- A sub-canopy structure for simulating oil palm in the Community Land Model (CLM-Palm):
 phenology, allocation and yield
- Yuanchao Fan^{1,2,*}, Olivier Roupsard^{3,4}, Martial Bernoux⁵, Guerric Le Maire³, Oleg Panferov⁶,
 Martyna M. Kotowska⁷, Alexander Knohl¹
- ⁵ ¹ University of Göttingen, Department of Bioclimatology, Büsgenweg 2, 37077 Göttingen,
- 6 Germany
- ² AgroParisTech, SIBAGHE (Systèmes int égr és en Biologie, Agronomie, G éosciences,
- 8 Hydrosciences et Environnement), 34093 Montpellier, France
- ³ CIRAD, UMR Eco&Sols (Ecologie Fonctionnelle & Biog éochimie des Sols et des Agro écosystèmes), 34060 Montpellier, France
- ⁴ CATIE (Tropical Agricultural Centre for Research and Higher Education), 7170 Turrialba,
 Costa Rica
- ⁵ IRD, UMR Eco&Sols, 34060 Montpellier, France
- ⁶ University of Applied Sciences Bingen, 55411 Bingen am Rhein, Germany
- ⁷ University of Gätingen, Department of Plant Ecology and Ecosystems Research, Untere
- 16 Karspüle 2, 37073 Gätingen, Germany
- 17 *Correspondence author. E-mail: yfan1@uni-goettingen.de
- 18

19 Abstract: In order to quantify the effects of forests to oil palm conversion occurring in the 20 tropics on land-atmosphere carbon, water and energy fluxes, we develop a new perennial crop 21 sub-model CLM-Palm for simulating a palm plant functional type (PFT) within the 22 framework of the Community Land Model (CLM4.5). CLM-Palm is tested here on oil palm 23 only but is meant of generic interest for other palm crops (e.g. coconut). The oil palm has monopodial morphology and sequential phenology of around 40 stacked phytomers, each 24 25 carrying a large leaf and a fruit bunch, forming a multilayer canopy. A sub-canopy 26 phenological and physiological parameterization is thus introduced, so that each phytomer has 27 its own prognostic leaf growth and fruit yield capacity but with shared stem and root 28 components. Phenology and carbon and nitrogen allocation operate on the different 29 phytomers in parallel but at unsynchronized steps, separated by a thermal period. An 30 important phenological phase is identified for the oil palm - the storage growth period of bud 31 and "spear" leaves which are photosynthetically inactive before expansion. Agricultural 32 practices such as transplanting, fertilization, and leaf pruning are represented. Parameters 33 introduced for the oil palm were calibrated and validated with field measurements of leaf area 34 index (LAI), yield and net primary production (NPP) from Sumatra, Indonesia. In calibration with a mature oil palm plantation, the cumulative yields from 2005 to 2014 matched notably 35 36 well between simulation and observation (mean percentage error = 3%). Simulated inter-37 annual dynamics of PFT-level and phytomer-level LAI were both within the range of field 38 measurements. Validation from eight independent oil palm sites shows the ability of the 39 model to adequately predict the average leaf growth and fruit yield across sites and 40 sufficiently represent the significant nitrogen and age related site-to-site variability in NPP 41 and yield. Results also indicate that seasonal dynamics of yield and remaining small-scale 42 site-to-site variability of NPP are driven by processes not yet implemented in the model or 43 reflected in the input data. The new sub-canopy structure and phenology and allocation 44 functions in CLM-Palm allow exploring the effects of tropical land use change, from natural ecosystems to oil palm plantations, on carbon, water and energy cycles and regional climate. 45

46 **1. Introduction**

47 Land-use changes in Southeast Asia have been accelerated by economy-driven expansion of 48 oil palm (Elaeis guineensis) agriculture since the 1990s (Miettinen et al., 2011). Oil palm is 49 currently one of the most rapidly expanding and high-yielding crops in the world (Carrasco et 50 al., 2014) and Indonesia as the largest global palm-oil producer has planned to double its oil-51 palm area from 9.7 million ha in 2009 to 18 million ha by 2020 (Koh and Ghazoul, 2010). 52 Since oil palms favor a tropical-humid climate with consistently high temperatures and 53 humidity, the plantations have converted large areas of rainforest in Indonesia in the past two 54 decades including those on carbon-rich peat soils (Carlson et al., 2012; Gunarso et al. 2013). 55 Undisturbed forests have long-lasting capacity to store carbon in comparison to disturbed or 56 managed vegetation (Luyssaert et al., 2008). Tropical forest to oil palm conversion has 57 significant implications on above- and belowground carbon stocks (Kotowska et al., 2015a). 58 However, the exact quantification of long-term and large-scale forest – oil palm replacement 59 effects is difficult as the greenhouse gas balance of oil palms is still uncertain due to 60 incomplete monitoring of the dynamics of oil palm plantations (including young development 61 stage), and lack of understanding of the carbon, nitrogen, water and energy exchange between 62 oil palms, soil and the atmosphere at ecosystem scale. Besides that, the assessment of these 63 processes in agricultural ecosystems is complicated by human activities e.g. crop management, 64 including planting and pruning, irrigation and fertilization, litter and residues management, 65 and yield outputs. One of the suitable tools for evaluating the feedback of oil palm expansion 66 is ecosystem modelling. Although a series of agricultural models exist for simulating the 67 growth and yield of oil palm such as OPSIM (van Kraalingen et al., 1989), ECOPALM 68 (Combres et al., 2013), APSIM-Oil Palm (Huth et al., 2014), PALMSIM (Hoffmann et al., 69 2014), these models did not aim yet at the full picture of carbon, water and energy exchanges 70 between land and atmosphere and remain to be coupled with climate models. Given the 71 current and potential large-scale deforestation driven by the expansion of oil palm plantations,

the ecosystem services such as yield, carbon sequestration, microclimate, energy and water
balance of this new managed monoculture landscape have to be evaluated in order to estimate

the overall impact of land-use change on environment including regional and global climate.

75 Land surface modelling has been widely used to characterize the two-way interactions 76 between climate and human activities in terrestrial ecosystems such as deforestation, 77 agricultural expansion, and urbanization (Jin and Miller, 2011; Oleson et al., 2004). A variety 78 of land models have been adapted to simulate land-atmosphere energy and matter exchanges 79 for major crops such as the Community Land Model (CLM, Oleson et al., 2013). CLM 80 represents the crop and naturally vegetated land units as patches of plant functional types 81 (PFTs) defined by their key ecological functions (Bonan et al., 2002). However, most of the 82 crops being simulated are annual crops such as wheat, corn, soybean, etc. Their phenological 83 cycles are usually represented as three stages of development from planting to leaf emergence, 84 to fruit-fill and to harvest, all within a year. Attempts were also made to evaluate the climate 85 effects of perennial crops, e.g. by extending the growing season of annuals (Georgescu et al., 86 2011). However the perennial crops such as oil palm, cacao, coffee, rubber, coconut, and 87 other fruiting trees and their long-term biophysical processes are not represented in the above 88 land models yet, despite the worldwide growing demand (FAO, 2013).

89 Oil palm is a perennial evergreen crop which can be described by the Corner's architectural 90 model (Hall éet al., 1978). A number of phytomers, each carrying a large leaf (frond) and 91 axillating a fruit bunch, emerge successively (nearly two per month) from a single meristem 92 (the bud) at the top of a solitary stem. They form a multilayer canopy with old leaves 93 progressively being covered by new ones, until being pruned at senescence. Each phytomer 94 has its own phenological stage and yield, according to respective position in the crown. The 95 oil palm is productive for more than 25 years, including a juvenile stage of around 2 years. In 96 order to capture the inter- and intra-annual dynamics of growth and yield and land-97 atmosphere energy, water and carbon fluxes in the oil palm system, a new structure and

98 dimension detailing the phytomer-level phenology, carbon (C) and nitrogen (N) allocation

and agricultural managements have to be added to the current integrated plant-level

100 physiological parameterizations in the land models. This specific refinement needs to remain

101 compliant with the current model structure though, and be simple to parameterize.

102 In this context, we develop a new CLM-Palm sub-model for simulating the growth, yield, and

103 energy and material cycling of oil palm within the framework of CLM4.5. It introduces a sub-

104 canopy phenological and physiological parameterization, so that multiple leaf and fruit

105 components operate in parallel but at delayed steps. A phytomer in the model is meant to

106 represent the average condition of an age-cohort of actual oil palm phytomers across the

107 whole plantation landscape. The overall gross primary production (GPP) by leaves and carbon

108 output by fruit harvests rely on the development trends of individual phytomers. The

109 functions implemented for oil palm combine the characteristics of both trees and crops, such

110 as the woody-like stem growth and turnover but the crop-like vegetative and reproductive

allocations which enable fruit C and N output. Agricultural practices such as transplanting,

112 fertilization, and leaf pruning are also represented.

