these processes if the specific application requires their representation (e.g. coastal ocean).

We included a sentence on this in the Section 2.2.3 "Nitrate and Ammonium" (pg. 12, lines 22-24)::
"*Anaerobic ammonium oxidation (anammox) is implicitly included in the model. The organic nitrogen released during denitrification is assumed to be directly oxidized with nitrite to N2 through a coupling between denitrification and anammox.*"

4. Comment:
The efficiency of binning procedure discussed in section 4.2.1 is doubtful. First of all, such binning assumes presence of STD bars on the plots. Also, I think that it would be more logical to group POC content into POC rain rate (RRPOC) classes rather than WD classes as RRPOC may significantly vary at different regions of the ocean of the same WD. Finally this binning gives a false impression of a good POC content fit. I realize that parameterization of multi-G model is beyond the scope of this sediment model development paper, therefore I suggest to use existing way to parameterize multi-G models and validate your model against the databases suggested in those studies (for example Stolpovsky et al., (submitted) https://www.biogeosciences-discuss.net/bg-2017-397/).

Response:
We thank the reviewer for the suggestion. We decided to follow suggestions from reviewer #1 and #3 (K. Wallmann) and shortened the cGENIE coupling section. Figure 12 (and its discussion) has been deleted from the manuscript. The R2 values in Figures 13 and 14 (as well as their discussion in the text) has been removed as well. Compare changes on pages 47-50.

The ranges of simulated SWI-fluxes from the stand-alone OMEN-SED model are already compared to the Stolpovsky et al., (2015) database in Figure 6.

If binned by RRPOC for uniform k-values, all grid-cells with same RRPOC have the same preservation in OMEN-SED. Therefore, this would not be very useful.

5. Comment:
POC is not a very good constraint, since measured POC is in large part the less reactive stuff that is left over after mineralization of the more reactive fractions. This was shown in Stolpovsky et al., 2015 paper (see the discussion in section 4.3). Fluxes at the SWI are believed to be a better constraint.

Response:
We shortened the coupling section of the manuscript (we removed the sensitivity analysis with the spatially uniform degradation rate constants, compare pages 47-50) and we will discuss an improved model-data analysis of the coupled model, using existing parameterizations and maps of SWI-fluxes, in a follow-up publication. Also compare response to comment 20 of reviewer #1:
 As stated in the manuscript (page 45, lines 23-26):
" *Our objective is not to perform and discuss a detailed calibration of the coupled models as this is beyond the scope of this sediment model development paper. Rather we want to showcase the feasibility of the model coupling, illustrate the range of results and thus information that can be generated with OMEN-SED and verify that model results capture the main observed global benthic biogeochemical*

features.”

Minor comments.

6. Comment:
Eq. 1: As a time and depth independent parameter, porosity should be moved out of differential in order to emphasize that it is constant: Porosity*dC/dt instead of d(Porosity*C)/dt.
Response:
This has been changed as indicated.

7. Comment:
P. 8, L. 1: It is not immediately clear that the authors are talking about water (not sediment) depth.
Response:
We agree that this is a bit misleading. We are actually talking about sediment depth. This has been changed in the revised manuscript to (pg. 8 lines 1-2):
*“All parameters in Eq. (1)**, apart from porosity and burial rate,** may vary with **sediment** depth and many reaction rate expressions depend on the concentration of other species. ”*

8. Comment:
Eq. 5: This representation sounds a bit odd. I think $z\infty$ should be replaced with zmax, as POC content at infinite depth believed to be zero.
Response:
The POC content of marine sediments does not tend to zero. A significant amount of POC is buried in marine sediments and enters the longterm C cycle (rock cycle). Without this imbalance between production and respiration, no O_2 would have accumulated in the atmosphere.
But we agree that the use of $z\infty$ is not ideal, as the sediment column in OMEN-SED is not modeled until infinite depth. We have replaced $z\infty$ with zmax in the entire manuscript.

9. Comment:
P. 9, L. 25: SWI is given without initial explanation.
Response:
The explanation has been added at this part in the manuscript: *“... **sediment-water interface (SWI)** …”*

10. Comment:
P. 25, L. 6 – 13: I agree that bioirrigation may enhance SWI fluxes of dissolved species, therefore I do not understand why this way of transport is technically ignored for all water depths (fir=1)?
Response:
In the following we repeat the answer given to Comment 10 of Reviewer #1:

The bioirrigation coefficient has been changed and is now represented by the empirical relationship with seafloor depth derived by Soetaert et al. (1996): fir = Min{1; 15.9 · z−0.43 }.
The text has been changed to (pg. 25 lines 21-23):
“Soetaert et al. (1996) derived an empirical relationship between fir and seafloor depth (fir = Min{1; 15.9 · z −0.43 }) based on observations from Archer and Devol (1992) and Devol and Christensen (1993) which is used in OMEN-SED“
And in the limitations section (pg. 54 lines 17-18):

"Also note that our 1-D diffusion/bioturbation model might not be appropriate to simulate non-accumulating permeable sands of the coastal ocean."

11. Comment:
P. 27, L. 28: PAWN is given without explanation.
Response:
As the name PAWN is derived from the authors names and not an acronym we do not think this information is of any value here.

12. Comment:
Fig. 7: Please add ticks and numbers to X-axis on H2S at 2213 and 4298m and NH4 at 108m. Some plates have very inconvenient ranges on horizontal axis, for example H2S at 4298m.
Response:
This has been changed as indicated.

13. Comment:
Sec. 3.3.2: I do not understand the rational for comparing OMEN-SED results with another model (Thullner et al. 2009). I would suggest comparing it to existing SWI flux database mentioned before (Stolpovsky et al., 2015). Also, reporting global denitrification rate modeled with OMEN-SED and its comparison with previous studies would support the model.
Response:
We evaluate the performance of OMEN-SED by comparing model results with data (section 3.2), as well as the results of a fully-formulated, numerical RTM (section 3.3). The comparison of OMEN-SED with the results of the numerically solved RTM allows evaluating to which extend simplifying assumptions (e.g. constant porosity, non-overlapping redox zones etc) affect simulation results and, thus, quantitatively test the performance of the computationally efficient OMEN-SED approach against the computationally expensive numerical approach.
The ranges of simulated SWI-fluxes from the stand-alone OMEN-SED model are already compared to the Stolpovsky et al., (2015) database in Figure 6.

14. Comment:
P. 55, L. 24 – 25: Bold assumption, I suggest to avoid such formulations. The major advantage of OMEN-SED is its tremendously low computation time which is so important for ESMs. As always, only two options of the following three can be true the same time: "quickly", "cheaply (super-computer is not needed)" or "qualitatively".

Response:
This is not an assumption, but the conclusion from the model-data and model-model comparison at the system scale. To clarify this, the sentence as been changed to (pg. 55 lines 5-6):
*"We have shown that the performance of OMEN-SED **at the system scale** is similar to that of a fully formulated, multi-component numerical model."*

Referee #3: K. Wallmann:

1. Comment:
The model ignores sulfide precipitation and pyrite formation. Consequently, dissolved sulfide produced by sulfate reduction and AOM at depth diffuses upward to be either oxidized by oxygen or released into ambient bottom waters. This is a very unrealistic set-up. In most sediments dissolved sulfide is removed from the pore water by pyrite precipitation while the remaining sulfide is oxidized with ferric iron, nitrate and nitrite before it can reach the oxic surface layer or the ocean. Aerobic sulfide oxidation is only important in highly reactive surface sediments where the diagenetic sequence is not maintained but several electron acceptors are used simultaneously. The model is based on the assumption that electron acceptors are used sequentially rather than simultaneously. Hence, it cannot simulate situations where aerobic sulfide oxidation is important but creates high rates of aerobic sulfide oxidation in geological settings where this process does in fact not occur. The authors should try to fix this problem. They could for example abandon the model parameter that defines the fraction of dissolved sulfide that escapes into bottom waters. In the modern ocean, sulfide leakage from sediments occurs only in very rare situations and it does not make sense to simulate these anoxic sediments with a model that ignores iron cycling, pyrite formation and sulfide precipitation. The authors could instead introduce a parameter that defines the fraction of sulfide that is precipitated as pyrite and update the alkalinity model accordingly.

Response:
We thank Prof. Wallmann for this very valid suggestion. We made the following changes to OMEN-SED:
When coupled to an ESM the γ_{H2S} value (fraction of H2S that is oxidised) becomes dependent on the bottom water oxygenation state. That is, γ_{H2S} = 1.0 for oxic bottom waters and a user defined value γ_{H2S} <1.0 for anoxic bottom waters. For simplicity this is still implemented as aerobic sulfide oxidation. In addition, we introduce another parameter (γ_{FeS}) representing the fraction of sulfide that is precipitated as pyrite (i.e. $0.0 <= \gamma_{FeS} < 1 - \gamma_{H2S}$) in the sulfate reduction zone. γ_{FeS} is an auxiliary parameter used as a fix until iron is explicitly represented (see pg. 54 lines 10-11, "... already planned future extensions of OMEN-SED include an explicit description of carbonate dissolution iron."). We thus assume that a user-defined fraction of the produced H2S precipitates as FeS(FeS2) in the sulfate reduction zone. If a user does not want to make any assumptions about FeS precipitation – it can be set it to 0.

The text, tables and equations (for O_2, SO_4, H_2S and alkalinity) are changed accordingly. The presented results have not been changed and we note that γ_{FeS} = 0.0 for all simulations, as we do not want to make any assumptions.

Changes made in:
2.2.2 Oxygen: Table 3 boundary condition 4.2 + Text pg. 12 lines 14-16:
" *It is assumed that respective fractions (γNH4 and γH2S) are directly reoxidised at the oxic/anoxic interface and the remaining fraction escapes reoxidation (or is precipitated as pyrite, γFeS).*"

2.2.4 Sulfate and Sulfide (pg. 14 lines 9-10): + Equations 23 + Table 5 boundary condition 5)
"In the sulfidic zone a defined fraction of sulfide, γFeS , can be precipitated as pyrite (in

the presented simulations γFeS = 0.0 as we do not want to make any assumptions about pyrite precipitation)."

2.2.7 Alkalinity: Table 8 boundary condition 5)
Text (pg. 20 lines 5-7):
*" In addition, the effect of secondary redox reactions, such as nitrification, sulfide and methane oxidation**, as well as pyrite precipitation,** are implicitly accounted for in the boundary conditions. "*

Text (pg. 20 lines 21-23):
*"The decrease of alkalinity due to oxidation of reduced species produced in the anoxic zones **and due to the precipitation of pyrite** (with stoichiometry ALK^{NIT} , ALK^{H2S} and **ALK^{FeS}**) is implicitly taken into account through the flux boundary condition at zox (Table 8 Eq. 5). "*

2.4.2 Stoichiometries and reaction parameters: (pg. 27 lines 19-23):
"However, when coupled to an ESM γH2S becomes dependent on the bottom water oxygenation state. That is, γH2 S = 1.0 for oxic bottom waters and a user defined value γH2S < 1.0 for anoxic bottom waters. The parameter γFeS represents the fraction of sulfide that is precipitated as pyrite in the sulfidic zone. The majority of H2S produced by sulfate reduction is reoxidised, but it is estimated that ∼ 10 − 25% is eventually buried as pyrite (Bottrell and Newton, 2006). However, this fraction can vary significantly over geological timescales (Berner, 1984). If a user does not want to make any assumptions about pyrite precipitation – it can be set to 0 (as in the results presented here)."

We also added pyrite precipiation to Table 1 and A1 in the Appendix.

2. Comment:
The authors use an empirical equation by Middleburg et al. (1997) to define burial velocity (w) as function of water depth (Eq. 46). Unfortunately, w is seriously overestimates by this equation. As an example, w at 1000 m water depth results as 160 cm kyr-1 applying Eq. 46 whereas the available data indicate global mean rates in the order of 10 – 20 cm kyr-1 for this water depth (Burwicz et al., 2011). The extremely high burial velocities derived from Eq. 46 compromise the TOC concentration and other model results especially when the model is applied at global scale.

Response:
The Middelburg et al. (1997) equation is just used in the stand-alone OMEN-SED version. When coupled to cGENIE we use the burial velocity of the ESM. In addition, the Burwicz et al. (2011) parameterisation is already added as an option in OMEN-SED (see pg. 24). We made it the default version for the stand-alone model. The sentence as been changed accordingly (compare pg. 24 line 28 – pg. 25 line 3).

3. Comment:
Comment 3.1: OMEN-SED is able to reproduce the strong down-core decrease in organic matter reactivity observed in marine sediments by using two or more organic matter fractions with widely different reactivity. This strength is nicely demonstrated in section 3.3 where the authors are able to show that typical pore water profiles are reproduced by the model applying kinetic constants (k1, k2) that span several orders of magnitude (Tab. 13). Subsequently, the authors try to reproduce the TOC distribution at the deep-

sea floor by coupling OMEN-SED to an earth system model. I think that TOC in surface sediments is not a good parameter to validate the model because almost the entire organic matter raining to the deep-sea floor is degraded in the surface sediment rather than preserved as sedimentary TOC. TOC concentrations in surface sediments at the deep-sea floor are governed by TOC rain rates, mass accumulation rates (burial velocity), adsorption of organic matter on mineral surfaces, and the kinetic properties of the very small refractory fraction that survives degradation (about 1 % of the total rain rate). The strength of OMEN-SED to degrade the reactive fractions in a meaningful way does not play out in this application.

Response:
We agree with the statement that TOC is not necessarily a good way to validate the coupled model and we would also favor fluxes or rates. However, we are not convinced that they give much better results if the database is limited. TOC in surface sediments was the data available on a global scale and also other ESM studies compare their results to it (e.g. HAMOCC, Palastanga et al. (2011)). As mentioned earlier, we will put in some more effort in a follow-up study where we compare calculated SWI-fluxes with observations.
In addition, as stated in the manuscript (page 45, lines 23-26):
" *Our objective is not to perform and discuss a detailed calibration of the coupled models as this is beyond the scope of this sediment model development paper. Rather we want to showcase the feasibility of the model coupling, illustrate the range of results and thus information that can be generated with OMEN-SED and verify that model results capture the main observed global benthic biogeochemical features.*"

Comment 3.2: Moreover, the model results are unrealistic. The best fit to the TOC data is apparently obtained assuming that the organic matter flux to the seabed is composed of two TOC fractions with very low reactivity in the order of 0.001 – 0.01 yr-1 (Fig. 12). This result is not consistent with the case study presented in section 3.3 that yields much higher k values (Tab. 13).

Response:
The low reactivities obtained for the global application (e.g. Fig. 12 – old manuscript) agree with published results (see Arndt et al., (2013)), as well as with the results obtained with HAMOCC using a 1G-model (they found kox=0.005 yr-1 & kanox=0.002 yr-1 for deep sea sediments, Palastanga et al. (2011)). In addition, our simulated oxygen penetration depths compare well with observations (see Fig. 16). Especially deep sea sites in the gyres are characterised by very low POC input and degradation rates which causes O2 to diffuse down to the basement of the sediments (Fischer et al., 2009; D'Hondt et al., 2015).
The sites used for the stand-alone case study in section 3.3 where not really deep sea sites (complete data sets from deep sea sites within gyres are difficult to obtain).

However, we decided to shorten the cGENIE coupling section (compare also response to your comment 3.3). The sensitivity analysis for the spatially uniform degradation rate constants (Figure 12) and it's discussion has been removed (compare pages 47-50).

Comment 3.2: Moreover, we have shown previously that this very low reactivity is not consistent with the benthic fluxes of oxygen and nitrate that have been measured at the seabed (Stolpovsky et al.,

2015). The error may be caused by the too high burial velocities applied in OMEN-SED (Eq. 46) and/or may be related to the rain rate and reactivity of organic matter calculated in GENIE.

Response:
The Stolpovsky et al. (2015) database is a very valuable source of information and we will compare our calculated fluxes using the coupled model with it in the follow-up study. The ranges of simulated SWI-fluxes from the stand-alone OMEN-SED model are already compared to the database in Figure 6. However, we would also argue that the Stolpovsky et al. (2015) database does not contain a representative amount of very deep ocean sites (e.g. within ocean gyres) characterised by very low SWI-fluxes (see e.g. Fischer et al., 2009; D'Hondt et al., 2015). D'Hondt el al. (2009) for instance found that the net rate of diagenetic degradation in the South Pacific Gyre is 1 to 3 orders of magnitude lower than at previously explored sites and they suggest that almost 50% of the worlds ocean may be characterised by these rates. In a more recent study D'Hondt et al. (2015) suggest: "...that oxygen and aerobic communities may occur throughout the entire sediment sequence in 15–44% of the Pacific and 9–37% of the global sea floor."

Comment 3.3: I would encourage the authors to delete the entire section 4 of the paper because it does not add useful information but presents rather misleading results. They should aim to present other more useful applications of their highly innovative analytical model in follow-up publications.

Response:
Here, we repeat parts of the response to comment 20 of reviewer #1:
We think that demonstrating how OMEN-SED can be coupled to an ESM and illustrating the type of output/information generated by OMEN-SED within such a coupling is a central aspect of the model description paper.
However, we agree trimming down this section (as in the re-submitted version). We will discuss an improved model-data analysis (also using observations of SWI-fluxes) .
Specifically, the sensitivity analysis for the spatially uniform degradation rate constants (Figure 12) and it's discussion has been removed (compare pages 47-50).

OMEN-SED ~~0.9~~1.0: A novel, numerically efficient organic matter sediment diagenesis module for coupling to Earth system models

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Abstract. We present the first version of OMEN-SED (Organic Matter ENabled SEDiment model), a new, one-dimensional analytical early diagenetic model resolving organic matter cycling and associated biogeochemical dynamics in marine sediments designed to be coupled to Earth system models. OMEN-SED explicitly describes organic matter (OM) cycling as well as associated dynamics of the most important terminal electron acceptors (i.e. O_2 , NO_3, SO_4) and methane (CH_4), related reduced substances (NH_4, H_2S), macronutrients (PO_4) and associated pore water quantities (ALK, DIC). Its reaction network accounts for the most important primary and secondary redox reactions, equilibrium reactions, mineral dissolution and precipitation, as well as adsorption and desorption processes associated with OM dynamics that affect the dissolved and solid species explicitly resolved in the model. To represent a redox-dependent sedimentary P cycle we also include a representation of the formation and burial of Fe-bound P and authigenic Ca-P minerals. Thus, OMEN-SED is able to capture the main features of diagenetic dynamics in marine sediments and, therefore, offers similar predictive abilities than a complex, numerical diagenetic model. Yet, its computational efficiency allows its coupling to global Earth system models and therefore the investigation of coupled global biogeochemical dynamics over a wide range of climate relevant timescales. This paper provides a detailed description of the new sediment model, an extensive sensitivity analysis, as well as an evaluation of OMEN-SED's performance through comprehensive comparisons with observations and results from a more complex numerical model. We find solid phase and dissolved pore water profiles for different ocean depths are reproduced with good accuracy and simulated terminal electron acceptor fluxes fall well within the range of globally observed fluxes. Finally, we illustrate its application in an Earth system model framework by coupling OMEN-SED to the Earth system model cGENIE and tune the OM degradation rate constants to optimise the fit of simulated benthic OM contents to global observations. We find simulated sediment characteristics of the coupled model framework, such as OM degradation rates, oxygen penetration depths and sediment-water interface fluxes are generally in good agreement with observations and in line with what one would expect on a global scale. Coupled to an Earth system model, OMEN-SED is thus a powerful tool that will not only help elucidate the role of benthic-pelagic exchange processes in the evolution and, in particular, the termination of a wide range of climate events, but will also allow a direct comparison of model output with the sedimentary record - the most important climate archive on Earth.

1 Introduction

Marine surface sediments are key components in the Earth system. They host the largest carbon reservoir within the surficial Earth system, provide the primary long term sink for atmospheric CO_2, recycle nutrients, and represent the most important geochemical archive used for deciphering past changes of biogeochemical cycles and climate (e.g. Berner, 1991; Archer and Maier-Reimer, 1994; Ridgwell and Zeebe, 2005; Arndt et al., 2013). Physical and chemical processes in sediments (i.e. diagenetic processes) depend on the water column and vice versa: Diagenesis is controlled by the external supply of solid material (e.g. organic matter, calcium carbonate, opal) from the water column and is affected by overlying bottom water concentrations of solutes. At the same time, sediments impact the water column directly either by short- and long-term storage of deposited material or diagenetic processing of deposited material and transport of terminal electron acceptors (e.g. O_2, SO_4) into the sediments, as well as metabolic products (e.g. nutrients, DIC) to the overlying bottom waters. This so-called benthic-pelagic coupling is essential for understanding global biogeochemical cycles and climate (e.g. Archer and Maier-Reimer, 1994; Archer et al., 2000; Soetaert et al., 2000; Mackenzie, 2005).

Biological primary production of organic matter (OM, generally represented in its simple form CH_2O in equation R1) and the reverse process of degradation can be written in a greatly simplified reaction as:

$$CO_2 + H_2O \rightleftharpoons CH_2O + O_2. \tag{R1}$$

On geological timescales, production of OM is generally greater than degradation, which results in some organic matter being buried in marine sediments and oxygen accumulating in the atmosphere. Thus, burial of OM deep into the sediment leads to net oxygen input to, and CO_2 removal from the atmosphere (Berner, 2004). On shorter timescales, the upper few meters of the sediments, where early diagenesis occurs are specifically important, as this zone controls whether a substance is recycled to the water column or buried for a longer period of time in the deeper sediments (Hensen et al., 2006). Most biogeochemical cycles and reactions in this part of marine sediments can be related either directly or indirectly to the degradation of organic matter (Middelburg et al., 1993; Arndt et al., 2013). Oxygen and nitrate for instance, the highest energy yielding electron acceptors, are preferentially consumed in the course of the degradation of organic matter, resulting in the release of ammonium and phosphorus to the pore water. As such, degradation of OM in the sediments can profoundly affect the oxygen and nutrient inventory of the ocean and thus primary productivity (Van Cappellen and Ingall, 1994; Lenton and Watson, 2000). Furthermore, organic matter degradation releases metabolic CO_2 to the pore water, causing it to have a lower pH and carbonate ion concentration, thus provoking the dissolution of calcium carbonate $CaCO_3$ (Emerson and Bender, 1981).

Benthic nutrient recycling from marine sediments has been suggested to play a key role for climate and ocean biogeochemistry throughout Earth history. For example, feedbacks between phosphorus storage and erosion from shelf sediments and marine productivity have been hypothesised to play an important role for glacial/interglacial atmospheric CO_2 changes (Broecker, 1982; Ruttenberg, 1993). Furthermore, benthic nutrient recycling from anoxic sediments has been invoked to explain the occurrence of more extreme events in Earth history, for instance Oceanic Anoxic Events (OAEs, e.g. Van Cappellen and Ingall, 1994; Mort et al., 2007; Tsandev and Slomp, 2009). OAEs represent severe disturbances of the global carbon, oxygen and nutrient cycles of the ocean and are usually characterized by widespread bottom water anoxia and photic zone eu-

xinia (Jenkyns, 2010). One way to explain the genesis and persistence of OAEs is increased oxygen demand due to enhanced primary productivity. Increased nutrient inputs to fuel primary productivity may in turn have come from marine sediments as the burial efficiency of phosphorus declines when bottom waters become anoxic (Ingall and Jahnke, 1994; Van Cappellen and Ingall, 1994). The recovery from OAE like conditions is thought to involve the permanent removal of excess CO_2 from the

5 atmosphere and ocean by burying carbon in the form of organic matter in marine sediments (e.g. Arthur et al., 1988; Jarvis et al., 2011), which is consistent with the geological record of widespread black shale formation (Stein et al., 1986). Models capable of simulating not only the expansion and intensification of oxygen minimum zones, but also of predicting how the underlying sediments interact are hence needed.

Quantifications of diagenetic processes in the sediments are possible through the application of idealised mathematical

10 representations, or so-called diagenetic models (see e.g. Berner, 1980; Boudreau, 1997). A plethora of different approaches have been developed, mainly following two distinct directions (see Arndt et al., 2013, for an overview). First, state-of-the art vertically resolved numerical models simulating the entire suite of essential coupled redox and equilibrium reactions within marine sediments (e.g. BRNS, Aguilera et al., 2005; CANDI, Boudreau, 1996; MEDIA, Meysman et al., 2003; MUDS, Archer et al., 2002; STEADYSED, Van Cappellen and Wang, 1996). These "complete", multi-component steady-state or non-steady-

15 state models, thus resolve the resulting characteristic redox-zonation of marine sediments through explicitly accounting for oxic OM degradation, denitrification, oxidation by manganese and iron (hydr)oxides, sulfate reduction and methanogenesis as well as the reoxidation of reduced byproducts (i.e. NH_4, Mn^{2+}, Fe^{2+}, H_2S, CH_4, see e.g. Regnier et al., 2011). Furthermore, they incorporate various mineral dissolution and precipitation reactions, as well as fast equilibrium sorption processes for example of NH_4, PO_4 and metal ions (i.e. Mn^{2+}, Fe^{2+} and Mg^{2+}, compare Van Cappellen and Wang, 1996; Meysman et al.,

20 2003). Modelled, depth-dependent, transport processes usually comprise advection, diffusion, bioturbation and bio-irrigation. This group of diagenetic models generally describes OM degradation via a so-called multi-G approach (Jørgensen, 1978; Berner, 1980), thus dividing the bulk organic matter pool into a number of compound classes that are characterised by different degradabilities k_i. Alternative approaches, so-called continuum models (Middelburg, 1989; Boudreau and Ruddick, 1991), assume a continuous distribution of reactive types but, although conceptually superior, are much less popular (Arndt et al.,

25 2013). These complex, multi-component models have a great potential for quantifying diagenetic dynamics at sites where comprehensive observational data sets are available to constrain its model parameters (see e.g. Boudreau et al., 1998; Wang and Van Cappellen, 1996; Thullner et al., 2009, for applications). However, due to the high degree of coupled processes and depth-varying parameters, the diagenetic equation needs to be solved numerically, thus resulting in a very high computational demand and consequently rendering their application in an Earth system model (ESM) framework with a large number of grid

30 points prohibitive. Additionally, their global applicability is seriously compromised by the restricted transferability of model parameters from one site to the global scale (Arndt et al., 2013).

The second group of diagenetic models emerged during the early days of diagenetic modelling when computing power was severely restricted (e.g. Berner, 1964). These models solve the diagenetic equation analytically, thus providing an alternative and computationally more efficient approach. Finding an analytical solution, especially when complex reaction networks are

35 to be considered, is not straightforward and analytical models are thus usually less sophisticated and comprehensive than nu-

merical models and generally require the assumption of steady state conditions. It has been shown that the complexity of the reaction network can be reduced by dividing the sediment column into distinct zones and accounting for the most pertinent biogeochemical processes within each zone, thus increasing the likelihood of finding an analytical solution without oversimplifying the problem. Analytical approaches with distinct biogeochemical zones were implemented and used in the seventies and eighties to describe observed pore water profiles (e.g. Vanderborght and Billen, 1975; Vanderborght et al., 1977; Billen, 1982; Goloway and Bender, 1982; Boudreau and Westrich, 1984) and later for inclusion into multi-box ecosystem models (e.g. Ruardij and Van Raaphorst, 1995; Gypens et al., 2008) and global Earth system models (Tromp et al., 1995). However, in addition to the oxic zone these models generally only describe one anoxic zone explicitly, either a denitrification (Vanderborght and Billen, 1975; Billen, 1982; Goloway and Bender, 1982; Ruardij and Van Raaphorst, 1995; Gypens et al., 2008) or a sulfate reduction zone (Boudreau and Westrich, 1984; Tromp et al., 1995). Furthermore, the approaches of Vanderborght and Billen (1975), Goloway and Bender (1982) and Tromp et al. (1995) do not explicitly account for reduced species (i.e. NH_4 and H_2S, respectively).

In most current ESMs sediment-water dynamics are either neglected or treated in a very simplistic way (Soetaert et al., 2000; Hülse et al., 2017). Most Earth system Models of Intermediate Complexity (EMICs) and also some of the higher resolution Earth system/climate models represent the sediment-water interface either as a reflective or a conservative/semi-reflective boundary (Hülse et al., 2017). Thus, all particulate material deposited on the seafloor is either instantaneously consumed (reflective boundary), or a fixed fraction is buried in the sediments (conservative/semi-reflective boundary). Both highly simplified approaches furthermore completely neglect the exchange of solute species through the sediment-water interface and, therefore, cannot resolve the complex benthic-pelagic coupling. However, due to their computational efficiency, both representations are often used in global biogeochemical models (e.g. Najjar et al., 2007; Ridgwell et al., 2007; Goosse et al., 2010). Analytical diagenetic models represent the most complex description of diagenetic dynamics in Earth system models. Examples of global ESMs employing a vertically resolved diagenetic model are NorESM (Tjiputra et al., 2013) and HAMOCC (Palastanga et al., 2011; Ilyina et al., 2013), both using a version of Heinze et al. (1999). None of the EMICs reviewed by Hülse et al. (2017) use such a sediment representation. DCESS (Shaffer et al., 2008) and MBM (Munhoven, 2007) are box models employing a vertically resolved diagenetic model. These analytic models account for the most important transport processes (i.e advection, bioturbation and molecular diffusion) through basic parametrizations and include fewer biogeochemical reactions which are generally restricted to the upper, bioturbated 10 cm of the sediments. Pore water species explicitly represented in DCESS (Shaffer et al., 2008) and the HAMOCC model of Heinze et al. (1999) and Palastanga et al. (2011) are restricted to DIC, TA, PO_4 and O_2. The MEDUSA model (Munhoven, 2007) considers CO_2, HCO_3^-, CO_3^{2-} and O_2. Other species produced or consumed during OM degradation are neglected. Thus, with oxygen being the only TEA explicitly modelled, the influence of reduced species is only implicitly included in the boundary conditions for O_2. A newer version of the HAMOCC model is a notable exception, as Ilyina et al. (2013) include NO_3 and denitrification explicitly. Furthermore, the version of Palastanga et al. (2011) represents an redox-dependent explicit sedimentary phosphorus cycle. Yet, reoxidation of reduced byproducts, so-called secondary redox reactions (e.g. oxidation of NH_4, H_2S or CH_4), or sorption processes are not included in any of the discussed models. Furthermore, these global models assume that the sedimentary organic matter pool is composed of just

a single compound class which is either degraded with a globally invariant degradation rate constant (Munhoven, 2007) or a fixed rate constant depending on local oxygen concentrations (Shaffer et al., 2008; Palastanga et al., 2011).