113 The main objectives of this paper are to: i) describe the development of CLM-Palm including

114 its phenology, carbon and nitrogen allocation, and yield output; ii) optimize model parameters

115 using field-measured leaf area index (LAI) and observed long-term monthly yield data from a

116 mature oil palm plantation in Sumatra, Indonesia; and iii) validate the model against

117 independent LAI, yield and net primary production (NPP) data from eight oil palm

118 plantations of different age in Sumatra, Indonesia.

119 2. Model development

For adequate description of oil palm functioning, we adapted the CLM crop phenology, allocation and vegetative structure subroutines to the monopodial morphology and sequential phenology of oil palm so that each phytomer evolves independently in growth and yield (Fig. 123 1). Their phenology sequence is determined by the phyllochron (the period in thermal time 124 between initiations of two subsequent phytomers) (Table A1). A maximum of 40 phytomers 125 with expanded leaves, each growing up to 7-m long, are usually maintained in plantations by 126 pruning management. There are also around 60 initiated phytomers developing slowly inside 127 the bud. The largest ones, already emerged at the top of the crown but unexpanded yet, are 128 named "spear" leaves (Fig. 1a). Each phytomer can be considered a sub-PFT component that 129 has its own prognostic leaf growth and fruit yield capacity but having 1) the stem and root 130 components that are shared by all phytomers, 2) the soil water content, nitrogen resources, 131 and resulting photosynthetic assimilates that are also shared and partitioned among all 132 phytomers, and 3) a vertical structure of the foliage, with the youngest at the top and the 133 oldest at the bottom of the canopy. Within a phytomer the fruit and leaf components do not 134 compete for growth allocation because leaf growth usually finishes well before fruit-fill starts. 135 However one phytomer could impact the other ones through competition for assimilates, 136 which is controlled by the C and N allocation subroutine according to their respective 137 phenological stages.

Here we describe only the new phenology, allocation and agricultural management functions developed for the oil palm. Photosynthesis, respiration, water and nitrogen cycles and other biophysical processes already implemented in CLM4.5 (Oleson et al., 2013) are not modified (except N retranslocation scheme) for the current study. The following diagram shows the new functions and their coupling with existing modules within the CLM4.5 framework (Fig. 2).

144 **2.1. Phenology**

145 Establishment of the oil palm plantation is implemented with two options: seed sowing or

transplanting of seedlings. In this study, the transplanting option is used. We design 7 post-

147 planting phenological steps for the development of each phytomer: 1) leaf initiation; 2) start

148 of leaf expansion; 3) leaf maturity; 4) start of fruit-fill; 5) fruit maturity and harvest; 6) start of

149 leaf senescence; and 7) end of leaf senescence and pruning (Fig. 1b). The first two steps 150 differentiate pre-expansion (heterotrophic) and post-expansion (autotrophic) leaf growth 151 phases. The other steps control leaf and fruit developments independently so that leaf growth 152 and maturity could be finished well before fruit-fill and leaf senescence could happen after 153 fruit harvest according to field observations. The modified phenology subroutine controls the 154 life cycle of each phytomer (sub-PFT level) as well as the planting, stem and root turnover, 155 vegetative maturity (start of fruiting) and final rotation (replanting) of the whole plant (PFT 156 level). Detailed description of oil palm phenology and nitrogen retranslocation during 157 senescence is in the Supplementary materials. The main phenological parameters are in Table 158 A1.

159 All phytomers are assumed to follow the same phenological steps, where the thermal length 160 for each phase is measured by growing degree-days (GDD; White et al., 1997). For oil palm, 161 a new GDD variable with 15 $^{\circ}$ C base temperature and 25 degree-days daily maximum (Corley 162 and Tinker, 2003; Goh, 2000; Hormaza et al., 2012) is accumulated from planting (abbr. 163 GDD_{15}). The phenological phases are signaled by respective GDD requirements, except that 164 pruning is controlled by the maximum number of expanded phytomers according to 165 plantation management (Table A1). Other processes in the model such as carbon and nitrogen 166 allocation for growth of new tissues respond to this phenology scheme at both PFT level and 167 phytomer level.

168 **2.2. Carbon and Nitrogen allocation**

169 In CLM, the fate of newly assimilated carbon from photosynthesis is determined by a coupled

170 C and N allocation routine. Potential allocation for new growth of various plant tissues is

171 calculated based on allocation coefficients and their allometric relationship (Table A2).

A two-step allocation scheme is designed for the sub-canopy phytomer structure and
according to the new phenology. First, available C (after subtracting respiration costs) is

174 partitioned to the root, stem, overall leaf, and overall fruit pools with respect to their relative 175 demands by dynamic allocation functions according to PFT-level phenology. The C:N ratios 176 for different tissues link C demand and N demand so that a N down-regulation mechanism is 177 enabled to rescale GPP and C allocation if N availability from soil mineral N pool and 178 retranslocated N pool does not meet the demand. Then, the actual C and N allocated to the 179 overall leaf or fruit pools are partitioned between different phytomers at the sub-PFT level 180 (Fig. 2). Details are described below.

181 2.2.1. PFT level allocation

182 C and N allocation at the PFT level is treated distinctly before and after oil palm reaches

vegetative maturity. At the juvenile stage before fruiting starts (i.e. $GDD_{15} < GDD_{min}$) all the allocation goes to the vegetative components. The following equations are used to calculate the allometric ratios for partitioning available C and N to the leaf, stem, and root pools.

186
$$A_{root} = a_{root}^i - (a_{root}^i - a_{root}^f) \frac{DPP}{Age_{max}}, \qquad (Eq. 1)$$

187
$$A_{leaf} = f_{leaf}^{i} \times (1 - A_{root})$$
(Eq. 2)

$$A_{stem} = 1 - A_{root} - A_{leaf}$$
(Eq. 3)

189 where $\frac{DPP}{Age_{max}} \le 1$, *DPP* is the days past planting, and Age_{max} is the maximum plantation age 190 (~25 years). a_{root}^{i} and a_{root}^{f} are the initial and final allocation coefficients for roots and f_{leaf}^{i} 191 is the initial leaf allocation coefficient before fruiting (Table A2). Root and stem allocation 192 ratios are calculated with Eqs. 1 and 3 for all ages and phenological stages of oil palm.

193 After fruiting begins, the new non-linear function is used for leaf allocation:

194
$$A_{leaf} = a_{leaf}^2 - (a_{leaf}^2 - a_{leaf}^f) \left(\frac{DPP - DPP_2}{Age_{max} \times d_{mat} - DPP_2}\right)^{d_{alloc}^{leaf}}$$
(Eq. 4)

where a_{leaf}^2 equals the last value of A_{leaf} calculated right before fruit-fill starts and DPP_2 is the days past planting right before fruit-fill starts. d_{mat} controls the age when the leaf allocation ratio approaches its final value a_{leaf}^f , while d_{alloc}^{leaf} determines the shape of change (convex when $d_{alloc}^{leaf} < 1$; concave when $d_{alloc}^{leaf} > 1$). A_{leaf} stabilizes at a_{leaf}^f when $DPP \ge$ $Age_{max}d_{mat}$. The equations reflect changed vegetative allocation strategy that shifts resources to leaf for maintaining LAI and increasing photosynthetic productivity when fruiting starts. The three vegetative allocation ratios A_{leaf} , A_{stem} and A_{root} always sum to 1.

At the reproductive phase a fruit allocation ratio A_{fruit} is introduced, which is relative to the total vegetative allocation unity. To represent the dynamics of reproductive allocation effort of oil palm, we adapt the stem allocation scheme for woody PFTs in CLM, in which increasing NPP results in increased allocation ratio for the stem wood (Oleson et al., 2013). A similar formula is used for reproductive allocation of oil palm so that it increases with increasing NPP:

208
$$A_{fruit} = \frac{2}{1 + e^{-b(NPP_{mon}-100)}} - a$$
 (Eq. 5)

where NPP_{mon} is the monthly sum of NPP from the previous month calculated with a runtime accumulator in the model. The number 100 (g C m⁻² mon⁻¹) is the base monthly NPP when the palm starts to yield (Kotowska et al., 2015a). Parameters *a* and *b* adjust the base allocation rate and the slope of change, respectively (Table A2). This function generates a dynamic curve of A_{fruit} increasing from the beginning of fruiting to full vegetative maturity, which is used in the allocation allometry to partition assimilates between vegetative and reproductive pools (Fig. 3).

216 2.2.2. Sub-PFT (phytomer) level allocation

217 Total leaf and fruit allocations are partitioned to the different phytomers according to their

218 phenological stages. Fruit allocation per phytomer is calculated with a sink size index:

219
$$S_p^{fruit} = \frac{GDD_{15} - H_p^{F,fill}}{H_p^{F,mat} - H_p^{F,fill}},$$
 (Eq. 6)

where p stands for the phytomer number, $H_p^{F,fill}$ and $H_p^{F,mat}$ are the phenological indices for 220 the start of fruit-fill and fruit maturity (with $H_p^{F.fill} \leq GDD_{15} \leq H_p^{F.mat}$). S_p^{fruit} increases 221 222 from zero at the beginning of fruit-fill to the maximum of 1 right before harvest for each 223 phytomer. This is because the oil palm fruit accumulates assimilates at increasing rate during 224 development until the peak when it becomes ripe and oil synthesis dominates the demand (Corley and Tinker, 2003). The sum of S_p^{fruit} for all phytomers gives the total reproductive 225 sink size index. Each phytomer receives a portion of fruit allocation by $\frac{S_p^{fruit}}{\sum_{n=1}^n S_n^{fruit}} \times A_{fruit}$, 226 227 where A_{fruit} is the overall fruit allocation by Eq. 5.

An important allocation strategy for leaf is the division of displayed versus storage pools for the pre-expansion and post-expansion leaf growth phases. These two types of leaf C and N pools are distinct in that only the displayed pools contribute to LAI growth, whereas the storage pools support the growth of unexpanded phytomers, i.e. bud & spear leaves, which remain photosynthetically inactive. Total C and N allocation to the overall leaf pool is divided to the displayed and storage pools by a fraction lf_{disp} (Table A2) according to the following equation:

235
$$A_{leaf}^{display} = lf_{disp} \times A_{leaf}$$
$$A_{leaf}^{storage} = (1 - lf_{disp}) \times A_{leaf}$$
(Eq. 7)

The plant level A^{display}_{leaf} and A^{storage}_{leaf} are then distributed evenly to expanded and
unexpanded phytomers, respectively, at each time step. When a phytomer enters the leaf

expansion phase, C and N from its leaf storage pools transfer gradually to the displayed pools
during the expansion period. Therefore, a transfer flux is added to the real-time allocation flux
and they together contribute to the post-expansion leaf growth.

LAI is calculated only for each expanded phytomer according to a constant specific leaf area (SLA) and prognostic amount of leaf C accumulated by phytomer *n*. In case it reaches the prescribed maximum (*PLAI_{max}*), partitioning of leaf C and N allocation to this phytomer becomes zero.

245 **2.3. Other parameterizations**

246

A part of N from senescent leaves and from the portion of live stem that turns dead is remobilized to a separate N pool that feeds plant growth or reproductive demand. Nitrogen of fine roots is all moved to the litter pool during root turnover. We do not consider N retranslocation from live leaves, stem and roots specifically during grain-fill that is designed for annual crops (Drewniak et al., 2013) because oil palm has continuous fruit-fill year around

Nitrogen retranslocation is performed exclusively during leaf senescence and stem turnover.

at different phytomers.

253 The fertilization scheme for oil palm is adapted to the plantation management generally

carried out in our study area, which applies fertilizer biannually, starting only 6 years after

255 planting, assuming each fertilization event lasts one day. Currently the CLM-CN

belowground routine uses an unrealistically high denitrification rate under conditions of

257 nitrogen saturation, e.g. after fertilization, which results in a 50% loss of any excess soil

258 mineral nitrogen per day (Oleson et al., 2013). This caused the simple biannual regular

259 fertilization nearly useless because peak N demand by oil palm is hard to predict given its

260 continuous fruiting and vegetative growth and most fertilized N is thus lost in several days.

261 The high denitrification factor has been recognized as an artifact (Drewniak et al., 2013; Tang

et al., 2013). According to a study on a banana plantation in the tropics (Veldkamp and Keller,

263	1997), around 8.5%	of fertilized N is lost	as nitrogen oxide (N ₂ O and NO). Accounting
-00	1 > > > , , ui o uii u o ie , o	01 101011111111111111111111111111111111	as ma sen on as	1.70 4110 1.0	

additionally for a larger amount of denitrification loss to gaseous N_2 , we modified the daily

denitrification rate from 0.5 to 0.001, which gives a 30% annual loss of N due to

denitrification that matches global observations (Galloway et al., 2004).

- 267 The irrigation option is turned off because oil palm plantations in the study area are usually
- 268 not irrigated. Other input parameters for oil palm such as its optical, morphological, and
- 269 physiological characteristics are summarized in Table A3. Most of them are generalized over
- the life of oil palm.

271 **3. Model evaluation**

272 **3.1. Site data**

273 Two oil palm plantations in the Jambi province of Sumatra, Indonesia provide data for 274 calibration. One is a mature industrial plantation at PTPN-VI (01 %1.6' S, 103 °23.5' E, 2186 ha) planted in 2002, which provides long-term monthly harvest data (2005 to 2014). Another 275 276 is a 2-year young plantation at a nearby smallholder site Pompa Air (01 '50.1' S, 103 '17.7' E, 277 5.7 ha). The leaf area and dry weight at multiple growth stages were measured by sampling 278 leaflets of phytomers at different ranks (+1 to +20) on a palm and repeating for 3 different 279 ages within the two plantations. The input parameter SLA (Table A2) was derived from leaf 280 area and dry weight (excluding the heavy rachis). The phytomer-level LAI was estimated 281 based on the number of leaflets (90-300) per leaf of a certain rank and the PFT-level LAI was 282 estimated by the number of expanded leaves (35-45) per palm of a certain age. In both cases, 283 a planting density of 156 palms per hectare ($8m \times 8m$ per palm) was used according to 284 observation.

Additionally, LAI, yield and NPP measurements from eight independent smallholder oil palm
plantations (50m × 50m each) were used for model validation. Four of these sites (HO1, HO2,
HO3, HO4, 11-18 years old) are located in the Harapan region nearby PTPN-VI, and another

288 four (BO2, BO3, BO4, BO5, 10-14 years old) are in Bukit Duabelas region (02 '04' S, 102 ° 289 47' E), both in Jambi, Sumatra. Fresh bunch harvest data were collected at these sites for a 290 whole year in 2014. Harvest records from both PTPN-VI and the 8 validation sites were 291 converted to harvested carbon (g C/m^2) with mean wet/dry weight ratio of 58.65 % and C 292 content 60.13 % per dry weight according to C:N analysis (Kotowska et al., 2015a). The oil 293 palm monthly NPP and its partitioning between fruit, leaf, stem and root were estimated 294 based on measurements of fruit yield (monthly), pruned leaves (monthly), stem increment 295 (every 6 month) and fine root samples (once in a interval of 6-8 month) at the eight validation 296 sites (Kotowska et al., 2015b).

The amount of fertilization at the industrial plantation PTPN-VI was 456 kg N ha⁻¹ yr⁻¹, 297 298 applied regularly twice per year since 6-year old. The smallholder plantations in Harapan (H 299 plots) and Bukit Duabelas (B plots) used much less fertilizer. From interview data, the H plots 300 had roughly regular N fertilization (twice per year), whereas among the B plots only BO3 301 indicated one fertilization event per year but the amount was unclear (applied chicken manure 302 in 2013) and the other plots had no N fertilization in 2013 and 2014 due to financial difficulty. 303 Fertilization history prior to 2013 is unavailable for all the smallholder plantations. Given the limited information, we consider two levels of fertilization for H plots (regular: 96 kg N ha⁻¹ 304 yr⁻¹, from 6-year old until 2014) and B plots (reduced: 24 kg N ha⁻¹ yr⁻¹, from 6-year old until 305 306 2012), respectively.

The mean annual rainfall (the Worldclim database: http://www.worldclim.org (Hijmans et al., 2005); average of 50 years) of the two investigated landscapes in Jambi Province was ~2567 mm y⁻¹ in the Harapan region (including PTPN-VI) and ~2902 mm y⁻¹ in the Bukit Duabelas region. In both areas, May to September represented a markedly drier season (30% less precipitation) in comparison to the rainy season between October and April. Air temperature is relatively constant throughout the year with an annual average of 26.7 °C. In both landscapes, the principal soil types are Acrisols: in the Harapan landscape loam Acrisols

- dominate, whereas in Bukit Duabelas the majority is clay Acrisol. Soil texture such as
- 315 sand/silt/clay ratios and soil organic matter C content were measured at multiply soil layers
- 316 (down to 2.5m) (Allen et al., 2015). They were used to create two sets of surface input data
- 317 for the two regions separately.