Obviously, such a simplification of the OM pool can neither account for the observed vast structural complexity in natural organic matter and its resulting different degradation rates nor for the rapid decrease in OM degradability in the uppermost centimetres of the sediments (Arndt et al., 2013). It has been suggested that at least a 3G approach is necessary to accurately represent organic matter dynamics in this part of the sediments where most OM is degraded (e.g. Soetaert et al., 1996). Even more restrictive is the use of O_2 as the only TEA and the complete absence of reduced substances and related secondary redox reactions. For the majority of the modern sediments (i.e. in the deep-ocean) O_2 is the primary electron acceptor, however, recent model and data studies have reported that sulfate reduction is the dominant degradation pathway on a global average (with contributions of 55-76% Canfield et al., 2005; Jørgensen and Kasten, 2006; Thullner et al., 2009). Oxygen becomes progressively less important as TEA with decreasing seafloor depth and sulfate reduction has been shown to account for 83% of OM degradation in coastal sediments (Krumins et al., 2013). In these environments most O_2 is used to reoxidise reduced substances produced during anaerobic degradation (Canfield et al., 2005; Thullner et al., 2009). Thus, the in situ production of e.g. NO_3 and SO_4 through oxidation of NH_4 and H_2S forms an important sink for O_2 which is entirely neglected in current sediment representations in global models. In addition, the lack of anoxic degradation pathways in these models limits their application to oxic oceans. Currently no analytical sediment model exists that can be used under anoxic conditions. Due to the lack of an appropriate sedimentary P cycle (with the exception of the HAMOCC version of Palastanga et al., 2011), no current global ESM is able to model the redox dependent P release from marine sediments and its implications for primary productivity, global biogeochemical cycles and climate. A sediment model suitable for the coupling to an ESM and enabling a wide range of paleo questions to be addressed has to provide a robust quantification of organic (and inorganic) carbon burial fluxes, benthic uptake/return fluxes of oxygen, growth-limiting nutrients and reduced species, as well as anoxic degradation pathways. As a consequence, the reaction network must account for the most important primary and secondary redox reactions, equilibrium reactions, mineral precipitation/dissolution and adsorption/desorption, resulting in a complex set of coupled reaction-transport equations.

Therefore, we developed the OrganicMatter ENabled SEDiment model (OMEN-SED), a new, one-dimensional, numerically efficient diagenetic model. OMEN-SED builds upon and stands in the tradition of earlier stand-alone, analytical diagenetic models (Vanderborght et al., 1977; Billen, 1982; Goloway and Bender, 1982; Boudreau, 1991), as well as of analytical diagenetic models developed for the coupling to regional scale ecosystem or global Earth system models (Ruardij and Van Raaphorst, 1995; Tromp et al., 1995; Heinze et al., 1999; Gypens et al., 2008).

OMEN-SED is the first analytical model to explicitly describe OM cycling as well as associated dynamics of the most important TEAs (i.e. O_2, NO_3, SO_4), related reduced substances (NH_4, H_2S), the full suite of secondary-redox reactions, macronutrients (PO_4) and associated pore water quantities (ALK, DIC). To represent a redox-dependent sedimentary P cycle we consider the formation and burial of Fe-bound P and authigenic Ca-P minerals. Thus, while OMEN-SED captures most of the features of a complex, numerical diagenetic model, its computational efficiency allows the coupling to global Earth system models and therefore the investigation of coupled global biogeochemical dynamics over different timescales. Here,

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the model is presented as a 2G-approach, however, OMEN-SED can be easily extended to a Multi-G approach. The first part of the paper provides a detailed description of OMEN-SED (Section 2). This includes descriptions of the general model approach (Section 2.1), of the conservation equations for all explicitly represented biogeochemical tracers (Section 2.2), as well as a summary of global relationships used to constrain reaction and transport parameters in OMEN-SED (Section 2.4). In addition, a generic algorithm is described which is used to match internal boundary conditions and to determine the integration constants for the analytical solutions (Section 2.3). In order to validate the stand-alone version of OMEN-SED, the second part of the paper performs an extensive sensitivity analysis for the most important model parameters and resulting sediment-water interface fluxes are compared with a global database (Section 3.1). In addition, results of the stand-alone model are compared with observed pore water profiles from different ocean depths (Section 3.2) and OMEN-SED simulations of TEA-fluxes along a typical ocean transect are compared with observations and results from a complete, numerical diagenetic model (Section 3.3). Thereafter, OMEN-SED is coupled to the carbon-centric version of the "GENIE" Earth system model (cGENIE, Ridgwell et al., 2007, Section 4.1). Sensitivity studies are carried out using this coupled model and modelled organic matter concentrations in the surface sediments are compared to a global database (Seiter et al., 2004, Section 4.2). We finally discuss potential applicabilities of OMEN-SED and critically analyse model limitations (Section 5).

2 Model Description

OMEN-SED is implemented as a FORTRAN version that can be easily coupled to any pelagic, biogeochemical model via the coupling routine **OMEN_SED_main**. In addition, OMEN-SED exists as a stand-alone version implemented in MATLAB and the entire model can be executed on a standard personal computer in less than 0.1 seconds. The source code of both, the FORTRAN and the MATLAB stand-alone version, as well as instructions for executing OMEN-SED and for plotting model results are available as a supplement to this paper.

The following section provides a detailed description of OMEN-SED and the fundamental equations underlying the model are highlighted. Tables 1 and A1 summarise the biogeochemical reaction network and Tables 9 and 10 provide a glossary of model parameters along with their respective units.

2.1 General Model Approach

In OMEN-SED, the calculation of benthic uptake, recycling and burial fluxes is based on the vertically resolved conservation equation for solid and dissolved species in porous media (e.g. Berner, 1980; Boudreau, 1997):

$$\frac{\partial \xi C_i}{\partial t} \xi \frac{\partial C_i}{\partial t} = -\frac{\partial}{\partial z}\left(-\xi D_i \frac{\partial C_i}{\partial z} + \xi w C_i\right) + \xi \sum_j R_i^j \tag{1}$$

where C_i is the concentration of biogeochemical species i, ξ equals the porosity ϕ for solute species and $(1-\phi)$ for solid species. The term z is the sediment depth, t denotes the time, D_i is the apparent diffusion coefficient of species i, w is the burial rate and $\sum_j R_i^j$ represents the sum of all biogeochemical rates j affecting species i.
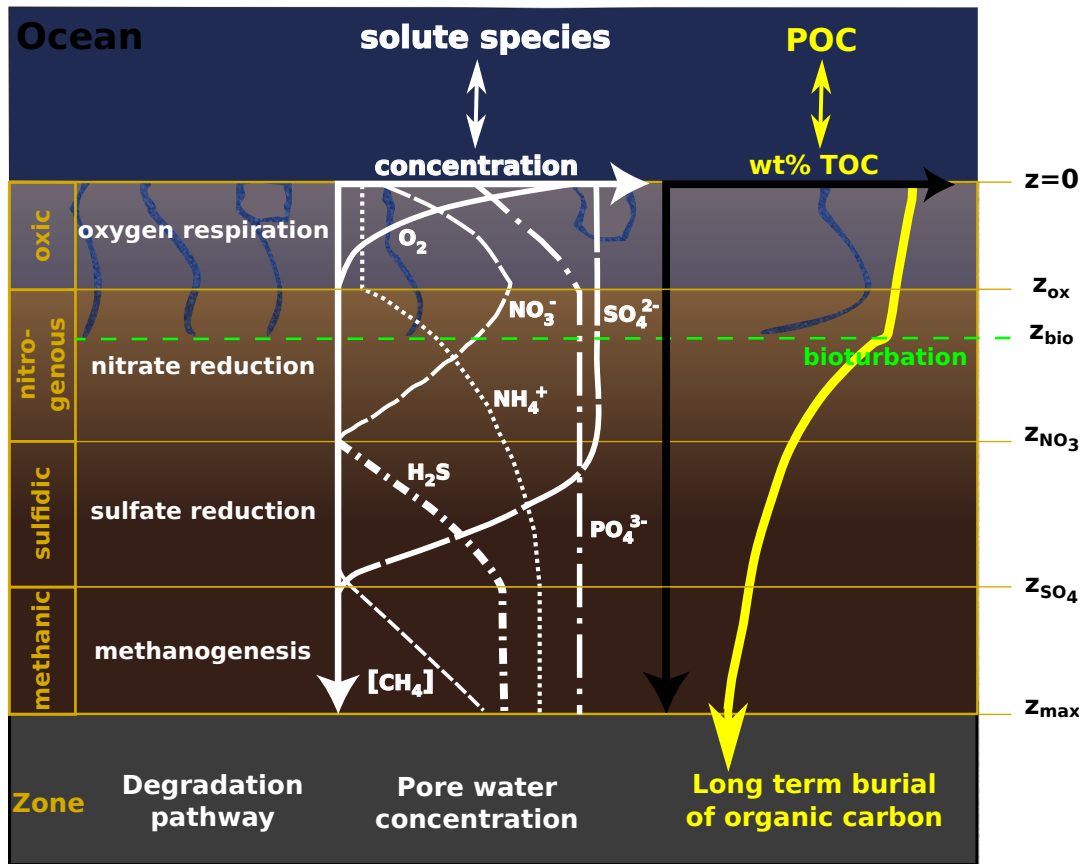
Figure 1. Schematic of the different modelled species and zones in OMEN-SED. Here showing the case $z_{ox} < z_{bio} < z_{NO_3} < z_{SO_4}$.

OMEN-SED accounts for both the advective, as well as the diffusive transport of solid and dissolved species. They are buried in the sediment according to a constant burial rate w, thus neglecting the effect of sediment compaction (i.e. $\frac{\partial \phi}{\partial z} = 0$) due to mathematical constraints. The molecular diffusion of dissolved species is described by Fick's law applying a species-specific apparent diffusion coefficient, $D_{\mathrm{mol},i}$. In addition, the activity of infaunal organisms in the bioturbated zone is simulated using a diffusive term (e.g. Boudreau, 1986), with a constant bioturbation coefficient D_{bio} in the bioturbated zone, while D_{bio} is set to zero below the maximum bioturbation depth, z_{bio}. The pumping activity by burrow-dwelling animals and the resulting ventilation of tubes, the so-called bioirrigation, is encapsulated in a factor f_{ir} that enhances the molecular diffusion coefficient (hence, $D_{i,0} = D_{\mathrm{mol},i} \cdot f_{ir}$, Soetaert et al., 1996). The reaction network of OMEN-SED accounts for the most important primary and secondary redox reactions, equilibrium reactions, mineral dissolution and precipitation, as well as adsorption and desorption processes associated with OM dynamics that affect the dissolved and solid species explicitly resolved in the model. Tables 1 and A1 provide a summary of the reactions and biogeochemical tracers considered in OMEN-SED together with their respective reaction stoichiometries.

Table 1. Reactions and biogeochemical tracers implemented in the reaction network of OMEN-SED. The primary and secondary redox reactions are listed in the sequence they occur with increasing sediment depth.

	Description
Primary redox reactions	Degradation of organic matter via aerobic degradation, denitrification, sulfate reduction, methanogenesis (implicit)
Secondary redox reactions	Oxidation of ammonium and sulfide by oxygen, anaerobic oxidation of methane by sulfate
Adsorption/Desorption	Ad-/Desorption of P on/from $Fe(OH)_3$, NH_4 adsorption, PO_4 adsorption
Mineral precipitation	Formation of authigenic P, pyrite precipitation (implicit)
Biogeochemical tracers	Organic matter (2-G or pseudo 3-G), oxygen, nitrate, ammonium, sulfate, sulfide (hydrogen sulfide), phosphate, Fe-bound P, DIC, ALK

All parameters in Eq. (1), apart from porosity and burial rate, may vary with sediment depth and many reaction rate expressions depend on the concentration of other species. Expressing Eq. (1) for a set of chemical species thus results in a non-linear, coupled set of equations that can only be solved numerically. However, OMEN-SED is designed for the coupling to Earth system models and, therefore, cannot afford a computationally expensive numerical solution. Instead, similar to early, analytical

5   diagenetic models, a computationally efficient analytical solution of Eq. (1) can be derived by 1) assuming steady state conditions (i.e. $\frac{\partial C_i}{\partial t} = 0$) and 2) reducing the vertical variability in parameters and reaction rate expressions by dividing the sediment column into a number of functional biogeochemical zones (Fig. 1, compare e.g. Billen, 1982; Goloway and Bender, 1982; Ruardij and Van Raaphorst, 1995; Tromp et al., 1995; Gypens et al., 2008, for similar solutions). More specifically, OMEN-SED follows Berner (1980) by dividing the sediment column into: I) a bioturbated and II) a non-bioturbated zone defined by an im-

10   posed, constant bioturbation depth z_{bio} (Fig. 1). Furthermore, it resolves the dynamic redox stratification of marine sediments by dividing the sediment into 1) an oxic zone delineated by the oxygen penetration depth z_{ox}; 2) a denitrification (or nitrogenous) zone situated between z_{ox} and the nitrate penetration depth z_{NO_3}; 3) a sulfate reduction zone situated between z_{NO_3} and the sulfate penetration depth z_{SO_4}; and 4) a methanogenic zone situated below z_{SO_4} (Fig. 1). Although in each of these zones Eq. (1) is applied with depth invariant parameters, parameter values may differ across zones. The biogeochemical zones are

15   linked by stating continuity in both concentrations and fluxes at the dynamic, internal boundaries ($z_b \in \{z_{bio}, z_{ox}, z_{NO_3}, z_{SO_4}\}$, compare e.g. Billen, 1982; Ruardij and Van Raaphorst, 1995). Note that these boundaries are dynamic because their depth varies in response to changing ocean boundary conditions and forcings (see Section 2.3.1 for details). Furthermore, the maximum bioturbation depth is not restricted to a specific biogeochemical zone, hence OMEN-SED allows bioturbation to occur in the anoxic zones of the sediment (here all zones $z > z_{ox}$ combined).

20   The formulation of the reaction term in Eq. (1) varies between zones and encapsulates the most pertinent reaction processes within the respective zone (see Section 2.2), thus simplifying the mathematical description of the reaction network while retaining most of its biogeochemical complexity. One such simplification is that the solid phase iron and manganese oxidants and its reductants are not considered in the reaction network. All consumption or production processes of dissolved species related to the degradation of organic matter are a function of the organic matter concentration. Because organic matter degradation is

described as a first-order degradation, these processes can be expressed as a series of exponential terms ($\sum_j \alpha_j \exp(-\beta_j z)$, see Eq. (2)). In addition, slow adsorption/desorption and mineral precipitation processes can be expressed as zero or first order (reversible) reaction (Q_m or $k_l \cdot C_i$, in Eq. (2)). Fast adsorption is described as an instantaneous equilibrium reaction using a constant adsorption coefficient K_i. The reoxidation of reduced substances is accounted for implicitly by adding a (consumption/production) flux to the internal boundary conditions (see Sections 2.2.2, 2.2.3 and 2.2.4). This simplification has been used previously by Gypens et al. (2008) for nitrate and ammonium and can be justified as it has been shown that the reoxidation mainly occurs within a thin layer at the oxic/anoxic interface (Soetaert et al., 1996). The general reaction-transport equation underlying OMEN-SED is thus given by:

$$\frac{\partial C_i}{\partial t} = 0 = \frac{D_i}{1+K_i}\frac{\partial^2 C_i}{\partial z^2} - w\frac{\partial C_i}{\partial z} - \frac{1}{1+K_i}\left(\sum_j \alpha_j \exp(-\beta_j z) + \sum_l k_l \cdot C_i - \sum_m Q_m\right) \tag{2}$$

where $1/\beta_j$ can be interpreted as the length scale and α_j as the relative importance (or the magnitude at $z=0$) of reaction j (Boudreau, 1997), k_l are generic first order reaction rate constants and Q_m are zero-order (or constant) reaction rates.

The analytical solution of Eq. (2) is of the general form:

$$C_i(z) = A \cdot \exp(az) + B \cdot \exp(bz) + \sum_j \frac{\alpha_j}{D\beta_j^2 - w\beta_j - \sum_l k_l} \cdot \exp(-\beta_j z) + \frac{\sum_m Q_m}{\sum_l k_l} \tag{3}$$

with

$$a = \frac{w - \sqrt{w^2 + 4 \cdot D \cdot \sum_l k_l}}{2 \cdot D}, \quad b = \frac{w + \sqrt{w^2 + 4 \cdot D \cdot \sum_l k_l}}{2 \cdot D} \tag{4}$$

where A and B are integration constants that can be determined by applying a set of internal boundary conditions (see Section 2.3) and $D = \frac{D_i}{1+K_i}$.

Based on Eq. (2) and its analytical solution Eq. (3), OMEN-SED returns the fraction of particulate organic carbon (POC) buried in the sediment, f_{POC}, as well as the benthic uptake/return fluxes F_{C_i} of dissolved species C_i (in mol cm^{-2} year^{-1}) in response to the dynamic interplay of transport and reaction processes under changing boundary conditions and forcings:

$$f_{\text{POC}} = \frac{\text{POC}(z_\infty)}{\text{POC}(0)}\frac{\text{POC}(z_{\text{max}})}{\text{POC}(0)} \tag{5}$$

$$F_{C_i} = \phi(0)\left(D_i \frac{\partial C_i(z)}{\partial z}\bigg|_{z=0} - w \cdot C_i(0)\right) \tag{6}$$

where w is the deposition rate, D_i is the diffusion coefficient and POC(0), ~~POC(z_∞)~~ POC(z_{max}), $C_i(0)$ denote the concentration of POC and dissolved species i at the ~~SWI~~ sediment-water interface (SWI) and at the lower sediment boundary, respectively.

2.2 Conservation Equations and Analytical Solution

The following sections provide a detailed description of the conservation equations and analytical solutions for each chemical species that is resolved in this version of OMEN-SED.

2.2.1 Organic Matter or Particulate Organic Carbon (POC)

5 In marine sediments, organic matter (or in the following called particulate organic carbon, POC) is degraded by heterotrophic activity coupled to the sequential utilisation of terminal electron acceptors (TEAs) according to the free energy gain of the half-reaction $(O_2 > NO_3^- > MnO_2 > Fe(OH)_3 > SO_4^{2-}$, e.g. Stumm and Morgan, 2012). Once all TEAs are depleted, organic matter is degraded via methanogenesis. Here, organic matter degradation is described via a multi G-model approach (Jørgensen, 1978), dividing the bulk OM into a number i of discrete compound classes POC_i characterised by class-specific first-order

10 degradation rate constants k_i. The conservation equation for organic matter dynamics is thus given by:

$$\frac{\partial POC_i}{\partial t} = 0 = D_{POC_i} \frac{\partial^2 POC_i}{\partial z^2} - w \frac{\partial POC_i}{\partial z} - k_i \cdot POC_i \tag{7}$$

with $D_{POC_i} = D_{bio}$ for $z \leq z_{bio}$ and $D_{POC_i} = 0$ for $z > z_{bio}$. Integration of equations (7) yields the following general solutions for the bioturbated and non-bioturbated layers:

I. Bioturbated zone ($z \leq z_{bio}$)

15 $$POC_i^I(z) = A_{1i} \cdot exp(a_{1i}z) + B_{1i} \cdot exp(b_{1i}z) \tag{8}$$

II. Non-bioturbated zone ($z_{bio} < z$)

$$POC_i^{II}(z) = A_{2i} \cdot exp(a_{2i}z) \tag{9}$$

where

$$a_{1i} = \frac{w - \sqrt{w^2 + 4 \cdot D_{POC_i} \cdot k_i}}{2 \cdot D_{POC_i}}, \quad b_{1i} = \frac{w + \sqrt{w^2 + 4 \cdot D_{POC_i} \cdot k_i}}{2 \cdot D_{POC_i}}, \quad a_{2i} = -\frac{k_i}{w} \tag{10}$$

20 Determining the integration constants ($A_{1,i}$, $B_{1,i}$, $A_{2,i}$) requires the definition of a set of boundary conditions (Table 2). For organic matter, OMEN-SED applies a known concentration ~~/flux~~ at the sediment-water interface and assumes continuity across the bottom of the bioturbated zone, z_{bio}. When OMEN-SED is coupled to an ESM, the POC depositional flux from the coupled ocean model is converted to a concentration by solving the flux divergence equation (51). The integration constants ($A_{1,i}$, $B_{1,i}$, $A_{2,i}$) are thus given by:

25 $$B_{1i} \overset{BC1)}{=} POC_{0i} - A_{1i} \tag{11}$$

$$A_{2i} \overset{BC2)}{=} \frac{A_{1i} \cdot exp(a_{1i}z_{bio}^-) + B_{1i} \cdot exp(b_{1i}z_{bio}^-)}{exp(a_{2i}z_{bio}^+)}$$

$$A_{1i} \overset{BC3)}{=} -\frac{B_{1i}b_{1i} \cdot exp(b_{1i}z_{bio}^-)}{a_{1i} \cdot exp(a_{1i}z_{bio}^-)}$$

See Section 2.3.1 for further details on how to find the analytical solution.

Table 2. OM Boundary conditions applied in OMEN-SED. For the boundaries we define: $z_{\mathrm{bio}}^- := \lim_{h\to 0}(z_{\mathrm{bio}} - h)$ and $z_{\mathrm{bio}}^+ :=$ $\lim_{h\to 0}(z_{\mathrm{bio}} + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$\mathrm{POC}_i(0) = \mathrm{POC}_{0i}$
$z = z_{\mathrm{bio}}$	continuity	2)	$\mathrm{POC}_i(z_{\mathrm{bio}}^-) = \mathrm{POC}_i(z_{\mathrm{bio}}^+)$
		3)	$-D_{\mathrm{bio}} \cdot \frac{\partial \mathrm{POC}_i}{\partial z}\big|_{z_{\mathrm{bio}}^-} = 0$

2.2.2 Oxygen

OMEN-SED explicitly accounts for oxygen consumption by the aerobic degradation of organic matter within the oxic zone, as well as the oxidation of reduced species (i.e. NH_4, H_2S) produced in the anoxic zones of the sediment. In the oxic zone ($z < z_{ox}$), the aerobic degradation consumes oxygen with a fixed $O_2 : C$ ratio (O_2C, Tab. 10). A predefined fraction, γ_{NH_4}, of the ammonium produced during the aerobic degradation of OM is nitrified to nitrate, consuming two moles of oxygen per mole of ammonium produced. In addition, OMEN-SED implicitly accounts for the oxygen consumption due to oxidation of reduced species (NH_4, H_2S) produced below the oxic zone through the flux boundary condition at the dynamically calculated oxygen penetration depth z_{ox} (see section 2.4.2 for details)~~oxygen penetration depth $z_{\overline{ox}}$~~. All oxygen consumption processes can thus be formulated as a function of organic matter degradation. The conservation equation for oxygen is given by:

$$\frac{\partial O_2}{\partial t} = 0 = D_{O_2}\frac{\partial^2 O_2}{\partial z^2} - w\frac{\partial O_2}{\partial z} - \frac{1-\phi}{\phi}\sum_i k_i \cdot [O_2C + 2\gamma_{NH_4}NC_i]\cdot POC_i(z) \tag{12}$$

For illustrative purposes, we here substitute the analytical solution for the POC depth profile and provide the analytical solution. The remaining paragraphs only outline the general equation, whose analytical solution can be derived in an identical manner. Substituting Eq. (8) and (9) for $POC_i(z)$ and Eq. (11) for B_{1i} gives:

I Bioturbated zone ($z \leq z_{\mathrm{bio}}$)

$$\frac{\partial O_2^I}{\partial t} = 0 \overset{8\&11}{=} D_{O_2}^I\frac{\partial^2 O_2}{\partial z^2} - w\frac{\partial O_2}{\partial z} - \frac{1-\phi}{\phi}\sum_i k_i \cdot [O_2C + 2\gamma_{NH_4}NC_i]\cdot\left(A_{1i}\cdot[exp(a_{1i}z) - exp(b_{1i}z)] + POC_{0i}\cdot exp(b_{1i}z)\right)$$

II Non-bioturbated zone ($z_{\mathrm{bio}} < z < z_{ox}$)

$$\frac{\partial O_2^{II}}{\partial t} = 0 \overset{9}{=} D_{O_2}^{II}\frac{\partial^2 O_2}{\partial z^2} - w\frac{\partial O_2}{\partial z} - \frac{1-\phi}{\phi}\sum_i k_i \cdot [O_2C + 2\gamma_{NH_4}NC_i]\cdot\left(A_{2i}\cdot exp(a_{2i}z)\right)$$

where $D_{O_2}^I$ and $D_{O_2}^{II}$ denote the O_2 diffusion coefficient for the bioturbated and non-bioturbated zone, respectively. The term $\frac{1-\phi}{\phi}$ accounts for the volume conversion from solid to dissolved phase and NC_i is the nitrogen to carbon ratio in POC. Integration yields the following analytical solution for each zone:

I Bioturbated zone ($z \leq z_{\mathrm{bio}}$):

$$O_2^I(z) = A_{O_2}^1 + B_{O_2}^1\cdot\exp(b_{O_2}^1 z) + \sum_i\Phi_{1,i}^I\cdot\exp(a_{1i}z) + \sum_i\Phi_{1,i}^{II}\cdot\exp(b_{1i}z) + \sum_i\Phi_{1,i}^{III}\cdot\exp(b_{1i}z) \tag{13}$$

11

II Non-bioturbated zone ($z_{\text{bio}} < z < z_{\text{ox}}$)

$$O_2^{II}(z) = A_{O_2}^2 + B_{O_2}^2 \cdot \exp(b_{O_2}^2 z) + \sum_i \Phi_{i,2}^I \cdot \exp(a_{2i} z) \tag{14}$$

with

$$b_{O_2}^1 = \frac{w}{D_{O_2}^I}, \quad b_{O_2}^2 = \frac{w}{D_{O_2}^{II}}$$

$$\Phi_{1,i}^I = \frac{1-\phi}{\phi} \cdot \frac{k_i \cdot (O_2C + 2\gamma_{NH_4} NC_i) \cdot A_{1i}}{D_{O_2}^I (-a_{1i})^2 - w \cdot (-a_{1i})}, \qquad \Phi_{1,i}^{II} = -\frac{1-\phi}{\phi} \cdot \frac{k_i \cdot (O_2C + 2\gamma_{NH_4} NC_i) \cdot A_{1i}}{D_{O_2}^I (-b_{1i})^2 - w \cdot (-b_{1i})}$$

$$\Phi_{1,i}^{III} = \frac{1-\phi}{\phi} \cdot \frac{k_i \cdot (O_2C + 2\gamma_{NH_4} NC_i) \cdot POC_{0i}}{D_{O_2}^I (-b_{1i})^2 - w \cdot (-b_{1i})}$$

$$\Phi_{i,2}^I := \frac{1-\phi}{\phi} \cdot \frac{k_i \cdot (O_2C + 2\gamma_{NH_4} NC_i) \cdot A_{2i}}{D_{O_2}^{II} (-a_{2i})^2 - w \cdot (-a_{2i})}$$

Determining the four integration constants ($A_{O_2}^1, B_{O_2}^1, A_{O_2}^2, B_{O_2}^2$, see Section 2.3 for details), as well as the *a priori* unknown oxygen penetration depth requires the definition of five boundary conditions (see Table 3). At the sediment-water interface, OMEN-SED applies a Dirichlet condition (i.e. known concentration) and assumes concentration and flux continuity across the bottom of the bioturbated zone, z_{bio}. The oxygen penetration depth z_{ox} marks the lower boundary and is dynamically calculated as the depth at which $O_2(z) = 0$. Therefore, OMEN-SED applies a Dirichlet boundary condition $O_2(z_{\text{ox}}) = 0$. In addition, a flux boundary is applied that implicitly accounts for the oxygen consumption by the partial oxidation of NH_4 and H_2S diffusing into the oxic zone from below (BC 4.2, Table 3). It is assumed that respective fractions (γ_{NH_4} and γ_{H_2S}) are directly reoxidised at the oxic/anoxic interface and the remaining fraction escapes reoxidation (or is precipitated as pyrite, γ_{FeS}). OMEN-SED iteratively solves for z_{ox} by first testing if there is oxygen left at $\cancel{z_\infty} z_{\text{max}}$ (i.e. $\cancel{O_2(z_\infty) > 0} O_2(z_{\text{max}}) \geq 0$). If that is not the case, it determines the root for the flux boundary condition 4.2 (Table 3). If $\cancel{z_{\text{ox}} = z_\infty} z_{\text{ox}} = z_{\text{max}}$, a zero diffusive flux boundary condition is applied as lower boundary condition.

2.2.3 Nitrate and Ammonium

Nitrogen dynamics in OMEN-SED are controlled by the metabolic production of ammonium, nitrification, denitrification as well as ammonium adsorption. Ammonium is produced by organic matter degradation in both the oxic and anoxic zones, while denitrification consumes nitrate in the denitrification zone with a fixed $NO_3 : C$ ratio (NO_3C, Tab. 10). Anaerobic ammonium oxidation (anammox) is implicitely included in the model. The organic nitrogen released during denitrification is assumed to be directly oxidized with nitrite to N_2 through a coupling between denitrification and anammox.

The adsorption of ammonium to sediment particles is formulated as an equilibrium process with constant equilibrium adsorption coefficient K_{NH_4}, thus assuming that the adsorption is fast compared to the characteristic time scales of transport processes (Wang and Van Cappellen, 1996). In addition, a defined fraction, γ_{NH_4}, of metabolically produced ammonium is directly nitrified to nitrate in the oxic zone, while the nitrification of upward diffusing ammonium produced in the sulfidic and methanic zones is implicitly accounted for in the boundary conditions. The conservation equations for ammonium and nitrate are thus given by:

Table 3. Boundary conditions for oxygen. For the boundaries we define: $z_{\text{bio}}^{-} := \lim_{h \to 0}(z_{\text{bio}} - h)$ and $z_{\text{bio}}^{+} := \lim_{h \to 0}(z_{\text{bio}} + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$O_2(0) = O_{20}$
$z = z_{\text{bio}}$	continuity	2)	$O_2(z_{\text{bio}}^{-}) = O_2(z_{\text{bio}}^{+})$
		3)	$-(D_{O_2,0} + D_{\text{bio}}) \cdot \frac{\partial O_2}{\partial z}\big|_{z_{\text{bio}}^{-}} = -D_{O_2,0} \cdot \frac{\partial O_2}{\partial z}\big|_{z_{\text{bio}}^{+}}$
$z = z_{\text{ox}}$	O_2 consumption	4)	**IF** ~~$(O_2(z_\infty) > 0)$~~ $(O_2(z_{\max}) > 0)$
($\tilde{z_{\text{ox}} = z_\infty}$ $z_{\text{ox}} = z_{\max}$)		4.1)	$\frac{\partial O_2}{\partial z}\big|_{z_{\text{ox}}} = 0$
			ELSE
($\tilde{z_{\text{ox}} < z_\infty}$ $z_{\text{ox}} < z_{\max}$)		4.2)	$O_2(z_{\text{ox}}) = 0$ and $-D_{O_2} \cdot \frac{\partial O_2}{\partial z}\big|_{z_{\text{ox}}} = F_{red}(z_{\text{ox}})$
	with		~~$F_{red}(z_{\text{ox}}) = \frac{1-\phi}{\phi} \cdot \int_{\tilde{z}_{\text{NO}_3}}^{\infty} \sum_i (2\gamma_{\text{NH}_4}\text{NC}_i + 2\gamma_{\text{H}_2\text{S}}\text{SO}_4\text{C}) k_i \text{POC}_i \, dz$~~ $F_{red}(z_{\text{ox}}) = \frac{1-\phi}{\phi} \cdot \int_{\tilde{z}_{\text{NO}_3}}^{z_{\max}}$

Note: $\tilde{z}_{\text{NO}_3} = z_{\text{ox}}$ as upper boundary here, as z_{NO_3} is not known at this point.

1. Oxic zone ($z \leq z_{\text{ox}}$)

$$\frac{\partial \text{NO}_3{}^{I}}{\partial t} = 0 = D_{\text{NO}_3} \frac{\partial^2 \text{NO}_3{}^{I}}{\partial z^2} - w \frac{\partial \text{NO}_3{}^{I}}{\partial z} + \gamma_{\text{NH}_4} \frac{1-\phi}{\phi} \cdot \sum_i \text{NC}_i \cdot k_i \cdot \text{POC}_i(z) \tag{15}$$

$$\frac{\partial \text{NH}_4{}^{I}}{\partial t} = 0 = \frac{D_{\text{NH}_4}}{1 + K_{\text{NH}_4}} \frac{\partial^2 \text{NH}_4{}^{I}}{\partial z^2} - w \frac{\partial \text{NH}_4{}^{I}}{\partial z} + \frac{1 - \gamma_{\text{NH}_4}}{1 + K_{\text{NH}_4}} \cdot \frac{1-\phi}{\phi} \cdot \sum_i \text{NC}_i \cdot k_i \cdot \text{POC}_i(z) \tag{16}$$

2. Denitrification (or nitrogenous) zone ($z_{\text{ox}} < z \leq z_{\text{NO}_3}$)

$$\frac{\partial \text{NO}_3{}^{II}}{\partial t} = 0 = D_{\text{NO}_3} \frac{\partial^2 \text{NO}_3{}^{II}}{\partial z^2} - w \frac{\partial \text{NO}_3{}^{II}}{\partial z} - \frac{1-\phi}{\phi} \text{NO}_3\text{C} \cdot \sum_i k_i \cdot \text{POC}_i(z) \tag{17}$$

$$\frac{\partial \text{NH}_4{}^{II}}{\partial t} = 0 = \frac{D_{\text{NH}_4}}{1 + K_{\text{NH}_4}} \frac{\partial^2 \text{NH}_4{}^{II}}{\partial z^2} - w \frac{\partial \text{NH}_4{}^{II}}{\partial z} \tag{18}$$

3. Sulfidic and methanic zone ($z_{\text{NO}_3} < z \leq z_{\max}$)

$$\frac{\partial \text{NH}_4{}^{III}}{\partial t} = 0 = \frac{D_{\text{NH}_4}}{1 + K_{\text{NH}_4}} \frac{\partial^2 \text{NH}_4{}^{III}}{\partial z^2} - w \frac{\partial \text{NH}_4{}^{III}}{\partial z} + \frac{1}{1 + K_{\text{NH}_4}} \cdot \frac{1-\phi}{\phi} \cdot \sum_i \text{NC}_i \cdot k_i \cdot \text{POC}_i(z) \tag{19}$$

where D_{NO_3} and D_{NH_4} denote the diffusion coefficients for NO_3 and NH_4 which depend on the bioturbation status of the respective geochemical zone (compare Section 2.3.1). Integration of Eq. (15) - (19) yields the analytical solutions, which are not further developed here but follow the procedure outlined in Section 2.2.2 for oxygen (also see Section 2.3.1 for more details on how to find the analytical solution). Table 4 summarises the boundary conditions applied in OMEN-SED to solve Eq. (15) - (19) and to find the *a priori* unknown nitrate penetration depth, z_{NO_3}. The model assumes known bottom water concentrations for both NO_3 and NH_4, the complete consumption of nitrate at the nitrate penetration depth (in case ~~$z_{\text{NO}_3} < z_\infty$~~ $z_{\text{NO}_3} < z_{\max}$) and no change in ammonium flux at ~~z_∞~~ $z_{\max}$. In addition, concentration and diffusive flux continuity across z_{bio} and z_{ox} is

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Table 4. Boundary conditions for nitrate and ammonium. For the boundaries we define: $z^-_\_ := \lim_{h\to 0}(z_\_ - h)$ and $z^+_\_ := \lim_{h\to 0}(z_\_ + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$NO_3(0) = NO_{30}$
$z = z_{bio}$	continuity	2)	$NO_3(z^-_{bio})=NO_3(z^+_{bio})$
		3)	$-(D_{NO_3,0} + D_{bio}) \cdot \frac{\partial NO_3}{\partial z}\big|_{z^-_{bio}} = -D_{NO_3,0} \cdot \frac{\partial NO_3}{\partial z}\big|_{z^+_{bio}}$
$z = z_{ox}$	continuity	4)	$NO_3(z^-_{ox})=NO_3(z^+_{ox})$
		5)	$-D_{NO_3} \cdot \frac{\partial NO_3}{\partial z}\big|_{z^-_{ox}} + \gamma_{NH_4} \cdot F_{NH_4}(z_{ox}) = -D_{NO_3} \cdot \frac{\partial NO_3}{\partial z}\big|_{z^+_{ox}}$
	where:		~~$F_{NH_4}(z_{ox}) = \frac{1}{1+K_{NH_4}} \cdot \frac{1-\phi}{\phi} \cdot \int_{z_{NO_3}}^{\infty} \sum_i NC_i \cdot k_i \cdot POC_i \, dz$~~ $\underline{F_{NH_4}(z_{ox}) = \frac{1}{1+K_{NH_4}} \cdot \frac{1-\phi}{\phi} \cdot}$
$z = z_{NO_3}$	NO_3 consumption	6)	**IF** ~~$(NO_3(z_\infty) > 0)$~~ $\underline{(NO_3(z_{max}) > 0)}$
(~~$z_{NO_3} = z_\infty$~~ $\underline{z_{NO_3} = z_{max}}$)		6.1)	$\frac{\partial NO_3}{\partial z}\big|_{z_{NO_3}} = 0$
			ELSE
(~~$z_{NO_3} < z_\infty$~~ $\underline{z_{NO_3} < z_{max}}$)		6.2)	$NO_3(z_{NO_3}) = 0$ and $\frac{\partial NO_3}{\partial z}\big|_{z_{NO_3}} = 0$
$z = 0$	known concentration	1)	$NH_4(0) = NH_{40}$
$z = z_{bio}$	continuity	2)	$NH_4(z^-_{bio})=NH_4(z^+_{bio})$
		3)	$-\frac{D_{NH_4,0}+D_{bio}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^-_{bio}} = -\frac{D_{NH_4,0}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^+_{bio}}$
$z = z_{ox}$	continuity	4)	$NH_4(z^-_{ox})=NH_4(z^+_{ox})$
		5)	$-\frac{D_{NH_4}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^-_{ox}} - \gamma_{NH_4} \cdot F_{NH_4}(z_{ox}) = -\frac{D_{NH_4}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^+_{ox}}$
	where:		~~$F_{NH_4}(z_{ox}) = \frac{1}{1+K_{NH_4}} \cdot \frac{1-\phi}{\phi} \cdot \int_{z_{NO_3}}^{\infty} \sum_i NC_i \cdot k_i \cdot POC_i \, dz$~~ $\underline{F_{NH_4}(z_{ox}) = \frac{1}{1+K_{NH_4}} \cdot \frac{1-\phi}{\phi} \cdot}$
$z = z_{NO_3}$	continuity	6)	$NH_4(z^-_{NO_3})=NH_4(z^+_{NO_3})$
	flux	7)	$-\frac{D_{NH_4}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^-_{NO_3}} = -\frac{D_{NH_4}}{1+K_{NH_4}} \cdot \frac{\partial NH_4}{\partial z}\big|_{z^+_{NO_3}}$
~~$z = z_\infty$~~ $\underline{z = z_{max}}$	zero NH_4 flux	8)	~~$\frac{\partial NH_4}{\partial z}\big|_{z_\infty} = 0$~~ $\underline{\frac{\partial NH_4}{\partial z}\big|_{z_{max}} = 0}$

considered for NO_3 and NH_4. Furthermore, the reoxidation of upward-diffusing reduced ammonium is accounted for in the oxic-anoxic boundary condition for nitrate and ammonium. OMEN-SED iteratively solves for z_{NO_3} by first testing if there is nitrate left at ~~z_∞~~ $\underline{z_{max}}$ (i.e. ~~$NO_3(z_\infty) > 0$~~ $\underline{NO_3(z_{max}) > 0}$) and, otherwise, by finding the root for the flux boundary condition 6.2 (Table 4).

2.2.4 Sulfate and Sulfide

Below the denitrification zone ($z > z_{NO_3}$), organic matter degradation is coupled to sulfate reduction, consuming sulfate and producing hydrogen sulfide with a fixed $SO_4 : C$ ratio (SO_4C, Tab. 10). In addition, the anaerobic oxidation of upward diffusing methane (AOM) produced below the sulfate penetration and the associated consumption of sulfate and production of sulfide; as well as the production of sulfate and consumption of sulfide through sulfide oxidation are implicitly accounted for through the boundary conditions (Table 5). In the sulfidic zone a defined fraction of sulfide, γ_{FeS}, can be precipitated as pyrite (in the presented simulations $\gamma_{FeS} = 0.0$ as we do not want to make any assumptions about pyrite precipitation). It can be safely assumed that almost all CH_4 is oxidized anaerobically in the sediments (e.g. Reeburgh, 2007, suggests up to 90%) - except for

active (very localized) sites and slope failure, which can, in theory, be accounted for through the γ_{CH_4} term. The conservation equations for sulfate and sulfide are thus given by:

1. Oxic and nitrogenous zone ($z \leq z_{NO_3}$)

$$\frac{\partial SO_4{}^I}{\partial t} = 0 = D_{SO_4} \frac{\partial^2 SO_4{}^I}{\partial z^2} - w \frac{\partial SO_4{}^I}{\partial z} \tag{20}$$

$$\frac{\partial H_2S^I}{\partial t} = 0 = D_{H_2S} \frac{\partial^2 H_2S^I}{\partial z^2} - w \frac{\partial H_2S^I}{\partial z} \tag{21}$$

2. Sulfidic zone ($z_{NO_3} < z \leq z_{SO_4}$)

$$\frac{\partial SO_4{}^{II}}{\partial t} = 0 = D_{SO_4} \frac{\partial^2 SO_4{}^{II}}{\partial z^2} - w \frac{\partial SO_4{}^{II}}{\partial z} - \frac{1-\phi}{\phi} \cdot \sum_i SO_4C \cdot k_i \cdot POC_i(z) \tag{22}$$

$$\frac{\partial H_2S^{II}}{\partial t} = 0 = D_{H_2S} \frac{\partial^2 H_2S^{II}}{\partial z^2} - w \frac{\partial H_2S^{II}}{\partial z} + \frac{1-\phi}{\phi} \cdot (1 - \gamma_{FeS}) \cdot \sum_i SO_4C \cdot k_i \cdot POC_i(z) \tag{23}$$

3. Methanic zone ($z_{SO_4} < z \leq z_{max}$)

$$\frac{\partial H_2S^{III}}{\partial t} = 0 = D_{H_2S} \frac{\partial^2 H_2S^{III}}{\partial z^2} - w \frac{\partial H_2S^{III}}{\partial z} \tag{24}$$

where D_{SO_4} and D_{H_2S} denote the diffusion coefficients for SO_4 and H_2S which depend on the bioturbation status of the respective geochemical zone (compare Section 2.3.1). Integration of Eq. (20) - (24) yields the analytical solution and Table 5 summarises the boundary conditions applied. OMEN-SED assumes known concentrations at the sediment-water interface and continuity across the bioturbation depth and the nitrate penetration depth. The reoxidation of reduced H_2S to SO_4 is accounted for implicitly via the oxic-anoxic boundary condition for both species, while reduction of SO_4 and the associated production of H_2S via AOM is accounted for through the respective boundary conditions at z_{SO_4}. In case ~~$z_{SO_4} < z_\infty$~~ $z_{SO_4} \leq z_{max}$, OMEN-SED assumes zero sulfate concentration at z_{SO_4} and its diffusive flux must equal the amount of methane produced below (with a methane to carbon ratio of MC); or, in case ~~$z_{SO_4} = z_\infty$~~ $z_{SO_4} = z_{max}$, a zero diffusive flux condition for sulfate is considered. OMEN-SED iteratively solves for z_{SO_4} by first testing if there is sulfate left at ~~z_∞~~ z_{max} (i.e. ~~$SO_4(z_\infty) > 0$~~ $SO_4(z_{max}) \geq 0$) and, otherwise, by finding the root for the flux boundary condition 8.2 (Table 5). At the lower boundary ~~z_∞~~ z_{max} zero diffusive flux of H_2S is considered.

2.2.5 Phosphate

The biogeochemical description of phosphorus (P) dynamics builds on earlier models developed by Slomp et al. (1996) and accounts for phosphorus recycling through organic matter degradation, adsorption onto sediments and iron(III) hydroxides (Fe-bound P), as well as carbonate fluorapatite (CFA or authigenic P) formation (see Figure 2 for a schematic overview of the sedimentary P cycle). In the oxic zone of the sediment, PO_4 liberated through organic matter degradation can adsorb to iron(III) hydroxides forming Fe-bound P (or FeP, Slomp et al., 1998). Below the oxic zone, PO_4 is not only produced via

Table 5. Boundary conditions for sulfate and sulfide. For the boundaries we define: $z^- := \lim_{h\to 0}(z_- - h)$ and $z^+_- := \lim_{h\to 0}(z_- + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$SO_4(0) = SO_{40}$
$z = z_{\mathrm{bio}}$	continuity	2)	$SO_4(z^-_{\mathrm{bio}}) = SO_4(z^+_{\mathrm{bio}})$
	flux	3)	$-(D_{SO_4,0} + D_{\mathrm{bio}}) \cdot \frac{\partial SO_4}{\partial z}\big|_{z^-_{\mathrm{bio}}} = -D_{SO_4,0} \cdot \frac{\partial SO_4}{\partial z}\big|_{z^+_{\mathrm{bio}}}$
$z = z_{\mathrm{ox}}$	continuity	4)	$SO_4(z^-_{\mathrm{ox}}) = SO_4(z^+_{\mathrm{ox}})$
	flux	5)	$-D_{SO_4} \cdot \frac{\partial SO_4}{\partial z}\big|_{z^-_{\mathrm{ox}}} + \gamma_{H_2S} \cdot F_{H_2S}(z_{\mathrm{ox}}) = -D_{SO_4} \cdot \frac{\partial SO_4}{\partial z}\big|_{z^+_{\mathrm{ox}}}$
	where:		~~$F_{H_2S}(z_{\mathrm{ox}}) = \frac{1-\phi}{\phi} \cdot \left(\int_{z_{NO_3}}^{SO_4} \sum_i SO_4 C \cdot k_i \cdot POC_i\, dz + \gamma_{CH_4} \cdot \int_{z_{SO_4}}^{\infty} \sum_i MC \cdot k_i \cdot POC_i\, dz \right)$~~ $F_{H_2S}\left(\int_{z_{NO_3}}^{SO_4} \sum_i SO_4 C \cdot k_i \cdot POC_i\, dz + \gamma_{CH_4} \cdot \int_{z_{SO_4}}^{z_{\max}} \sum_i MC \cdot k_i \cdot POC_i\, dz \right)$
$z = z_{NO_3}$	continuity	6)	$SO_4(z^-_{NO_3}) = SO_4(z^+_{NO_3})$
	flux	7)	$-D_{SO_4} \cdot \frac{\partial SO_4}{\partial z}\big|_{z^-_{NO_3}} = -D_{SO_4} \cdot \frac{\partial SO_4}{\partial z}\big|_{z^+_{NO_3}}$
$z = z_{SO_4}$	SO$_4$ consumption	8)	**IF** ~~$(SO_4(z_\infty) > 0)$~~ $(SO_4(z_{\max}) > 0)$
($z_{SO_4} = z_\infty$ ~~$z_{SO_4} = z_{\max}$~~)		8.1)	$\frac{\partial SO_4}{\partial z}\big|_{z_{SO_4}} = 0$
			ELSE
(~~$z_{SO_4} < z_\infty$~~ $z_{SO_4} \leq z_{\max}$)		8.2)	$SO_4(z_{SO_4}) = 0$ and $-D_{SO_4} \cdot \frac{\partial SO_4}{\partial z}\big|_{z_{SO_4}} = \gamma_{CH_4} \cdot F_{CH_4}(z_{SO_4})$
	with		~~$F_{CH_4}(z_{SO_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{SO_4}}^{\infty} \sum_i MC \cdot k_i \cdot POC_i\, dz$~~ $F_{CH_4}(z_{SO_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{SO_4}}^{z_{\max}} \sum_i MC \cdot k_i \cdot P$
$z = 0$	known concentration	1)	$H_2S(0) = H_2S_0$
$z = z_{\mathrm{bio}}$	continuity	2)	$H_2S(z^-_{\mathrm{bio}}) = H_2S(z^+_{\mathrm{bio}})$
	flux	3)	$-(D_{H_2S,0} + D_{\mathrm{bio}}) \cdot \frac{\partial H_2S}{\partial z}\big|_{z^-_{\mathrm{bio}}} = -D_{H_2S,0} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^+_{\mathrm{bio}}}$
$z = z_{\mathrm{ox}}$	continuity	4)	$H_2S(z^-_{\mathrm{ox}}) = H_2S(z^+_{\mathrm{ox}})$
	flux	5)	$-D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^-_{\mathrm{ox}}} - \gamma_{H_2S} F_{H_2S}(z_{\mathrm{ox}}) = -D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^+_{\mathrm{ox}}}$
	where:		~~$F_{H_2S}(z_{\mathrm{ox}}) = \frac{1-\phi}{\phi} \cdot \left(\int_{z_{NO_3}}^{SO_4} \sum_i SO_4 C \cdot k_i \cdot POC_i\, dz + \gamma_{CH_4} \cdot \int_{z_{SO_4}}^{\infty} \sum_i MC \cdot k_i \cdot POC_i\, dz \right)$~~ $F_{H_2S}\left(\int_{z_{NO_3}}^{SO_4} \sum_i SO_4 C \cdot k_i \cdot POC_i\, dz + \gamma_{CH_4} \cdot \int_{z_{SO_4}}^{z_{\max}} \sum_i MC \cdot k_i \cdot POC_i\, dz \right)$
$z = z_{NO_3}$	continuity	6)	$H_2S(z^-_{NO_3}) = H_2S(z^+_{NO_3})$
	flux	7)	$-D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^-_{NO_3}} = -D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^+_{NO_3}}$
$z = z_{SO_4}$	continuity	8)	$H_2S(z^-_{SO_4}) = H_2S(z^+_{SO_4})$
	flux (with AOM)	9)	$-D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^-_{SO_4}} + \gamma_{CH_4} \cdot F_{CH_4}(z_{SO_4}) = -D_{H_2S} \cdot \frac{\partial H_2S}{\partial z}\big|_{z^+_{SO_4}}$
	where:		~~$F_{CH_4}(z_{SO_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{SO_4}}^{\infty} \sum_i MC \cdot k_i \cdot POC_i\, dz$~~ $F_{CH_4}(z_{SO_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{SO_4}}^{z_{\max}} \sum_i MC \cdot k_i \cdot P$
~~$z = z_\infty$~~ $z = z_{\max}$	zero H$_2$S flux	10)	~~$\frac{\partial H_2S}{\partial z}\big|_{z_\infty} = 0$~~ $\frac{\partial H_2S}{\partial z}\big|_{z_{\max}} = 0$

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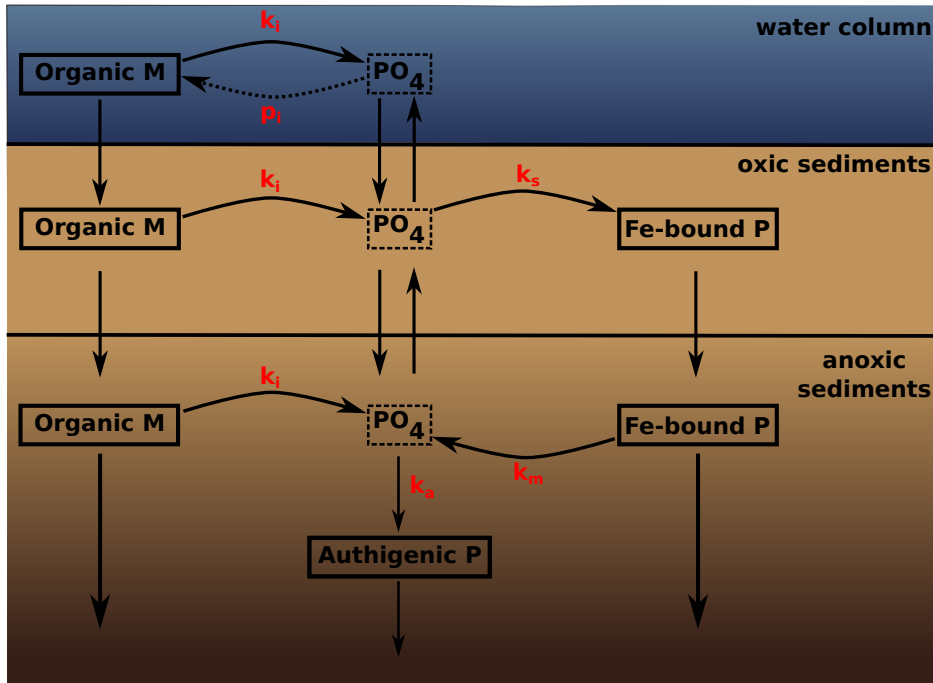
Figure 2. A schematic of the sedimentary P cycle in OMEN-SED. Red numbers represent kinetic rate constants for phosphorus dynamics (compare Table 10; p_i represents uptake rate of PO_4 via primary production in shallow environments). Adapted from Slomp et al. (1996).

organic matter degradation but can also be released from the Fe-bound P pool due to the reduction of iron(III) hydroxides under anoxic conditions. Furthermore, in these zones phosphate concentrations build up and pore waters can thus become supersaturated with respect to carbonate fluorapatite, thus triggering the authigenic formation of CFA (Van Cappellen and Berner, 1988). Phosphorus bound in these authigenic minerals represents a permanent sink for reactive phosphorus (Slomp
5 et al., 1996). As for ammonium, the adsorption of P to the sediment matrix is treated as an equilibrium processes, parameterised with dimensionless adsorption coefficients for the oxic and anoxic zone, respectively ($K_{PO_4}^{ox}$, $K_{PO_4}^{anox}$ Slomp et al., 1998). The sorption and desorption of P to iron(III) hydroxides as well as the authigenic fluorapatite formation are described as first-order reactions with rate constants k_s, k_m and k_a, respectively (Table 10). The rate of the respective process is calculated as the product of the rate constant and the difference between the current concentration (of PO_4 and FeP) and an equilibrium
10 or asymptotic concentration Slomp et al. (1996). The asymptotic Fe-bound P concentration is FeP^∞ and the equilibrium concentration for P sorption and authigenic fluorapatite formation are $PO_4{}^s$ and $PO_4{}^a$, respectively (Table 10). The last term in Eq. (25) and (26) represents sorption of PO_4 to FeP in the oxic zone, the last term in Eq. (27) and (28) is the release of PO_4 from the FeP pool and the 4th term in Eq. (28) represents the permanent loss of PO_4 to authigenic fluorapatite formation. The conservation equations for phosphate and Fe-bound P are thus given by:

Table 6. Boundary conditions for phosphate and Fe-bound P (FeP). For the boundaries we define: $z_-^- := \lim_{h\to 0}(z_- - h)$ and $z_-^+ := \lim_{h\to 0}(z_- + h)$.

Boundary	Condition	
$z = 0$	known concentration	1) $PO_4(0)=PO_{40}$
$z = z_{\text{bio}}$	continuity	2) $PO_4(z_{\text{bio}}^-)=PO_4(z_{\text{bio}}^+)$
	flux	3) $(D_{PO_4,0} + D_{\text{bio}}) \cdot \frac{\partial PO_4}{\partial z}\big|_{z_{\text{bio}}^-} = D_{PO_4,0} \cdot \frac{\partial PO_4}{\partial z}\big|_{z_{\text{bio}}^+}$
$z = z_{\text{ox}}$	continuity	4) $PO_4(z_{\text{ox}}^-)=PO_4(z_{\text{ox}}^+)$
	flux	5) $-\frac{D_{PO_4}}{1+K_{PO_4}^{ox}} \cdot \frac{\partial PO_4}{\partial z}\big|_{z_{\text{ox}}^-} = -\frac{D_{PO_4}}{1+K_{PO_4}^{anox}} \cdot \frac{\partial PO_4}{\partial z}\big|_{z_{\text{ox}}^+}$
~~$z = z_\infty$~~ $z = z_{\max}$	flux	10) ~~$\frac{\partial PO_4}{\partial z}\big|_{z_\infty} = 0$~~ $\frac{\partial PO_4}{\partial z}\big|_{z_{\max}} = 0$
$z = 0$	known concentration	1) $FeP(0) = FeP_0$
$z = z_{\text{bio}}$	continuity	2) $FeP(z_{\text{bio}}^-)=FeP(z_{\text{bio}}^+)$
	flux	3) $\frac{\partial FeP}{\partial z}\big|_{z_{\text{bio}}^-} = \frac{\partial FeP}{\partial z}\big|_{z_{\text{bio}}^+}$
$z = z_{\text{ox}}$	continuity	4) $FeP(z_{\text{ox}}^-)=FeP(z_{\text{ox}}^+)$
	flux	5) $\frac{\partial FeP}{\partial z}\big|_{z_{\text{ox}}^-} = \frac{\partial FeP}{\partial z}\big|_{z_{\text{ox}}^+}$
~~$z = z_\infty$~~ $z = z_{\max}$	asymptotic concentration	10) ~~$FeP(z_\infty) = FeP_\infty$~~ $FeP(z_{\max}) = FeP^\infty$

1. Oxic zone ($z \le z_{\text{ox}}$)

$$\frac{\partial PO_4^I}{\partial t} = \frac{D_{PO_4}}{1+K_{PO_4}^{ox}} \frac{\partial^2 PO_4^I}{\partial z^2} - w\frac{\partial PO_4^I}{\partial z} + \frac{1-\phi}{\phi}\frac{1}{1+K_{PO_4}^{ox}}\sum_i (PC_i \cdot k_i \cdot POC_i(z)) - \frac{k_s}{1+K_{PO_4}^{ox}}(PO_4^I - PO_4^s) \quad (25)$$

$$\frac{\partial FeP^I}{\partial t} = D_{FeP}\frac{\partial^2 FeP^I}{\partial z^2} - w\frac{\partial FeP^I}{\partial z} + \frac{\phi}{1-\phi}k_s(PO_4^I - PO_4^s) \quad (26)$$

2. Anoxic zones ($z_{\text{ox}} < z \le z_{\max}$)

$$\frac{\partial FeP^{II}}{\partial t} = D_{FeP}\frac{\partial^2 FeP^{II}}{\partial z^2} - w\frac{\partial FeP^{II}}{\partial z} - k_m(FeP^{II} - FeP^\infty) \quad (27)$$

$$\frac{\partial PO_4^{II}}{\partial t} = \frac{D_{PO_4}}{1+K_{PO_4}^{anox}} \frac{\partial^2 PO_4^{II}}{\partial z^2} - w\frac{\partial PO_4^{II}}{\partial z} + \frac{1-\phi}{\phi}\frac{1}{1+K_{PO_4}^{anox}}\sum_i (PC_i \cdot k_i \cdot POC_i(z))$$
$$- \frac{k_a}{1+K_{PO_4}^{anox}}(PO_4^{II} - PO_4^a) + \frac{(1-\phi)}{\phi}\frac{k_m}{1+K_{PO_4}^{anox}}(FeP^{II} - FeP^\infty) \quad (28)$$

where D_{PO_4} denotes the diffusion coefficient for PO_4 which depends on the bioturbation status of the respective geochemical zone and $D_{FeP} = D_{\text{bio}}$ for $z \le z_{\text{bio}}$ and $D_{FeP} = 0$ for $z > z_{\text{bio}}$ (compare Section 2.3.1). Integration of Eq. (25) - (28) yields the analytical solution and Table 6 summarises the boundary conditions applied in OMEN-SED. The model assumes known bottom water concentrations and equal concentrations and diffusive fluxes at z_{bio} and z_{ox} for both species. Additionally OMEN-SED considers no change in phosphate flux and an asymptotic Fe-bound P concentration at ~~z_∞.~~ $z_{\max}$.