318 **3.2. Model setup**

319 The model modifications and parameterizations were implemented according to CLM4.5 320 standards. A new sub-PFT dimension called *phytomer* was added to all the new variables so 321 that the model can output history tapes of their values for each phytomer and prepare restart 322 files for model stop and restart with bit-for-bit continuity. Simulations were set up in point 323 mode (a single 0.5×0.5 degree grid) at every 30-min time step. A spin-up procedure (Koven et 324 al., 2013) was followed to get a steady-state estimate of soil C and N pools, with the CLM-325 CN decomposition cascade and broadleaf evergreen tropical forest PFT. The soil C and N 326 pools were rescaled to match the average field observation at two reference lowland rainforest 327 sites in Harapan and Bukit Duabelas regions (Allen et al., 2015; Guillaume et al., 2015), 328 which serve as the initial conditions. The forest was replaced with the oil palm at a specific 329 year of plantation establishment (2002 for PTPN-VI and 1996, 1997, 1999, 2000, 2001, 2002, 330 2003, 2004 for HO3, HO1, HO2, BO2, BO3, BO4, HO4, BO5, respectively). The oil palm 331 functions were then turned on and simulations continued until 2014. The 3-hourly ERA 332 Interim climate data (Dee et al., 2011) were used as atmospheric forcing.

333 **3.3. Calibration of key parameters**

A simulation from 2002 to 2014 at the PTPN-VI site was used for model calibration. Both the PFT level and phytomer level LAI development were calibrated with field observations in 2014 from a chronosequence approach (space for time substitution) using oil palm samples of three different age and multiple phytomers of different rank (section 3.1). Simulated yield outputs (around twice per month) were calibrated with monthly harvest records of PTPN-VI plantation from 2005 to 2014. Cumulative yields were compared because the timing of
harvest in the plantations was largely uncertain and varied depending on weather and other
conditions.

342 To simplify model calibration, we focused on parameters related to the new phenology and 343 allocation processes. Phenological parameters listed in Table A1 were determined according 344 to field observations and existing knowledge about oil palm growth phenology (Combres et 345 al., 2013; Corley and Tinker, 2003) as well as plantation management in Sumatra, Indonesia. 346 Allocation coefficients in Table A2 were more uncertain and they were the key parameters to 347 optimize in order to match observed LAI and yield dynamics according to the following 348 sensitivity analysis. Measurements of oil palm NPP and its partitioning between fruit, canopy, 349 stem, and root from the eight sites (section 3.1) were used as a general reference when 350 calibrating the allocation coefficients.

Leaf C:N ratio and *SLA* were determined by field measurements. Other C:N ratios and optical and morphological parameters in Table A3 were either fixed by field observations or adjusted in-between trees and crops.

354 **3.4. Sensitivity analysis**

355 Performing a full sensitivity analysis of all parameters used in simulating oil palm (more than 356 100 parameters, though a majority are shared with natural vegetation and other crops) would 357 be a challenging work. As with calibration, we limited the sensitivity analysis to a set of 358 parameters introduced for the specific PFT and model structure designed for oil palm (Tables 359 A1 and A2). Among the phenological parameters, *mxlivenp* (maximum number of expanded 360 phytomers) and *phyllochron* (Table A1) are closely related to pruning frequency but they 361 should not vary widely for a given oil palm breed and plantation condition. Therefore, they were fixed at the average level for the study sites in Jambi, Sumatra. Parameter PLAI_{max} is 362 363 only meant for error controlling, although in our simulations phytomer-level LAI never

364 reached *PLAI_{max}* (see Fig. 5 in results) because environmental constraints and nitrogen down-

365 regulation already limited phytomer leaf growth well within the range. *GDD*_{init} was kept to

366 zero because only the transplanting scenario was considered for seedling establishment.

367 We tested two hypotheses of phytomer level leaf development based on the other

368 phenological parameters: 1) considering the leaf storage growth period, that is, the bud &

369 spear leaf phase is explicitly simulated with the GDD parameters in Table A1 and $lf_{disp} = 0.3$

in Table A2; 2) excluding the storage growth period by setting $GDD_{exp} = 0$ and $lf_{disp} = 1$ so

that leaf expands immediately after initiation and leaf C and N allocation all goes to the

372 photosynthetic active pools.

The sensitivity of allocation and photosynthesis parameters in Table A2 were tested by adding or subtracting 10% or 30% to the baseline values (calibrated) one-by-one and calculating their effect on final cumulative yield at the end of simulation (December 2014). In fact, all the allocation parameters are interconnected because they co-determine photosynthesis capacity and respiration costs as partitioning to the different vegetative and reproductive components varies. This simple approach provides a starting point to identify sensitive parameters, although a more sophisticated sensitivity analysis is needed in the future.

380 3.5. Validation

381 In this study, we only validated the model structure and model behavior on simulating

aboveground C dynamics and partitioning as represented by LAI, fruit yield and NPP.

383 Independent leaf measurement, yield and monthly NPP data collected in 2014 from the eight

mature oil palm sites (H and B plots) were compared with the eight simulations using the

385 same model settings and calibrated parameters, except that two categories of climate forcing,

386 surface input data (for soil texture) and fertilization (regular vs. reduced) were prescribed for

the H plots and B plots, respectively.

388 **4. Results**

389 4.1. Calibration with LAI and yield

390 In calibration with the industrial PTPN-VI plantation, the PFT-level LAI dynamics simulated 391 by the model incorporating the pre-expansion phase matches well with the LAI measurements 392 for three different ages (Fig. 4). Simulated LAI for the PFT increases with age in a sigmoid 393 relationship. The dynamics of LAI is also impacted by pruning and harvest events because oil 394 palms invest around half of their assimilates into fruit yield. Oil palms are routinely pruned by 395 farmers to maintain the maximum number of expanded leaves around 40. Hence, when yield 396 begins 2-3 years after planting, LAI recurrently shows an immediate drop after pruning and 397 then quickly recovers. The pruning frequency decreases with age because the phyllochron 398 increases to 1.5 times at 10-year old (Supplementary materials). Simulations without the pre-399 expansion storage growth phase show an unrealistic fast increase of LAI before 3 years old, 400 much higher than observed in the field. At older age after yield begins, LAI drops drastically 401 and recovers afterwards. Although the final LAI could stabilize at a similar level, the initial 402 jump and drop of LAI at young stage do not match field observations and cannot be solved by 403 adjusting parameters other than GDD_{exp} . Hereafter, all simulations were run using the pre-404 expansion phase.

405 The phytomer level LAI development is comparable with leaf samples from the field (Fig. 5). 406 The two leaf samples at rank 5 (LAI = 0.085) and rank 20 (LAI = 0.122) of a mature oil palm 407 in PTPN-VI (the two black triangles for 2014) are within the range of simulated values. The 408 other sample at rank 25 (LAI = 0.04, for 2004) of a young oil palm in Pompa Air (smallholder 409 plantation) is lower than the simulated value. Each horizontal color bar clearly marks the 410 post-expansion leaf phenology cycle, including gradual increment of photosynthetic LAI 411 during phytomer development and gradual declining during senescence. The pre-expansion 412 phase is not included in the figure but model outputs show that roughly 60-70% of leaf C in a 413 phytomer is accumulated before leaf expansion, which is co-determined by the allocation

414 ratio lf_{disp} and the lengths of two growth phases set by GDD_{exp} and $GDD_{L.mat}$. This is

415 comparable to observations on coconut palm that dry mass of the oldest unexpanded leaf

416 accounts for 60% of that of a mature leaf (Navarro et al., 2008). Only when the palm becomes

417 mature, phytomer LAI could come closer to the prescribed *PLAI_{max}* (0.165). However, during

418 the whole growth period from 2002 to 2014 none of the phytomers have reached $PLAI_{max}$,

419 which is the prognostic result of the carbon balance simulated by the model.

420 The cumulative yield of baseline simulation has overall high consistency with harvest records

421 (Fig. 6). The mean percentage error (MPE) is only 3%. The slope of simulated curve

422 increases slightly after 2008 when the LAI continues to increase and NPP reaches a high level

423 (Fig. 3). The harvest records also show the same pattern after 2008 when heavy fertilization 424 began (456 kg N ha⁻¹ vr⁻¹).

425 The per-month harvest records exhibit strong zig-zag pattern (Fig. 7). One reason is that oil 426 palms are harvested every 15-20 days and summarizing harvest events by calendar month 427 would result in uneven harvest times per month, e.g. two harvests fall in a previous month and 428 only one in the next month. Yet it still shows that harvests at PTPN-VI plantation dominated 429 from October to December whereas in the earlier months of each year harvest amounts were 430 significantly lower. The simulated monthly yield has less seasonal fluctuation, but it responds 431 to the fluctuation of precipitation (Fig. 7). A slight positive linear correlation exists between 432 simulated yield and the mean precipitation of a 60-day period (corresponds to the main fruit-433 filling and oil synthesis period) before each harvest event (Pearson's r = 0.15). Examining the 434 longer term year-to-year variability, a clear increasing trend of yield with increasing 435 plantation age is captured by the model, largely matching field records since the plantation 436 began to yield in 2005.

437 **4.2. Sensitivity analysis**

438 The leaf nitrogen fraction in Rubisco (F_{LNR}) is shown to be the most sensitive parameter (Fig. 439 8), because it determines the maximum rate of carboxylation at 25 $\,^{\circ}$ C (V_{cmax25}) together with 440 SLA (also sensitive), foliage nitrogen concentration (CN_{leaf}, Table A3) and other constants. 441 Given the fact that F_{LNR} should not vary widely in nature for a specific plant, we constrained 442 this parameter within narrow boundaries to get a V_{cmax25} around 100.7, the same as that shared 443 by all other crop PFTs in CLM. We fixed SLA to 0.013 by field measurements. The value is only representative of the photosynthetic leaflets. The initial root allocation ratio (a_{root}^i) has 444 445 considerable influence on yield because it modifies the overall respiration cost along the gradual declining trend of fine root growth across 25 years (Eq. 1). The final ratio (a_{root}^{f}) has 446 447 limited effects because its baseline value (0.1) is set very low and thus the percentage changes are insignificant. The leaf allocation coefficients $(f_{leaf}^{i}, a_{leaf}^{f})$ are very sensitive parameters 448 449 because they determine the magnitudes of LAI and GPP and consequently yield. The coefficients d_{mat} and d_{alloc}^{leaf} control the nonlinear curve of leaf development (Eq. 4) and 450 hence the dynamics of NPP and that partitioned to fruits. Increased F_{stem}^{live} results in higher 451 452 proportion of live stem throughout life, given the fixed stem turnover rate (Supplementary 453 materials), and therefore it brings higher respiration cost and lower yield. The relative 454 influence of fruit allocation coefficients a and b on yield is much lower than the leaf 455 allocation coefficients because of the restriction of A_{fruit} by NPP dynamics (Eq. 5). 456 Parameters lf_{disp} and transplant have negligible effects. lf_{disp} has to work together with the 457 phenological parameter GDD_{exp} to give a reasonable size of spear leaves before expansion 458 according to field observation. The sensitivity of GDD_{exp} is shown in Fig. 4. Varying the size 459 of seedlings at transplanting by 10% or 30% does not alter the final yield, likely because the 460 initial LAI is still within a limited range $(0.1 \sim 0.2)$ given the baseline value 0.15.

461 **4.3. Model validation with independent dataset**

The LAI development curves for the eight oil palm sites follow similar patterns since field
transplanting in different years, except that the B plots (BO2, BO3, BO4) are restrained in

464 LAI growth after 11 years old because of reduced fertilization (Fig. 9a). The field data in 465 2014 also shows the check by N limitation and even exhibits a decreasing trend of LAI with 466 increasing plantation age at B plots except BO5 which is under 10 years old (Fig. 9b). In 467 general, the modelled LAI has a positive relationship with plantation age under regularly 468 fertilized condition and it stabilizes after 15-year old (site HO3) as controlled by d_{mat} (Eq. 4). This age-dependent trend is observed in the field with a notable deviation by site HO1. The 469 470 average LAI of the eight sites from the model is comparable with field measurement in 2014 471 (MPE = 13%). There are large uncertainties in field LAI estimates because we did not 472 measure LAI at the plot level directly but only sampled leaf area and dry weight of individual

473 phytomers and scaled the values up.

474 The simulated annual yields match closely with field observations in 2014 at both the H plots 475 (MPE = 2%) and B plots (MPE = 2%); Fig. 10). With regular fertilization in the H plots, both 476 the modelled and observed yield are slightly higher in the older plantations (HO2, HO1, and HO3) than the younger one (H04) but stabilize around 1280 g C m⁻² yr⁻¹ past the age of 15 477 years. In contrast, the B plots have significantly lower yield because of reduced N input and 478 479 the model is able to capture the N limitation effect on both NPP and yield, i.e. the declining 480 trend with increasing age, which is consistent with field observation. The model simulates 481 slightly higher NPP than field estimates at 7 smallholder sites (MPE = 10%) using the input 482 parameters calibrated and optimized only for LAI and yield at the industrial PTPN-VI 483 plantation. Field measured leaf NPP only includes leaf litter production but does not account 484 for the growing size of canopy (i.e. increasing LAI).

485 **5. Discussion**

Calibration and validation with multiple site data demonstrate the utility of CLM-Palm and its
sub-canopy structure for simulating the growth and yield of the unique oil palm plantation
system within a land surface modeling context.

489 The pre-expansion phenological phase is proved necessary for simulating both phytomer-490 level and PFT-level LAI development in a prognostic manner. The leaf C storage pool 491 provides an efficient buffer to support phytomer development and maintain overall LAI 492 during fruiting. It also avoids an abnormally fast increase of LAI in the juvenile stage when C 493 and N allocation is dedicated to the vegetative components. Without the leaf storage pool, the 494 plant's canopy develops unrealistically fast at young age and then enters an emergent drop 495 once fruit-fill begins (Fig. 4). This is because the plant becomes unable to sustain leaf growth 496 just from its current photosynthetic assimilates when a large portion is allocated to fruits. 497 The model well simulates year-to-year variability in yield (Fig. 7), in which the increasing 498 trend is closely related to the fruit allocation function (Fig. 3) and LAI development (Fig. 4). 499 The seasonal variability in simulated yield corresponds to the precipitation data but it is 500 difficult to interpret the difference with monthly harvest records due to the artificial zig-zag 501 pattern. The harvest records from plantations do not necessarily correspond to the amount of 502 mature fruits along a phenological time scale due to varying harvest arrangements, e.g. fruits 503 are not necessarily harvested when they are ideal for harvest, but when it is convenient. 504 Observations of mature fruits on a tree basis (e.g. Navarro et al., 2008 on coconut) would be 505 more suitable to compare with modeled yield, but such data are not available at our sites. 506 Some studies have also demonstrated important physiological mechanisms on oil palm yield 507 including inflorescence gender determination and abortion rates that both respond to seasonal 508 climatic dynamics although with a time lag (Combres et al., 2013; Legros et al., 2009). The 509 lack of representation of such physiological traits might affect the seasonal dynamics of yield 510 simulated by our model, but these mechanisms are rarely considered in a land surface 511 modelling context. Nevertheless, the results correspond generally to the purpose of our 512 modelling which is focused on the long-term climatological effects of oil palm agriculture. 513 The correct representation of multi-year trend of carbon balance which we did reach is more 514 important than the correct prediction of each yield. For latter the more agriculturally-oriented 515 models should be used.

516 Resource allocation patterns for perennial crops are more difficult to simulate than annual 517 crops. For annuals, the LAI is often assumed to decline during grain-fill (Levis et al., 2012). 518 However, the oil palm has to sustain a rather stable leaf area while partitioning a significant 519 amount of C to the fruits. The balance between reproductive and vegetative allocations is 520 crucial. The dynamics of A_{fruit} as a function of monthly NPP is proved useful to capture the 521 increasing yield capacity of oil palms during maturing at favorable conditions (Fig. 6, 7) and 522 also able to adjust fruit allocation and shift resources to the vegetative components under stress conditions (e.g. N limitation, Fig. 9 and 10). The value of A_{fruit} increased from 0.5 to 523 524 1.5 (Fig. 3), resulting more than a half partitioning of NPP to the reproductive pool at mature 525 stage which matched closely with field observations (Fig. 10; Kotowska et al., 2015a; 526 Kotowska et al., 2015b). Our experiments (not shown here) confirmed that the dynamic 527 function is more robust than a simple time-dependent or vegetation-size-dependent allocation 528 function.