Table 7. Boundary conditions for DIC. For the boundaries we define: $z_-^- := \lim_{h\to 0}(z_- - h)$ and $z_-^+ := \lim_{h\to 0}(z_- + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$\mathrm{DIC}(0) = \mathrm{DIC}_0$
$z = z_{\mathrm{bio}}$	continuity	2)	$\mathrm{DIC}(z_{\mathrm{bio}}^-)=\mathrm{DIC}(z_{\mathrm{bio}}^+)$
	flux	3)	$-(D_{\mathrm{DIC},0} + D_{\mathrm{bio}}) \cdot \frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_{\mathrm{bio}}^-} = -D_{\mathrm{DIC},0} \cdot \frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_{\mathrm{bio}}^+}$
$z = z_{\mathrm{SO}_4}$	continuity	4)	$\mathrm{DIC}(z_{\mathrm{SO}_4}^-)=\mathrm{DIC}(z_{\mathrm{SO}_4}^+)$
	flux (with AOM)	5)	$-D_{\mathrm{DIC}} \cdot \frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_{\mathrm{SO}_4}^-} + \gamma_{\mathrm{CH}_4} \cdot F_{\mathrm{CH}_4}(z_{\mathrm{SO}_4}) = -D_{\mathrm{DIC}} \cdot \frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_{\mathrm{SO}_4}^+}$
	where:		~~$F_{\mathrm{CH}_4}(z_{\mathrm{SO}_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{\mathrm{SO}_4}}^{\infty} \sum_i \mathrm{MC} \cdot k_i \cdot \mathrm{POC}_i \, dz$~~ $F_{\mathrm{CH}_4}(z_{\mathrm{SO}_4}) = \frac{1-\phi}{\phi} \cdot \int_{z_{\mathrm{SO}_4}}^{z_{\max}} \sum_i \mathrm{MC} \cdot k_i \cdot \mathrm{POC}_i$
~~$z = z_\infty$~~ $z = z_{\max}$	zero DIC flux	6)	~~$\frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_\infty} = 0$~~ $\frac{\partial \mathrm{DIC}}{\partial z}\big|_{z_{\max}} = 0$

2.2.6 Dissolved Inorganic Carbon (DIC)

OMEN-SED accounts for the production of dissolved inorganic carbon (DIC) through organic matter degradation, as well as methane oxidation. Organic matter degradation produces dissolved inorganic carbon with a stoichiometric DIC : C ratio of 1:2 in the methanic zone and 1:1 in the rest of the sediment column ($\mathrm{DICC}^{\mathrm{II}}$ and $\mathrm{DICC}^{\mathrm{I}}$ respectively). DIC production through methane oxidation is implicitly taken into account through the boundary condition at z_{SO_4}. A mechanistic description of DIC production from CaCO_3 dissolution would lead to significant mathematical problems and is therefore not included in the current version of OMEN-SED. The conservation equations for DIC are thus given by:

1. Oxic, nitrogenous and sulfidic zone ($z \leq z_{\mathrm{SO}_4}$)

$$\frac{\partial \mathrm{DIC}^I}{\partial t} = 0 = D_{\mathrm{DIC}} \frac{\partial^2 \mathrm{DIC}^I}{\partial z^2} - w \frac{\partial \mathrm{DIC}^I}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i \mathrm{DICC}^{\mathrm{I}} \cdot k_i \cdot \mathrm{POC}_i(z) \tag{29}$$

2. Methanic zone ($z_{\mathrm{SO}_4} < z \leq z_{\max}$)

$$\frac{\partial \mathrm{DIC}^{II}}{\partial t} = 0 = D_{\mathrm{DIC}} \frac{\partial^2 \mathrm{DIC}^{II}}{\partial z^2} - w \frac{\partial \mathrm{DIC}^{II}}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i \mathrm{DICC}^{\mathrm{II}} \cdot k_i \cdot \mathrm{POC}_i(z) \tag{30}$$

where D_{DIC} denotes the diffusion coefficient for DIC (taking the values for HCO_3^- from Schulz (2006)) which depends on the bioturbation status of the respective geochemical zone. Integration of Eq. (29) and (30) yields the analytical solution and Table 7 summarises the boundary conditions applied in OMEN-SED. A Dirichlet condition is applied at the sediment-water interface. In addition, the model assumes a zero diffusive flux through the lower boundary ~~z_∞~~ $z_{\max}$ and continuity across the bottom of the bioturbated zone, as well as the sulfate penetration depth. An additional flux boundary condition at z_{SO_4}, implicitly accounts for DIC production through anaerobic oxidation of methane (Table 7 Eq. 5).

2.2.7 Alkalinity

Organic matter degradation and secondary redox reactions exert a complex influence on alkalinity (e.g. Jourabchi et al., 2005; Wolf-Gladrow et al., 2007; Krumins et al., 2013). To model alkalinity, OMEN-SED divides the sediment column ~~is~~ into four

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geochemical zones, where different equations describe the biogeochemical processes using variable stoichiometric coefficients (compare values in Table 10). Above z_{ox}, the combined effects of NH_4 and P release due to aerobic OM degradation increases alkalinity according to ALK^{OX} whereas nitrification decreases alkalinity with stoichiometry ALK^{NIT}. In the remaining three zones anaerobic OM degradation generally results in an increase in alkalinity, with the exact magnitude depending on the nature of the terminal electron acceptor used (i.e. ALK^{DEN}, ALK^{SUL}, ALK^{MET}). In addition, the effect of secondary redox reactions, such as nitrification, sulfide and methane oxidation, as well as pyrite precipitation, are implicitly accounted for in the boundary conditions. Note that the alkalinity description in the current version of OMEN-SED does not account for $CaCO_3$ dissolution/precipitation due to the mathematical complexity of the problem. In OMEN-SED, the conservation equations for alkalinity are thus given by:

1. Oxic zone ($z \leq z_{\text{ox}}$)

$$\frac{\partial ALK^I}{\partial t} = 0 = D_{ALK} \frac{\partial^2 ALK^I}{\partial z^2} - w \frac{\partial ALK^I}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i \left(ALK^{NIT} \cdot \frac{\gamma_{NH_4}}{1 + K_{NH_4}} NC_i + ALK^{OX} \right) \cdot k_i \cdot POC_i(z) \tag{31}$$

2. Denitrification or nitrogenous zone ($z_{\text{ox}} < z \leq z_{NO_3}$)

$$\frac{\partial ALK^{II}}{\partial t} = 0 = D_{ALK} \frac{\partial^2 ALK^{II}}{\partial z^2} - w \frac{\partial ALK^{II}}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i ALK^{DEN} \cdot k_i \cdot POC_i(z) \tag{32}$$

3. Sulfidic zone ($z_{NO_3} < z \leq z_{SO_4}$)

$$\frac{\partial ALK^{III}}{\partial t} = 0 = D_{ALK} \frac{\partial^2 ALK^{III}}{\partial z^2} - w \frac{\partial ALK^{III}}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i ALK^{SUL} \cdot k_i \cdot POC_i(z) \tag{33}$$

4. Methanic zone ($z_{SO_4} < z \leq z_{\max}$)

$$\frac{\partial ALK^{IV}}{\partial t} = 0 = D_{ALK} \frac{\partial^2 ALK^{IV}}{\partial z^2} - w \frac{\partial ALK^{IV}}{\partial z} + \frac{1-\phi}{\phi} \cdot \sum_i ALK^{MET} \cdot k_i \cdot POC_i(z) \tag{34}$$

where D_{ALK} denotes the diffusion coefficient for alkalinity (taking the values for HCO_3^- from Schulz (2006)) which depends on the bioturbation status of the respective geochemical zone. Integration of Eq. (31) - (34) yields the analytical solution and Table 8 summarises the boundary conditions applied in OMEN-SED. A Dirichlet boundary condition is applied at the sediment-water interface. The decrease of alkalinity due to oxidation of reduced species produced in the anoxic zones and due to the precipitation of pyrite (with stoichiometry ALK^{NIT}~~and~~, ALK^{H_2S} and ALK^{FeS}) is implicitly taken into account through the flux boundary condition at z_{ox} (Table 8 Eq. 5). Furthermore, the oxidation of methane by sulfate reduction increases alkalinity with stoichiometry ALK^{AOM} which is accounted for through the flux boundary condition at z_{SO_4} (Table 8 Eq. 9). At the lower boundary ~~z_∞~~$z_{\max}$ a zero diffusive flux condition is applied.

2.3 Determination of Integration Constants

The integration constants of all general analytical solutions derived above change in response to changing boundary conditions. Thus, OMEN-SED has to re-determine integration constants for each dynamic zone (i.e. $z_{\text{ox}}, z_{\text{bio}}, z_{NO_3}$ and z_{SO_4}) at every time

Table 8. Boundary conditions for alkalinity. For the boundaries we define: $z^-_{\_} := \lim_{h\to 0}(z_{\_} - h)$ and $z^+_{\_} := \lim_{h\to 0}(z_{\_} + h)$.

Boundary	Condition		
$z = 0$	known concentration	1)	$\mathrm{ALK}(0) = \mathrm{ALK}_0$
$z = z_{\mathrm{bio}}$	continuity	2)	$\mathrm{ALK}(z^-_{\mathrm{bio}}) = \mathrm{ALK}(z^+_{\mathrm{bio}})$
	flux	3)	$-(D_{\mathrm{ALK},0} + D_{\mathrm{bio}}) \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^-_{\mathrm{bio}}} = -D_{\mathrm{ALK},0} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^+_{\mathrm{bio}}}$
$z = z_{\mathrm{ox}}$	continuity	4)	$\mathrm{ALK}(z^-_{\mathrm{ox}}) = \mathrm{ALK}(z^+_{\mathrm{ox}})$
	flux	5)	$-D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^-_{\mathrm{ox}}} + F_{\mathrm{ALK}}(z_{\mathrm{ox}}) = -D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^+_{\mathrm{ox}}}$
	where:		~~$F_{\mathrm{ALK}}(z_{\mathrm{ox}}) = \frac{1-\phi}{\phi} \cdot \left(\mathrm{ALK}^{\mathrm{H_2S}} \cdot \gamma_{\mathrm{H_2S}} \int_{z_{\mathrm{NO_3}}}^{\mathrm{SO_4}} \sum_i \mathrm{SO_4C} \cdot k_i \cdot \mathrm{POC}_i \, dz\right)$~~ $F_{\mathrm{ALK}}(z_{\mathrm{ox}}) = \frac{1-\phi}{\phi} \cdot \big(\dots$
			~~$+ \frac{1-\phi}{\phi} \cdot \left(\mathrm{ALK}^{\mathrm{NIT}} \frac{\gamma_{\mathrm{NH_4}}}{1+k_{\mathrm{NH_4}}} \int_{z_{\mathrm{NO_3}}}^{\infty} \sum_i \mathrm{NC}_i \cdot k_i \cdot \mathrm{POC}_i \, dz\right)$~~ $\dots \big(\int$
			$+ \frac{1-\phi}{\phi} \cdot \left(\mathrm{ALK}^{\mathrm{NIT}} \frac{\gamma_{\mathrm{NH_4}}}{1+k_{\mathrm{NH_4}}} \int_{z_{\mathrm{NO_3}}}^{z_{\mathrm{max}}} \sum_i \mathrm{NC}_i \cdot k_i \cdot \mathrm{POC}_i \, dz\right)$
$z = z_{\mathrm{NO_3}}$	continuity	6)	$\mathrm{ALK}(z^-_{\mathrm{NO_3}}) = \mathrm{ALK}(z^+_{\mathrm{NO_3}})$
	flux	7)	$-D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^-_{\mathrm{NO_3}}} = -D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^+_{\mathrm{NO_3}}}$
$z = z_{\mathrm{SO_4}}$	continuity	8)	$\mathrm{ALK}(z^-_{\mathrm{SO_4}}) = \mathrm{ALK}(z^+_{\mathrm{SO_4}})$
	flux (with AOM)	9)	$-D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^-_{\mathrm{SO_4}}} + F_{\mathrm{ALK}}(z_{\mathrm{SO_4}}) = -D_{\mathrm{ALK}} \cdot \frac{\partial \mathrm{ALK}}{\partial z}\big|_{z^+_{\mathrm{SO_4}}}$
	where:		~~$F_{\mathrm{ALK}}(z_{\mathrm{SO_4}}) = \frac{1-\phi}{\phi} \cdot \left(\mathrm{ALK}^{\mathrm{AOM}} \gamma_{\mathrm{CH_4}} \int_{z_{\mathrm{SO_4}}}^{\infty} \sum_i k_i \cdot \mathrm{POC}_i \, dz\right)$~~ $F_{\mathrm{ALK}}(z_{\mathrm{SO_4}}) = \frac{1-\phi}{\phi} \cdot \big(\mathrm{Al}\dots$
~~$z = z_\infty$~~ $z = z_{\mathrm{max}}$	zero ALK flux	10)	~~$\frac{\partial \mathrm{ALK}}{\partial z}\big|_{z_\infty} = 0$~~ $\frac{\partial \mathrm{ALK}}{\partial z}\big|_{z_{\mathrm{max}}} = 0$

step for all biogeochemical species. The bioturbation boundary poses a particular challenge as it can theoretically occur in any of the dynamic geochemical zones (Fig. 3). Therefore, in order to generalise and simplify this recurring boundary matching problem, an independent, generic algorithm (Generic Boundary Condition Matching) is implemented (rather than using multiple fully-worked-out algebraic solutions for each possible case and every biogeochemical species). As a consequence, the algorithm only has to solve a two-simultaneous-equation problem.

2.3.1 Generic Boundary Condition Matching (GBCM)

As discussed in Section 2.1, the solution of the general steady-state transport-reaction equation (Eq. (2)) for a generic species C is of the general form:

$$C(z) = A \cdot \exp(az) + B \cdot \exp(bz) + \sum_j \frac{\alpha_j}{D\beta_j^2 - w\beta_j - k} \cdot \exp(-\beta_j z) + \frac{Q}{k} \tag{35}$$

and can therefore be expressed as:

$$C(z) = A \cdot E(z) + B \cdot F(z) + G(z) \tag{36}$$

where $E(z)$, $F(z)$ are the homogeneous solutions of the ODE, G(z) the particular integral (collectively called the basis functions), and A, B are the integration constants that must be determined using the boundary conditions (shown in Fig. 3 for the whole sediment column).

Each internal boundary matching problem (i.e. excluding $z = 0$ and ~~$z = z_\infty$~~ $z = z_{\max}$) involves matching continuity and flux for the two solutions of the respective reaction-transport equation above, $C_U(z)$ (= 'upper'), and below, $C_L(z)$ (= 'lower'), the dynamic boundary at $z = z_b$:

$$C_U(z) = A_U \cdot E_U(z) + B_U \cdot F_U(z) + G_U(z) \tag{37}$$

$$C_L(z) = A_L \cdot E_L(z) + B_L \cdot F_L(z) + G_L(z). \tag{38}$$

OMEN-SED generally applies concentration continuity and flux boundary conditions at its internal, dynamic boundaries: Continuity (where for generality we allow a discontinuity V_b)

$$C_U(z_b) = C_L(z_b) + V_b \tag{39}$$

Flux

$$D_U C'_U(z_b) + w C_U(z_b) = D_L C'_L(z_b) + w C_L(z_b) + F_b \tag{40}$$

where w is advection, D are the diffusion coefficients and F_b is any flux discontinuity (e.g. resulting from secondary redox reactions).

Considering that the advective flux above and below the boundary is equal (i.e. $w C_U(z_b) = w C_L(z_b)$) and substituting the general ODE solutions (37), (38), the boundary conditions can be represented as two equations connecting the four integration constants:

$$\begin{pmatrix} E_U & F_U \\ D_U E'_U & D_U F'_U \end{pmatrix} \begin{pmatrix} A_U \\ B_U \end{pmatrix} = \begin{pmatrix} E_L & F_L \\ D_L E'_L & D_L F'_L \end{pmatrix} \begin{pmatrix} A_L \\ B_L \end{pmatrix} + \begin{pmatrix} G_L - G_U + V_b \\ D_L G'_L - D_U G'_U + F_b - w V_b \end{pmatrix} \tag{41}$$

where the ODE solutions E, F, G are all evaluated at z_b.

Equation (41) can now be solved to give A_U and B_U as a function of the integration constants from the layer below (A_L and B_L), thereby constructing a piecewise solution for both layers, with just two integration constants (this is implemented in the function **benthic_utils.matchsoln** of OMEN-SED):

$$\begin{pmatrix} A_U \\ B_U \end{pmatrix} = \begin{pmatrix} c_1 & c_2 \\ c_3 & c_4 \end{pmatrix} \begin{pmatrix} A_L \\ B_L \end{pmatrix} + \begin{pmatrix} d_1 \\ d_2 \end{pmatrix}. \tag{42}$$

Using Eq. (42), $C_U(z)$ in (37) can now be rewritten as a function of A_L and B_L (implemented in **benthic_utils.xformsoln**)):

$$C_U(z) = (c_1 A_L + c_2 B_L + d_1) \cdot E_U(z) + (c_3 A_L + c_4 B_L + d_2) \cdot F_U(z) + G_U(z) \tag{43}$$

and hence define the "transformed" basis functions $E^*_U(z)$, $F^*_U(z)$, $G^*_U(z)$ such that:

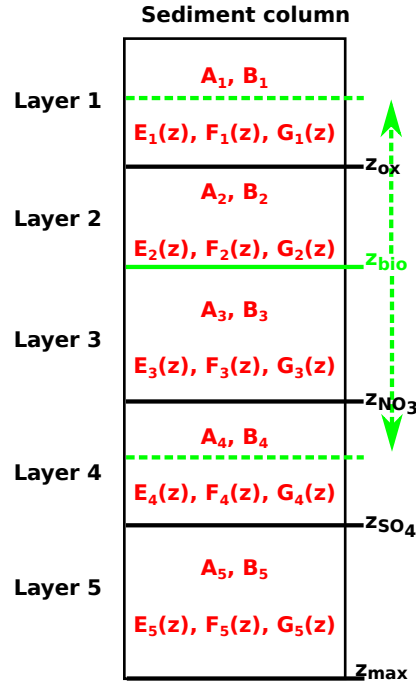$$C_U(z) = A_L \cdot E^*_U(z) + B_L \cdot F^*_U(z) + G^*_U(z) \tag{44}$$

where

$$E_U^*(z) = c_1 E_U(z) + c_3 F_U(z)$$
$$F_U^*(z) = c_2 E_U(z) + c_4 F_U(z) \tag{45}$$
$$G_U^*(z) = G_U(z) + d_1 E_U(z) + d_2 F_U(z)$$

Figure 3. Schematic of the generic boundary condition matching (GBCM) problem. Showing the resulting integration constants (A_i, B_i) and ODE solutions (E_i, F_i, G_i) for the different sediment layers and the ~~variable~~ bioturbation boundary (possible locations are indicated by the green vertical arrow).

Equations (42), (44) and (45) can now be consecutively applied for each of the dynamic biogeochemical zone boundaries (Fig. 3), starting at the bottom of the sediment column. The net result is a piecewise solution of the whole sediment column with just two integration constants (coming from the lowest layer), which can then be solved for by applying the boundary conditions at the sediment-water interface and the bottom of the sediments.

2.3.2 Abstracting out the bioturbation boundary

The bioturbation boundary affects the diffusion coefficient of the modelled solutes, as well as the conservation equation of organic matter (and thereby the exact form of each reaction-transport equation). This boundary is particularly inconvenient as it can, in principle, occur in the middle of any of the dynamically shifting biogeochemical zones and therefore generate multiple

cases (Fig. 3). The GBCM algorithm described above is thus not only used to construct a piecewise solution of the whole sediment column, but also to abstract out the bioturbation boundary. For each biogeochemical zone the "bioturbation-status" is initially tested (i.e. fully bioturbated, fully non-bioturbated, or crossing the bioturbation boundary). Therefore, the upper and lower boundaries for the different zones (e.g. for the nitrogenous zone: $z_U = z$ox, $z_L = z_{NO_3}$), as well as the respective reactive terms and diffusion coefficients (bioturbated and non-bioturbated) are passed over to the routine **zTOC.prepfg_l12** where the bioturbation-status is determined. In case the bioturbation depth is located within this zone (i.e. $z_U < z_{bio} < z_L$) a piecewise solution for this layer is constructed. Therefore, the reactive terms and diffusion coefficients are handed over to the routines **zTOC.calcfg_l1** and **zTOC.calcfg_l2** which calculate the basis functions (E_U, F_U, G_U and E_L, F_L, G_L) and their derivatives for the bioturbated and the non-bioturbated part of this specific geochemical zone. The concentration and flux for both solutions at z_{bio} are matched and the coefficients $c_1, c_2, c_3, c_4, d_1, d_2$ (as in Eq. (42)) are calculated by the routine **benthic_utils.matchsoln**. These coefficients and the "bioturbation-status" of the layer are passed back to the main GBCM algorithm where they can be used by the routine **benthic_utils.xformsoln** to calculate the "transformed" basis functions ($E_U^*(z)$, $F_U^*(z)$, $G_U^*(z)$) such that both layers are expressed in the same basis (compare Eq. (43 - 45)).

For instance, in the case of sulfate, **zTOC.prepfg_l12** is called three times before the actual profile is calculated (once per zone: oxic, nitrogenous, sulfidic) and hands back the information about the "bioturbation-status" of the three layers and the coefficients $c_1, c_2, c_3, c_4, d_1, d_2$ for the biogeochemical zone including the bioturbation depth. When calculating the complete piecewise solution for the sediment column, this information is passed to the function **zTOC.calcfg_l12** which sorts out the correct solution type to use. The main GBCM algorithm therefore never needs to know whether it is dealing with a piecewise solution (i.e. matched across the bioturbation boundary) or a "simple" solution (i.e. the layer is fully bioturbated or fully non-bioturbated).

2.4 Model Parameters

The following section provides a summary of global relationships used to constrain reaction and transport parameters in OMEN-SED. Table 9 synthesises sediment and transport parameters, while Tab. 10 provides an overview of all biogeochemical parameters used in OMEN-SED.

2.4.1 Transport Parameters

The burial of sediments and pore water is directly related to the accumulation of new material on the seafloor (i.e. sedimentation, Burdige, 2006). This results in a downward advective flux of older sediment material and pore water in relation to the sediment-water interface. When coupled to an ocean model, its sedimentation flux can be readily used in OMEN-SED. The stand-alone version of OMEN-SED uses the empirical global relationship between sediment accumulation rate (w in cm yr^{-1}) and seafloor depth (z in m) of Middelburg et al. (1997) :

$$w = 3.3 \cdot 10^{-0.87478367 - 0.00043512 \cdot z}.$$

As an option we include the parameterisation of Burwicz et al. (2011) Burwicz et al. (2011) :

$$w = \frac{w_1}{1 + (\frac{z}{z1})^{c1}} + \frac{w_2}{1 + (\frac{z}{z2})^{c2}} \tag{46}$$

with parameter values as found in the original study (i.e. $w_1 = 0.117\,\text{cm yr}^{-1}$, $w_2 = 0.006\,\text{cm yr}^{-1}$, $z1 = 200\,\text{m}$, $z2 = 4000\,\text{m}$, $c1 = 3$, $c2 = 10$). As an option we include the parameterisation of Middelburg et al. (1997)

$$w = 3.3 \cdot 10^{-0.87478367 - 0.00043512 \cdot z} \tag{47}$$

As mentioned before (Section 2.1), the diffusion coefficient of species i is calculated as $D_i = D_{i,0} + D_{\text{bio}} = D_{\text{mol},i} \cdot f_{ir} + D_{\text{bio}}$ for dissolved species and $D_i = D_{\text{bio}}$ for solid species. The bioturbation coefficient D_{bio} ($\text{cm}^2\,\text{yr}^{-1}$) is constant in the bioturbated zone and also follows the empirical relationship by Middelburg et al. (1997):

$$D_{\text{bio}} = 5.2 \cdot 10^{0.76241122 - 0.00039724 \cdot z} \tag{48}$$

Observations indicate that bioturbation is largely restricted to the upper 10 cm of the sediments and is only marginally related to seafloor depth (e.g. Boudreau, 1998; Teal et al., 2010). Therefore, OMEN-SED imposes a globally invariant bioturbation depth z_{bio} of 10 cm. In case the bottom water oxygen concentration is low (here $<4.5\,\text{nmol cm}^{-3}$ which is often used to define suboxic waters, e.g. Morrison et al., 1999; Karstensen et al., 2008) infaunal activity is assumed to cease and $z_{\text{bio}} = 0.01$ cm. We choose a low value unequal to zero in order to simplify the implementation of the model. This approach ensures that the sediment column always consists of a bioturbated (even though very small for the low oxygen condition) and a non-bioturbated zone, thus the same GBCM algorithm can be used to solve the conservation equations. Furthermore, when OMEN-SED is coupled to an Earth system model the same method can be used to convert the POC depositional flux into a SWI concentration (i.e. the flux needs to be converted assuming bioturbation, see Section 4.1).

Bioirrigation (i.e the pumping activity by burrow-dwelling animals) exchanges burrow water with overlying water and may enhance the SWI-flux of solutes (Aller, 1984, 1988). Several approaches exist to incorporate this into a 1-D diagenetic model, for instance as a non-local transport/exchange process (Boudreau, 1984; Emerson et al., 1984) or as an enhancement factor of the molecular diffusion coefficient (Devol and Christensen, 1993; Soetaert et al., 1996). In OMEN-SED the latter approach is applied and the apparent "bio-diffusion" coefficient is calculated as $D_{i,0} = D_{\text{mol},i} \cdot f_{ir}$. Soetaert et al. (1996) derived an empirical relationship between f_{ir} and seafloor depth ($f_{ir} = \text{Min}\{1; 15.9 \cdot z^{-0.43}\}$) based on observations from Archer and Devol (1992) and Devol and Christensen (1993) . As this relationship just varies for depth above ~623 m (with a maximum value of 3 at ~50 m) a constant value of $f_{ir} = 1$ which is used in the default OMEN-SED configuration. The specific molecular diffusion coefficients $D_{\text{mol},i}$ are corrected for sediment porosity ϕ, tortuosity F and are linearly interpolated for an ambient temperature T (in °C) using zero-degree coefficients D_i^0 and temperature-dependent diffusion coefficients D_i^T (Soetaert et al., 1996):

$$D_{\text{mol},i} = (D_i^0 + D_i^T \cdot T) \cdot \frac{1}{\phi \cdot F}.$$

Tortuosity can be expressed in terms of porosity as $F = \frac{1}{\phi^m}$ (Ullman and Aller, 1982) with the exponent m varying according to the type of sediment (here $m = 3$ is used representing muddy sediments with high porosity). Values for D_i^T and D_i^0 are summarised in Table 9 and are adapted from Li and Gregory (1974), Schulz (2006) and Gypens et al. (2008).

Table 9. Sediment characteristics and transport parameters.

Parameter	Unit	Value	Description/Source
ρ_{sed}	g cm^{-3}	2.6	Sediment density
w	cm yr^{-1}	Fct. of seafloor depth or from ESM	Advection/Sediment accumulation rate (Middelburg et al., 1997) Middelburg et al. (1997)
z_{bio}	cm	10 or 0.01	Bioturbation depth (Boudreau, 1998; Teal et al., 2010) Boudreau (1998); Teal et al. (2010)
D_{bio}	$\text{cm}^2\,\text{yr}^{-1}$	Fct. of seafloor depth	Bioturbation coefficient (Middelburg et al., 1997) Middelburg et al. (1997)
ϕ	-	0.85	Porosity
F	-	$\frac{1}{\phi^m}$	Tortuosity, here m=3
f_{ir}	-	1 Fct. of seafloor depth	Irrigation factor Soetaert et al. (1996)
Diffusion coefficients (Li and Gregory, 1974; Schulz, 2006; Gypens et al., 2008)			
$D_{\text{O}_2}^0$	$\text{cm}^2\,\text{yr}^{-1}$	348.62	Molecular diffusion coefficient of oxygen at 0°C
$D_{\text{O}_2}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	14.09	Diffusion coefficient for linear temp. dependence of oxygen
$D_{\text{NO}_3}^0$	$\text{cm}^2\,\text{yr}^{-1}$	308.42	Molecular diffusion coefficient of nitrate at 0°C
$D_{\text{NO}_3}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	12.26	Diffusion coefficient for linear temp. dependence of nitrate
$D_{\text{NH}_4}^0$	$\text{cm}^2\,\text{yr}^{-1}$	309.05	Molecular diffusion coefficient of ammonium at 0°C
$D_{\text{NH}_4}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	12.26	Diffusion coefficient for linear temp. dependence of ammonium
$D_{\text{SO}_4}^0$	$\text{cm}^2\,\text{yr}^{-1}$	157.68	Molecular diffusion coefficient of sulfate at 0°C
$D_{\text{SO}_4}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	7.88	Diffusion coefficient for linear temp. dependence of sulfate
$D_{\text{H}_2\text{S}}^0$	$\text{cm}^2\,\text{yr}^{-1}$	307.48	Molecular diffusion coefficient of sulfide at 0°C
$D_{\text{H}_2\text{S}}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	9.64	Diffusion coefficient for linear temp. dependence of sulfide
$D_{\text{PO}_4}^0$	$\text{cm}^2\,\text{yr}^{-1}$	112.91	Molecular diffusion coefficient of phosphate at 0°C
$D_{\text{PO}_4}^T$	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	5.59	Diffusion coefficient for linear temp. dependence of phosphate
D_{DIC}^0	$\text{cm}^2\,\text{yr}^{-1}$	151.69	Molecular diffusion coefficient of DIC at 0°C
D_{DIC}^T	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	7.93	Diffusion coefficient for linear temp. dependence of DIC
D_{ALK}^0	$\text{cm}^2\,\text{yr}^{-1}$	151.69	Molecular diffusion coefficient of ALK at 0°C
D_{ALK}^T	$\text{cm}^2\,\text{yr}^{-1}\,^\circ\text{C}^{-1}$	7.93	Diffusion coefficient for linear temp. dependence of ALK

Note: DIC and ALK coefficients are the values of HCO_3^- from Schulz (2006).

2.4.2 Stoichiometries and reaction parameters

The first-order organic matter degradation constants of compound class i, k_i (yr^{-1}), are assumed invariant along the sediment column and therefore independent of the nature of the terminal electron acceptor. The rate constants can be altered manually to fit observed sediment profiles (compare modelled profiles in Section 3.2) or related to a master variable provided by a coupled Earth system model (e.g. sedimentation rate, see Section 4.2). The partitioning of the bulk OM pool into reactivity classes (f_i) needs to be specified manually in the stand-alone version or can be provided by the ESM. Organic matter degradation releases N, P and DIC to the pore water using Redfield molar ratios (Redfield, 1963) and consumes TEA with specific stoichiometries (O_2C, NO_3C, SO_4C) as summarised in Table 10. Table A1 in the appendix provides a list of reactions and their stoichiometries as implemented in OMEN-SED. The effect of OM degradation~~and~~, secondary redox reactions and pyrite precipitation on total alkalinity is also accounted for via reaction specific stoichiometries representing the release of NH_4, H_2S and P and is based on Jourabchi et al. (2005). In reality, the reoxidation of reduced substances produced during OM degradation may be incomplete. Yet, in OMEN-SED we have to assume their complete, instantaneous reoxidation at z_{ox} to allow for an analytical solution. In order to relax this assumption, for cases where it can be justified, we include a "switch" to allow part of the NH_4, H_2S and CH_4 flux to escape reoxidation. The secondary redox parameters (i.e. γ_{NH_4}, γ_{H_2S}, γ_{CH_4}) therefore account for the fraction of reduced substances that are reoxidised and would be ideally parameterised for instance in relation to bottom water oxygen concentration or oxygen penetration depth (z_{ox}). Gypens et al. (2008) for example expressed γ_{NH_4} as a function of oxygen penetration depth ($\gamma_{NH_4} = 0.243 \cdot \ln(z_{ox}) + 1.8479$) based on a fitting exercises to a numerical model and showed that the fraction varies between 0.2 for $z_{ox} = 0.1$cm and 1.0 for $z_{ox} > 3$cm. Due to mathematical constraints in OMEN-SED for finding an analytical solution to the model equations these fractions take constant values generally representing oxygenated deep sea conditions. However, when coupled to an ESM γ_{H_2S} becomes dependent on the bottom water oxygenation state. That is, $\gamma_{H_2S} = 1.0$ for oxic bottom waters and a user defined value $\gamma_{H_2S} < 1.0$ for anoxic bottom waters. The parameter γ_{FeS} represents the fraction of sulfide that is precipitated as pyrite in the sulfidic zone. The majority of H_2S produced by sulfate reduction is reoxidised, but it is estimated that $\sim 10-25\%$ is eventually buried as pyrite (Bottrell and Newton, 2006). However, this fraction can vary significantly over geological timescales (Berner, 1984). If a user does not want to make any assumptions about pyrite precipitation – it can be set to 0 (as in the results presented here). The instantaneous equilibrium adsorption coefficients of NH_4 and PO_4 (K_{NH_4}, $K_{PO_4}^{ox}$, $K_{PO_4}^{anox}$) are based on Wang and Van Cappellen (1996) and Slomp et al. (1998), respectively. The first order rate constants for sorption of PO_4 to Fe oxides (k_s), release of PO_4 from Fe-bound P due to Fe-oxide reduction (k_m) and authigenic CFA precipitation (k_a), as well as the pore water equilibrium concentrations for P sorption and CFA precipitation (PO_4^s, PO_4^a) and the asymptotic concentration for Fe-bound P (FeP^∞) are taken from Slomp et al. (1996). See Table 10 for a complete summary of the parameters and their values.

Table 10. Values for biogeochemical parameters used in OMEN-SED. The variables x, y and z denote the elemental ratio of carbon, nitrogen and phosphorus of the degrading organic matter (here set to $C : N : P = 106 : 16 : 1$).

Parameter/Variable	Unit	Value	Description
Stoichiometric factors and molecular ratios			
NC_i	mol/mol	$\frac{y}{x} = \frac{16}{106}$	Nitrogen to carbon ratio
PC_i	mol/mol	$\frac{z}{x} = \frac{1}{106}$	Phosphorus to carbon ratio
MC	mol/mol	0.5	Methane to carbon ratio
			produced during methanogenesis
$DICC^I$	mol/mol	1.0	DIC to carbon ratio until z_{SO_4}
$DICC^{II}$	mol/mol	0.5	DIC to carbon ratio below z_{SO_4}
O_2C	mol/mol	$\frac{x+2y}{x} = \frac{138}{106}$	Oxygen to carbon ratio
NO_3C	mol/mol	$\frac{4x+3y}{5x} = \frac{94.4}{106}$	Nitrate to carbon ratio
SO_4C	mol/mol	$\frac{106}{212}$	Sulfate to carbon ratio
ALK^{OX}	mol/mol	$\frac{y-2z}{x} = \frac{14}{106}$	ALK from aerobic degradation
ALK^{NIT}	mol/mol	-2	ALK from nitrification
ALK^{DEN}	mol/mol	$\frac{4x+3y-10z}{5x} = \frac{92.4}{106}$	ALK from denitrification
ALK^{SUL}	mol/mol	$\frac{x+y-2z}{x} = \frac{120}{106}$	ALK from sulfate reduction
ALK^{MET}	mol/mol	$\frac{y-2z}{x} = \frac{14}{106}$	ALK from methanogenesis
ALK^{H_2S}	mol/mol	-2	ALK from H_2S oxidation
ALK^{FeS}	mol/mol	-2	ALK from pyrite precipitation
ALK^{AOM}	mol/mol	2	ALK from AOM
Secondary reaction parameters			
γ_{NH_4}	-	0.9	Fraction of NH_4 that is nitrified
γ_{H_2S}	-	1.0	Fraction of H_2S that is oxidised (oxic bottom waters)
	-	0.95	Fraction of H_2S that is oxidised (anoxic bottom waters)
γ_{FeS}	-	0.0	Fraction of H_2S that is precipitated as pyrite
γ_{CH_4}	-	0.99	Fraction of CH_4 that is oxidised
Adsorption coefficients (Wang and Van Cappellen, 1996; Slomp et al., 1998)			
K_{NH_4}	-	1.4	NH_4 adsorption coefficient
$K_{PO_4}^{ox}$, $K_{PO_4}^{anox}$	-	200.0, 2.0	PO_4 adsorption coefficient (oxic, anoxic)
P related parameters (Slomp et al., 1996)			
k_s	yr^{-1}	94.9	Rate constant for PO_4 sorption
k_m	yr^{-1}	0.193	Rate constant for Fe-bound P release
k_a	yr^{-1}	0.365	Rate constant for authigenic CFA precipitation
$PO_4{}^s$	$mol\,cm^{-3}$	$1 \cdot 10^{-9}$	Equilibrium conc. for P sorption
FeP^∞	$mol\,cm^{-3}$	$1.99 \cdot 10^{-10}$	Asymptotic concentration for Fe-bound P
$PO_4{}^a$	$mol\,cm^{-3}$	$3.7 \cdot 10^{-9}$	Equilibrium conc. for authigenic P precipitation

3 Stand-alone sensitivity analysis and case studies

### 3.1 Sensitivity Analysis

3.1.1 Methodology

Model parameters implicitly account for processes that are not explicitly resolved. They are notoriously difficult to constrain and thus a primary source of uncertainty for numerical and analytical models - in particular on the global scale and/or in data-poor areas. A comprehensive sensitivity analysis can help quantify this uncertainty and identify the most sensitive parameters. More specifically, sensitivity analysis is used to investigate how the variations in the outputs (y_1, ..., y_N) of a model can be attributed to variations in the different input parameters (x_1, ..., x_M, Pianosi et al., 2016). Different types of sensitivity indices, which quantify the relative influence of parameter x_i on output y_j with a scalar $S_{i,j}$ (for $i \in \{1, ..., M\}$ and $j \in \{1, ..., N\}$), can be calculated, ranging from simple one-at-a-time methods to statistical evaluations of the output distribution (e.g. variance-based or density-based approaches Pianosi et al., 2016). The latter indices take values between zero and one ($S_{i,j} \in [0,1]$), where zero indicates a non-influential parameter and a higher value a more influential parameter. Here, sensitivity analysis is used mainly to identify which parameters have the largest impact on the different model outputs and therefore require more careful calibration. As the probability density functions of our model outputs (i.e. the resulting SWI-fluxes) are generally highly-skewed towards extreme organic matter degradation rates (not shown) variance-based sensitivity indices may not be a suitable proxy for output uncertainty (Pianosi et al., 2016). Hence, instead the density-based PAWN method by Pianosi and Wagener (2015) is employed which considers the entire conditional and unconditional Cumulative Distribution Function (CDF) of the model output rather than its variance only. The unconditional CDF, $F_y(y)$, of output y is obtained when all uncertain parameters (x_1, ..., x_M) are varied simultaneously, and the conditional CDFs, $F_{y|x_i}(y)$, are obtained when all inputs but the i-th parameter are varied (i.e. x_i is fixed to a so-called conditioning value). The sensitivity index of parameter i is measured by the distance between the two CDFs using the Kolmogorov-Smirnov statistic (Kolmogorov, 1933; Smirnov, 1939), i.e.:

$$S_i = \max_{x_i} \max_y |F_y(y) - F_{y|x_i}(y)|. \tag{49}$$

Since $F_{y|x_i}(y)$ accounts for what happens when the variability due to x_i is removed, the distance between the two CDFs provides a measure of the effects of x_i on the output y. Due to the model complexity it is impossible to compute the sensitivity indices analytically. Therefore, they are approximated from a Latin-Hypercube sampling of parameter inputs and calculated outputs. For a brief description of the methodology, see Fig. 4. For more details, we refer the interested reader to Pianosi and Wagener (2015).

The PAWN method, as implemented within the Sensitivity Analysis for Everyone (SAFE) matlab toolbox (Pianosi et al., 2015), is used to investigate M = 11 model parameters for ranges as specified in Table 11. Sensitivity indices for all resulting SWI-fluxes for two idealised sediment conditions (i.e. anoxic at 400 m and oxic at 4000 m, see Table 12) are calculated. We use NU = 200 samples to estimate the unconditional CDF, NC = 100 samples to estimate the conditional CDFs and n = 10
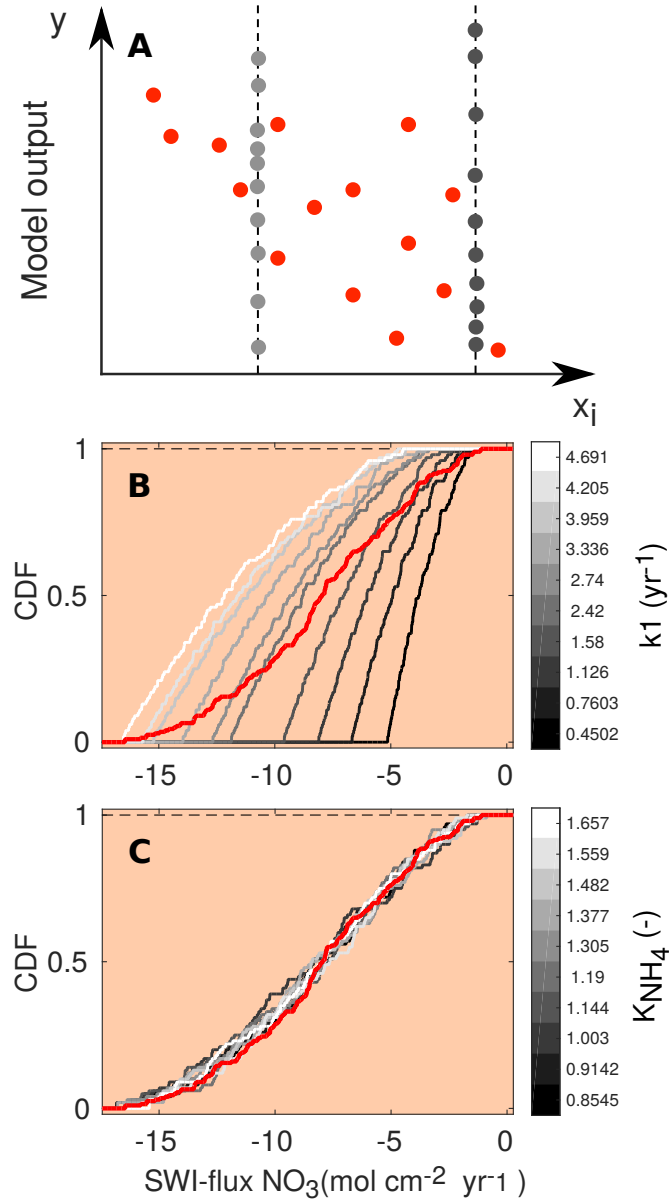
Figure 4. A: Schematic of the PAWN method, plotting an uncertain parameter (x_i) against a generic model output (y). Red dots represent points for calculating the unconditional CDF (NU, here 15), grey dots are points for calculating each conditional CDF (NC, here 10) with n = 2 conditioning points as an example. The user can change the values of NU, NC and n. The number of model evaluations equals N_{eval} =NU + n·NC·M, where M is the number of uncertain input parameters. B + C: Two examples of CDFs of the model calculated SWI-flux of NO_3 using NU = 200, NC = 100 and n = 10. The red lines are the unconditional distribution functions $F_y(NO_3)$ and the grey lines are the conditional distribution functions $F_{y|x_i}(NO_3)$ at different fixed values of the input parameters k_1 (B) and K_{NH_4} (C). As the maximal distance between conditional CDFs and unconditional CDF is greater for k_1, this parameter is more influential for the model output (here SWI-flux of NO_3, compare Fig. 5).

Table 11. Range of model parameters used for sensitivity analysis of model predicted output.

Parameter	Description	Units	Minimum	Maximum	Source
k_1	labile OM degradation constant	yr^{-1}	$1e^{-4}$	5.0	(1)
$\widetilde{k}_2$	order of refractory OM degradation constant ($k_2 = \widetilde{k}_2 \cdot k_1$)	-	$1e^{-4}$	$1e^{-1}$	(1)
f_1	fraction of labile OM	-	0.02	0.98	-
$K_{\mathrm{NH_4}}$	Adsorption coefficient	-	0.8	1.7	(2)
$\gamma_{\mathrm{NH_4}}$	NH_4 fraction oxidised		0.5	1.0	-
$\gamma_{\mathrm{H_2S}}$	H_2S fraction oxidised		0.5	1.0	-
$K_{\mathrm{PO_4}}^{\mathrm{ox}}$	Adsorption coeff. oxic	-	100.0	400.0	(3)
$K_{\mathrm{PO_4}}^{\mathrm{anox}}$	Adsorption coeff. anoxic	-	1.3	2.0	(3)
k_s	kinetic P sorption	yr^{-1}	0.1	100.0	(4, 5)
k_m	Fe-bound P release	yr^{-1}	0.015	0.02	(4, 5)
k_a	authigenic P formation	yr^{-1}	0.001	10.0	(4, 6)

Sources: (1) Arndt et al. (2013); (2): Van Cappellen and Wang (1996); (3): Krom and Berner (1980)

(4): Gypens et al. (2008); (5): Slomp et al. (1996); (6): Van Cappellen and Berner (1988)

Table 12. Model boundary conditions for the two idealised sediment conditions used for the sensitivity analysis (Fig. 5 and 6). All solute concentrations are in nmol cm^{-3}.

Depth (m)	Temp. (°C)	OC (wt%)	O_2	NO_3	SO_4	PO_4	z_{bio} (cm)
400	8.0	2.0	0.0	40.0	28,000	40.0	0.001
4000	1.5	1.0	300.0	20.0	28,000	40.0	10.0

conditioning points. Thus as $N_{\mathrm{eval}} = 200 + 100 \cdot 10 \cdot 11$, 11200 model evaluations are performed for each sediment condition. The resulting indices are then translated into a colour code and summarised in a pattern plot to simplify comparison (Fig. 5).

3.1.2 Results

~~Fig.~~ Figure 5 summarises results of the sensitivity analysis as a colour map. Results indicate that generally the most significant
5 parameters for all model outputs are the degradation rate constant for the labile OM pool (k_1) and the fraction of this pool to the total OM stock (f_1). Other parameters play a minor role for the SWI-fluxes, with the exception of the secondary redox parameters (i.e. $\gamma_{\mathrm{NH_4}}$, $\gamma_{\mathrm{H_2S}}$) in the oxic scenario. Here, NH_4, SO_4 and H_2S fluxes are very sensitive to changes in $\gamma_{\mathrm{NH_4}}$ and $\gamma_{\mathrm{H_2S}}$, as these parameters determine how much of the respective TEA is produced in situ via reoxidation, thus affecting the resulting SWI-fluxes. For the oxic scenario, the reoxidation of H_2S produced in the sulfidic layer also has a strong influence
10 on alkalinity ($\gamma_{\mathrm{H_2S}}$, Table 8 Eq. 5) as it decreases alkalinity by 2 moles per mole of S oxidized (ALK$^{\mathrm{H_2S}}$, Table 10). However, these high sensitivities are partially caused by the wide range of allowed values ($\gamma_{\mathrm{NH_4}}$, $\gamma_{\mathrm{NH_4}} \in [0.5; 1.0]$). Yet, for oxic deep

sea conditions it is more likely that reduced substances are almost completely reoxidised (e.g. Hensen et al., 2006). For the anoxic scenario the secondary redox parameters are essentially non-influential as no O_2 is available for the reoxidation of reduced substances. Especially for the oxic condition the PO_4 SWI-flux appears to be insensitive to P-related parameters (i.e. $K_{PO_4}^{ox}$, $K_{PO_4}^{anox}$, k_s, k_m, k_a) as the majority is absorbed to Fe-oxides. The sensitivities change if other PO_4 related equilibrium concentrations $PO_4{}^s$, $PO_4{}^a$ and FeP^∞ are used (not shown). Overall the results of the sensitivity analysis are in line with what one would expect from a diagenetic model and thus provide ground to confirm that OMEN-SED provides sensible results. The parameterisation of the organic matter pools (f_1) and their degradation rate constants (k_1, k_2) is critical especially when the model is used in a global Earth system model framework, as these parameters, as well as the γ-parameters, can have a very important influence on the flux of dissolved species through the SWI. At the same time these are the weakest constrained parameters. Thus, one should rather choose γ-values close to 1 and consider carefully where a relaxation of the "all reoxidised" assumption is appropriate. In contrast, the importance of the OM degradation rate constants can not be overemphasised. Therefore, much care should be given to how these are parameterised in coupled simulations and a range of different plausible scenarios should be tested to quantify uncertainty.

Because of the strong sensitivity of model results on OM degradation rate parameters, we further explore the sensitivity of simulated sediment-water exchange fluxes to variations in organic matter degradation parameters by varying k_1, f_1 and $\widetilde{k_2}$ while all other model parameters are set to their default values (Tables 9 and 10). Minimum and maximum values for k_1, $\widetilde{k_2}$ and f_1 in the shallow ocean are as in Table 11. For the deep sea condition we account for the presence of more refractory OM by sampling $f_1 \in [0.02, 0.3]$, whereas the variation of k_1 and $\widetilde{k_2}$ is as in the shallow ocean. The parameter space is sampled using another Latin-Hypercube approach with sample sizes of $N = 3500$ for each idealised sediment condition. Figure 6 summarises the results of the sensitivity study and the ranges of observed O_2 and NO_3 sediment-water interface fluxes extracted from a global database (Stolpovsky et al., 2015) are indicated on the colour scale. The colour patterns in Figure 6 A and B reveal the complex interplay between the amount of labile OM f_1 and its degradation rate k_1 for the resulting SWI-fluxes of NO_3 in anoxic sediments and O_2 in aerobic sediments. In general, a higher degradation rate in combination with more labile OM available leads to a higher SWI-flux. However, higher fluxes extend over a larger range of k_1-values when the amount of labile OM f_1 is high. The absence of a colour pattern in Figure 6 C highlights the limited interaction of the two model parameters for NO_3 SWI-fluxes under oxic conditions. Figure 6 shows that SWI-fluxes can vary widely over the range of plausible organic matter degradation parameters and that simulated fluxes generally fall within the range of observed SWI-fluxes. However, a large number of different k-f combinations can result in SWI-fluxes that fall within the observed ranges reported by Stolpovsky et al. (2015) further emphasising the care that should be devoted to constraining OM degradation parameters.

3.2 Case study: Simulations of sediment cores

3.2.1 Methodology

In order to illustrate the capabilities of OMEN-SED, comprehensive datasets from the Santa Barbara Basin (Reimers et al., 1996), as well as from the Iberian margin and the Nazaré Canyon (Epping et al., 2002) are modelled. Modelled profiles are
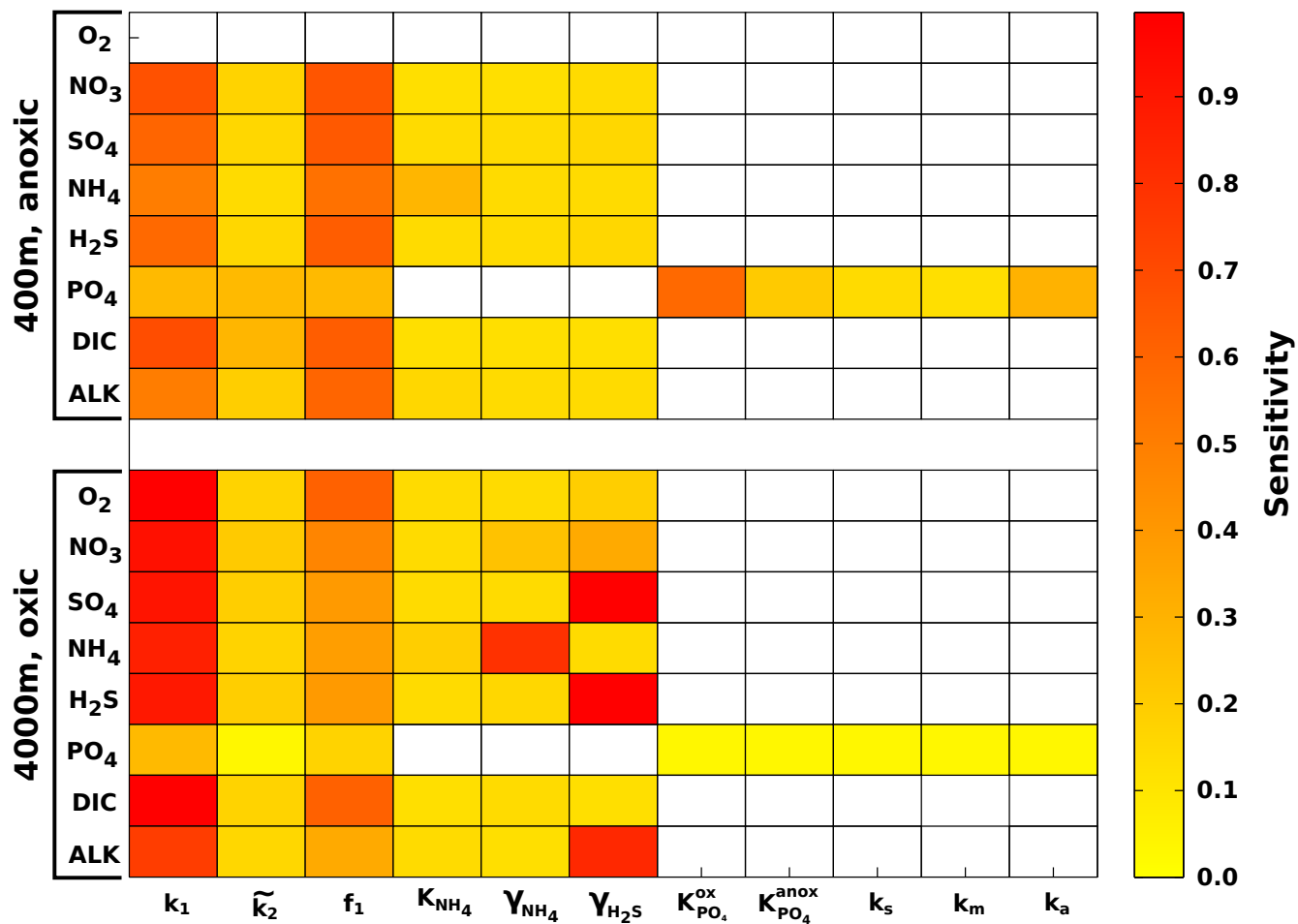
Figure 5. Pattern plot, showing the output sensitivity for each SWI flux (i.e. the chemical compounds on the vertical axis) and each input factor (i.e. the model parameters on the horizontal axis) for two idealised sediment cores. White patterns are assigned where the SWI flux is independent of the specific parameter.
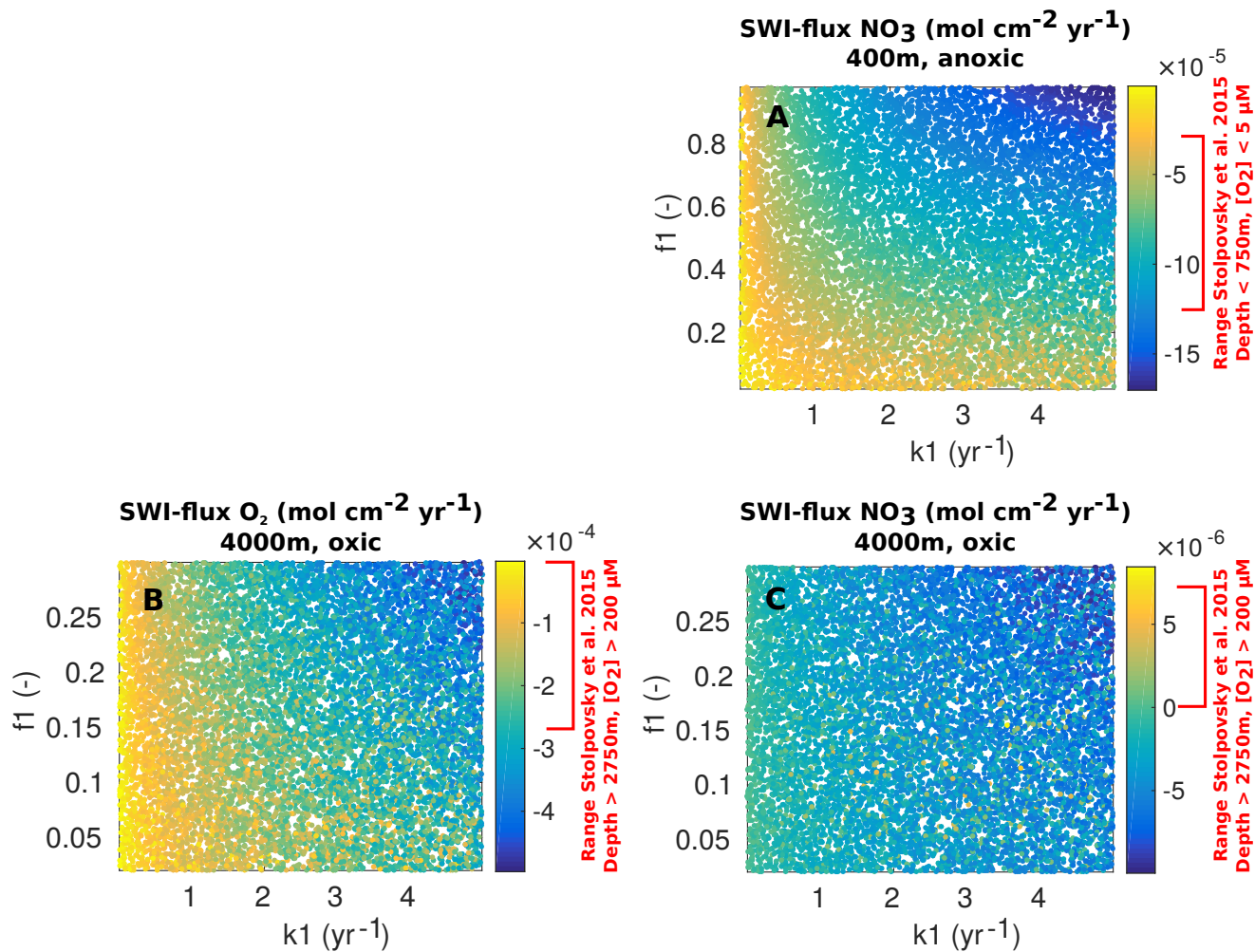
Figure 6. Scatter plots (k_1 vs f_1) of resulting OMEN-SED SWI-fluxes for the 400m anoxic (A: NO_3) and 4000m oxic (B: O_2, C: NO_3) scenario. Negative values represent a flux from the water column into the sediments. Ranges indicated in red on the colour scale correspond to observed benthic fluxes as reported in the global database of Stolpovsky et al. (2015).

compared with measured pore water data from different depths including the continental shelf (108 m) and the lower slope (2213 m) located at the Iberian margin, the upper slope (585 m) from the Santa Barbara Basin, and a deep sea site (4298 m) in the Nazaré Canyon. The Santa Barbara Basin is characterised by anoxic bottom waters, high POC concentrations and varved sediments (Reimers et al., 1990), therefore the depth of bioturbation in OMEN-SED is restricted to the upper 0.01 cm. In the uppermost sediments iron(III) hydroxides are reduced, releasing Fe^{2+} which reacts with sulfide to form iron sulfides. Thus, the Fe cycle exerts a strong control on sulfide concentrations in the sediments of this basin (Reimers et al., 1996). In addition, the sediments are generally supersaturated with respect to carbonate fluorapatite by and below 2 cm (Reimers et al., 1996). The Iberian margin, situated in the northeastern Atlantic, generally belongs to the more productive regions of the global ocean (Longhurst et al., 1995), however, seasonal changes in upwelling creates a strong temporal variability in primary productivity and organic carbon deposition. Submarine canyons in this area (like the Nazaré Canyon) may deliver organic carbon from the shelf to the ocean interior (van Weering et al., 2002; Epping et al., 2002). For a more detailed description of the study areas and the experimental work, the interested reader is referred to the publications by Reimers et al. (1996) and Epping et al. (2002).