529 The phenology and allocation processes in land surface models are usually aimed to represent 530 the average growth trend of a PFT at large spatial scale (Bonan et al., 2002; Drewniak et al., 531 2013). We made a step forward by comparing point simulations with multiple specific site 532 observations. The model predicts well the average LAI development and yield as well as NPP 533 of mature plantations across two different regions. Site-to-site variability in yield and NPP at 534 the Harapan and Bukit Duabelas plots under contrasting conditions (regular vs. reduced 535 fertilization) is largely captured by the model. The decreasing trend of yield and pause of LAI 536 growth in B plots after 10 years old (Fig. 9, 10) reflect reduced N availability observed in the 537 clay Acrisol soil in Bukit Duabelas (Allen et al., 2015) with very limited C and N return from 538 leaf litter because of pruning and piling of highly lignified leaves (Guillaume et al., 2015). 539 Yet there remains small-scale discrepancy in LAI, NPP or yield in some sites which is 540 possibly due to the fact that microclimate, surface input data and the amount and timing of 541 fertilization were only prescribed as two categories for H and P plots, respectively. Field data 542 show the proportion of NPP allocated to yield is significantly higher in plot HO1 (70%) than

543 in other plots (50% to 65%) which could explain the low LAI of HO1. This is not reflected in 544 the model as the same parameters are used in the fruit allocation function (Eq. 5) across sites. 545 The deviation in allocation pattern is likely due to difference in plantation management (e.g. 546 harvest and pruning cycles), which has been shown to be crucial for determining vegetative 547 and reproductive growth (Euler et al., 2015). Other factors such as insects, fungal infection, 548 and possibly different oil palm progenies could also result in difference in oil palm growth 549 and productivity, but they are typically omitted in land surface models. Generalized input 550 parameterization across a region is usually the case when modeling with a PFT, although a 551 more complex management (e.g. dynamic fertilization) scheme could be devised and 552 evaluated thoroughly with additional field data, which we lack at the moment.

553 Overall, the sub-canopy phytomer-based structure, the extended phenological phases for a 554 perennial crop PFT and the two-step allocation scheme of CLM-Palm are distinct from 555 existing functions in land surface models. The phytomer configuration is similar to the one 556 already implemented in other oil palm growth and yield models such as the APSIM-Oil Palm 557 model (Huth et al., 2014) or the ECOPALM yield prediction model (Combres et al., 2013). 558 But the implementation of this sub-canopy structure is the first attempt among land surface 559 models. CLM-Palm incorporates the ability of an agricultural model for simulating growth 560 and yield, beside that it allows the modeling of biophysical and biogeochemical processes as a 561 land model should do, e.g. what is the whole fate of carbon in plant, soil and atmosphere if 562 land surface composition shifts from a natural system to the managed oil palm system? In a 563 following study, a fuller picture of the carbon, nitrogen, water and energy fluxes over the oil 564 palm landscape are examined with CLM-Palm presented here and evaluated with Eddy 565 Covariance flux observation data. We develop this palm sub-model in the CLM framework as it allows coupling with climate models so that the feedbacks of oil palm expansion to climate 566 567 can be simulated in future steps.

568 6. Conclusions

569 The development of CLM-Palm including canopy structure, phenology, and carbon and 570 nitrogen allocation functions was proposed for modeling an important agricultural system in 571 the tropics. This paper demonstrates the ability of the new palm module to simulate the inter-572 annual dynamics of vegetative growth and fruit yield from field planting to full maturity of 573 the plantation. The sub-canopy-scale phenology and allocation strategy are necessary for this 574 perennial evergreen crop which yields continuously on multiple phytomers. The pre-575 expansion leaf storage growth phase is proved essential for buffering and balancing overall vegetative and reproductive growth. Average LAI, yield and NPP are satisfactorily simulated 576 577 for multiple sites, which fulfills the main mission of a land surface modeling approach, that is, 578 to represent the average conditions and dynamics of large-scale processes. On the other hand, 579 simulating small-scale site-to-site variation ($50m \times 50m$ sites) requires detailed input data on 580 site conditions (e.g. microclimate) and plantation managements that are often not available 581 thus limiting the applicability of the model at small scale. Nevertheless, the CLM-Palm model 582 sufficiently represents the significant region-wide variability in oil palm NPP and yield driven 583 by nutrient input and plantation age in Jambi, Sumatra. The point simulations here provide a 584 starting point for calibration and validation at large scales.

585 To be run in a regional or global grid, the age class structure of plantations needs to be taken 586 into account. This can be achieved by setting multiple replicates of the PFT for oil palm, each 587 planted at a point of time at a certain grid. As a result, a series of oil palm cohorts developing 588 at different grids could be configured with a transient PFT distribution dataset, which allows 589 for a quantitative analysis of the effects of land-use changes, specifically rainforest to oil palm 590 conversion, on carbon, water and energy fluxes. This will contribute to the land surface 591 modeling community for simulating this structurally unique, economically and ecologically 592 sensitive, and fast expanding oil palm land cover.

593 Acknowledgements:

594 This study was funded by the European Commission Erasmus Mundus FONASO Doctorate 595 fellowship. Field trips were partly supported by the Collaborative Research Centre 990

- 596 (Ecological and Socioeconomic Functions of Tropical Lowland Rainforest Transformation
- 597 Systems (Sumatra, Indonesia)) funded by the German Research Foundation (DFG). We are
- 598 grateful to Kara Allen (University of Göttingen, Germany), Dr. Bambang Irawan (University
- 599 of Jambi, Indonesia) and the PTPN-VI plantation in Jambi for providing field data on oil palm.
- 600 The source code of the post-4.5 version CLM model was provided by Dr. Samuel Levis from
- 601 National Center for Atmospheric Research (NCAR), Boulder, CO, USA.
- This open-access publication was funded by the University of Gättingen.



Growing degree-days (base 15°C)

A phyllochron

Fig. 1. (a) New sub-canopy phytomer structure of CLM-Palm. P^1 to P^n indicate expanded phytomers and P^{-1} to P^{-n} at the top indicate unexpanded phytomers packed in the bud. Each phytomer has its own phenology, represented by different colors corresponding to: (b) the phytomer phenology: from initiation to leaf expansion, to leaf maturity, to fruit-fill, to harvest, to senescence and to pruning. Phytomers initiate successively according to the phyllochron (the period in heat unit between initiations of two subsequent phytomers). Detailed phenology description is in Supplementary materials.

613



614

Fig. 2. Original and modified structure and functions for developing CLM-Palm in the
framework of CLM4.5. Original functions from CLM4.5 are represented in black or grey.
New functions designed for CLM-Palm are represented in red, including phenology,
allocation, pruning, fruit harvest and export, as well as the sub-canopy (sub-PFT) structure.



Fig. 3. Time course of reproductive allocation rate (blue line) in relation to monthly NPP from the previous month (*NPP_{mon}*, green line) according to Eq. 5. A_{fruit} is relative to the vegetative unity ($A_{leaf} + A_{stem} + A_{root} = 1$ and $0 \le A_{fruit} \le 2$). The *NPP_{mon}* was simulated with calibrated parameters for the PTPN-VI site.



Fig. 4. PFT-level LAI simulated by CLM-Palm, with and without the pre-expansion growth
phase in the phytomer phenology and compared to field measurements used for calibration.
The initial sudden increase at year 1 represents transplanting from nursery. The sharp drops
mark pruning events.

Fig. 5. Simulated phytomer level LAI dynamics (horizontal color bar) compared with field observations (black triangles with measured LAI value). The newly expanded phytomer at a given point of time has a rank of 1. Each horizontal bar represents the life cycle of a phytomer after leaf expansion. Phytomers emerge in sequence and the y-axis gives the total number of phytomers that have expanded since transplanting in the field. Senescent phytomers are pruned.

Fig. 6. Simulated PFT-level yield compared with monthly harvest data (2005-2014) from the

calibration site PTPN-VI in Jambi, Sumatra. CLM-Palm represents multiple harvests from
 different phytomers (about twice per month). The cumulative harvest amounts throughout

643 time are compared.

Fig. 7. Simulated and observed monthly yield at PTPN-VI compared with monthly

646 precipitation dynamics (mean: 206 mm per month). The modeled yield outputs are per harvest

647 event (every 15-20 days depending on the phyllochron), while harvest records are the

summary of harvest events per month. The model output is thus rescaled to show the monthly

trend of yield that matches the mean of harvest records, given that the cumulative yields are

almost the same between simulation and observation as shown in Fig. 6.

Fig. 8. Sensitivity analysis of key allocation parameters in regard of the cumulative yield atthe end of simulation, with two magnitudes of change in the value of a parameter one-by-one

while others are hold at the baseline values in Table A2.

Fig. 9. Validation of LAI with 8 independent oil palm sites (sequence in plantation age) fromthe Harapan (regular fertilization) and Bukit Duabelas (reduced fertilization) regions: (a)

659 shows the LAI development of each site simulated by the model since planting; (b) shows the 660 comparison of field measured LAI in 2014 with model.

Fig. 10. Validation of yield and NPP with 8 independent oil palm sites from the Harapan (H)
and Bukit Duabelas (B) regions with different fertilization treatments. Field data were
collected in 2014.

666 Appendix A

667 Summary of main parameters

Table A1. Summary of new phenological parameters introduced for the phenology subroutine of CLM-Palm. The default values were determined by

calibration and with reference to field observations and literatures on oil palm (Combres et al., 2013; Corley and Tinker, 2003; Hormaza et al., 2012; Legros
 et al., 2009).

Parameter	Default	Min	Max	Explanation (Unit)
GDD _{init}	0	0	1500	GDD needed from planting to the first phytomer initiation (°days). Initiation refers to the start of active accumulation of leaf C. A value 0 implies transplanting.
GDD_{exp}	1550	0	8000	GDD needed from leaf initiation to start of leaf expansion for each phytomer (pre-expansion) (days)
$GDD_{L.mat}$	1250	500	1600	GDD needed from start of leaf expansion to leaf maturity for each phytomer (post-expansion) (days)
$GDD_{F.fill}$	3800	3500	4200	GDD needed from start of leaf expansion to beginning of fruit-fill for each phytomer (days)
$GDD_{F.mat}$	5200	4500	6500	GDD needed from start of leaf expansion to fruit maturity and harvest for each phytomer (days)
$GDD_{L.sen}$	6000	5000	8000	GDD needed from start of leaf expansion to beginning of senescence for each phytomer (days)
GDD_{end}	6650	5600	9000	GDD needed from start of leaf expansion to end of senescence for each phytomer ('days)
GDD_{min}	7500	6000	10000	GDD needed from planting to the beginning of first fruit-fill (days)
Age_{max}	25	20	30	Maximum plantation age (productive period) from planting to final rotation /replanting (years)
PLAI _{max}	0.165	0.1	0.2	Maximum LAI of a single phytomer $(m^2 m^{-2})$
mxlivenp	40	30	50	Maximum number of expanded phytomers coexisting on a palm
phyllochron	130	100	160	Initial phyllochron (=plastochron): the period in heat unit between the initiations of two successive phytomers. The value increases to 1.5 times, i.e. 195, at 10-year old (days)

Table A2. Summary of parameters involved in C and N allocation. The default values were determined by calibration and with reference to field
 measurements (Kotowska et al., 2015a).

Parameter	Defaults	Min	Max	Explanation (Unit)
*lf _{disp}	0.3	0.1	1	Fraction of C and N allocated to the displayed leaf pool
*transplant	0.15	0	0.3	Initial total LAI assigned to existing expanded phytomers at transplanting. Value 0 implies planting as seeds.
f_{leaf}^i	0.15	0	1	Initial value of leaf allocation coefficient before the first fruit-fill
a_{root}^i	0.3	0	1	Initial value of root allocation coefficient before the first fruit-fill
a_{leaf}^{f}	0.28	0	1	Final value of leaf allocation coefficient after vegetative maturity
a_{root}^{f}	0.1	0	1	Final value of root allocation coefficient after vegetative maturity
F_{stem}^{live}	0.15	0	1	Fraction of new stem allocation that goes to live stem tissues, the rest to metabolically inactive stem tissues
d_{mat}	0.6	0.1	1	Factor to control the age when the leaf allocation ratio stabilizes at a_{leaf}^{f} according to Eq. 4
d^{leaf}_{alloc}	0.6	0	5	Factor to control the nonlinear function in Eq. 4. Values < 1 give a convex curve and those > 1 give a concave curve. Value 1 gives a linear function.
*a	0.2	0	1	Parameter <i>a</i> for fruit allocation coefficient A_{fruit} in Eq. 5
*b	0.02	0	1	Parameter <i>b</i> for fruit allocation coefficient A_{fruit} in Eq. 5
SLA	0.013	0.01	0.015	Specific leaf area (m ² g ^{-1} C)
F_{LNR}	0.1005	0.05	0.1	Fraction of leaf nitrogen in Rubisco enzyme. Used together with SLA to calculate V_{cmax25} (g N Rubisco g ⁻¹ N)

673 *New parameters introduced for oil palm. Others are existing parameters in CLM but mostly are redefined or used in changed context.

Parameter	Value	Definition (Unit)	Comments
<i>CN</i> _{leaf}	33	Leaf carbon-to-nitrogen ratio (g C g^{-1} N)	By leaf C:N analysis
CN _{root}	42	Root carbon-to-nitrogen ratio (g C g^{-1} N)	Same as all other PFTs
CN _{livewd}	50	Live stem carbon-to-nitrogen ratio (g C g^{-1} N)	Same as all other PFTs
CN_{deadwd}	500	Dead stem carbon-to-nitrogen ratio (g C g^{-1} N)	Same as all other PFTs
<i>CN</i> _{lflit}	60	Leaf litter carbon-to-nitrogen ratio (g C g^{-1} N)	Same as other tree PFTs
CN _{fruit}	75	Fruit carbon-to-nitrogen ratio (g C $g^{-1}N$)	Higher than the value 50 for other crops because of high oil content in palm fruit

Stem reflectance in the visible or near-infrared bands

Leaf transmittance in the visible or near-infrared bands

Stem transmittance in the visible or near-infrared bands

Number of palms per hectare (stems ha^{-2})

Leaf reflectance in the visible (VIS) or near-infrared (NIR) bands

Table A3. Other optical, morphological, and physiological parameters for oil palm.

r^{leaf} vis/nir

r^{stem} r_{vis/nir}

τ^{leaf} v_{vis/nir}

 $au_{vis/nir}^{stem}$

stocking

0.09/0.45

0.16/0.39

0.05/0.25

0.001/

0.001

150

χL	-0.4	Leaf angle distribution index for radiative transfer ($0 =$ random leaves; $1 =$ horizontal leaves; $-1 =$ vertical leaves)	Estima -0.4	ated by field $\leq \chi_L \leq 0.6$	observa	ation. In	CLM,
taper	50	Ratio of stem height to radius-at-breast-height	Field	observation.	Used	togethe	r with

Field observation. Used together with *stocking* and *dwood* to calculate canopy top and bottom heights.

Values adjusted in-between trees and crops

Field observation. Used to calculate stem area index (SAI) by: $SAI = 0.05 \times LAI \times stocking$.

dwood	100000	Wood density (gC m^{-3})	Similar as coconut palm (O. Roupsard, personal communication)
R_{z0m}	0.05	Ratio of momentum roughness length to canopy top height	T. June, personal communication
R_d	0.76	Ratio of displacement height to canopy top height	T. June, personal communication