In OMEN-SED sediment characteristics and boundary conditions are set to the observed values where available (Table 13). Other sediment characteristics (e.g. sedimentation rate, porosity, density), stoichiometric factors and secondary reaction parameters are set to the default value (see Tables 9 and 10). Organic matter is modelled as two fractions, with different first-order degradation rate constants. The POC and pore water profiles were manually fitted by optimizing the POC partitioning into the fast and slow degrading pool and their respective first-order degradation rate constants (priority is given to reproduce the POC and O_2 profiles). For phosphorus the equilibrium concentration for authigenic P formation (PO_4^a) was adjusted to fit the PO_4 concentration at $\cancel{z_\infty}z_{max}$.

3.2.2 Results

Figure 7 compares modelled and observed sediment profiles for the Santa Barbara Basin and the Iberian margin. Results show that OMEN-SED is able to capture the main diagenetic features across a range of different environments without changing model parameters (other than the 4 we tuned, i.e. k_1, k_2, f_1 and PO_4^a) to site specific conditions. For the two open Iberian margin stations (108 and 2213 m) OMEN-SED fits all observations well. OMEN-SED does especially well at seafloor depth (SFD) 2213 m by reproducing the deep O_2 penetration and the subsurface maximum in NO_3 concentration due to the nitrification of NH_4 (note, that NH_4 is overestimated at this SFD). For the anoxic Santa Barbara Basin (585 m) the decrease in SO_4 and the increase in ALK concentration with sediment depth is well represented, indicating the importance of sulfate reduction as the primary pathway of OM degradation at this site (compare with Meysman et al., 2003). However, a misfit is observed for H_2S and PO_4 in the upper 20 cm of this sediment core. The discrepancy for H_2S can be explained by high iron(III) hydroxide concentrations, which is reduced to degrade organic matter (especially in the $2-4$ cm depth interval), therefore placing the beginning of the sulfate reduction zone and the production of H_2S to the deeper sediments (Reimers et al., 1996). Iron processes are currently not dynamically represented in OMEN-SED. In addition, produced dissolved Fe reacts with H_2S to form iron sulfides (e.g. pyrite, FeS_2) and thus further inhibits the rise of H_2S (Reimers et al., 1990). The iron cycle also plays a critical role for phosphorus, as the reduction of iron(III) hydroxides in the surface sediments releases sorbed phosphate,

Table 13. ~~Model boundary~~ Boundary conditions for ~~the~~ simulated sediment profiles ~~in~~ at the ~~Santa Barbara basin~~ Iberian margin (108 and 2213 m)~~and Iberian margin~~, the Santa Barbara basin (585 m) and the Nazaré Canyon (4298 m) reported in Figure 7. For all sites a DIC bottom water concentration of 2,400 nmols cm^{-3} is assumed.

Sediment characteristics:

Depth (m)	Temp. (°C)	z_{bio} (cm)	D_{bio} (cm^{2}yr^{-1})	POC$_1$ (wt%)	POC$_2$ (wt%)	k_1 (yr^{-1})	k_2 (yr^{-1})	PO$_4^a$ (nmol cm^{-3})
108	12.50	1.00	0.02	2.64	1.8	0.650	$1.0e^{-5}$	15.0
585	5.85	0.01	0.02	2.00	3.5	0.200	$8.0e^{-4}$	90.0
2213	3.20	10.00	0.17	0.45	0.5	0.100	$4.0e^{-4}$	5.0
4298	2.50	4.20	0.18	0.83	1.2	0.052	$1.0e^{-5}$	5.0

Bottom water concentrations of solutes (all in nmol cm^{-3}):

Depth	O$_2$	NO$_3$	SO$_4$	NH$_4$	H$_2$S	PO$_4$	Alkalinity
108	210.0	9.6	28,000	0.40	0.0	0.0	2,400
585	10.0	25.0	28,000	0.00	0.0	50.0	2,480
2213	250.0	25.0	28,000	0.60	0.0	0.0	2,400
4298	243.0	30.1	28,000	0.22	0.0	0.0	2,400

leading to pore waters around and below 2 cm which are supersaturated with respect to fluorapatite, thus initiating CFA precipitation. Reimers et al. (1996) could even show that the accumulation of CFA is mainly restricted to the near-surface sediments (~ 5 cm) instead of throughout the sediment column. As OMEN-SED currently does not include an iron-cycle, and Fe-bound P and CFA processes are highly parameterised, the model is not able to capture these complex, non-steady state phosphorus dynamics at this specific site. For the Nazaré Canyon station (4298 m) satisfactory fits could be realised apart from NH$_4$. However, also Epping et al. (2002) could not obtain a better fit using the diagenetic model OMEXDIA. They suggested non-local solute exchange resulting from bioirrigation being responsible for the higher NH$_4$ concentrations at this site which is neglected in their model, as well as in OMEN-SED. Furthermore, the fractured POC profile (indicating episodic depositional events through the canyon) could have been approximated using a different partitioning of the bulk POC into labile and refractory pool with different degradation rate constants, thus potentially leading to a better fit of the NH$_4$ profile. In general, better approximations of the data could have potentially been acquired by applying a sensitivity study using different NC-ratios (e.g. Epping et al., 2002, report different ratios from Redfield stoichiometry) and exploring the parameter space for the secondary reaction parameters (γ_{NH_4}, γ_{H_2S}). However, considering these generalisations and our assumption of steady-state, which might not be valid, particularly for the complex Santa Barbara basin, the shallow core and the Nazaré Canyon, which are affected by seasonality and biology, OMEN-SED generally reproduces the observed pore water trends and hence captures the main diagenetic processes.
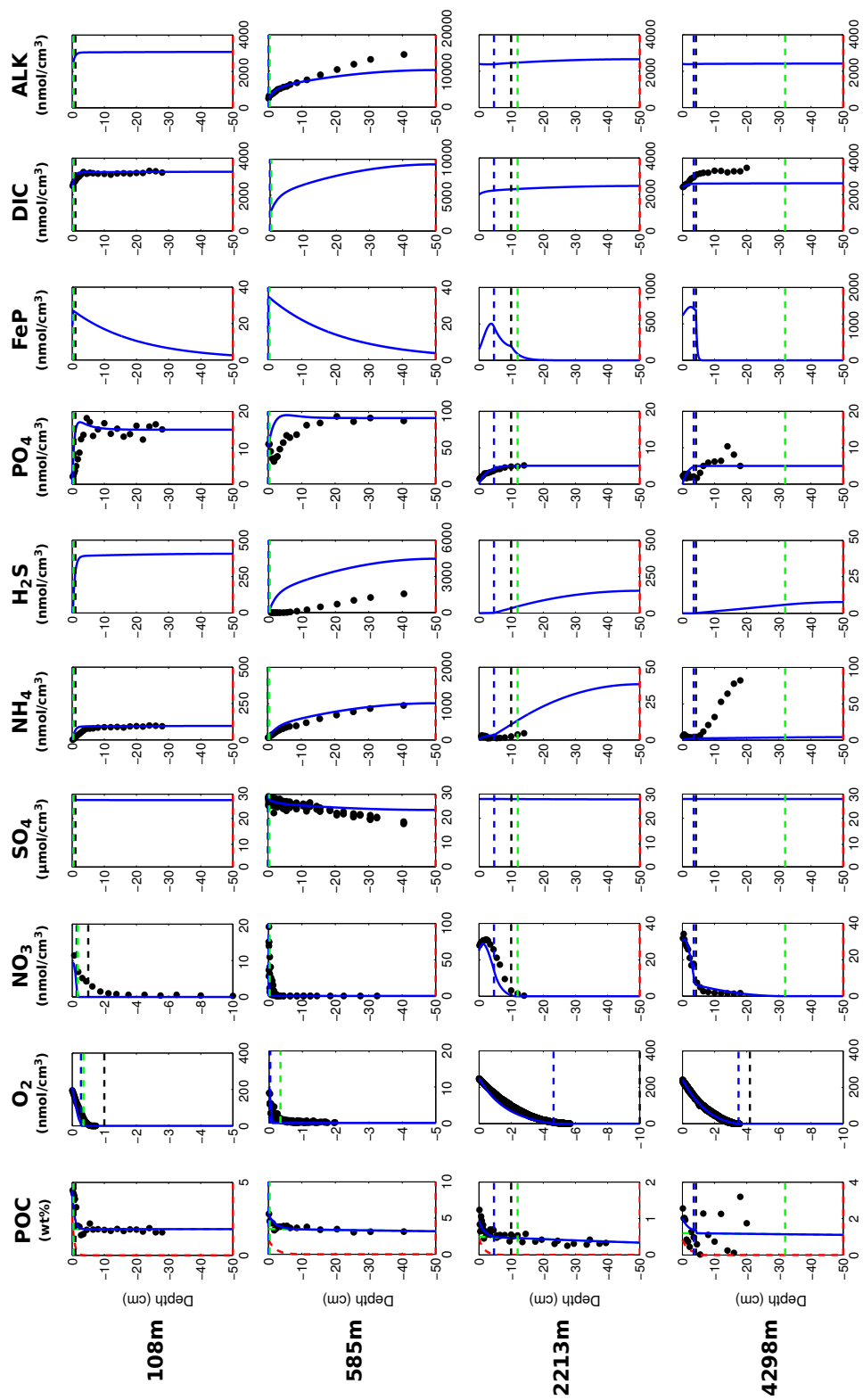
Figure 7. Modelled (curves) and measured (filled dots) solid phase and dissolved pore water profiles for four different sediment cores. Note that different scales are used for different stations. The blue POC curve represents the sum of the refractory (green) and labile (red) POC fraction. The horizontal dashed lines in each panel indicate the bioturbation depth (black) and the penetration depths of oxygen (blue), nitrate (green) and sulfate (red) as calculated by OMEN-SED.

3.3 Case study: Stand-alone simulations of global ocean transect

3.3.1 Methodology

In this section we explore to what degree OMEN-SED is capable of capturing the dynamics of organic matter degradation pathways and related TEA-fluxes as simulated with a more complete and complex numerical diagenetic model (Thullner et al., 2009). Therefore, we reproduce the simulations of typical conditions along a global ocean hypsometry of Thullner et al. (2009) and compare our modelled TEA-fluxes with the results of the complex model as well as with observations from Middelburg et al. (1996). To explore the global degradation of OM in the seafloor Thullner et al. (2009) quantified various diagenetic processes using the Biogeochemical Reaction Network Simulator (BRNS, Aguilera et al., 2005), a flexible simulation environment suitable for reactive transport simulations of complex biogeochemical problems (e.g. Jourabchi et al., 2005; Thullner et al., 2005). Thullner et al. (2009) used seafloor depth (SFD) as the master variable and calculated model parameters, such as w, D_{bio} and ϕ, from existing empirical relationships (e.g. Van Cappellen and Wang, 1995; Middelburg et al., 1997). Organic matter degradation was described with a 1G approach, thus assuming a single pool of organic matter of uniform reactivity. The first order rate constant was related to the burial velocity, w (cm year^{-1}), following the empirical relationship of Boudreau (1997):

$$k = 0.38 \cdot w^{0.59}. \tag{50}$$

This rate constant can be assumed as the mean reactivity of the organic matter fractions which are degraded in the upper, bioturbated $10-20$ cm of the sediments. Thus, more reactive fractions (degraded during days/weeks close to the SWI) and more refractory fractions (degraded on longer time scales deeper in the sediments) are not captured by this relationship (Boudreau, 1997). BRNS simulations were performed using boundary conditions and parameters for depths representative for shelf, slope and deep sea sediments (i.e. SFD of 100m, 200m, 500m, 1000m, 2000m, 3500m and 5000m). In order to reproduce these results, OMEN-SED is configured here as a 1G model and boundary conditions and model parameters are defined as in Thullner et al. (2009, see Table 14). As OMEN-SED assumes a fixed fraction (i.e. γ_{NH_4}, γ_{H_2S}) of reduced substances to be reoxidised, which exerts a large impact on the resulting SWI-fluxes (compare Section 3.1), two sets of simulations are performed in order to show the range of possible model outputs. In the first setup 95% of the reduced substances are reoxidised (i.e. $\gamma_{NH_4} = \gamma_{H_2S} = 0.95$) and in the second, less realistic case, only 5% are reoxidised (all other model parameters and boundary conditions are equal).

3.3.2 Results

Figure 8 compares simulated SWI-fluxes of TEAs (i.e. O_2, NO_3 and SO_4) along the global hypsometry using OMEN-SED (black lines) with the results of Thullner et al. (2009) (red lines). Observations for O_2 and NO_3 fluxes are taken from Middelburg et al. (1996). Due to the applied empirical relations organic matter flux to the seafloor decreases by 2 orders of magnitude from 100 to 5000 m and its degradation rate constant by 1 order of magnitude (Table 14). Therefore, the rate of organic matter degradation is about 50 times greater at 100 m than at 5000 m (compare Thullner et al., 2009), thus resulting in a decrease

38

Table 14. Seafloor depth dependency of key model parameters and boundary conditions (adapted from Thullner et al., 2009).

	Seafloor depth						
	100 m	200 m	500 m	1000 m	2000 m	3500 m	5000 m
Model parameters							
w^a (cm yr^{-1})	3.98×10^{-1}	3.60×10^{-1}	2.67×10^{-1}	1.62×10^{-1}	5.94×10^{-2}	1.32×10^{-2}	2.94×10^{-3}
D_{bio}^a (cm^2 yr^{-1})	27.5	25.1	19.0	12.1	4.83	1.23	0.310
ϕ^b	0.85	0.85	0.80	0.80	0.80	0.80	0.80
T^c (°C)	10.3	9.7	8.1	5.8	3.0	1.5	1.4
ρ_{sed}^c (g cm^{-3})	2.5	2.5	2.5	2.5	2.5	2.5	2.5
k^d (yr^{-1})	0.221	0.208	0.174	0.130	0.0718	0.0296	0.0122
Upper boundary conditions							
$\text{POC}_{\text{flux}}^a$ (μmol cm^{-2} yr^{-1})	510	467	357	228	93.0	24.3	6.33
POC^e (wt%)	0.79	0.78	0.55	0.50	0.42	0.32	0.25
$O_{2,0}^c$ (nmol cm^{-3})	132	129	121	114	116	135	141
$NO_{3,0}^c$ (nmol cm^{-3})	17.3	18.6	22.1	26.5	31.0	31.6	31.6
$SO_{4,0}^b$ (nmol cm^{-3})	28,000	28,000	28,000	28,000	28,000	28,000	28,000

[a] Derived from Middelburg et al. (1997).

[b] Derived from Van Cappellen and Wang (1995).

[c] Derived from Conkright et al. (2002).

[d] Derived from Boudreau (1997).

[e] Calculated with OMEN-SED from POC$_{\text{flux}}$.

of TEA-fluxes along the hypsometry (Figure 8). The 95%-reoxidation experiments (dots) show proportionally higher O_2 in-fluxes than the 5%-reoxidation experiments (triangles) because more O_2 is utilised for in situ production of NO_3 and SO_4 in the sediments. This is also mirrored by the increased NO_3 out-flux and decreased SO_4 in-flux for shallower SFDs. This is in line with the results of Thullner et al. (2009) which showed that in situ production is an important pathway of SO_4 supply in the sediment, which is responsible for $\sim$80% of the total OM degradation at depths between 100 and 2000 m (in our results SO_4 is not used for OM degradation in OMEN-SED below 2000m). In general, Figure 8 shows that OMEN-SED captures the main trends in observed and numerically simulated TEA fluxes well. Results also confirm that higher γ-values better represent SWI-fluxes for most of the global hypsometry. A slight overestimation of shallow ocean SWI-fluxes (SFD < 200 m) for the high γ scenario indicates that slightly lower γ-values would better capture SWI-fluxes in these areas, where rapid oxygen consumption favours the escape of reduced species across the SWI.

In addition, observed O_2 fluxes in the upper 2000m are generally encompassed by our total range in predicted OMEN-SED fluxes. Oxygen fluxes for the deep-sea sediments, however, are slightly underestimated. These deviations can presumably be related to the assumed 1G description of organic matter degradation, which neglects the more labile OM pool. This highly reactive pool is degraded close to the sediment surface, thus promoting higher aerobic degradation rates and higher O_2 fluxes. Nitrate fluxes in the upper 500m of the Atlantic Ocean are well predicted. However, as in Middelburg et al. (1996) the direction of calculated nitrate fluxes in the upper 1000m of the Pacific Ocean differ from the observations. Middelburg et al. (1996) related these discrepancies to the globally averaged model parameters and the applied boundary conditions. They could reduce the disagreements significantly by using more representative bottom water concentrations for the eastern Pacific and a higher flux of labile organic matter for their 2G model. By changing the boundary conditions and the N:C elemental ratio of organic matter for the whole hypsometry, it is possible to obtain a better model-data fit with OMEN-SED for the shallow Pacific Ocean (green line in Fig. 8B). Bohlen et al. (2012) report that the elemental N:C ratio strongly deviates from Redfield stoichiometry (0.151) with specifically lower values for the East Pacific Ocean. The use of their globally averaged value of 0.067 allows reconciling modelled and observed values provided that bottom water conditions are also changed to the low oxygen/high nitrate levels more likely to be found in the shallow Pacific Ocean (O_2 = 10 nmol cm^{-3} and NO_3 = 80 nmol cm^{-3}).

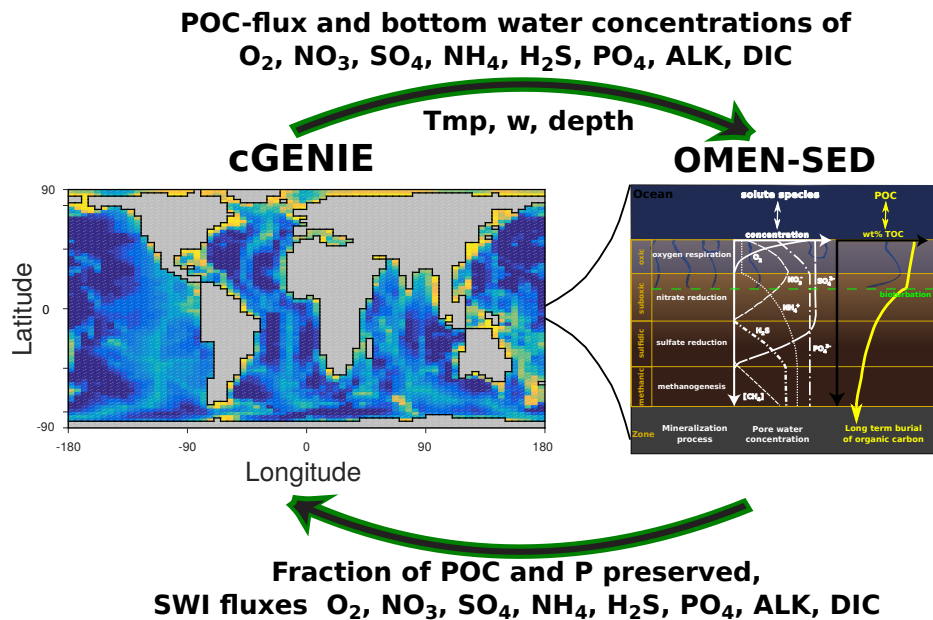## 4 Coupled pre-industrial Earth system model simulations

4.1 Coupling to the cGENIE Earth system model

In a final step, we couple OMEN-SED to the carbon-centric version of the "GENIE" Earth system model (cGENIE, Ridgwell et al., 2007) in order to illustrate how a fully coupled ocean-sediment system can be configured and applied. We start by providing a brief description of cGENIE and the coupling procedure (Fig. 9).

cGENIE is a model of Intermediate Complexity based on the efficient climate model "C-GOLDSTEIN" of Edwards and Marsh (2005), featuring a frictional-geostrophic 3D-ocean circulation model coupled to a fast Energy-Moisture Balance 2D-atmosphere together with a dynamic-thermodynamic sea-ice component. The version of cGENIE used here includes the marine geochemical cycling of carbon, oxygen, phosphorus and sulfur (Ridgwell et al., 2007), preservation of carbonates in deep-sea
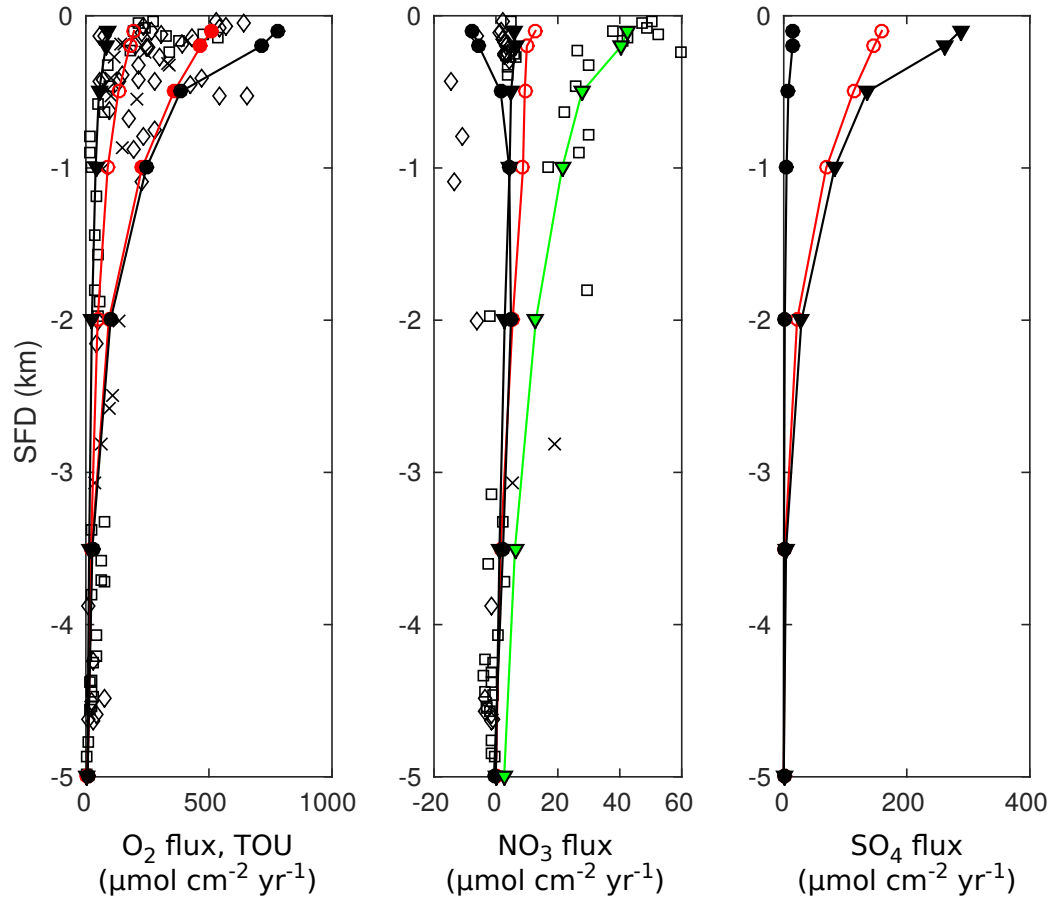
Figure 8. Fluxes of O_2, NO_3 and SO_4 to the sediment along the global hypsometry. Red lines (with open symbols) are modelled fluxes from Thullner et al. (2009) using BRNS; black lines are results from OMEN-SED ($\bullet$: $\gamma_{NH_4} = \gamma_{H_2S} = 0.95$; $\blacktriangledown$: $\gamma_{NH_4} = \gamma_{H_2S} = 0.05$). Observations of TEA fluxes are taken from Middelburg et al. (1996) ($\Diamond$: Atlantic, $\square$: Pacific, $\times$: Arctic/Indian Ocean). Also plotted in Figure (A) are the total oxygen uptake (TOU) estimates of Thullner et al. (2009) (filled red symbols). The green line indicates OMEN-SED results for low oxygen/high nitrate levels and the lower NC-ratio. Positive fluxes are directed from the ocean into the sediments.

POC-flux and bottom water concentrations of
O$_2$, NO$_3$, SO$_4$, NH$_4$, H$_2$S, PO$_4$, ALK, DIC

Tmp, w, depth

cGENIE   **OMEN-SED**

Fraction of POC and P preserved,
SWI fluxes O$_2$, NO$_3$, SO$_4$, NH$_4$, H$_2$S, PO$_4$, ALK, DIC

Figure 9. Schematic of the relationship between OMEN-SED and cGENIE. Arrows represent the information transferred between models.

sediments (SEDGEM, Ridgwell and Hargreaves, 2007) and terrestrial weathering (Colbourn et al., 2013). The ocean model is implemented on a 36×36 equal-area horizontal grid with 16 vertical levels using the pre-industrial continental configuration and bathymetry as in Archer et al. (2009). A finer grid (72×72) is used for the sediments (see Fig. 11C and Ridgwell and Hargreaves, 2007) and OMEN-SED is called by SEDGEM for each wet ocean grid point.

5   In our Earth system model set-up, we prescribe the burial sediment fluxes of detrital material, opal and $CaCO_3$, while leaving OMEN-SED to calculate organic matter preservation. This assumption serves two purposes. First, the run-time of the model is minimized as steady-state conditions are reached earlier, compared to the ca. 20-50 kyr adjustment time for surface sediment $CaCO_3$ (Ridgwell and Hargreaves, 2007). Second, invariant flux fields remove feedbacks between OMEN-SED and the calculation of $CaCO_3$ preservation (changes in organic matter preservation affect $CaCO_3$ dissolution and hence burial rates

10   which in turn affects weight percent of organic matter in the sediments) that would not only lengthen the sediment adjustment time but also make it impossible to carry out unbiased comparisons between different assumptions regarding organic matter reactivity in OMEN-SED.

We derive these fields form the data compilation of Archer (1996) as follows. First, we re-grid the Archer (1996) interpolated non carbonate mass accumulation rate field (NC_{flux}) to the 72×72 cGENIE sediment grid. This field includes detrital material

15   plus opal (plus a minor contribution from organic matter). We could then directly calculate $\sum_{flux}$ (total burial flux of all components or total sediment accumulation rate) from this plus measurements of coretop wt% $CaCO_3$ (C_{wtpct}) (Archer, 1996) as $\sum_{flux} = NC_{flux} \cdot (1 - \frac{C_{wtpct}}{100})^{-1}$. However, some of the Archer (1996) database C_{wtpct} values are both close to

42

100% and associated with high NC_{flux}, and hence would lead to unrealistically high values for $\sum_{flux}$. We therefore impose a plausibility filter, by also re-gridding coretop wt% opal (O_{wtpct}) and quartz (Q_{wtpct}) and for grid points in which more than one component is reported and the sum exceeds 100 wt%, normalizing the individual components ~~. (Note that~~ (for grid points with only a single solid component, no change is made~~.))~~. We then calculate the individual solid component burial fluxes, and

5 sum them up. To interpolate between the grid points associated with data, we iteratively average nearest (adjoining) neighbours. The distribution of the total burial flux $\sum_{flux}$ (in $g\,cm^{-2}\,kyr^{-1}$) is shown in Figure C1 in the Appendix.

Depending on the configuration of the overlying biogeochemical ocean model, processes can be included or excluded in OMEN-SED and stoichiometric factors (Tab. 10) need to be matched between models to ensure preservation of mass. As nitrogen is not modelled explicitly in the employed cGENIE configuration, NC_i, ALK^{NIT} and ALK^{DEN} in OMEN-SED are set

10 to zero. cGENIE, however, implicitly includes the effects of NH_4 release and its complete nitrification on alkalinity but neglects the impact of P release. Therefore, alkalinity stoichiometries for aerobic degradation, sulfate reduction and methanogenesis are changed to $ALK^{OX} = -16/106$, $ALK^{SUL} = 122/106$ and $ALK^{MET} = -16/106$, respectively (compare to default in Table 10).

Various biogeochemical tracers and parameters are transferred from SEDGEM to OMEN-SED (see Fig. 9) and are converted

15 into the required units. Bottom water concentrations of solutes are converted from $mol\,kg^{-1}$ to $mol\,cm^{-3}$ and the depositional flux of POC (POC_{flux}) is converted from $cm^3\,cm^{-2}\,yr^{-1}$ to $mol\,cm^{-2}\,yr^{-1}$ assuming an average density of POC of $1.0\,g\,cm^{-3}$. Within the water column in cGENIE, POC is partitioned into two fractions with different degradation length scales of $\sim590\,m$ and $1000000\,m$, respectively. The labile pool thus degrades while sinking through the water column, whereas the refractory pool is assumed relatively unreactive (Ridgwell et al., 2007). Thus, depending on seafloor depth, the partitioning of bulk

20 POC reaching the sediments is different (Fig. 10A+B). This information is used by OMEN-SED to define the parameter f_1. Other parameters used from cGENIE are seafloor depth and local temperature. The advection/burial rate (w) is taken from the previous time-step of cGENIE, however, it is assured that w is not smaller than the detrital flux (Det_{flux}) to the sediments (e.g. $w < 0$ can occur if initially carbonate rich sediments are eroded during the spin-up of cGENIE). In case ($w \leq Det_{flux}$ & $Det_{flux} = 0.0$) all POC is remineralised at the ocean floor. Furthermore, a minimum value of $w = 0.4\,cm\,kyr^{-1}$ is imposed

25 as OMEN-SED tends to be less stable for lower values. For comparison, this threshold is crossed for seafloor depths below 7000 m when applying the relationship between burial rate and water depth of Middelburg et al. (1997) and below 5200 m for the Burwicz et al. (2011) parameterisation. The bulk POC_{flux} is separated into the labile and refractory component and the routine to find the steady-state solution for POC is called. Here, the two POC depositional fluxes are first converted into SWI concentrations ($POC_i(z = 0)$, in $mol\,cm^{-3}$) by solving the flux divergence equation:

30 $$\frac{\partial F}{\partial z} = -\frac{\partial}{\partial z}\left(-\xi D_i \frac{\partial POC_i}{\partial z} + \xi w POC_i\right) \qquad (51)$$

for z=0. OMEN-SED then computes the fraction of POC preserved in the sediment (f_{POC}, see Eq. (5)) and subsequently calls the routines to find the steady-state solutions for the solute substances. Note, that in this initial coupling the calculated benthic uptake/return fluxes F_{C_i} of dissolved species C_i (compare Eq. (6)) are adjusted for the advective loss at the lower sediment

boundary ($\cancel{w \cdot C_i(z_\infty)} \underline{w \cdot C_i(z_{max})}$) to assure the conservation of mass in the coupled model:

$$F_{C_i} = \phi(0) \left(D_i \left. \frac{\partial C_i(z)}{\partial z} \right|_{z=0} - w \left[C_i(0) - C_i(z_{\cancel{\infty} \underline{max}}) \right] \right).$$ (52)

In case OMEN-SED computes unrealistic results for POC preservation (i.e. $f_{POC} < 0.0$ or $f_{POC} > 1.0$) we discard the results of OMEN-SED and all POC is remineralised at the ocean floor. For the modern ocean set-up, using the adjustments for w described above, this has not occurred and is just installed as a safety check. Finally, f_{POC} and the SWI-fluxes of solutes (F_{C_i}, in $\mathrm{mol\,cm^{-2}\,yr^{-1}}$) are returned to cGENIE. In case no POC is deposited on the seafloor (i.e. $POC_{flux} = 0$), OMEN-SED is not executed and f_{POC} and F_{C_i} for all i are set to zero. In order to reduce memory requirements, the sediment profiles (e.g. as shown in Fig. 7) are not calculated in the FORTRAN version of OMEN-SED, however, the boundary conditions are saved and sediment profiles for specific grid-cells, ocean basins and ocean transects can be plotted at the end of each experiment using the stand-alone MATLAB version of OMEN-SED.

4.2 Parameterising the OM degradation rate constants in a global model
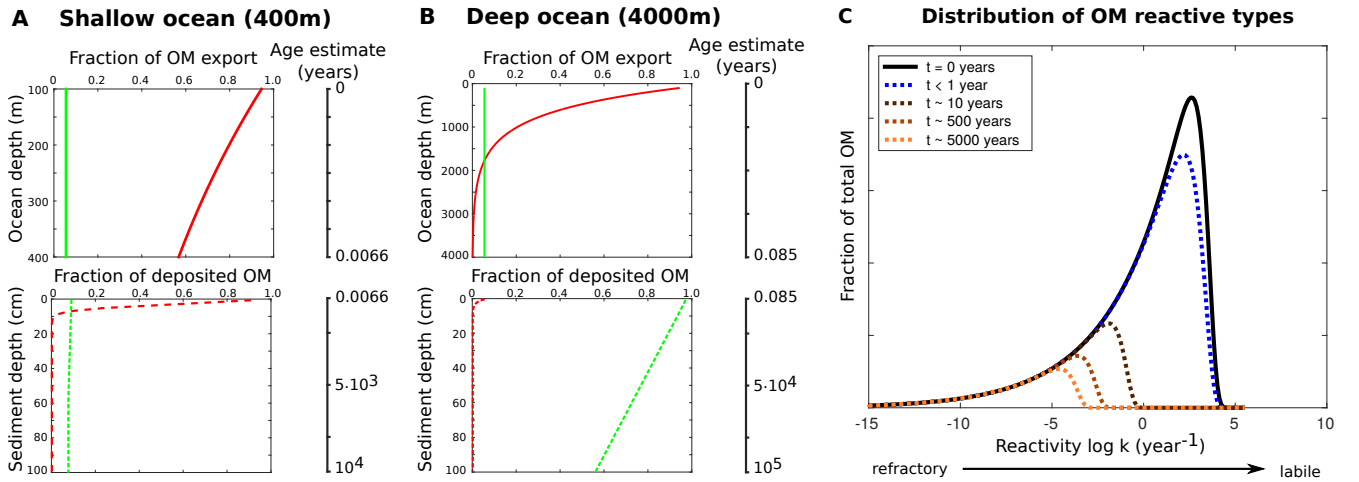
Figure 10. Idealised relationship of organic matter decomposition during remineralisation in the water column and the sediments. **A+B - Upper panels:** Water column development of the two organic matter fractions as represented in cGENIE for two ocean depths (red: labile OM with degradation length scale of 589m; green: refractory OM which is unreactive in the water column). The values are normalised to OM export at 100m. Age estimates for the OM since its export from the euphotic zone are calculated using a sinking velocity of 125m/day. **A+B - Lower panels:** Schematic representation of the development of the two OM fractions in the sediments (normalised to OM deposited on the seafloor). For the age estimates in the sediment column an advection rate of 0.01 and 0.001cm/yr is assumed, respectively. **C:** Idealised distribution functions of OM reactive types during remineralisation for different OM ages assuming a reactive continuum model for OM degradation. The initial distribution (at $t = 0$) represents fresh OM when it is exported from the euphotic zone (characterised by $a = 3e^{-4}$ $\mathrm{yr^{-1}}$ and $\nu = 0.125$ Boudreau et al., 2008).

As shown in our sensitivity analysis (Section 3.1) and discussed by Arndt et al. (2013), the degradation rate constants for OM (k_i) are the most influential parameters and exert a dominant control on the SWI-flux of redox-sensitive elements as well as the preservation of organic matter. Yet, their spatial variability is unknown at the global scale and reported rate constants in the sediments can vary by about 10 orders of magnitude or more (Middelburg et al., 1993; Arndt et al., 2013). Furthermore, when OMEN-SED is coupled to cGENIE, very different timescales have to be considered for OM degradation in the sediments compared to the water column (Fig. 10A+B) and thus the diagenetic rate constants cannot be easily implied by the assumed water column POC flux profiles in cGENIE. To illustrate this, lets consider the degradation of fresh, marine organic matter as it is transported and degraded along the ocean-sediment continuum. The bulk material is composed of a complex mixture of different organic carbon compounds that can be described by a reactivity continuum. Microbes preferentially degrade the more reactive organic matter compounds first (Emerson and Hedges, 1988; Wakeham et al., 1997; Lee et al., 2000), resulting in the preferential preservation of more unreactive compounds, rendering the remaining mixture less and less reactive with time. Thus depending on the age of OM (or depth in the water and sediment column) the reactivity distribution of its compounds changes significantly (Fig. 10C) and the multi-G (2G in this case) approximation of this continuum has to take this shift into account. Fig. 10 illustrates these changes in the original reactivity distribution within a ocean-sediment framework. The reactivity distribution $t < 1$ year represents the organic matter mixture after it settled through the water column (Fig. 10C). Only the most reactive OM compounds are remineralised. This explains why the POC flux in the ocean can be represented with a 1G or pseudo 2G degradation model. In the sediments, however, much longer timescales have to be considered and a wider range of more unreactive compounds are degraded. As a consequence, significant changes in the reactivity distribution already take place in the upper millimetres of the sediments ($t \sim 10$ years, Fig. 10C). Therefore, a broader range of OM reactive types must be represented by the degradation model to capture the reactivity spectrum of OM in surface sediments, explaining why at least a pseudo 3G model ~~(including two degradable and one refractory fraction Soetaert et al., 1996; Boudreau, 1997; Stolpovsky et al., 2015)~~ (including two degra required. To complicate the situation even further, different sediment depths can represent very different timescales. For instance, half a meter of sediment can be deposited within a year in a coastal setting, while it will represent thousands of years (if not more) of sedimentation in a deep ocean setting. Therefore, residence times and thus degradation rate timescales (or OM age) are mainly controlled by advection rates. For instance, assuming an advection rate of 0.01 cm/yr for the shallow ocean, OM at 5cm depth is has been degraded for approximately 500 years, whereas a deep ocean advection rate of 0.001 cm/year allowed for OM degradation of approximately 5000 years at the same depth. As a consequence, organic matter degradation in deep ocean sediments affects a much wider range of the reactivity continuum and our simple pseudo 3G approximation of the complex OM mixture needs to reflect this by allowing for different k and f values (Fig. 10C). Furthermore, by assuming steady-state in OMEN-SED we assume that deposition fluxes of OM are constant over the characteristic timescales of the reaction-transport processes.

Thus defining appropriate OM degradation rate constants is a major challenge and source of uncertainty for diagenetic models. The rate constants in models are either determined through profile fitting for a specific site or, for global applications, they are related to a single, readily available characteristic (or master variable) of the local environmental conditions. For instance, considerable effort has been expended to relate the apparent rate constant for oxic and anoxic OM degradation to sedimentation

rate (w) and various empirical relations have been proposed ~~(Toth and Lerman, 1977; Tromp et al., 1995; Boudreau, 1997; Stolpovsky et al.~~ ~~Nevertheless, these relationships are generally based and/or tested on limited data sets and their global applicability, especially~~ ~~under past or projected future environmental conditions is questionable (Arndt et al., 2013) .~~ (Toth and Lerman, 1977; Tromp et al., 1995; F Stolpovsky et al. (2015, 2017) suggested empirically derived approaches to constrain degradation rate constants in a 2G model

5   on a global scale. These approaches are derived from present-day observations and might help constrain parameters for present-day applications. However, the problem of constraining 2G degradation model parameters remains for largely different environmental conditions encountered in the past that could also prevail in the future (Arndt et al., 2013) . We hence test several alternative schemes in the coupled OMEN-cGENIE framework. Our objective is not to perform and discuss a detailed calibration of the coupled models as this is beyond the scope of this sediment model development paper. Rather we want to

10   showcase the feasibility of the model coupling, illustrate the range of results and thus information that can be generated with OMEN-SED and verify that model results capture the main observed global benthic biogeochemical features.

4.2.1   Methodology

In this section we compare modelled mean POC weight percentages (wt%) in the upper 5cm of the sediments (POC_{5cm}) to the global distribution pattern of POC content in surface sediments (< 5cm sediment depth) of Seiter et al. (2004) using

15   different parameterisations for the degradation rate constants k_1 and k_2. For our observational target we take the original POC distribution pattern in $1° \times 1°$ grid resolution (interpolated from > 5500 measurements, compare Seiter et al., 2004) and transform it onto the 72×72 SEDGEM grid (Figure 11). The regridding of the original POC distribution obviously affects the resolution of the data, especially for the continental margin, as some sites with higher POC wt% are lost in the regridding process (compare e.g. maximum values for the East Pacific and upwelling waters of the Namibian shelf, Figure 11A + B).

20   The colour of the points in Figures ~~??~~ 12 - 13 indicates the seafloor depth (SFD) of the respective cGENIE grid-cell. As the individual data-points are highly scattered and in order to see if a certain relation between k_1 and k_2 performs better for specific ocean depths, the data-points are binned into 6 uniform depth-classes of 1000m each (respective mean POC wt% and SFD are represented by the triangles). The regression line ~~(and the corresponding R^2-value)~~ is calculated for the 6 bin-classes and included in the figures.

25   To parameterise the reactivity of organic matter in OMEN-SED two different schemes are tested and compared. First, spatially uniform degradation rate constants k_1 and k_2 are assumed. By simulating two different pools of POC in the water-column characterised by different degradation length scales (Ridgwell et al., 2007), cGENIE implicitly accounts for the decrease in mean POC reactivity with water-depth. The rate constants for the more refractory OM pool, k_2, is systematically varied ~~between~~ ~~0.004 and 0.006 year^{-1}~~ and the more labile OM component, described by k_1, is assumed to degrade a multiple times faster~~(i.e.~~

30   ~~$x \in \{1.1, 1.2, 1.3, 1.5, 2\}$). However, although~~ . Values for these parameters are selected in accordance to the best fit to the global surface POC observations (Seiter et al., 2004) . Although accounting for the decrease in mean POC reactivity with seafloor depth, this approach does not take into account the change in distribution of organic matter reactivity types caused by different burial velocities and thus different residence time scales in the sediments (Fig. 10). Therefore, the second approach uses the empirical relationship proposed by Boudreau (1997), which relates the apparent OM degradation rate constant in the
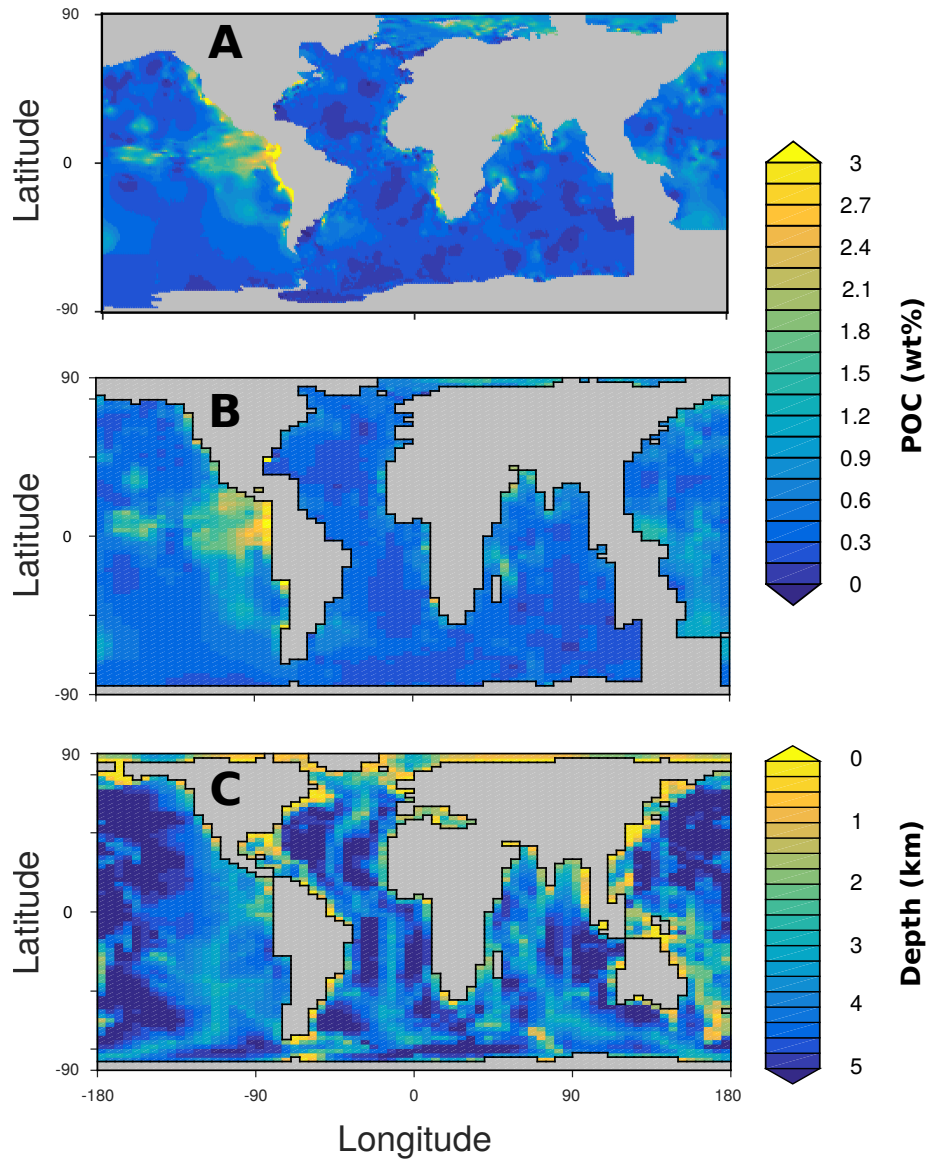
Figure 11. Observed distribution of sediment surface (< 5cm) POC wt% (A, B) and cGENIE bathymetry (C). (A) Original global distribution of POC wt% interpolated on a $1° \times 1°$ grid from more than 5500 individual data points (compare Seiter et al., 2004, for the interpolation procedure). (B) Observed POC wt% data transformed onto the 72×72 SEDGEM grid. Grid points without any observations are left blank (grey). (C) Gridded continental configuration and ocean bathymetry of the 16-level, 72×72 equal-area cGENIE grid.

upper sediments to the burial velocity, w (cm year^{-1}, see also Section 3.3):

$$k_{\text{app}} = 0.38 \cdot w^{0.59}. \tag{53}$$

Following Boudreau (1997) and Stolpovsky et al. (2015) it can be assumed that k_{app} represents the mean OM reactivity within the upper 10-20cm of the sediments. The following assumptions are made in order to calculate the two degradation rate

5   constants for OMEN-SED:

$$k_{\text{app}} = f_1 \cdot k_1 + (1 - f_1) \cdot k_2 \tag{54}$$

$$k_1 = x \cdot k_2 \tag{55}$$

where x describes the relation between k_1 and k_2 and is subject to sensitivity experiments (with values of ~~$x \in \{2, 5, 8, 10, 12, 15, 20, 25\}$~~ $x \in$:

$20, 25\}$). Note that the differences between k_1 and k_2 using this approach is significantly larger as in the globally uniform ap-

10   proach. As the fractions of labile and refractory OM reaching the sediments f_1 is known from cGENIE, k_1 and k_2 can be calculated independently for each grid-cell.

     To simulate steady state sediment composition we configure the model as a "closed" system, i.e., one in which there is no loss of $CaCO_3$ through burial. The redox dependent P-cycle in OMEN-SED is not used in these experiments and all organic phosphorus is returned at the seafloor. To speed up the calculation and to assure that ocean redox changes caused by OMEN-

15   SED do not impact the sediment composition of $CaCO_3$, we use the prescribed solid fields as described earlier. Apart from the prescribed fields and the 72×72 sediment grid the model is configured as in Archer et al. (2009) and atmospheric CO_2 is restored to a pre-industrial value of 278 ppmv. First a 20,000 year spinup is performed without OMEN-SED being coupled. All presented coupled cGENIE-OMEN simulations are run for 10,000 years to steady state from this spinup. OMEN-SED is called for each grid-cell in every time step, feeding back the resulting SWI-fluxes and the fraction of POC preserved in the

20   sediments to cGENIE.

4.2.2   Results

~~Crossplots comparing modelled and observed mean POC wt$$ in the upper 5 cm of the sediments using spatially uniform degradation rate constants k_1 and k_2. Data-points are binned into 6 uniform depth-classes of 1000m as in Fig. 12, each class is represented by a triangle. Grid-points with more than 4.0 POC wt$$ are not shown.Figure ?? presents results for the spatially~~

25   ~~uniform degradation rate experiments. In general, using spatially uniform degradation rate constants 5 of the 6 bin-classes are located closer to the 1:1 line as in the experiments using the Boudreau (1997) relation (Fig. 12). Also the slope of some regression lines is close to 1.0 (e.g. $(k_2, x) \in \{(0.004, 1.5), (0.0045, 1.3), (0.005, 1.2), (0.005, 1.3), (0.0055, 1.1), (0.0055, 1.2), (0.006, 1.1)$ indicating that the simpler parameterisation adequately captures the relationship between between depth and observed POC wt$$ by bin-class. The reason for this is that BIOGEM provides a depth dependent POC flux and partitioning between the~~

30   ~~two fractions (Fig. 10). The shallowest bin-class (between 0 and 1000m) represents an exception, as OMEN-SED tends to overestimate POC preservation for this depth class. However, this could also be related to the regridding of the original POC distribution pattern of (Seiter et al., 2004) on to the SEDGEM grid, as some data grid-cells with higher POC wt$$ on the~~
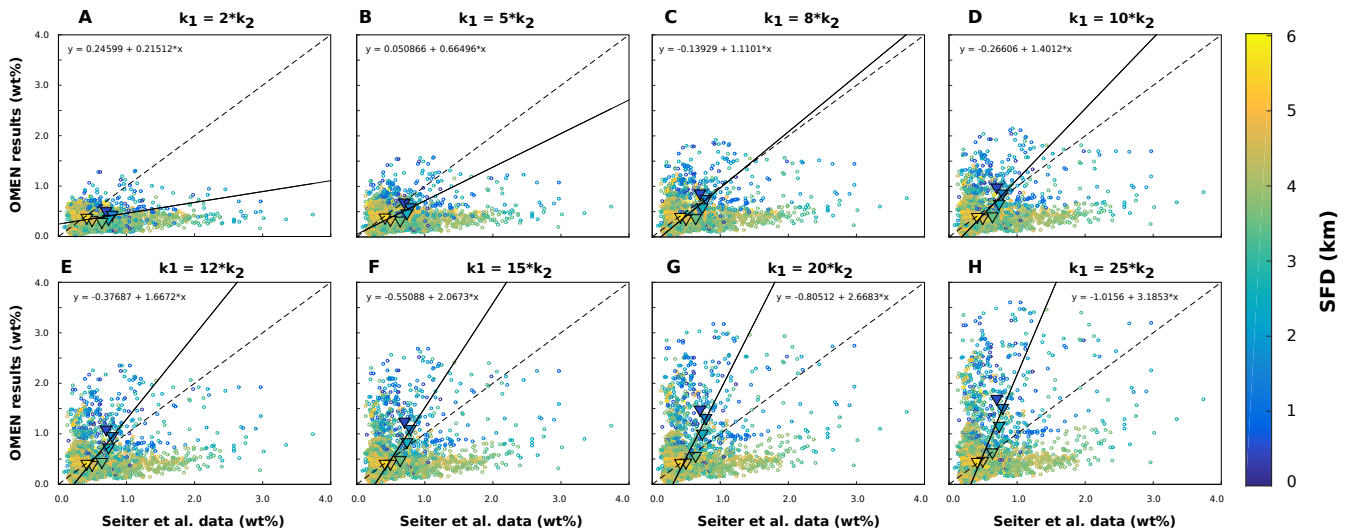
Figure 12. Crossplots comparing modelled and observed mean POC wt% in the upper 5 cm of the sediments using the relationship of Boudreau (1997) and the assumptions of Eq. (54) and (55) to calculate k_1 and k_2. Data-points are binned into 6 uniform depth-classes of 1000m, each class is represented by a triangle. Grid-points with more than 4.0 POC wt% are not shown.

~~continental margin are lost due to the restricted SEDGEM resolution (compare Section 4.2). Overall, using this parameterisation, a relationship where the labile POC fraction degrades not more than 1.5 times faster than the refractory fraction fits the Seiter et al. (2004) data better than a larger spread between both POC pools (i.e. $x > 2.0$). We discuss later the implications of collapsing the k values in this specific calibration for long-term, geological carbon preservation.~~

5   ~~Next~~ Figure 12 presents results for the relationship of Boudreau (1997) ~~and~~ using the assumptions of Eq. (54) and (55) ~~are used~~ to calculate k_1 and k_2. ~~In Figure 12 (A-H)~~ Here, the relation between the two degradation rate constants (Eq. (55)) is changed globally, thus independent of the seafloor depth. The crossplots show that it is not possible to achieve a solution where all bin-classes fall onto, or close to, the 1:1 line. Also, the slope of the regression lines are generally much larger or smaller than 1.0 (with the exception of Figure 12C), indicating that the relationship between depth and observed POC wt% by bin-class

10   is not adequately represented by the model. ~~The R^2 values are strictly monotonically increasing for increasing x because a depth-dependency is artificially imposed for the modelled POC wtthrough the relation between k_1 and k_2.~~ When looking at the individual bin-classes it can be seen that shallow ocean depths are better represented by smaller differences between k_1 and k_2 (e.g. $k_1 = 5 \cdot k_2$ for SFD < 1000m, Figure 12B), and the deep ocean by a larger spread (e.g. $k_1 = 25 \cdot k_2$ for SFD > 3000m, Figure 12H). These results reflect the preferential degradation of more reactive organic matter types (Wakeham et al., 1997;

15   Lee et al., 2000) and thus the change in the distribution functions of OM reactive types for different OM ages (Fig. 10C). In the shallow ocean bulk POC consists of fresher organic matter types on average and is therefore generally more reactive overall (i.e. higher k_{app} due to higher w in the model) as in the deep ocean. In addition, OM at 5cm sediment depth in the deep ocean

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is generally older as in the shallow ocean due to lower burial rates, therefore more reactive types are affected by degradation and a larger spread between k-values is needed to capture these dynamics (compare Fig. 10C).

Departing from our theoretical considerations (see discussion of Fig. 10C), we use these observations to create a depth dependent relationship between the two degradation rate constants, where x in Eq. (55) is a function of SFD and takes values of $x = 5$ for SFD < 1000m, $x = 8$ for 1000m $\leq$ SFD < 2000m, $x = 12$ for 2000m $\leq$ SFD < 3000m and $x = 25$ for SFD ≥ 3000m for the 6 SFD bin-classes, respectively. ~~In~~ Figure 13 compares this depth dependent approach with the best model using spatially uniform degradation rate constants ($k_2 = 0.005$, $k_1 = 1.3 \cdot k_2$). In the depth dependent approach all bin-classes are close to the 1:1 line and ~~the resulting regression model accounts for 92.6of the variance of the modelled~~ the slope of the regression line (0.9662) indicates that the relationship between depth and observed POC wt% ~~around the observed mean of~~ for the bin-classes ~~(Figure 13B). Furthermore, the slope of the regression line (0.9662)indicates that~~ is well represented by the model (Fig. 13A). Using spatially uniform degradation rate constants 5 of the 6 bin-classes are located close to the 1:1 line (Fig. 13B), indicating that also the simpler parameterisation adequately captures the relationship between between depth and observed POC wt% ~~for the bin-classes is well represented by the model~~by bin-class. The reason for this is that BIOGEM provides a depth dependent POC flux and partitioning between the two fractions (Fig. 10). The shallowest bin-class (between 0 and 1000m) represents an exception, as OMEN-SED tends to overestimate POC preservation for this depth class. However, this could also be related to the regridding of the original POC distribution pattern of (Seiter et al., 2004) on to the SEDGEM grid, as some data grid-cells with higher POC wt% on the continental margin are lost due to the restricted SEDGEM resolution (compare Section 4.2). The histograms (Fig. 13C+D) visualize the difference between modelled and observed mean POC concentrations and demonstrate the high density of data points close to the 1:1 line. For the depth dependent approach, 92.5% of the cGENIE grid-cells show a difference between modelled and observed POC concentration of less than 1.0 POC wt%; in 79.9% of the grid-cells, the difference is less than 0.5 POC wt% (for the globally uniform approach the percentages are 95.37% and 70.95%, respectively).