675 References

- Allen, K., Corre, M. D., Tjoa, A., and Veldkamp, E.: Soil nitrogen-cycling responses to
 conversion of lowland forests to oil palm and rubber plantations in Sumatra, Indonesia,
 PLoS ONE, 10(7), e0133325, doi:10.1371/journal.pone.0133325, 2015
- Bonan, G. B., Levis, S., Kergoat, L., and Oleson, K. W.: Landscapes as patches of plant
 functional types: An integrated concept for climate and ecosystem models, Global
 Biogeochemical Cycles, 16 (2), 1021-1051, 2002.
- 682 Carlson, K. M., Curran, L. M., Asner, G. P., Pittman, A. M., Trigg, S. N., and Adeney, J. M.:
 683 Carbon emissions from forest conversion by Kalimantan oil palm plantations, Nature
 684 Clim. Change, 3(3), 283–287, doi:10.1038/nclimate1702, 2012.
- Carrasco, L. R., Larrosa, C., Milner-Gulland, E. J., and Edwards, D. P.: A double-edged
 sword for tropical forests, Science, 346(6205), 38-40, 2014.
- Combres, J.-C., Pallas, B., Rouan, L., Mialet-Serra, I., Caliman, J.-P., Braconnier, S., Soulie,
 J.-C., and Dingkuhn, M.: Simulation of inflorescence dynamics in oil palm and
 estimation of environment-sensitive phenological phases: a model based analysis,
 Functional Plant Biology, 40(3), 263-279, 2013.
- 691 Corley R. H. V. and Tinker, P. B. (Eds.): The oil palm, 4th edition, Blackwell Science,
 692 Oxford, 2003.
- Dee, D. P., Uppala, S. M., Simmons, A. J., Berrisford, P., Poli, P., Kobayashi, S., ... and
 Vitart, F.: The ERA-Interim reanalysis: Configuration and performance of the data
 assimilation system, Quarterly Journal of the Royal Meteorological Society, 137(656),
 553-597, 2011.
- Drewniak, B., Song, J., Prell, J., Kotamarthi, V. R., and Jacob, R.: Modeling agriculture in the
 community land model, Geoscientific Model Development, 6(2), 495-515,
 doi:10.5194/gmd-6-495-2013, 2013.
- Euler, M.: Oil palm expansion among Indonesian smallholders adoption, welfare
 implications and agronomic challenges, Ph.D. thesis, University of Göttingen, Germany,
 145 pp., 2015.
- FAO. FAOSTAT Database, Food and Agriculture Organization of the United Nations, Rome,
 Italy, available at: http://faostat.fao.org/site/339/default.aspx (last access: 17 June 2015),
 2013.
- Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S.
 P., ... and V össmarty, C. J.: Nitrogen cycles: past, present, and future, Biogeochemistry,
 708 70(2), 153-226, 2004.
- Georgescu, M., Lobell, D. B., and Field, C. B.: Direct climate effects of perennial bioenergy
 crops in the United States, Proceedings of the National Academy of Sciences, 108(11),
 4307-4312, 2011.
- Goh K. J.: Climatic requirements of the oil palm for high yields, in: Managing oil palm for
 high yields: agronomic principles, Goh K.J. (Eds.), pp. 1–17, Malaysian Soc. Soil Sci.
 and Param Agric. Surveys, Kuala Lumpur, 2000.
- Guillaume, T., Damris, M., and Kuzyakov, Y.: Losses of soil carbon by converting tropical
 forest to plantations: erosion and decomposition estimated by δ13C, Global change
 biology, 21, 3548–3560, doi: 10.1111/gcb.12907, 2015.
- Gunarso, P., Hartoyo, M. E., Agus, F., and Killeen, T. J.: Oil Palm and Land Use Change in
 Indonesia, Malaysia, and Papua New Guinea. In: Killeen T, Goon J, editors. Reports
 from the Science Panel of the Second GHG Working Group of the Rountable for
 Sustainable Palm Oil (RSPO). Kuala Lumpur, 2013.
- Hall é, F., Oldeman, R. A. A. and Tomlinson, P. B.: Tropical trees and forests. An
 architectural analysis. Springer-Verlag, Berlin, 441 pp., 1978.
- Hijmans, R. J., Cameron, S. E., Parra, J. L., Jones, P. G., and Jarvis, A.: Very high resolution
 interpolated climate surfaces for global land areas, International journal of climatology,
 25(15), 1965-1978, 2005.

- Hoffmann, M. P., Vera, A. C., Van Wijk, M. T., Giller, K. E., Oberth ür, T., Donough, C., and
 Whitbread, A. M.: Simulating potential growth and yield of oil palm (Elaeis guineensis)
 with PALMSIM: Model description, evaluation and application, Agricultural Systems,
 131, 1-10, 2014.
- Hormaza, P., Fuquen, E. M., and Romero, H. M.: Phenology of the oil palm interspecific
 hybrid Elaeis oleifera × Elaeis guineensis, Scientia Agricola, 69(4), 275-280, 2012.
- Huth, N. I., Banabas, M., Nelson, P. N., and Webb, M.: Development of an oil palm cropping
 systems model: lessons learned and future directions, Environ. Modell. Softw., 62, 411–
 419, doi:10.1016/j.envsoft.2014.06.021, 2014.
- Jin, J. M. and Miller, N. L.: Regional simulations to quantify land use change and irrigation
 impacts on hydroclimate in the California Central Valley, Theoretical and Applied
 Climatology, 104, 429-442, 2011.
- Koh, L. P. and Ghazoul, J.: Spatially explicit scenario analysis for reconciling agricultural
 expansion, forest protection, and carbon conservation in Indonesia, P. Natl. Acad. Sci.
 USA, 107, 11140–11144, doi: 10.1073/pnas.1000530107, 2010.
- Kotowska, M. M., Leuschner, C., Triadiati T., Selis M., and Hertel, D.: Quantifying aboveand belowground biomass carbon loss with forest conversion in tropical lowlands of
 Sumatra (Indonesia), Global Change Biol., 21, 3620-3634, doi: 10.1111/gcb.12979,
 2015a.
- Kotowska, M. M., Leuschner, C., Triadiati, T., and Hertel, D.: Conversion of tropical lowland
 forest lowers nutrient return with litterfall, and alters nutrient use efficiency and
 seasonality of net primary productivity, Oecologia, submitted, 2015b.
- Koven, C. D., Riley, W. J., Subin, Z. M., Tang, J. Y., Torn, M. S., Collins, W. D., Bonan, G.
 B., Lawrence, D. M., and Swenson, S. C.: The effect of vertically resolved soil
 biogeochemistry and alternate soil C and N models on C dynamics of CLM4,
 Biogeosciences, 10(11), 7109-7131, doi:10.5194/bg-10-7109-2013, 2013.
- Legros, S., Mialet-Serra, I., Caliman, J. P., Siregar, F. A., Clement-Vidal A., and Dingkuhn,
 M.: Phenology and growth adjustments of oil palm (Elaeis guineensis) to photoperiod and climate variability, Annals of Botany 104, 1171–1182. doi:10.1093/aob/mcp214,
 2009.
- Levis, S., Bonan, G., Kluzek, E., Thornton, P., Jones, A., Sacks, W., and Kucharik, C.:
 Interactive crop management in the Community Earth System Model (CESM1):
 Seasonal influences on land-atmosphere fluxes, J. Climate, 25, 4839-4859,
 DOI:10.1175/JCLI-D-11-00446.1., 2012.
- Luyssaert, S., Schulze, E. D., Börner, A., Knohl, A., Hessenmöller, D., Law, B. E., Ciais, P.,
 and Grace, J.: Old-growth forests as global carbon sinks, Nature, 455(7210), 213-215,
 2008.
- Miettinen, J., Shi, C. H. and Liew, S. C.: Deforestation rates in insular Southeast Asia
 between 2000 and 2010, Global Change Biology, 17, 2261-2270, 2011.
- Navarro, M. N. V., Jourdan, C., Sileye, T., Braconnier, S., Mialet-Serra, I., Saint-Andre, L., ...
 and Roupsard, O.: Fruit development, not GPP, drives seasonal variation in NPP in a
 tropical palm plantation, Tree physiology, 28(11), 1661-1674, 2008.
- Oleson, K. W., Bonan, G. B., Levis, S., and Vertenstein, M.: Effects of land use change on
 North American climate: impact of surface datasets and model biogeophysics, Climate
 Dynamics, 23, 117-132, 2004.
- Oleson, K., Lawrence, D., Bonan, G., Drewniak, B., Huang, M., Koven, C., Levis, S., Li, F.,
 Riley, W., Subin, Z., Swenson, S., Thornton, P., Bozbiyik, A., Fisher, R., Heald, C.,
 Kluzek, E., Lamarque, J.-F., Lawrence, P., Leung, L., Lipscomb, W., Muszala, S.,
 Ricciuto, D., Sacks, W., Sun, Y., Tang, J., and Yang, Z.-L.: Technical description of
 version 4.5 of the Community Land Model (CLM), National Center for Atmospheric
 Research, Boulder, Colorado, USA, 420 pp., doi:10.5065/D6RR1W7M, 2013.
- Tang, J. Y., Riley, W. J., Koven, C. D., and Subin, Z. M.: CLM4-BeTR, a generic
 biogeochemical transport and reaction module for CLM4: model development,
 evaluation, and application, Geosci. Model Dev., 6, 127-140. doi:10.5194/gmd-6-1272013, 2013.

- van Kraalingen, D. W. G., Breure, C. J., and Spitters, C. J. T.: Simulation of oil palm growth
 and yield, Agricultural and forest meteorology, 46(3), 227-244, 1989.
- Veldkamp, E., and Keller, M.: Nitrogen oxide emissions from a banana plantation in the
 humid tropics, Journal of Geophysical Research: Atmospheres (1984–2012), 102(D13),
 15889-15898, 1997.
- White, M. A., Thornton, P. E., and Running, S. W.: A continental phenology model for
 monitoring vegetation responses to interannual climatic variability, Global Biogeochem.
 Cycles, 11, 217-234, 1997.