Both experiments reproduce minimal POC concentrations in the subtropical gyres and generally higher concentrations along the continental margins (Fig. 13E+F). Both experiments, however, underestimate mean POC wt% in the surface sediments of the equatorial east Pacific and overestimate POC concentrations in the North Pacific and Southern Oceans (Fig. 13G+H). The depth dependent approach of Boudreau (1997) shows more spatial variability in POC preservation than the other parameter-isation. In general, implementing lower, anaerobic degradation rate constants when bottom water oxygen concentrations fall below a threshold value could potentially improve the simulation of higher POC concentrations in areas with high POC input to the sediments (Palastanga et al., 2011).

4.3 Modelled fluxes and sediment characteristics

For the depth dependent Boudreau (1997) approach modelled SWI-fluxes and sediment characteristics are shown in Figure 14. Modelled total POC degradation (POC_{degr}) rates in the upper sediments decrease from the shelves to the deep sea by up to 2 orders of magnitude (Fig. 14B). This is in agreement with data from the literature (e.g. Middelburg et al., 1993, 1997; Burdige, 2007) and other model results (e.g. Thullner et al., 2009) which indicate that the highest degradation rates in marine
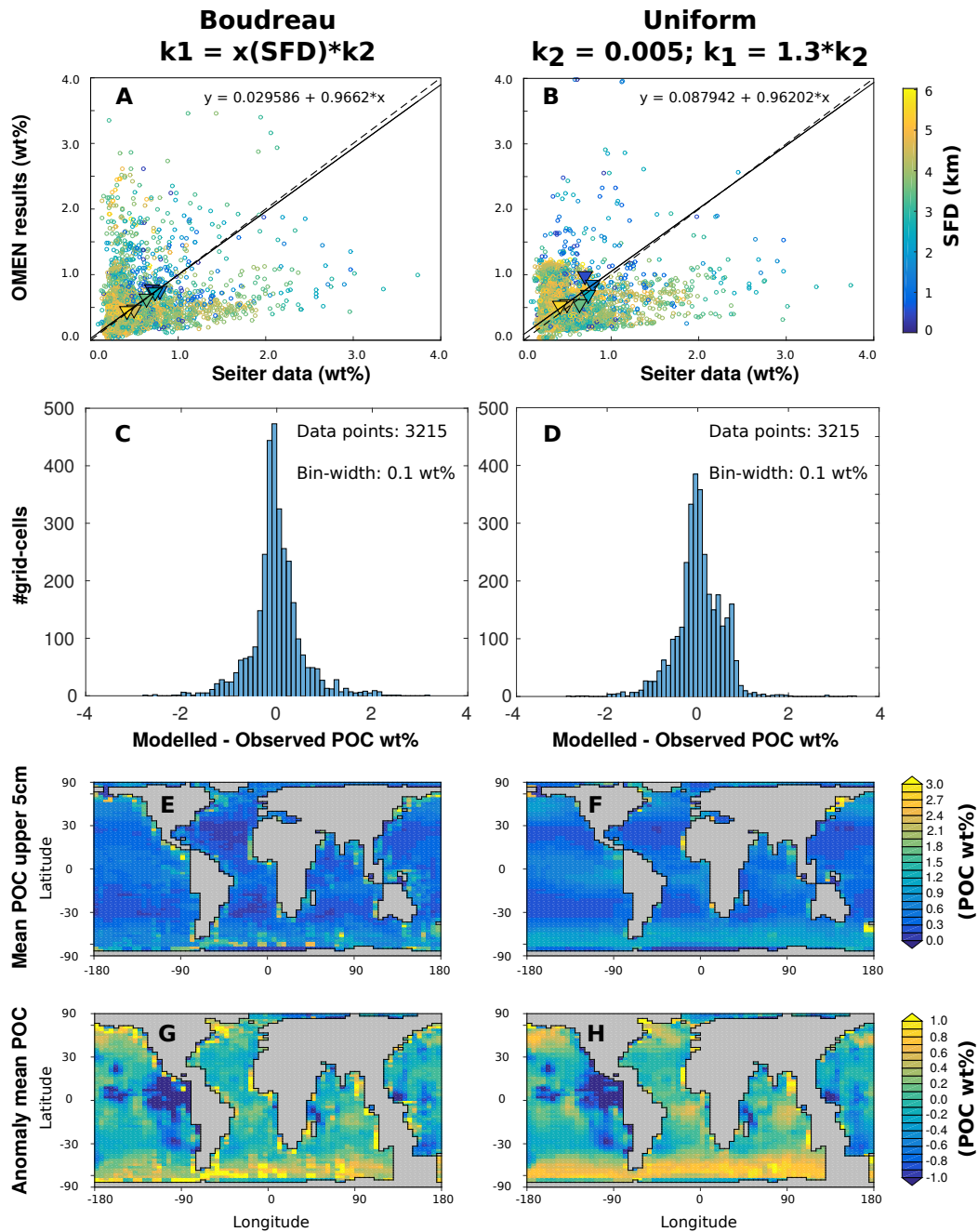
Figure 13. Mean POC concentrations in the upper 5cm of the sediments (POC_{5cm}) using the depth dependent parameterisation $k_1 = x(SFD) \cdot k_2$ adapted from Boudreau (1997, left) and the globally uniform model ($k_2 = 0.005$, $k_1 = 1.3 \cdot k_2$, right)~~and the depth dependent parameterisation $k_1 = x(SFD) \cdot k_2$ adapted from Boudreau (1997)~~. A+B: Crossplots as shown before in Fig. ~~??~~ and 12. C+D: Histograms of the residuals of modelled minus observed POC_{5cm}. E+F: POC_{5cm} as calculated with OMEN-SED. G+H: Difference map of POC_{5cm} as calculated with OMEN-SED and interpolated data from Seiter et al. (2004).

Boudreau $k_1 = x(\text{SFD}) * k_2$
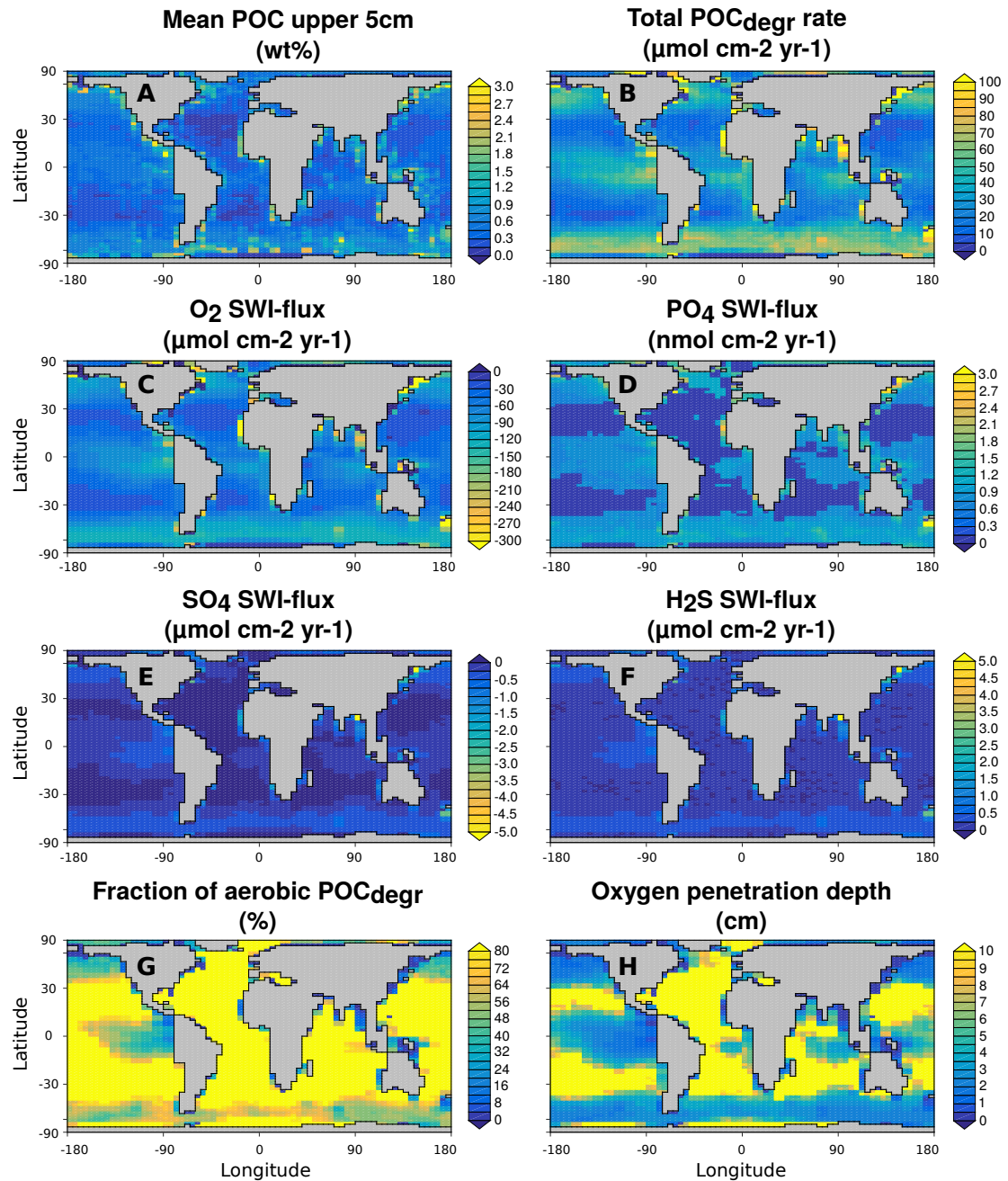
Figure 14. Sediment characteristics related to POC degradation and oxygen consumption for the depth dependent paramaterisation after Boudreau (1997) with $k_1 = x(\text{SFD}) \cdot k_2$. Total POC_{degr} rate and fraction of aerobic POC_{degr} are the respective values for the first 5cm in the sediments.

sediments are found in the coastal ocean (SFD < 200 m). Oxygen fluxes into the sediments (Fig. 14C) range from 0.0 for the deep ocean and sites without OM deposition to values of about 300 μmol cm^{-2}yr^{-1} for the shallow ocean with the highest POC degradation rates. Influx of SO$_4$ into the sediments is rather low (between 0.0 and 23.9 μmol cm^{-2}yr^{-1}) because in OMEN-SED 95% of produced H$_2$S is reoxidised to SO$_4$, therefore sulfate reduction is mainly driven by in situ sulfide oxidation.

5 However, in general the coupled model fluxes fall well within the ranges predicted by the stand-alone global hypsometry experiments (O$_2$ between 0.0 and 800 μmol cm^{-2}yr^{-1} and SO$_4$ between 0.0 and about 300 μmol cm^{-2}yr^{-1}, compare Section 3.3). In accordance with the total POC degradation rates the release of PO$_4$ shows a maximum value of 8.12 nmol cm^{-2}yr^{-1} on the shelves (Fig. 14D). The relative contribution of aerobic POC degradation in the upper sediments increases from the shelves to the deep sea (Fig. 14G) which is also consistent with estimates from Thullner et al. (2009) who found that oxygen is

10 responsible for less than 10% of POC$_{degr}$ at 100 m SFD and for more than 80% in the deep sea. The oxygen penetration depth in OMEN-SED increases from values below 1cm at the shelves to more than 10cm in the deep ocean (Fig. 14H and 15). Small oxygen penetration depths of a few millimetres are typical for bioturbated sediments in the coastal ocean (e.g. Wenzhöfer and Glud, 2002) and the oxygen penetration depth has been shown to increase rapidly with SFD to more than 10 cm in the deep sea (Meile and Van Cappellen, 2003; Glud, 2008). Fischer et al. (2009) and D'Hondt et al. (2015) even found cores along a transect

15 in the South Pacific gyre being oxygenated over their entire length (up to 8 m or even 75 m, respectively) which is consistent with our model results (not shown). Simulated mean oxygen penetration depths for the 6 depth bin-classes also agree well with observations compiled by Glud (2008) and Meile and Van Cappellen (2003, Fig. 15).

5 Scope of applicability and model limitations

Because of the high computational cost associated with resolving benthic dynamics, most Earth system Models of Intermediate

20 Complexity (EMICs) and also some of the higher resolution Earth system models either completely neglect or merely include a highly simplified representation of benthic-pelagic exchange processes (Hülse et al., 2017). However, benthic-pelagic coupling plays an important role for carbon cycling and the lack of its representation in EMICs compromises our ability to assess the response and recovery of the Earth system to major past, present and future carbon cycle and climate perturbations. As a consequence, there is a need for benthic biogeochemical models that are able to capture the main features of benthic biogeochemical

25 dynamics, but that are, at the same time, computationally efficient enough to allow for a direct, dynamic coupling to ocean biogeochemical model. Therefore, we have developed OMEN-SED, a, one-dimensional analytical early diagenetic model that offers a predictive ability similar to complex, numerical diagenetic models at a significantly reduced computational cost.

OMEN-SED is thus not problem-specific. Its reaction network resolves the most pertinent benthic biogeochemical species as well as the most important processes that control their cycling and burial in marine sediments. OMEN-SED can thus be coupled

30 to a wide range of regional to global ocean biogeochemical models, as well as EMICs and higher resolution Earth system models to investigate a wide range of research questions associated with past, present or future carbon and macro-nutrient cycling. For instance, OMEN-SED can be used to i) quantify benthic macro-nutrient recycling from the shallow coastal to the deep, open ocean, ii) investigate the role of benthic-pelagic coupling in the development of past or future ocean anoxia/euxinia,
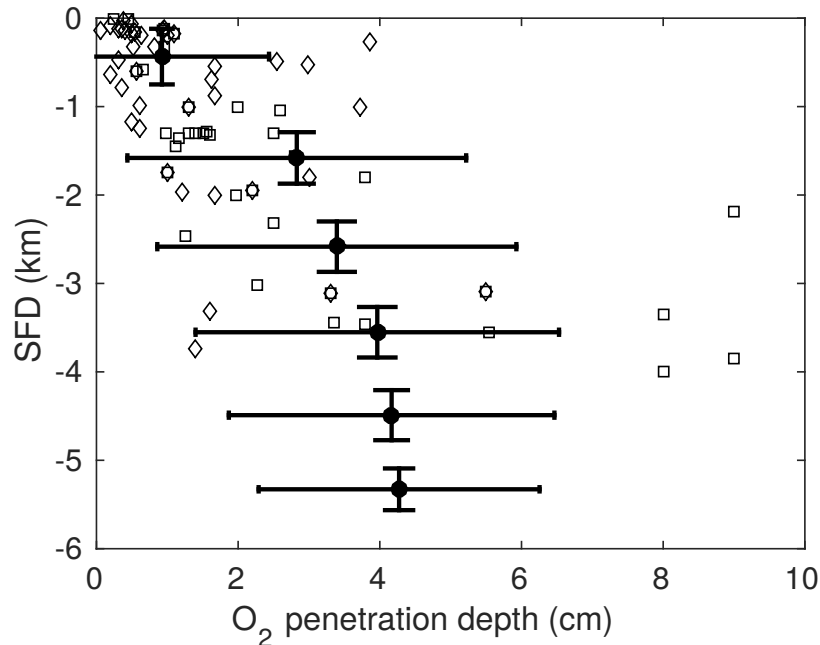
Figure 15. Seafloor depth versus O_2 penetration depth for the depth dependent paramaterisation after Boudreau (1997) with $k_1 = x(\text{SFD}) \cdot k_2$. Diamonds represent observations compiled by Meile and Van Cappellen (2003) and squares observations from Glud (2008). Circles are the mean model results for the 6 SFD bin-classes (with standard deviations). Grid-cells where the entire sediment column is oxic (i.e. $z_{\text{ox}} = 100\,\text{cm}$) are not considered in these statistics (17, 32, 102, 300, 477 and 307 cells for the 6 bin-classes, respectively).

or to iii) estimate global organic carbon burial in marine sediments. In theory, its scope of applicability thus ranges from the regional to the global and from the seasonal to the millennial time-scale. In order to simulate organic matter preservation in the deeper sediments and thus addressing questions concerning long-term, geological carbon burial the degradation rate constant for the refractory OM pool has to be scaled down. The resulting larger difference between degradation rate constants can be

5   interpreted as being needed to capture the broader range of OM reactivities degraded over the entire sediment column (see Fig. 10). Instead, more collapsed degradation rate constants are needed to model OM degradation in the upper sediments, such as the first 5 centimetres as shown in Section 4.2.2. In addition, the computational efficiency of OMEN-SED allows calculating quantitative sensitivity indices requiring large sample sizes such as variance- or density-based approaches. Therefore, OMEN-SED can also help quantitatively investigate sensitivity of benthic model output to systematic variations in model parameters

10   when the model has been tuned to a site-specific problem.

However, OMEN-SED is inevitably associated with a certain degree of simplifications that may compromise the applicability of the model in its current version under certain circumstances. First, we have assumed steady state conditions to allow for an analytical solution of the coupled diagenetic equations. This steady-state assumption is only valid if the variability in boundary conditions and fluxes is generally longer than the characteristic timescales of the reaction-transport processes. As a conse-

quence, OMEN-SED is well suited for the coupling to EMICs and the investigation of long-term dynamics in sediment-water exchange fluxes, for instance, during past extreme climate events. Yet, in its current version, OMEN-SED is not able to predict the transient response of benthic process rates and fluxes to short-term or seasonal variations of boundary conditions. Yet, future versions of OMEN-SED, could approximate non-steady state conditions by incorporating a time-step dependent relaxation be-

5   tween different steady states, similar to the schemes used in Ruardij and Van Raaphorst (1995) and Arndt and Regnier (2007). Such a pseudo-transient approach would enable the application of OMEN-SED to systems characterised by high-frequency fluctuations in boundary conditions, such as the coastal ocean or estuaries. Furthermore, by their very nature, analytical models do not allow for overlapping biogeochemical zones or depth dependent porosity, which introduces a certain error to simulation results. However, the energy yield dependent sequence of oxidants is generally valid (e.g. Hensen et al., 2006) and the good

10   agreement between OMEN-SED and the results obtained with a fully formulated numerical RTM (allowing for overlapping TEA use and depth dependent porosity, Section 3.3) shows that these are not critical limitations of OMEN-SED - even for shallow sediments.

      ~~Second, although~~ Although the model explicitly simulates DIC and alkalinity production and, thus, has the potential to predict pH profiles within the sediment, a major limitation at this stage is the lack of an explicit description for $CaCO_3$

15   precipitation/dissolution coupled to OM decomposition, which also controls the inorganic carbon system (Krumins et al., 2013). In addition, the current version of OMEN-SED does not yet explicitly resolve iron and manganese dynamics (although note, that iron is implicitly accounted for in the PO4 equation). This lack currently limits the applicability of OMEN-SED to iron- and manganese rich environments, such as coastal marine environments, upwelling regions or ferruginous oceans. In addition, it also compromises the ability of OMEN-SED in predicting H_2S fluxes in Fe-rich anoxic environments, where high

20   iron pore water concentrations can deplete pore water H_2S by iron-sulfide mineral precipitation (e.g. Meyers, 2007). Therefore, already planned future extensions of OMEN-SED include an explicit description of ~~iron.~~ carbonate dissolution and iron. Also note that our 1-D diffusion/bioturbation model might not be appropriate to simulate non-accumulating permeable sands of the coastal ocean.

      Finally, just as all global models, the global application of OMEN-SED is complicated by the lack of an objective, global

25   framework for biogeochemical process parameterisation. The sensitivity study presented here shows that this lack is particularly critical for OM degradation rate parameters (k_i, f_i) and the γ-values describing the completeness of secondary redox reactions. A comparison between simulated OM contents and observations indicates that a depth dependent k-f ~~relationships provide~~ relationship provides the best fit (Section 4.2.2)~~, confirming more theoretical considerations regarding the different time and reactivity scales that need to be considered (see Section 4.2~~. These results confirm that reducing the continuous distribution

30   of organic matter reactivities into two distinct reactivity classes (2G Model) requires different k-f values for shallow vs deep ocean sediments because of the largely different reaction timescales involved (also see Fig. 10). With respect to γ-values, model simulations along the global hypsometry (Section 3.3) have shown that high γ-values generally capture the main SWI-flux features, but have also highlighted that slightly lower γ-values would result in a better fit of SWI-fluxes to observations of the shallow ocean.

6 Conclusions

Here, we have described in detail and tested OMEN-SED, a new, analytical early diagenetic model resolving organic matter cycling and associated biogeochemical dynamics. OMEN-SED has been explicitly designed for the coupling to EMICs and combines biogeochemical complexity with computational efficiency. It is the first analytical diagenetic model to explicitly rep-
5 resent oxic degradation, denitrification, sulfate reduction and implicitly methanogenesis, as well as the reoxidation of reduced substances, adsorption/desorption, as well as mineral precipitation/dissolution. Explicitly resolved pore water species include O_2, NO_3, NH_4, SO_4, H_2S, DIC and ALK and the solid phase includes two degradable fractions of organic matter, Fe-bound P and authigenic Ca-P minerals.

An extensive sensitive analysis, based on the density-based PAWN method (Pianosi and Wagener, 2015) emphasizes the
10 importance of OM degradation rate parameters (k_i, f_i) and thus highlights the need for the development of an objective, global framework to parameterize OM degradation rate parameters. We have shown that the performance of OMEN-SED at the system scale is similar to that of a fully formulated, multi-component numerical model. The new analytical model is able to reproduce observed pore water profiles across a wide range of depositional environments and captures observed global patterns of SWI-fluxes, oxygen penetration depths, biogeochemical reaction rates, as well as surface sediment organic matter contents.
15 Coupled to EMICs or higher resolution Earth system models, OMEN-SED is thus well suited to examine the role of sediments in global biogeochemical cycles in response to a wide range of past or future carbon cycle or climate perturbations over various timescales.

Competing interests. The authors declare that they have no conflict of interest.

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Appendix A: Reaction Network

Table A1. Primary pathways of organic matter degradation, secondary redox reactions and stoichiometries implemented in the reaction network.

Pathway	Stoichiometry
	Primary Redox reactions
Aerobic degradation	$(CH_2O)_x(NH_3)_y(H_3PO_4)_z + (x+2y)O_2 + (y+2z)HCO_3^- \rightarrow (x+y+2z)CO_2 + yNO_3^- + zHPO_4^{2-} + (x+2y+2z)H_2O$
Denitrification	$(CH_2O)_x(NH_3)_y(H_3PO_4)_z + \frac{(4x+3y)}{5}NO_3^- \rightarrow \frac{(2x+4y)}{5}N_2 + \frac{(x-3y+10z)}{5}CO_2 + \frac{(4x+3y-10z)}{5}HCO_3^- + zHPO_4^{2-} + \frac{(3x+6y+10z)}{5}H_2O$
Sulfate reduction	$(CH_2O)_x(NH_3)_y(H_3PO_4)_z + \frac{x}{2}SO_4^{2-} \rightarrow (y-2z)CO_2 + \frac{x}{2}H_2S + (x+y-2z)HCO_3^- + yNH_4^+ + zHPO_4^{2-}$
Methanogenesis	$(CH_2O)_x(NH_3)_y(H_3PO_4)_z + (y-2z)H_2O \rightarrow \frac{x}{2}CH_4 + \frac{x-2y+4z}{2}CO_2 + (x-2z)HCO_3^- + yNH_4^+ + zHPO_4^{2-}$
	Secondary Redox reactions
Nitrification	$NH_4^+ + 2O_2 + 2HCO_3^- \rightarrow NO_3^- + 2CO_2 + 3H_2O$
Sulfide oxidation	$H_2S + 2O_2 + 2HCO_3^- \rightarrow SO_4^{2-} + 2CO_2 + 2H_2O$
AOM	$CH_4 + CO_2 + SO_4^{2-} \rightarrow 2HCO_3^- + H_2S$
	Adsorption reactions and mineral precipitation
NH_4 adsorption	$NH_4^+ \xrightarrow{K_{NH_4}} NH_4^+ (ads)$
P ad-/desorption	$PO_4^{2-} \xrightarrow{K_{PO_4}^{I,II}} PO_4^{2-}(ads);$ $\qquad PO_4^{2-} \xrightarrow{k_s} Fe-bound\ P \xrightarrow{k_m} PO_4^{2-}$
CFA precipitation	$PO_4^{2-} \xrightarrow{k_a} CFA$
pyrite precipitation	$Fe^{2+} + HS^- \rightarrow FeS + H^+, FeS + H_2S \rightarrow H_2 + FeS_2$

Appendix B: Sensitivity Analysis
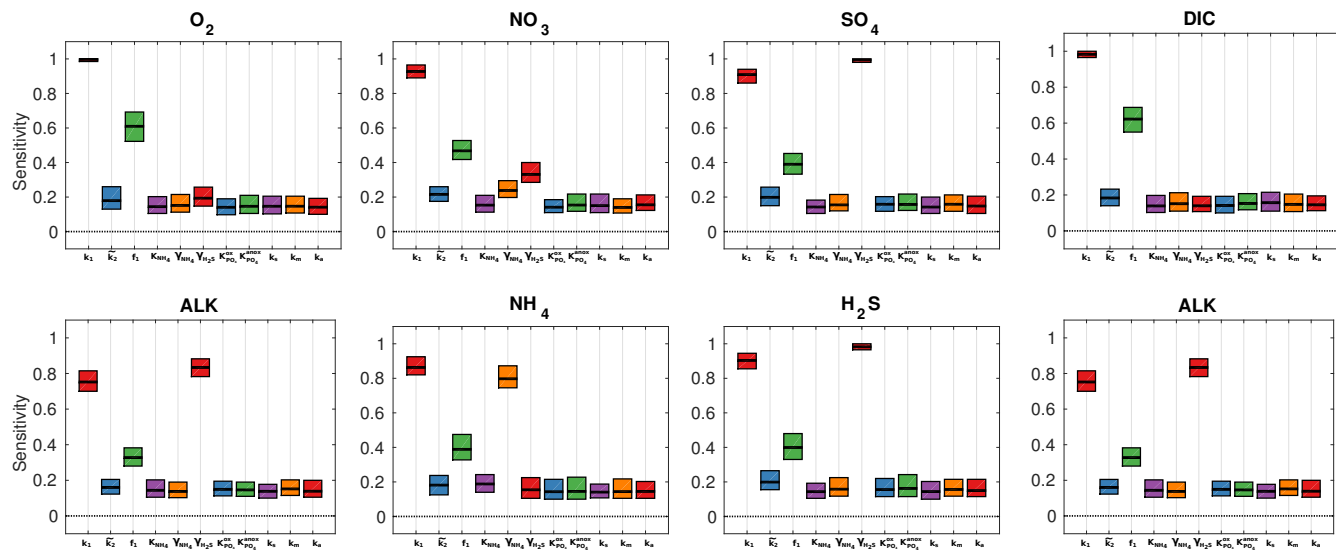
Figure B1. Box plot of parameter sensitivities for the calculated SWI-fluxes for the 4000m oxic condition. Average sensitivities (black lines) and 90% confidence intervals using $N = 11200$ model evaluations and $Nboot = 100$ bootstrap resamples.

Appendix C: Prescribed burial flux fields
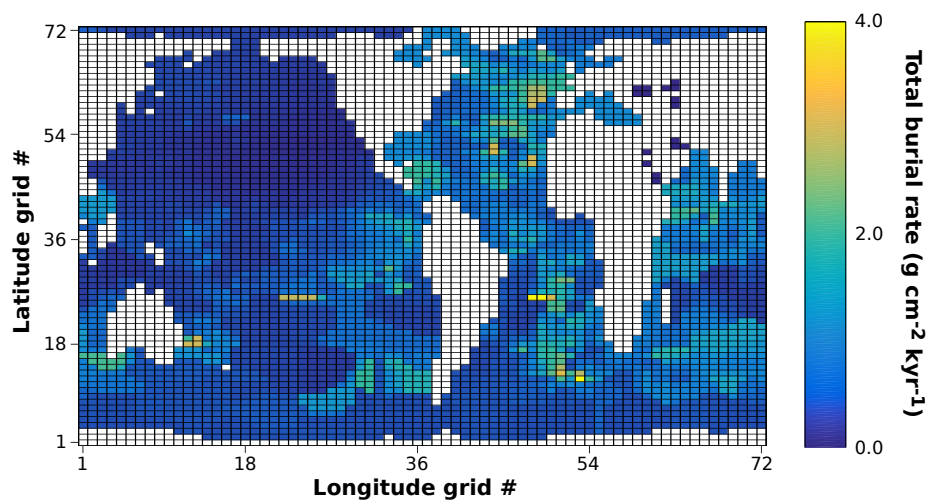
Figure C1. Distribution of prescribed total burial fluxes of detrital material, opal and $CaCO_3$ (in g cm^{-2} kyr^{-1}), re-gridded from the data compilation of Archer (1996) using a method explained in the text. Note, latitude and longitude are shown in cGENIE grid-cells and not in degrees.

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